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Translational Therapeutics

Folate receptor alpha in ovarian cancer tissue and patient serum is associated with disease burden and treatment outcomes

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BACKGROUND: Survival rates for ovarian cancer remain poor, and monitoring and prediction of therapeutic response may benefit from additional markers. Ovarian cancers frequently overexpress Folate Receptor alpha (FRα) and the soluble receptor (sFRα) is measurable in blood. Here we investigated sFRα as a potential biomarker.

METHODS: We evaluated sFRα longitudinally, before and during neo-adjuvant, adjuvant and palliative therapies, and tumour FRα expression status by immunohistochemistry. The impact of free FRα on the efficacy of anti-FRα treatments was evaluated by an antibody-dependent cellular cytotoxicity assay.

RESULTS: Membrane and/or cytoplasmic FRα staining were observed in 52.7% tumours from 316 ovarian cancer patients with diverse histotypes. Circulating sFRα levels were significantly higher in patients, compared to healthy volunteers, specifically in patients sampled prior to neoadjuvant and palliative treatments. sFRα was associated with FRα cell membrane expression in the tumour. sFRα levels decreased alongside concurrent tumour burden in patients receiving standard therapies. High concentrations of sFRα partly reduced anti-FRα antibody tumour cell killing, an effect overcome by increased antibody doses.

CONCLUSIONS: sFRα may present a non-invasive marker for tumour FRα expression, with the potential for monitoring patient response to treatment. Larger, prospective studies should evaluate FRα for assessing disease burden and response to systemic treatments.

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BACKGROUND

Ovarian cancer is the third most common but most lethal gynaecological malignancy worldwide. Non-specific symptomatology and lack of validated screening tools often result in late-stage diagnosis [1]. Whilst a combination of surgical debulking and platinum/taxane-based chemotherapy followed by maintenance PARP inhibitors and/or bevacizumab [2–5] is the mainstay of treatment for advanced disease, most women subsequently relapse, resulting in 5-year survival rates of 13–27% for patients with late-stage (III–IV) disease [6].

Currently, CA125 is the most widely used serum biomarker for monitoring ovarian cancer. However, intra-tumour heterogeneity, a key feature of ovarian tumours [7], suggests the presence of distinct sub-clonal populations within a tumour, possibly harbouring different marker phenotypes. Therefore, incorporating several complementary biomarkers that may account for the molecular heterogeneity of ovarian cancer sub-clones as well as different histotypes could improve surveillance and evaluating response to treatment [8].

FRα is a glycosylphosphatidylinositol (GPI)-anchored cell-surface glycoprotein encoded by the *FOLR1* gene. FRα binds with high

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affinity to folic acid and its derivatives [9] and mediates cellular processes, including cell division, proliferation, and tissue growth via signalling cascades and components of the folate cycle [10–12]. The distribution of FRa across non-malignant tissues is limited; however, elevated expression has been observed in several cancers, including ovarian, endometrial, lung and breast cancer subsets [13–19]. Up to 90% of ovarian cancers overexpress FRa [20, 21]; however, differential levels of expression have been observed across different ovarian cancer histotypes [22]. FRa can be shed from tumour cells and is detected in the circulation in soluble form (sFRa) [23–25]. Given that FRa is highly expressed in ovarian cancer, and is readily measurable in the serum, sFRa may have the potential to be leveraged as an adjunct clinical biomarker for the disease.

In addition to its potential role as a clinical biomarker, there has been significant interest in using FRa as a therapeutic target in patients with ovarian cancer, with several FRa-targeted therapeutics, such as farletuzumab and mirvetuximab soravtansine, in late-phase trials [26–31]. Despite promising results, FRa-targeted agents are yet to be approved for use in patients with cancer. This may in part be attributed to the challenge of selecting patients who are likely to respond. Immunohistochemical staining of archival tumour samples is commonly used to determine FRa protein expression levels for the inclusion of patients in clinical trials [32]. Additionally, sFRa-based assessment of FRa tumour expression might complement the selection of patients for FRa-targeted therapeutics. Presently, understanding the correlations of circulating levels of sFRa with tumour volume and/or the degree of tumoural FRa expression, and whether sFRa levels are influenced by treatments, are still required before this approach can be implemented clinically.

In this study, we performed immunohistochemical staining for FRa expression in tumours to investigate the relationship between sFRa and tumour FRa status and to evaluate the possible application of sFRa testing in future clinical studies of FRa-targeted therapies. We investigated sFRa levels in ovarian cancer patients longitudinally, before and during treatment with neo-adjuvant, adjuvant and palliative systemic therapies, as well as in relation to concurrent tumour burden. Tumour burden was defined by a bespoke scoring system designed for this study, with the aim of capturing the volume of disease in its entirety. Furthermore, we asked whether free FRa could impact the efficacy of anti-FRa treatments.

METHODS

Analysis of *FOLR1* expression in normal tissues and ovarian cancer

Gene expression of *FOLR1* (encoding FRa) by various cancer cell lines, tumours and normal tissues were interrogated using the Cancer Cell Line Encyclopedia (CCLE) online tool (www.broadinstitute.org/ccle), The Pathology Atlas of The Human Protein Atlas online tool [33, 34] (<https://www.proteinatlas.org/humanproteome/pathology>), and Xenabrowser online tool [35] (Xenabrowser.net), respectively. Gene expression of *FOLR1* and molecules in the folate/FRa signalling pathway: MTHFR, methylene tetrahydrofolate reductase; FOLH1, glutamate carboxypeptidase; TYMS, thymidylate synthase; DHFR, dihydrofolate reductase, were studied in normal ovary and ovarian cancer tissues using the Gene Expression Profiling Interactive Analysis (GEPIA) online tool [36] (<http://gepia.cancer-pku.cn/index.html>).

Sample collection pathway to evaluate FRa in tumour tissues and patient blood

Sample collection for this study was reviewed and approved by the Guy's Research Ethics Committee (Reference 09/H0804/45 and 16/LO/0366) and performed at Guy's Hospital in London UK. Ovarian cancer patients and healthy volunteers were enrolled by written informed consent. Ovarian cancer patients who had undergone screening for potential inclusion in an anti-FRa clinical trial provided archival tumour tissue ($N = 316$); of these, $N = 55$ also provided blood samples. An additional cohort of $N = 88$

patients was approached during outpatient clinic appointments and provided blood samples only.

To study tumour-expressed and sFRa in ovarian cancer patients, we established a protocol to collect and analyse tumour tissue ($N = 316$ patients) and serum ($N = 143$ patients) samples (Fig. 1). Where possible, longitudinal serum samples were collected at defined timepoints throughout standard treatment regimens. Furthermore, these were evaluated with reference to patient characteristics from clinical databases; namely tumour histotype, germline BRCA1/2 mutation status (Supplementary Table 1), and tumour burden score (calculated from concurrent imaging as outlined in Supplementary Table 2). Sera from healthy volunteers ($N = 61$) were also collected and studied as controls.

Serum samples were prepared by centrifugation of clotted blood in SST Clot Activator and Polymer Gel Hemogard Closure Blood Tubes (BD) at 2500 RPM for 15 min at 4 °C and stored at 80 °C until analysis.

Tumour expression of FRa by immunohistochemistry (IHC)

Immunohistochemistry (IHC) was used to evaluate the membrane and cytoplasmic expression of FRa by formalin-fixed, paraffin-embedded normal tissue sections in tissue microarrays and tumour sample sections from ovarian cancer patients [15, 37]. Novocastra™ Liquid mouse anti-human FRa primary antibody (clone BN3.2, Leica) was applied to formalin-fixed, paraffin-embedded tumour sections (from primary debulking surgery) for 32 min at room temperature at 1/500 dilution, followed by detection with Ultra Universal 3,3'-diaminobenzidine (DAB) detection kit (Ventana Medical Systems Inc.) and then Haematoxylin II applied for 8 min. Scoring of tissue sections was performed by pathologists. FRa positivity was determined as $\geq 5\%$ staining in the membrane and/or cytoplasm (at any intensity). Scoring is also presented as staining on the membrane of 0–<5, 5–<25, 25–<50 and $\geq 50\%$ tumour cells (at any intensity), and as *H* scores (calculated by multiplying the % of cells with membrane expression with the intensity of staining scored 0–3; maximum *H* score of 300) [38].

Serum FRa and anti-FRa autoantibodies

ELISAs were performed to evaluate the concentrations of circulating sFRa and anti-FRa IgG autoantibodies in serum samples from ovarian cancer patients and healthy volunteers [37, 39]. MaxiSORP™ plates (Nunc) were coated with 100 μ l/well of 2 μ g/ml monoclonal mouse anti-human FRa IgG1 antibody (clone 548908) or 1 μ g/ml recombinant FRa, respectively (both R&D Systems). Following incubation at 4 °C overnight, 250 μ l/well SuperBlock™ (Perbio Science Ltd.) was added for 2 h at room temperature (RT). Serum samples were diluted to 20% in a 50:50 solution of SuperBlock™ and PBS-0.05% Tween® 20 solution (Severn Biotech and Sigma, respectively). Standard curves of recombinant FRa (R&D Systems), or anti-FRa human IgG1 monoclonal antibody (prepared in house) were diluted in SuperBlock™-PBS-0.05% Tween® 20, supplemented with 20% human serum albumin (type AB male, Sigma). Samples and standards were added 50 μ l/well, in triplicate, and incubated for 2 h at RT. FRa was detected by 50 μ l/well biotinylated polyclonal goat anti-human FRa IgG1 antibody (R&D Systems, diluted to 25 ng/ml in SuperBlock™-PBS-0.05% Tween® 20) for 2 h at RT, followed by 50 μ l/well streptavidin-peroxidase conjugate (Pierce, diluted 1/22,000 in SuperBlock™-PBS-0.05% Tween® 20) for 30 min at RT. Anti-FRa IgG autoantibodies were detected by 50 μ l/well HRP-conjugated polyclonal goat anti-human Fc γ -specific F(ab')₂ fragment (Jackson Immuno Research, diluted 1/500 in SuperBlock™-PBS-0.05% Tween® 20) for 45 min at RT. Plates were developed with 50 μ l/well OPD (Sigma) diluted to 0.5 mg/ml in stable peroxidase substrate buffer (Pierce), for 5–10 min at RT, in darkness, followed by 50 μ l/well 1 M HCl solution (Sigma). Washing at each step was performed with 250 μ l/well PBS-0.05% Tween® 20 4 times. A Fluostar Omega microplate reader (BMG LABTECH) was used with an absorbance of 492 nm, and correction wavelength of 650 nm. Standard curves were fitted using a 4-point variable curve-fitting program using a minimum of 6 points (MARS software, BMG LABTECH). The lower levels of quantification (LLOQ) were 6.25 and 3.125 ng/ml, respectively. Values below LLOQ are reported as 0 ng/ml. Each sample was analysed in technical triplicates and data are reported as the mean of two independent experiments.

Tumour burden score

Tumour burden scores were calculated from concurrent imaging as outlined in Supplementary Table 2. Briefly, total scores were determined by the sum of points calculated according to the number of sites of disease, the number of metastases per site, and the maximum metastasis diameter per site.

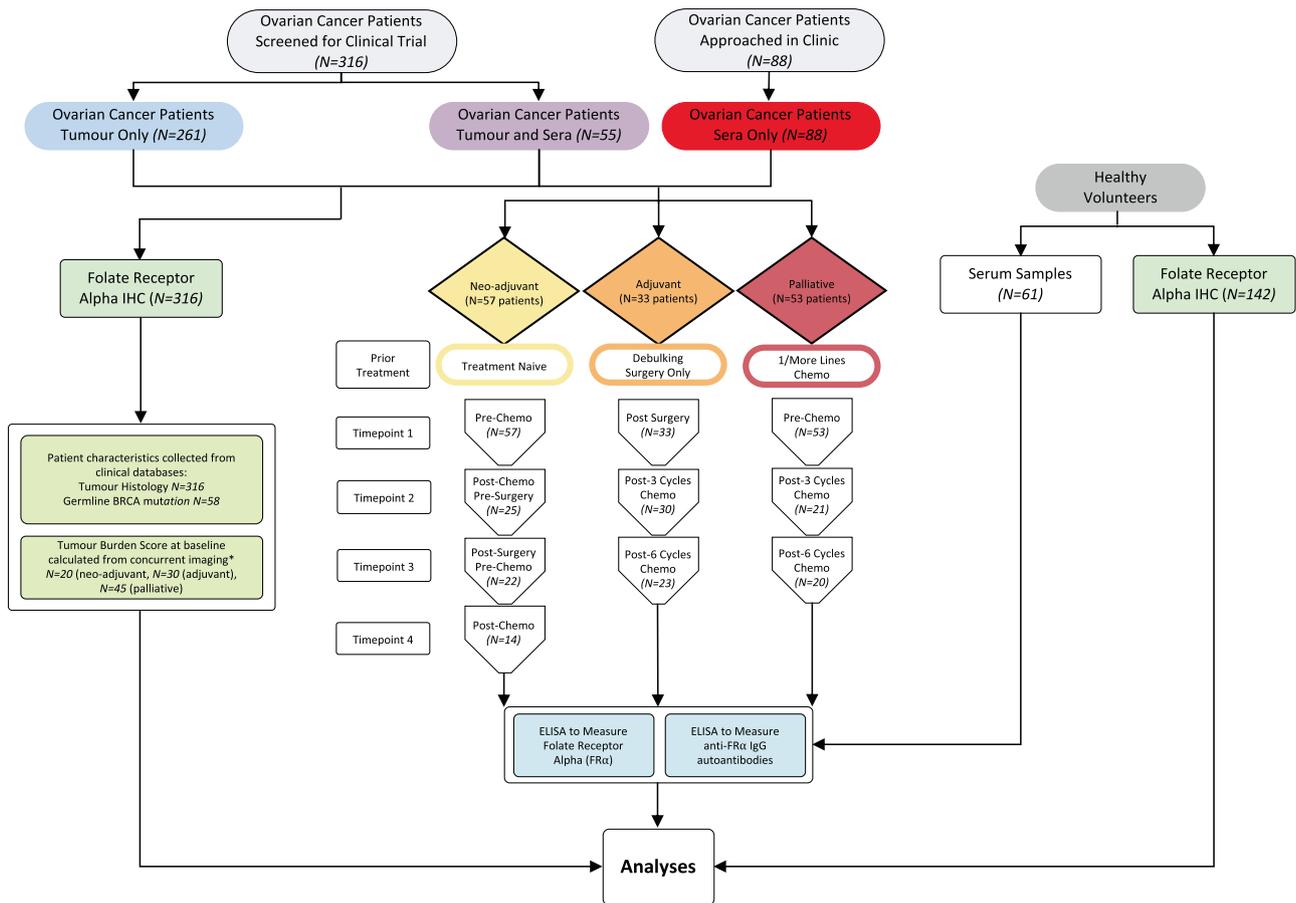


Fig. 1 Study design and sample workflow. Serum samples from ovarian cancer patients were collected at timepoint 1 and up to 3 sequential treatment-related time points (e.g., for the treatment-naïve group: pre-chemo, post-chemo/pre-surgery, post-surgery/pre-chemo, and post-chemo). These were collected alongside serum samples from healthy volunteers. Serum samples were studied for sFR α and anti-FR α autoantibodies, and FR α protein expression in patient tumours and normal tissue microarrays was evaluated by immunohistochemistry. Patient characteristics, including tumour histotype, were collected from clinical databases (see Supplementary Table 1 for ovarian patients in the longitudinal study). Patient tumour burden scores were calculated from concurrent imaging (*see Supplementary Table 2 for a scoring system). *N* numbers are indicated in the figure.

In vitro efficacy of FR α -targeting therapy

Cell-surface binding of an FR α -specific IgE therapeutic candidate, MOv18 IgE, to FR α -expressing IGROV1 ovarian cancer cells (CVCL_1304, Sigma) was confirmed by incubation of 1×10^5 cells with indicated concentrations of MOv18 IgE for 30 min at 4 °C, followed by FITC-conjugated polyclonal goat anti-human IgE antibody (1/50 dilution; Vector Laboratories) for 20 min at 4 °C. An established flow cytometric antibody-dependent cellular cytotoxicity (ADCC) and phagocytosis (ADCP) assay was used, as previously described [40, 41], to evaluate the efficacy of a FR α -specific IgE therapeutic candidate, MOv18 IgE, in the presence of soluble FR α . IGROV1 cancer cells were incubated with human peripheral blood mononuclear cells (PBMCs) or U937 human monocytic cells (CRL-1593.2, ATCC) (at ratios of 1:10 and 1:1.5, respectively), together with indicated concentrations of MOv18 IgE (in-house) and recombinant FR α antigen (R&D Systems). Cell lines were routinely tested for mycoplasma contamination by PCR. Flow cytometric acquisitions were performed on a FACS Canto II using FACSDiva software, and analyses were conducted using the FlowJo software (FlowJo LLC, BD). Each condition was analysed in technical triplicates and data are reported as the mean of four independent experiments.

Statistical analyses

For in vitro efficacy assays, with equal sample size, and assuming 90% power and a significance level of 5%, we calculated that at least 3 independent experiments are required for each group to detect a difference between conditions. For analyses of patient serum and tumour tissues, a minimum of 5 samples were compared (90% power and a significance level of 5%). No samples were excluded from the analysis. Descriptive statistical analyses of the data were performed to calculate EC50, as well as group comparisons

using non-parametric Mann–Whitney *T* test, one-way analysis of variance with non-parametric Kruskal–Wallis multiple comparisons and Chi-square test. Beta coefficient estimates for change in sFR α and tumour burden score between longitudinal timepoints were calculated using linear regression. Linear regression analyses were also performed to assess whether the variation in sFR α over the treatment course, was associated with the variation of the tumour burden score. Receiver operating characteristic (ROC) curve analyses were performed to assess the prediction capabilities of: (i) sFR α for tumour expression of FR α , and (ii) sFR α for high tumour burden score. The correlation between concurrent tumour burden score and sFR α levels was assessed by Pearson's correlation coefficient. All the analyses were conducted using GraphPad Prism 9 or IBM SPSS statistics 27. Test significance (*P* value) is represented as follows: **P* < 0.05, ***P* < 0.01, ****P* < 0.001, *****P* < 0.0001. Error bars represent the standard error of mean (SEM).

Reporting summary

Further information on research design is available in the Nature Research Reporting Summary linked to this article.

RESULTS

High FR α gene expression in ovarian cancer compared with other tumour types and normal tissues

We confirmed the expression of FR α across ovarian cancer cell lines and tissue specimens, several other cancer types, and normal tissues, by interrogating publicly available data [33–35]. *FOLR1* (encoding FR α) was highly expressed in human ovarian cancer cell

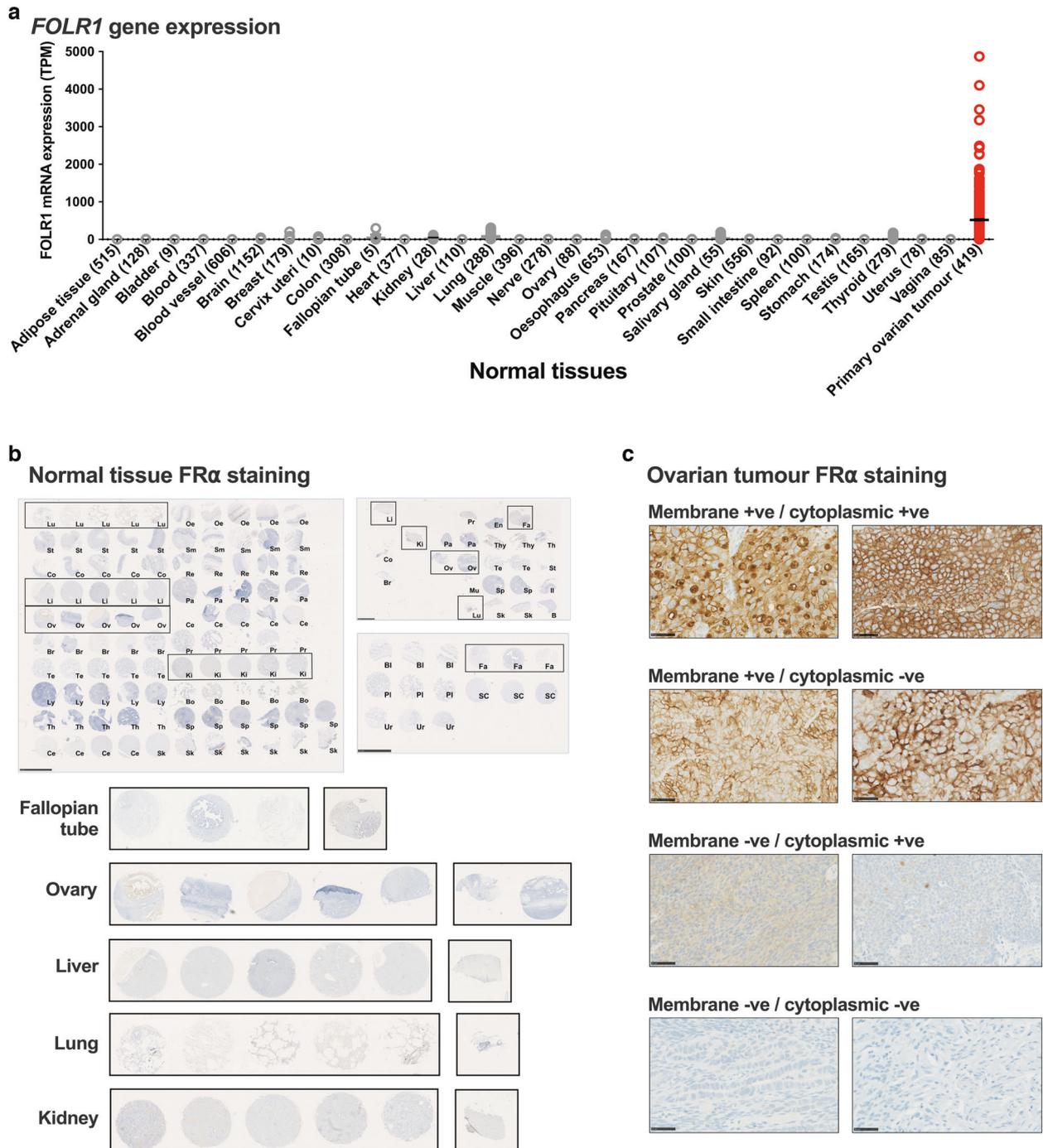


Fig. 2 Transcriptomic and immunohistochemical analyses of FR α expression in normal tissues and ovarian tumours. **a** Low *FOLR1* gene expression in normal tissues (grey) compared to primary ovarian tumours (red) (N numbers indicated in parentheses; The Human Protein Atlas online tool Xenabrowser online tool [35] (Xenabrowser.net)). **b** No membrane or cytoplasmic FR α protein expression on a broad range of normal tissues ($N = 142$; scale bars = 2.5 mm). Key normal tissue types; fallopian tube (Fa), ovary (Ov), liver (Li), lung (Lu) and kidney (Ki) are highlighted with black boxes and zoomed images shown. **c** Representative images of ovarian tumour FR α staining ($\times 40$ magnification; scale bar = 50 μm). Bl bladder, Bo bone marrow, B brain, Br breast, Cb cerebrum; Ce cervix; Co colon; En endometrium, Fa fallopian tube, Il ileum, Ki kidney, Li liver, Lu lung, Ly lymph node, Mu muscle, striated, Oe oesophagus, Ov ovary, Pa pancreas, Pl placenta, Pr prostate, Re rectum, Sk skin, Sm small intestine, SC spinal cord, Sp spleen, St stomach, Te testis, Th thymus, Thy thyroid, Ur ureter.

lines and ovarian cancer specimens, compared to other cell lines and cancers (Supplementary Fig. 1A, B). In line with the known roles of folate/FR α pathway in cell growth, proliferation and survival [11, 12] (Supplementary Fig. 1C), we found dysregulated expression of folate pathway genes in ovarian cancer [36]: gene expression of *FOLR1*, *TYMS* (thymidylate synthase) *DHFR*

(dihydrofolate reductase) were significantly-greater, and gene expression of *MTFHR* (methylene tetrahydrofolate reductase) was lower in tumours ($N = 426$), compared to normal ovary ($N = 88$) (Supplementary Fig. 1D). *FOLR1* expression was markedly higher in primary ovarian tumour samples compared to a range of normal tissues [35] (Fig. 2a). These data suggest that *FOLR1* is highly

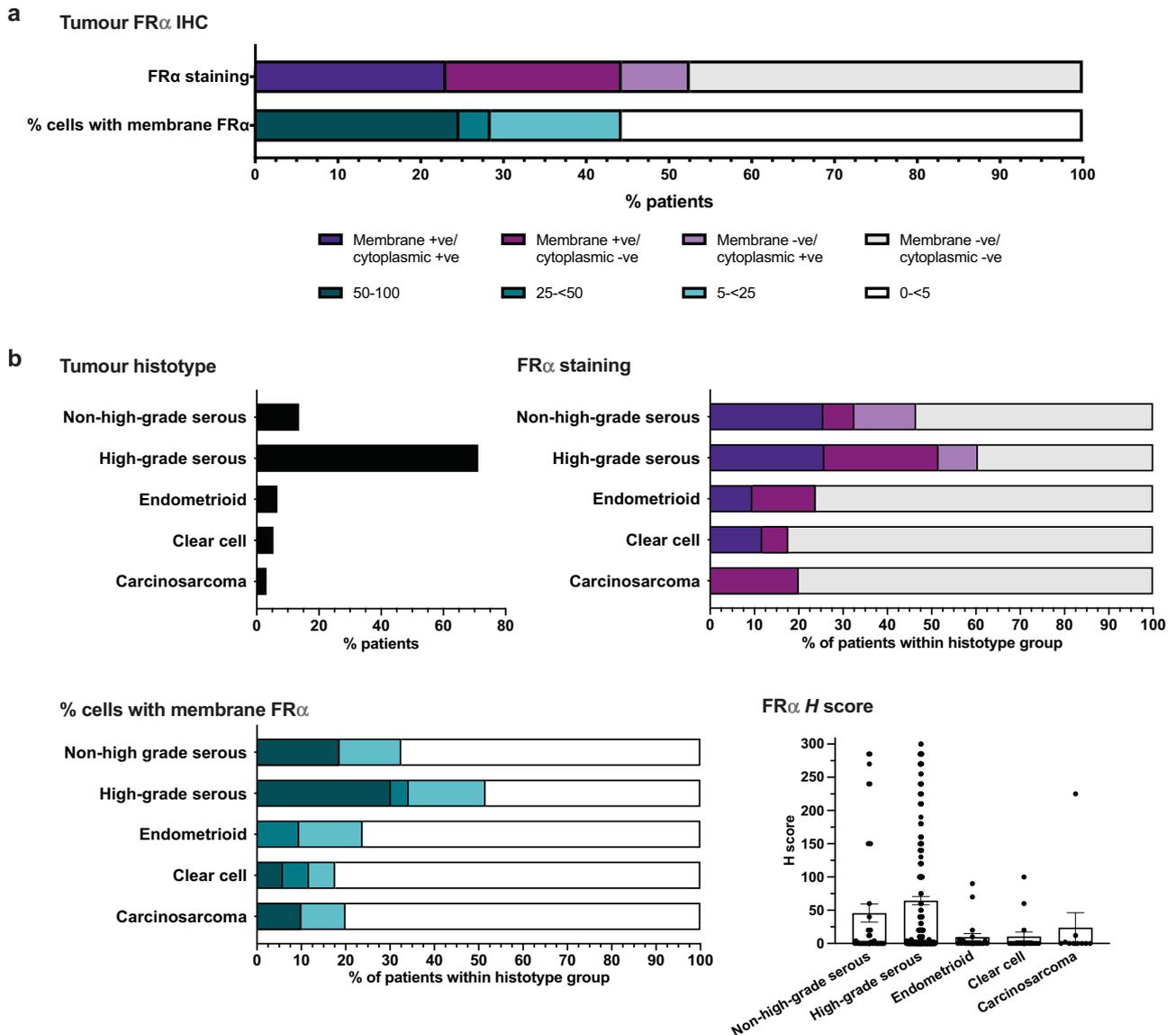


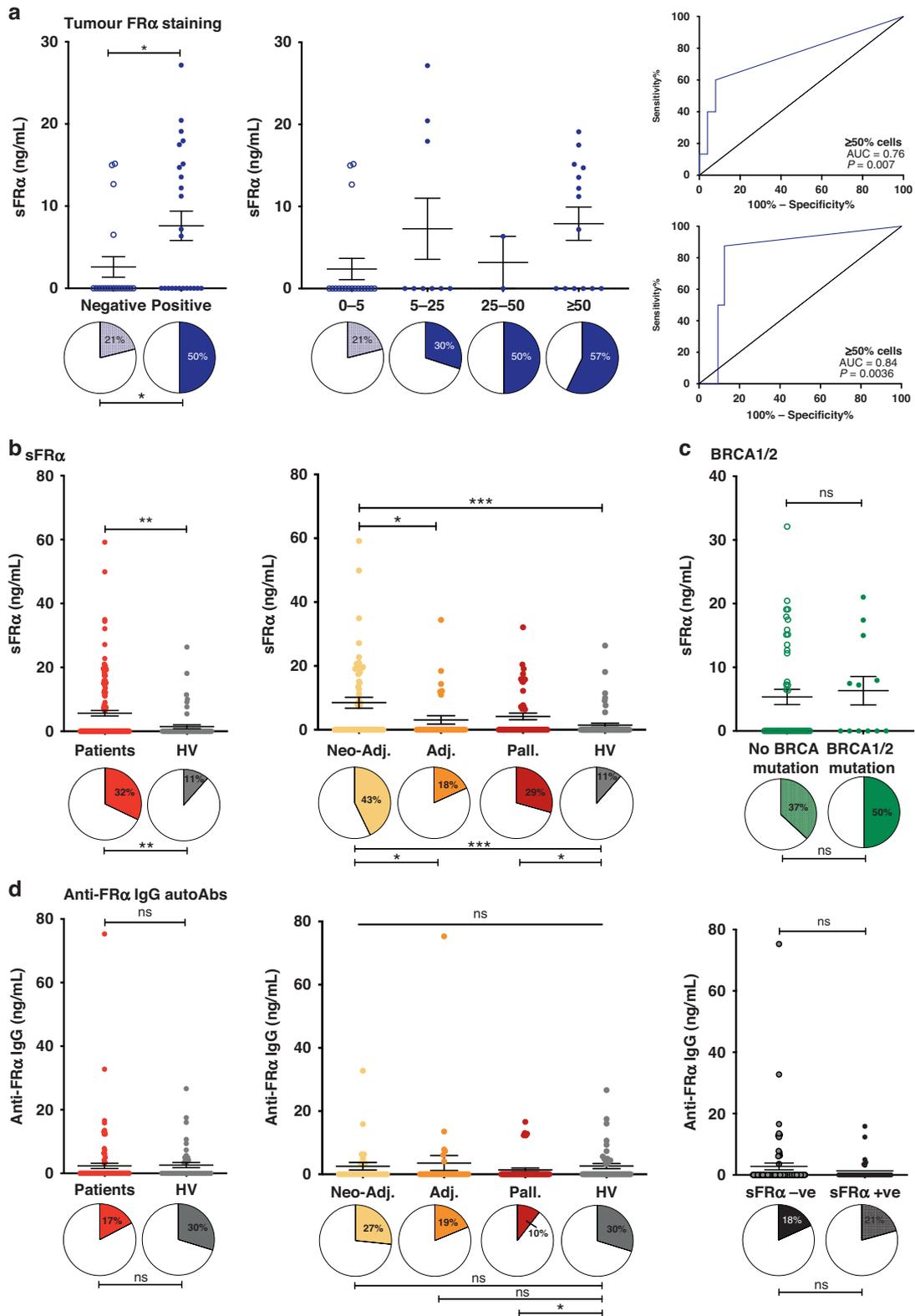
Fig. 3 Immunohistochemical analyses reveal a mixture of cell surface and cytoplasmic FR α protein expression in ovarian cancer tissues. **a** Percentage of patients with tumours expressing FR α on the membrane, in the cytoplasm, both or neither, and with membrane staining on 0–<5, 5–<25, 25–<50 or \geq 50% of tumour cells. **b** Top left: Overall percentage of patients with each tumour histotype; Top right: Percentage of patients with membrane and cytoplasmic positivity within each histotype subgroup; Bottom left: Percentage of patients within each histotype subgroup with different proportions of membrane positive tumours cells; Bottom right: Immunohistochemical *H* scores calculated in each histotype subgroup.

expressed in ovarian and other cancers compared with normal tissue.

Immunohistochemical analyses reveal a mixture of cell surface and cytoplasmic FR α protein expression in ovarian cancer tissues

Having established high levels of *FOLR1* gene expression in ovarian cancer, we evaluated FR α protein expression in normal tissues and ovarian tumours by immunohistochemistry (Figs. 1 and 2; $N = 142$ normal tissues; $N = 316$ ovarian tumours). No membrane or cytoplasmic FR α expression was observed across a range of normal tissues, including normal fallopian tube, ovary, lung, kidney and liver, where FR α expression has been previously reported at luminal surfaces [22, 42] (Fig. 2b). Comparatively, 52.7% of tumours from this diverse patient cohort were positive for either membrane or cytoplasmic staining, or both (Figs. 2c and 3a). Membrane staining of \geq 5%

tumour cells was observed in 44.2% of patients (5–<25% cells in 15.8% patients, 25–<50% cells in 3.8% patients, and \geq 50% cells in 24.6% patients) (Fig. 3a). In our cohort, 71% of tumours were known to have high-grade serous histotype, with other histotypes represented less frequently (Fig. 3b, top left). Sixty percent of the high-grade serous tumours were positive for either membrane or cytoplasmic FR α protein staining in \geq 5% of cells (Fig. 3b, top right). In each sample, FR α expression on the membrane was not always associated with expression in the cytoplasm and vice versa, apart from 20% of the tumour samples that demonstrated both membrane and cytoplasmic FR α expression (data not shown). Of the fifty percent of high-grade serous tumours with \geq 5% of tumour cells showing membrane FR α expression, 17.3% of tumours showed expression on 5–<25% cells, 4% of tumours on 25–<50% cells and 30.2% of tumours on \geq 50% cells (Fig. 3b, bottom left). Furthermore, the immunohistochemical *H* score (calculated from the % of stained



cells and staining intensity) was greater for high-grade serous tumours compared to other histotypes (Fig. 3b, bottom right). These findings confirm expression of FR α by ovarian cancers, particularly those with high-grade serous histotype.

Soluble FR α but not anti-FR α autoantibodies were elevated in the circulation of ovarian cancer patients compared with healthy subjects

We next measured sFR α and anti-FR α IgG autoantibodies in the circulation of ovarian cancer patients at timepoint 1 (prior to the start of the treatment regimen, Fig. 1), compared with healthy

Fig. 4 Soluble FR α but not anti-FR α autoantibodies were elevated in the circulation of ovarian cancer patients compared with healthy subjects. sFR α and anti-FR α autoantibodies were measured in three patient cohorts (neo-adjuvant, adjuvant, palliative) at timepoint 1 (see Fig. 1). **a** sFR α levels were significantly higher in patients with FR α tumour cell membrane expression, compared to patients with FR α -negative tumours (left) and with trend for higher levels in patients with tumours showing FR α cell membrane expression in a greater proportion of tumour cells (middle). In neo-adjuvant treatment-naïve patients, baseline sFR α concentration was predictive of FR α cell membrane expression in both $\geq 5\%$ and $\geq 50\%$ of tumour cells (right top and bottom, respectively). **b** Significantly higher sFR α levels were measured in ovarian cancer patients compared to healthy volunteers, and between patient cohorts (proportion of samples with detectable sFR α indicated by filled pie chart sections below). **c** sFR α levels, or proportions of samples with detectable sFR α , were not associated with the patient's germline BRCA1/2 mutational status. **d** Comparable levels of anti-FR α autoantibodies were detected in serum samples from ovarian cancer patients and healthy volunteers (left), across all patient cohorts (middle), and irrespective of detectable sFR α (right). *N* numbers in Fig. 1; * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$, **** $P \leq 0.0001$. Statistical tests: *t* test, one-way ANOVA with Kruskal–Wallis multiple comparisons, receiver operating characteristic (ROC) curve analyses and Chi square test. Error bars represent the standard error of mean (SEM).

subjects (Fig. 4). In patients where IHC analysis showed tumour cell positivity for FR α ($\geq 5\%$ tumour cells in the sample), the concentration of sFR α and proportion of samples with detectable levels, were significantly greater (7.6 ± 1.8 ng/ml; detectable in 50%), compared to patients with FR α -negative tumours (2.6 ± 1.2 ng/ml; detectable in 21%) (Fig. 4a, left). Furthermore, the proportion of samples with detectable sFR α was greatest in patients with tumours where $\geq 50\%$ cells expressed FR α on the membrane, and a trend for higher concentration of sFR α was observed in this group, compared to patients with 0– $<25\%$ positive tumour cells in tumour lesions (Fig. 4a, middle). Although there was no correlation between sFR α levels and % tumour cells expressing FR α or the IHC *H* score (data not shown), receiver operating characteristic (ROC) curve analysis revealed that the baseline level of circulating sFR α in neo-adjuvant treatment-naïve patients was predictive of positive tumour cell membrane expression of FR α (AUC = 0.76, $P = 0.007$ for predicting tumours with $\geq 5\%$ positive cells and AUC = 0.84, $P = 0.0036$ for predicting tumours with $\geq 50\%$ positive cells; Fig. 4a, right, top and bottom, respectively). On the other hand, sFR α concentration was not predictive of cytoplasmic FR α expression in the patient's tumour (AUC = 0.71, $P = 0.21$; Supplementary Fig. 2).

We found significantly higher concentrations of sFR α in patient (5.7 ± 0.9 ng/ml (mean \pm SEM); $N = 143$), compared with healthy volunteer sera (1.4 ± 0.6 ng/ml; $N = 61$), and a higher proportion of samples with detectable levels of sFR α in patients compared to healthy individuals (32 and 11%, respectively) (Fig. 4b, left). sFR α levels, and the proportion of samples with detectable sFR α , were significantly raised at the start of neo-adjuvant chemotherapy (8.5 ± 1.7 ng/ml; detectable in 43%) compared to prior to adjuvant chemotherapy, where patients had recently undergone debulking surgery (3.1 ± 1.3 ng/ml; detectable in 18%) (Fig. 4b, right). At the start of palliative treatment, sFR α also showed signs of elevation (4.2 ± 1.1 ng/ml; detectable in 29%). Furthermore, regardless of the presence of confirmed germline BRCA mutations, sFR α levels were not significantly different, suggesting no clear association with these pathogenic genomic alterations (Fig. 4c).

As elevated levels of sFR α were detected in ovarian cancer, we considered whether patients mounted a humoral response to FR α -expressing tumours and/or circulating sFR α . Anti-FR α IgG autoantibodies were detected in 17% of serum samples from ovarian cancer patients (2.3 ± 0.8 ng/ml), and in 30% from healthy volunteers (2.6 ± 0.8 ng/ml) (Fig. 4d, left). We found no significant differences in anti-FR α autoantibody levels and proportion of samples with detectable anti-FR α autoantibodies across patient cohorts (Fig. 4d, middle), and no differences between patients with or without detectable circulating sFR α (Fig. 4d, right).

Variations in sFR α are associated with tumour burden

Having confirmed that soluble FR α levels were elevated in the circulation of ovarian cancer patients, we explored whether changes in sFR α levels were indicative of changes in the patients' disease burden as a response to treatment (Fig. 5). First, we devised a tumour burden scoring system for the purpose of this

study, in order to apply a numerical score to the entirety of disease observed by routine clinical imaging at each timepoint in a patient's treatment regimen (Supplementary Table 2). Tumour burden score was used to capture the volume of disease at all sites, instead of selectively focusing on only the target lesions as captured by RECIST score [43]. Using this scoring system, as expected disease burden was significantly higher in patients at the start of neo-adjuvant and palliative treatment, than in the adjuvant cohort (Fig. 5a). The tumour burden score markedly decreased over the course of neo-adjuvant treatment, with a significant reduction measured between timepoints 1 (treatment naïve) and 2 (post-neo-adjuvant chemotherapy, but before debulking surgery). Although a similar trend was observed over the course of palliative treatment, the change in tumour burden score was not significant (Fig. 5b).

Focussing on patients with detectable levels of circulating sFR α at the start of treatment (timepoint 1), we next evaluated the concentration of sFR α in patient serum samples collected over the course of treatment (timepoints outlined in Fig. 1). Similarly to tumour burden score, sFR α levels decreased over the course of neo-adjuvant and palliative treatment, with significant changes between timepoints 1 and 2, and a downward trend was observed in adjuvant patients (Fig. 5c).

In neo-adjuvant patients, we compared concurrent sFR α and tumour burden scores. For sFR α the greatest decrease occurred between timepoints 1 and 2, whereas the decrease in the tumour burden score appeared to be more gradual over the entire course of treatment (Fig. 5d, left). In order to determine whether a reduction in sFR α was associated with a reduction in tumour burden score, we analysed its change between timepoints over the course of treatment. The beta-coefficient estimates of the linear regression of the marker (Fig. 5d, right) confirmed the pattern of change observed in Fig. 5d left and described above. A modest correlation was observed between tumour burden score and the level of circulating of sFR α (Supplementary Fig. 3; $R^2 = 0.14$, $P = 0.004$). Receiver operating characteristic (ROC) curve analysis showed that levels of circulating sFR α in neo-adjuvant treatment-naïve patients were associated with high values of tumour burden score (upper quartile: ≥ 75 th percentile) (AUC = 0.74, $P = 0.005$) (Fig. 5e).

These data suggest that longitudinal sFR α levels may be reflective of the patients' disease burden and changes in disease burden following standard therapies.

Soluble FR α may partly reduce the anti-tumour functions of a FR α -targeted therapeutic antibody candidate

We next evaluated whether the efficacy of anti-FR α treatments may be inhibited by the presence of free FR α in the circulation (Fig. 6). The FR α -specific IgE therapeutic candidate, MOv18 IgE [41, 44–47], bound to FR α -expressing IGROV1 ovarian cancer cells in a concentration dependent manner (EC₅₀ = 0.53 μ g/ml), but not to non-FR α -expressing A2058 and SKBR3 melanoma and breast cancer cells (Fig. 6a). We then explored the impact of circulating FR α on anti-FR α IgE antibody-dependent cellular

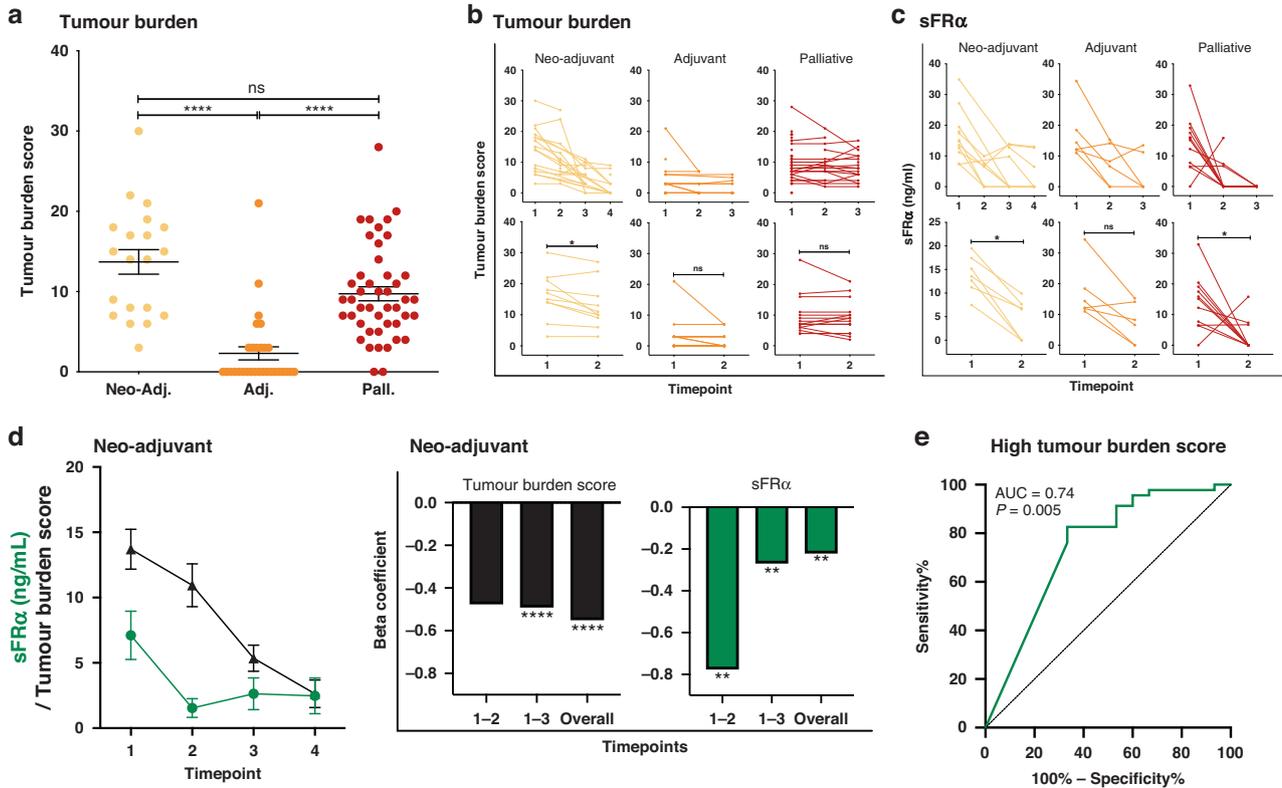


Fig. 5 Serum markers and tumour burden score change during patient treatment. **a** Patient tumour burden score at the start of treatment (calculated as described in Supplementary Table 2) was significantly greater in the neo-adjuvant and palliative ovarian cancer patient cohorts, compared to the adjuvant patient cohort. **b, c** Changes in tumour burden score (**b**) and levels of sFR α (**c**) were measured across treatment timepoints for all ovarian cancer patient cohorts. **d** (Left) Comparison of changes in serum sFR α and tumour burden score over time in patients receiving standard neo-adjuvant treatment regimen. (Right) Beta coefficients illustrate the variation in sFR α among the different time points, namely timepoints 1 and 2, timepoints 1 and 3, and overall across the earliest and latest timepoint for which data were collected. *N* numbers and treatment timepoints as indicated in Fig. 1. **e** Receiver operating characteristic (ROC) curve analysis evaluating the capacity sFR α to predict high values of tumour burden scores in the neo-adjuvant treatment-naive patient group. sFR α levels were predictive of high tumour burden scores (high defined as the upper quartile: ≥ 75 th percentile of tumour burden scores). * $P \leq 0.05$, ** $P \leq 0.01$, **** $P \leq 0.0001$. Statistical tests: *t* test, one-way ANOVA with Kruskal–Wallis multiple comparisons, receiver operating characteristic (ROC) curve analyses and linear regression analysis. Error bars represent the standard error of mean (SEM).

cytotoxicity (ADCC) of cancer cells (Fig. 6b). ADCC of FR α -expressing IGROV1 cancer cells by human peripheral blood mononuclear cells (PBMCs) mediated by low concentrations of MOv18 IgE below or at the EC₅₀ (0.2 or 0.6 $\mu\text{g/ml}$), were significantly-reduced by high (20 ng/ml), but not by low (4 ng/ml), recombinant FR α concentrations (≥ 20 ng/ml being the serum concentration measured in the top 8% of our patients) (Fig. 6c). In contrast, ADCC mediated by MOv18 IgE at levels close to target saturation (1.7 $\mu\text{g/ml}$) was somewhat, but not significantly perturbed. Furthermore, at saturating levels of IgE (5 $\mu\text{g/ml}$), ADCC by both PBMCs (Fig. 5c) and U937 human monocytic cells (Fig. 6d) was not reduced by the presence of even the highest levels of soluble FR α (20 ng/ml and 200 ng/ml, respectively).

These data suggest that the efficacy of anti-FR α antibodies may be impaired by high concentrations of circulating free FR α antigen (as measured in the 8% of ovarian cancer patients with the highest levels of sFR α ; see Fig. 4a). However these partial blocking effects of free FR α were overcome by higher doses of the therapeutic candidate.

DISCUSSION

It has been previously reported that FR α may be a potential therapeutic target for ovarian cancer, and that this tumour-associated antigen is also shed from the tumour into the patient circulation [14, 25, 48, 49]. However, extensive studies to identify

the levels of this marker in relation to disease burden and treatment have been lacking. In this study we evaluated FR α tumour expression and levels of sFR α in the circulation, with the aim of further characterising the clinico-pathologic correlates of this potential clinical biomarker, and of exploring the role of sFR α as a surrogate for tumoural FR α expression, tumour burden and response to treatment.

We first confirmed that gene expression of *FOLR1* (for FR α) and of members of the folate signalling pathway (*FOLH1*, *TYMS* and *DHFR*), were elevated in ovarian cancer cell lines and ovarian cancer tissues, compared to normal tissues (Fig. 2 and Supplementary Fig. 1). We furthermore evaluated FR α in normal tissues ($N = 142$), ovarian tumours ($N = 316$), patient serum ($N = 143$), and serum from healthy volunteers ($N = 61$) (Fig. 1). While no membrane or cytoplasmic FR α staining was observed in a broad range of normal tissues evaluated by IHC (Fig. 2), around half of ovarian tumour samples expressed FR α , either in the cytoplasm, on the cell membrane, or both, with 60% of the high-grade serous tumours being positive for FR α expression (Figs. 2 and 3). These data are consistent with previously published studies reporting FR α expression by 75–85% of high-grade serous samples [21, 22, 50]. As previously observed [14, 25], sFR α levels were significantly raised in patients, compared to healthy subjects, supporting the potential of sFR α as a serum marker for disease (Fig. 4). To explore the clinical correlates of FR α , we considered the levels of sFR α in different patient cohorts. The concentration of

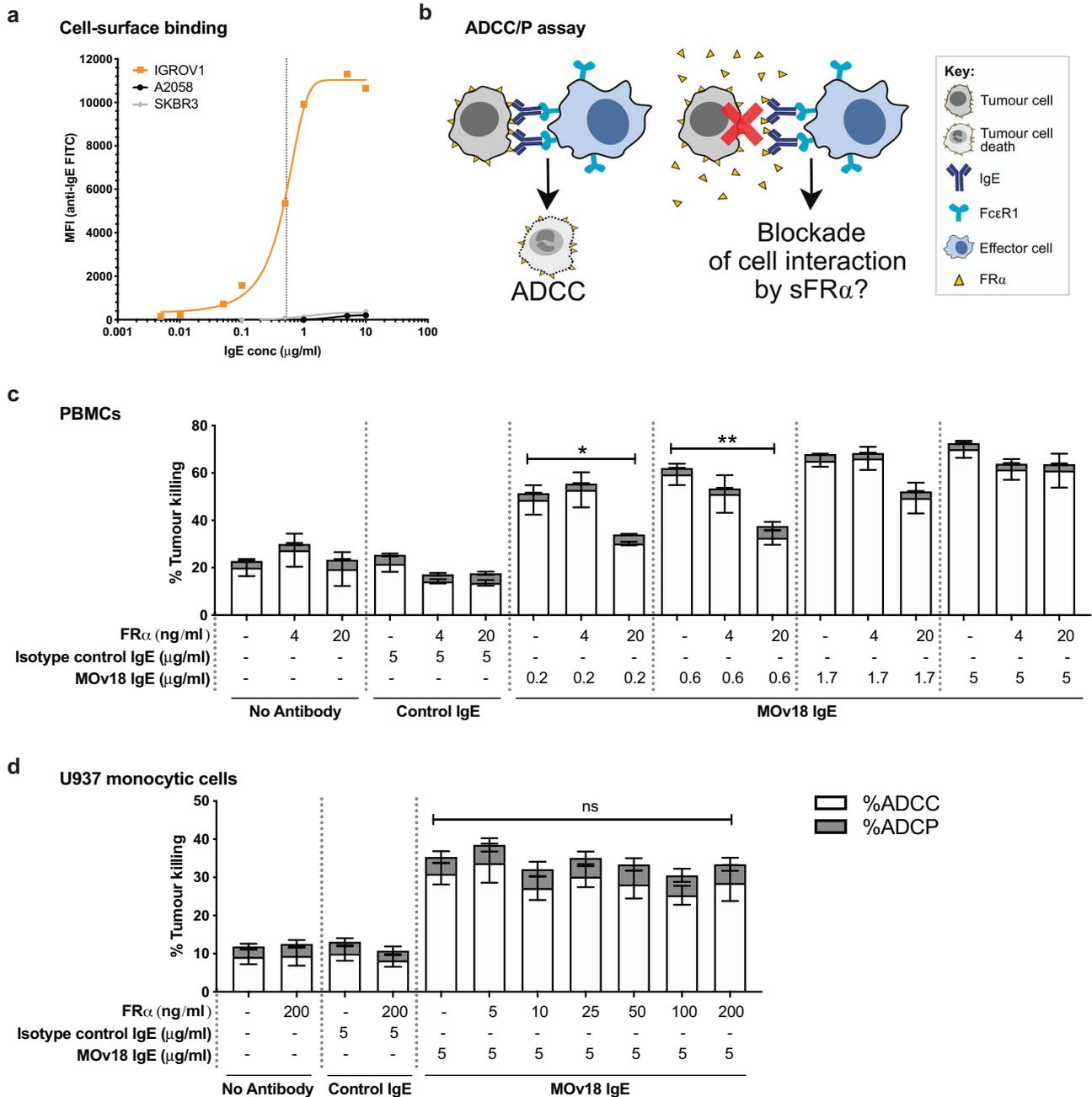


Fig. 6 Potential blockade of efficacy of a FR α -targeted therapeutic antibody candidate by soluble FR α . **a** Binding of MOv18 IgE to FR α -expressing IGROV1 ovarian, but not to melanoma (A2058) or breast (SKBR3), cancer cells ($EC_{50} = 0.53 \mu\text{g/ml}$ for binding to IGROV1 cells, indicated by vertical dotted line). **b** Schematic of potential blockade of MOv18 IgE anti-tumour function by sFR α . **c** Using PBMCs, the level of antibody-dependent cellular cytotoxicity (ADCC) mediated by the FR α -specific antibody MOv18 IgE was significantly reduced by a high concentration of FR α antigen (20 ng/ml) where MOv18 IgE was introduced at non-saturating concentrations of cancer cell-associated FR α (0.2 and 0.6 $\mu\text{g/ml}$; below or at the EC_{50} as shown in **a**). High concentrations of FR α antigen (20 ng/ml) did not block ADCC mediated by concentrations of MOv18 IgE above the EC_{50} (see **a**). **d** Similarly, with U937 monocytic cells, ADCC mediated by a saturating concentration of MOv18 IgE (5 $\mu\text{g/ml}$; see **a**) was not blocked by supraphysiological concentrations of FR α antigen (200 ng/ml). Antibody-dependent cellular phagocytosis (ADCP) was not mediated by MOv18 IgE. $N = 6$ and 4 independent experiments, respectively. * $P \leq 0.05$, ** $P \leq 0.01$, ns not significantly different. Statistical test: one-way ANOVA with Kruskal–Wallis multiple comparisons. Error bars represent the standard error of mean (SEM).

sFR α and the proportion of patients with detectable sFR α were greater in patients at the start of neo-adjuvant or palliative treatments, compared to patients commencing adjuvant chemotherapy, in whom debulking surgery had recently been performed.

Since we measured higher levels of sFR α in ovarian cancer patient cohorts likely to have a higher disease burden (namely those commencing neo-adjuvant and palliative chemotherapy), we developed a scoring system to quantify the burden of disease as observed in concurrent imaging (Fig. 5). As expected, tumour

burden scores were significantly higher at baseline timepoints for patients in the neo-adjuvant and palliative cohorts, compared to the adjuvant cohort. In addition, in the neo-adjuvant and palliative cohorts, the tumour burden score showed a downward trend across longitudinal treatment timepoints. Circulating sFRa levels were also observed to fall across sequential treatment timepoints, however beta coefficients demonstrated that the changes were more marked between early treatment timepoints, compared to a more delayed decline in tumour burden score. Furthermore, ROC curve analysis showed that sFRa levels were found to be associated with tumour burden score (Fig. 5e). Taken together, our findings suggest that sFRa should be further explored as a useful non-invasive marker, in evaluating disease burden and measuring response to treatment in some patients, as well as selection of patients for potential FRa-targeting therapies. Since our analyses were performed on a limited number of patient samples, future studies in larger data sets are required.

In addition to exploring the clinical correlates of sFRa, we examined the possible role of sFRa as a surrogate for tumour FRa expression. Levels of sFRa were significantly higher in patients with tumours expressing FRa on the cell membrane, compared to patients with tumours negative for FRa. Detectable sFRa in the patient circulation was predictive of FRa-positive tumour cell membrane expression (Fig. 4a). These data are in concordance with the hypothesis that tumours expressing FRa can shed this antigen into the circulation [25, 48].

A number of FRa-targeting therapeutics are currently in pre-clinical and clinical development [12, 26, 31], including our own FRa-specific IgE antibody (MOv18 IgE) [41, 44–47, 51]. Here we demonstrate that the patients with FRa-expressing tumours, which could be targeted with such therapies, are likely to have circulating sFRa. Therefore, we evaluated whether this soluble tumour-associated antigen could act as a decoy to block cancer cell recognition and consequently treatment efficacy (Fig. 6). High concentrations of FRa, equivalent to those measured in sera from 8% of patients, perturbed the ADCC of FRa-expressing tumour cells by immune effector cells mediated by low concentrations of MOv18 IgE. However, at higher antibody concentrations, the anti-tumoural potency of MOv18 IgE was unaltered, suggesting that the number of antigen-binding sites on the antibody outweighed any blocking effects of free FRa antigen and were thus sufficient to trigger cancer cell cytotoxicity. To overcome any decoy functions of sFRa, in future it may be necessary to increase the required therapeutic dose of anti-FRa therapies in the minority of patients with the greatest levels of sFRa. In the case of MOv18 IgE, this limitation may be overcome by the kinetic properties of this antibody class: once engaged with FcεRI-expressing immune effector cells, MOv18 IgE may be trafficked away from the circulation into the tumour where the majority of FRa may be present on the target tumour cells rather than in soluble form. Predictions for the safe administration of FRa-specific IgE antibody in patients with detectable sFRa (and/or anti-FRa autoantibodies) in the circulation have been previously made using ex vivo mast cell degranulation and basophil activation test (BAT) approaches [37, 39]. The associations between MOv18 IgE efficacy and safety, with levels of sFRa, will be studied in the ongoing Phase I clinical trial of this therapeutic candidate (ClinicalTrials.gov Identifier NCT02546921).

CONCLUSIONS

We demonstrate that FRa overexpression, at the cell membrane and in the cytoplasm, is associated with ovarian cancer, presenting a promising therapeutic target. sFRa shed into the circulation of ovarian cancer patients was most markedly observed at the start of neo-adjuvant and palliative treatment and was predictive of tumour cell surface but not of cytoplasmic FRa expression. Furthermore, sFRa was reflective of patients' disease burden and

response to standard treatment, as measured with our devised tumour burden score. Therefore, sFRa may present a useful and dynamic non-invasive marker for tumour FRa expression, which could be used to monitor patient response to treatment, and which may reflect potential for resistance to FRa-targeting therapies or a requirement for increased drug administration if detected at high levels. The potential utility of sFRa should be evaluated further in prospective studies of large patient cohorts.

DATA AVAILABILITY

The data that support the findings of this study are available from the corresponding author upon reasonable request.

REFERENCES

- Menon U, Gentry-Maharaj A, Burnell M, Singh N, Ryan A, Karpinskyj C, et al. Ovarian cancer population screening and mortality after long-term follow-up in the UK Collaborative Trial of Ovarian Cancer Screening (UKTOCS): a randomised controlled trial. *Lancet*. 2021;397:2182–93.
- Gonzalez-Martin A, Pothuri B, Vergote I, DePont Christensen R, Graybill W, Mirza MR, et al. Niraparib in patients with newly diagnosed advanced ovarian cancer. *N Engl J Med*. 2019;381:2391–402.
- Moore K, Colombo N, Scambia G, Kim BG, Oaknin A, Friedlander M, et al. Maintenance olaparib in patients with newly diagnosed advanced ovarian cancer. *N Engl J Med*. 2018;379:2495–505.
- Perren TJ, Swart AM, Pfisterer J, Ledermann JA, Pujade-Lauraine E, Kristensen G, et al. A phase 3 trial of bevacizumab in ovarian cancer. *N Engl J Med*. 2011;365:2484–96.
- Ray-Coquard I, Pautier P, Pignata S, Perol D, Gonzalez-Martin A, Berger R, et al. Olaparib plus bevacizumab as first-line maintenance in ovarian cancer. *N Engl J Med*. 2019;381:2416–28.
- Cancer Research UK. <https://www.cancerresearchuk.org/health-professional/cancer-statistics/statistics-by-cancer-type/ovarian-cancer/survival#heading=Three>. Accessed February.
- Lee JY, Yoon JK, Kim B, Kim S, Kim MA, Lim H, et al. Tumor evolution and intratumor heterogeneity of an epithelial ovarian cancer investigated using next-generation sequencing. *BMC Cancer*. 2015;15:85.
- Bast RC Jr., Lu Z, Han CY, Lu KH, Anderson KS, Drescher CW, et al. Biomarkers and strategies for early detection of ovarian cancer. *Cancer Epidemiol Biomark Prev*. 2020;29:2504–12.
- Luhrs CA, Slomiany BL. A human membrane-associated folate binding protein is anchored by a glycosyl-phosphatidylinositol tail. *J Biol Chem*. 1989;264:21446–9.
- Sierra EE, Goldman ID. Recent advances in the understanding of the mechanism of membrane transport of folates and antifolates. *Semin Oncol*. 1999;26:11–23.
- Cheung A, Opzoozer J, Ilieva KM, Gazinska P, Hoffmann RM, Mirza H, et al. Anti-folate receptor alpha-directed antibody therapies restrict the growth of triple-negative breast cancer. *Clin Cancer Res*. 2018;24:5098–111.
- Cheung A, Bax HJ, Josephs DH, Ilieva KM, Pellizzari G, Opzoozer J, et al. Targeting folate receptor alpha for cancer treatment. *Oncotarget*. 2016;7:52553–74.
- Yang R, Kolb EA, Qin J, Chou A, Sowers R, Hoang B, et al. The folate receptor alpha is frequently overexpressed in osteosarcoma samples and plays a role in the uptake of the physiologic substrate 5-methyltetrahydrofolate. *Clin Cancer Res*. 2007;13:2557–67.
- O'Shannessy DJ, Somers EB, Palmer LM, Thiel RP, Oberoi P, Heath R, et al. Serum folate receptor alpha, mesothelin and megakaryocyte potentiating factor in ovarian cancer: association to disease stage and grade and comparison to CA125 and HE4. *J Ovarian Res*. 2013;6:29.
- O'Shannessy DJ, Somers EB, Maltzman J, Smale R, Fu YS. Folate receptor alpha (FRA) expression in breast cancer: identification of a new molecular subtype and association with triple negative disease. *Springerplus*. 2012;1:22.
- Bueno R, Appasani K, Mercer H, Lester S, Sugarbaker D. The alpha folate receptor is highly activated in malignant pleural mesothelioma. *J Thorac Cardiovasc Surg*. 2001;121:225–33.
- Allard JE, Risinger JI, Morrison C, Young G, Rose GS, Fowler J, et al. Overexpression of folate binding protein is associated with shortened progression-free survival in uterine adenocarcinomas. *Gynecol Oncol*. 2007;107:52–7.
- Shi H, Guo J, Li C, Wang Z. A current review of folate receptor alpha as a potential tumour target in non-small-cell lung cancer. *Drug Des Dev Ther*. 2015;9:4989–96.
- Boogerd LS, Boonstra MC, Beck AJ, Charehbili A, Hoogstins CE, Prevoo HA, et al. Concordance of folate receptor-alpha expression between biopsy, primary tumor and metastasis in breast cancer and lung cancer patients. *Oncotarget*. 2016;7:17442–54.

20. Markert S, Lassmann S, Gabriel B, Klar M, Werner M, Gitsch G, et al. Alpha-folate receptor expression in epithelial ovarian carcinoma and non-neoplastic ovarian tissue. *Anticancer Res.* 2008;28:3567–72.
21. Kalli KR, Oberg AL, Keeney GL, Christianson TJ, Low PS, Knutson KL, et al. Folate receptor alpha as a tumor target in epithelial ovarian cancer. *Gynecol Oncol.* 2008;108:619–26.
22. Kobel M, Madore J, Ramus SJ, Clarke BA, Pharoah PD, Deen S, et al. Evidence for a time-dependent association between FOLR1 expression and survival from ovarian carcinoma: implications for clinical testing. *An Ovarian Tumour Tissue Analysis consortium study.* *Br J Cancer.* 2014;111:2297–307.
23. Leung F, Dimitromanolakis A, Kobayashi H, Diamandis EP, Kulasingam V. Folate-receptor 1 (FOLR1) protein is elevated in the serum of ovarian cancer patients. *Clin Biochem.* 2013;46:1462–8.
24. Yang XY, Mackins JY, Li QJ, Antony AC. Isolation and characterization of a folate receptor-directed metalloprotease from human placenta. *J Biol Chem.* 1996;271:11493–9.
25. Basal E, Eghbali-Fatourech GZ, Kalli KR, Hartmann LC, Goodman KM, Goode EL, et al. Functional folate receptor alpha is elevated in the blood of ovarian cancer patients. *PLoS ONE.* 2009;4:e6292.
26. Scaranti M, Cojocaru E, Banerjee S, Banerji U. Exploiting the folate receptor alpha in oncology. *Nat Rev Clin Oncol.* 2020;17:349–59.
27. Vergote I, Armstrong D, Scambia G, Teneriello M, Sehouli J, Schweizer C, et al. A randomized, double-blind, placebo-controlled, phase III study to assess efficacy and safety of weekly farletuzumab in combination with carboplatin and taxane in patients with ovarian cancer in first platinum-sensitive relapse. *J Clin Oncol.* 2016;34:2271–8.
28. Sato S, Itamochi H. Profile of farletuzumab and its potential in the treatment of solid tumors. *Oncotargets Ther.* 2016;9:1181–8.
29. Moore K, Oza A, Colombo N, Oaknin A, Scambia G, Lorusso D, et al. FORWARD I (GOG 3011): a phase III study of mirvetuximab soravtansine, a folate receptor alpha (FRA)-targeting antibody-drug conjugate, versus chemotherapy in patients with platinum-resistant ovarian cancer [abstract 9920]. *Ann Oncol.* 2019;30:v403–434.
30. Oza A, Vergote I, Gilbert LG, Ghatage P, Lisyankaya A, Ghamande S, et al. A randomized double-blind phase III trial comparing vintafolide (EC145) and pegylated liposomal doxorubicin (PLD/Doxil®/Caelyx®) in combination versus PLD in participants with platinum-resistant ovarian cancer (PROCEED). *Gynecol Oncol.* 2015;137:436–45.
31. Matulonis UA, Oaknin A, Pignata S, Hannelore D, Colombo N, Van Gorp T, et al. Abstract 5512: Mirvetuximab soravtansine (MIRV) in patients with platinum-resistant ovarian cancer with high folate receptor alpha (FRα) expression: characterization of antitumor activity in the SORAYA study. *J Clin Oncol.* 2022;40:5512–5512.
32. Ebel W, Routhier EL, Foley B, Jacob S, McDonough JM, Patel RK, et al. Preclinical evaluation of MORAb-003, a humanized monoclonal antibody antagonizing folate receptor-alpha. *Cancer Immun.* 2007;7:6.
33. Györfy B, Lanczky A, Szallasi Z. Implementing an online tool for genome-wide validation of survival-associated biomarkers in ovarian-cancer using microarray data from 1287 patients. *Endocr Relat Cancer.* 2012;19:197–208.
34. Uhlen M, Zhang C, Lee S, Sjostedt E, Fagerberg L, Bidkhori G, et al. A pathology atlas of the human cancer transcriptome. *Science.* 2017;357:eaan2507.
35. Goldman MJ, Craft B, Hastie M, Repecka K, McDade F, Kamath A, et al. Visualizing and interpreting cancer genomics data via the Xena platform. *Nat Biotechnol.* 2020;38:675–8.
36. Tang Z, Li C, Kang B, Gao G, Li C, Zhang Z. GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses. *Nucleic Acids Res.* 2017;45:W98–102.
37. Bax HJ, Khiabany A, Stavrika C, Pellizzari G, Chan Wah Hak C, Robinson A, et al. Basophil activation test in cancer patient blood evaluating potential hypersensitivity to an anti-tumor IgE therapeutic candidate. *Allergy.* 2020;75:2069–73.
38. Christoph DC, Asuncion BR, Hassan B, Tran C, Maltzman JD, O'Shannessy DJ, et al. Significance of folate receptor alpha and thymidylate synthase protein expression in patients with non-small-cell lung cancer treated with pemetrexed. *J Thorac Oncol.* 2013;8:19–30.
39. Rudman SM, Josephs DH, Cambrook H, Karagiannis P, Gilbert AE, Dodev T, et al. Harnessing engineered antibodies of the IgE class to combat malignancy: initial assessment of FcγεR-mediated basophil activation by a tumour-specific IgE antibody to evaluate the risk of type I hypersensitivity. *Clin Exp Allergy.* 2011;41:1400–13.
40. Bracher M, Gould HJ, Sutton BJ, Dombrowicz D, Karagiannis SN. Three-colour flow cytometric method to measure antibody-dependent tumour cell killing by cytotoxicity and phagocytosis. *J Immunol Methods.* 2007;323:160–71.
41. Josephs DH, Bax HJ, Dodev T, Georgouli M, Nakamura M, Pellizzari G, et al. Anti-folate receptor-alpha IgE but not IgG recruits macrophages to attack tumors via TNFα/MCP-1 signaling. *Cancer Res.* 2017;77:1127–41.
42. Buist MR, Molthoff CF, Kenemans P, Meijer CJ. Distribution of OV-TL 3 and MOv18 in normal and malignant ovarian tissue. *J Clin Pathol.* 1995;48:631–6.
43. Schwartz LH, Litiere S, de Vries E, Ford R, Gwyther S, Mandrekas S, et al. RECIST 1.1-update and clarification: from the RECIST committee. *Eur J Cancer.* 2016;62:132–7.
44. Gould HJ, Mackay GA, Karagiannis SN, O'Toole CM, Marsh PJ, Daniel BE, et al. Comparison of IgE and IgG antibody-dependent cytotoxicity in vitro and in a SCID mouse xenograft model of ovarian carcinoma. *Eur J Immunol.* 1999;29:3527–37.
45. Karagiannis SN, Bracher MG, Hunt J, McCloskey N, Beavil RL, Beavil AJ, et al. IgE-antibody-dependent immunotherapy of solid tumors: cytotoxic and phagocytic mechanisms of eradication of ovarian cancer cells. *J Immunol.* 2007;179:2832–43.
46. Karagiannis SN, Wang Q, East N, Burke F, Riffard S, Bracher MG, et al. Activity of human monocytes in IgE antibody-dependent surveillance and killing of ovarian tumor cells. *Eur J Immunol.* 2003;33:1030–40.
47. Mantovani LT, Miotti S, Menard S, Canevari S, Raspagliesi F, Bottini C, et al. Folate binding protein distribution in normal tissues and biological fluids from ovarian carcinoma patients as detected by the monoclonal antibodies MOv18 and MOv19. *Eur J Cancer.* 1994;30A:363–9.
48. Kurosaki A, Hasegawa K, Kato T, Abe K, Hanaoka T, Miyara A, et al. Serum folate receptor alpha as a biomarker for ovarian cancer: Implications for diagnosis, prognosis and predicting its local tumor expression. *Int J Cancer.* 2016;138:1994–2002.
49. Toffoli G, Cernigoi C, Russo A, Gallo A, Bagnoli M, Boiocchi M. Overexpression of folate binding protein in ovarian cancers. *Int J Cancer.* 1997;74:193–8.
50. Crane LM, Arts HJ, van Oosten M, Low PS, van der Zee AG, van Dam GM, et al. The effect of chemotherapy on expression of folate receptor-alpha in ovarian cancer. *Cell Oncol.* 2012;35:9–18.
51. Spicer J, Basu B, Montes A, Banerji U, Kristeleit R, Veal GJ. Abstract CT141: phase 1 trial of MOv18, a first-in-class IgE antibody therapy for cancer. *Cancer Res.* 2020;80:CT141.

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AUTHOR CONTRIBUTIONS

HJB, SNK and DHJ conceived and designed the study, and developed the methodology. HJB, JC, CS, A Santaolalla, GO, AK, MG, JL-A, LCGFP, CCWH, AR, AP, NW, CN-L, S Malas, JEMC, MN, RL, ST, KFG, CG, SP, MVH and DHJ acquired the data or helped with the data analysis. AB, SL, A Sayasneh, RK, SG, AM, JS and DHJ provided clinical support to recruit patients and volunteers. HJB, JC, CS, MN, S Mele, S Crescioli and DHJ acquired samples. S Canevari and MF provided key reagents and expertise. HJB, JC, CS, JS, SNK and DHJ discussed and interpreted the data, and wrote and edited the manuscript. SNK and DHJ supervised the study.

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COMPETING INTERESTS

SNK and JS are founders and shareholders of Epsilogen Ltd. KFG is an employee of Epsilogen Ltd. HJB, MG, JL-A and LCGFP are employed through a fund provided by Epsilogen Ltd. SNK, JS, DHJ, HJB and KFG are inventors of patents on antibody technologies. All other authors have declared no conflict of interest.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Sample collection for this study has been reviewed and approved by the Guy's Research Ethics Committee (References 09/H0804/45 and 16/LO/0366). Ovarian cancer patients and healthy volunteers were enrolled by written, informed consent.

CONSENT FOR PUBLICATION

Not applicable.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41416-022-02031-x>.

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