Check for updates

CORRESPONDENCE **OPEN** Real-world impact of bridging therapy on outcomes of ide-cel for myeloma in the U.S. Myeloma Immunotherapy Consortium

© The Author(s) 2024

Blood Cancer Journal (2024)14:63; https://doi.org/ 10.1038/s41408-024-00993-0

Idecabtagene vicleucel (ide-cel) the first FDA-approved gene therapy for relapsed refractory multiple myeloma (RRMM). However, its administration presents challenges in logistical management, selecting bridging therapy (BT), and customizing T-cell manufacturing, a complex process spanning several weeks [1]. In the KarMMa trial, BT was allowed but limited to specific prior drug classes (e.g., dexamethasone, cyclophosphamide (Cy), daratumumab, carfilzomib, bortezomib, or pomalidomide) [2]. This study delves into the impact of different BT on outcomes for RRMM patients undergoing standard of care (SOC) ide-cel treatment, aiming to clarify their role in CAR T therapy outcomes.

This retrospective multicenter study observed RRMM patients receiving ide-cel treatment at 11 U.S. medical centers within the U.S. Myeloma Immunotherapy Consortium. All RRMM patients treated with ide-cel from 5/2021 to 5/2022, were included. BT was defined as systemic treatment between leukapheresis and CAR-T infusion, categorized into: Selinexor (Selinexor-containing regimens); alkylator (alkylator-based); PI combos (sole or combined proteasome inhibitor (PI) therapy); IMiD +/- mAb combos (steroids with/without immunomodulatory (IMiDs) and/or Monoclonal Antibodies (mAb).

High-risk cytogenetics included del (17p), t(4;14), and t(14;16) pre-CAR-T. Cytokine Release Syndrome (CRS) and Immune Effector Cell-Associated Neurotoxicity Syndrome (ICANS) were graded by American Society for Transplantation and Cellular Therapy criteria [3]. Hematologic toxicity was assessed using Common Terminology Criteria for Adverse Events version 5.0 [4]. Disease response followed the revised International Myeloma Working Group criteria [5].

Subgroup comparisons employed chi-square/Fisher's exact tests for categorical, t-tests/ANOVA for continuous variables. Survival analysis used Kaplan-Meier/Cox proportional hazards model. Stepwise Cox regression identified significant variables associated with overall survival (OS) and progression free survival (PFS), based on p-values < 0.2 from initial analysis. SAS (v9.4) and IBM SPSS (v29.0) conducted all statistical analyses.

Of 214 ide-cel patients, 170 (79%) underwent BT, encompassing 12% Selinexor, 45% alkylator, 15% PI combos, 18% IMiD +/mAb combos, and 11% other therapies (e.g., belantamab mafodotin, focal radiation). Forty-four patients (21%) did not receive BT (no-BT). The median BT duration was 1 month (range, 1-7). While most had 1–2 months, one patient had 7 months with 4 treatments. BT patients showed poorer eastern Cooperative Oncology Group Performance Status (ECOG), higher Revised International Staging System (R-ISS) (2-3), elevated ferritin (>300 ng/mL), and C-reactive protein (CRP) (>5 mg/L) at lymphodepleting chemotherapy (Supplementary 1). IMiD +/mAb combos had less extramedullary disease (EMD) (p = 0.052), and alkylator groups often received alkylator as last preapheresis treatment (p < 0.001). No differences in ECOG, R-ISS, ferritin, CRP, or high-risk cytogenetics among BT subgroups emerged. Median prior therapy count, and penta-refractory status were similar between BT and no-BT groups, including BT subgroups.

No significant differences in incidence/severity of CRS between BT and no-BT groups were noted, consistent across BT subgroups. Although ICANS incidence was higher in the BT, it lacked statistically significance (any grade: 21% vs. 14%, p = 0.070; grade \geq 2: 13% vs. 2%, p = 0.070). Within the BT subgroup, Selinexor showed notably higher ICANS (grade ≥ 2) at 38% compared to others (alkylator 9%, PI combos 0%, IMiD +/- mAb combos 17%; p = 0.023). Notably, patients receiving Selinexor had no known central nervous system pathology. The reason is unclear, but endothelial dysfunction and increased blood-brain barrier permeability [6] might contribute to the higher ICANS rate in this subaroup.

BT patients had longer hospital stays than no-BT (median: 10 vs. 8 days; p < 0.001). Alkylator and Selinexor had the lengthiest stays (median: 11 and 10.5 days, respectively), followed by PI combos (9 days) and IMiD±mAb combos (9 days).

At day 7 post-infusion, no cytopenia differences emerged between BT and no-BT groups. However, Selinexor caused highergrade anemia (p < 0.001) and thrombocytopenia (p = 0.043).

At 3 months post-infusion, BT showed higher rates of any-grade neutropenia (47% vs. 27.5%, p = 0.030), and anemia (79% vs. 50%, p < 0.001), with no significant differences in severe cases (≥ 3). Thrombocytopenia didn't differ between groups or subgroups (supplementary 2).

At 3 months post-CAR-T, response didn't differ between BT and no-BT (p = 0.802 for overall response rate (\geq partial response (PR); p = 0.208 for complete response (CR), consistent across BT subgroups (supplementary 3).

Median follow-up: 9.7 months (range: 0.2-19.5). Median PFS: 8.16 months (95% confidence interval (Cl): 6.61-9.31), OS: not reached (NR). 1-year PFS and OS rates: 36 and 63%, respectively. BT patients showed inferior PFS (6.68 vs. 11.48 months in no-BT, p = 0.007) and OS (13.85 vs. NR months in no-BT, p = 0.002) , (Table 1, Fig. 1A, B).

IMiD±mAb combos showed comparable PFS to no-BT (median PFS: 12.01 months (95% CI, 5.79-NR) vs. 11.48 months (95% CI, 9.05–17.73); p = 0.56). Other therapies exhibited varying PFS durations in comparison with IMiD±mAb combos: Selinexor (9.77 months, 95% Cl, 4.11–13.88; p = 0.48), Pl combos (6.41 months, 95% Cl, 2.70–12.50; p = 0.24), and alkylator (6.51 months, 95% Cl, 4.18–8.16; p = 0.030) (Fig. 1C).

Alkylator use resulted in inferior OS (median OS: 11.97 months, 95% CI: 8.91-15.53) compared to others (NR) (p = 0.001) (Fig. 1D). Seventy deaths occurred: 49 diseaserelated, 4 due to CRS/ICANS, 1 CAR-T myocarditis, 10 infectionrelated, rest unrelated.

Received: 22 October 2023 Revised: 15 January 2024 Accepted: 17 January 2024 Published online: 12 April 2024

Table 1. Outcome based on Bridging therapy.	ased on	Bridging therapy.									
	No.	Bridging therapy (BT)	r (BT)		No.	Bridging therapy type	ype				
Outcome		Yes (n, %)	No (<i>n</i> , %)	P-value		No BT (N = 44)	Selinexor (N = 21)	Alkylator (N = 76)	PI combos (N = 25)	IMiD Combo (N = 30)	P-value
Best CR or better at 3 months	172	54 (41%)	21 (52.5%)	0.208	157	21 (52.5%)	5 (26%)	22 (42%)	7 (37%)	12 (44%)	0.412
Best ORR at 3 months	172	111 (84%)	35 (87.5%)	0.802	157	35 (87.5%)	15 (79%)	45 (87%)	15 (79%)	26 (96%)	0.387
MRD negativity at 3 months	95	52 (72%)	20 (87%)	0.175	119	20 (89%)	8 (61.5%)	13 (56.5%)	5 (83%)	18 (90%)	0.042
Best CR or better at 6 months	126	44 (47%)	17 (53%)	0.547	119	17 (53%)	4 (33%)	18 (43%)	6 (54.5%)	12 (54.5%)	0.666
Best ORR at 6 months	126	76 (81%)	30 (94%)	0.099	85	30 (94%)	10 (83%)	31 (74%)	10 (91%)	19 (86%)	0.199
PFS	213			0.007							0.010
Median PFS (95% CI), months		6.68 (5.79–8.49)	11.48 (9.05–17.73)			11.48 (9.05–17.73)	9.77 (4.11–13.88)	6.51 (4.18–8.16)	6.41 (2.70–12.50)	12.01 (5.79- NR)	
SO	213			0.002							0.001
Median OS (95% Cl), months		13.85 (11.97- NR)	NR			NR-NR	NR-NR	11. <i>97</i> (8.91–15.53)	NR-NR	NR-NR	
Best response: \geq complete response, best overall response rate \geq partial response, <i>Cl</i> Confidence interval, <i>NR</i> not reached. <i>PFS</i> progression-free survival, <i>OS</i> Overall survival, <i>BT</i> bridging therapy, Selinexor containing Selinexor as part of regimen, Alkylator containing alkylator as part of regimen, <i>Pl</i> combos containing proteasome inhibitor alone or in combination, <i>IMID</i> combos containing steroid \pm immunomodulator (IMIDs) \pm monoclonal antibodies (MoA), no BT no bridging therapy, <i>P</i> -values \leq 0.05 are shown in bold.	plete res as part c liDs) ± m	ponse, best overall of regimen, Alkylati onoclonal antibodie	response rate ≥ par or containing alkyla ≘s (MoA), no BT no b	tial response, (tor as part of ridging therap	<i>CI</i> Confic f regime y, <i>P</i> -valu	tence interval, NR no n, Pl combos conta- ies ≤ 0.05 are shown	t reached. <i>PFS</i> progr ining proteasome in in bold.	tial response, Cl Confidence interval, NR not reached. PFS progression-free survival, OS Overall survival, BT bridging therapy, Selinexor tor as part of regimen, Pl combos containing proteasome inhibitor alone or in combination, <i>IMiD</i> combos containing steroid \pm oridging therapy, P-values \leq 0.05 are shown in bold.	05 Overall survival, <i>BT</i> combination, <i>IMiD</i> co	⁻ bridging therapy, mbos containing	Selinexor steroid ±

The univariate Cox regression for OS and PFS confirmed inferior outcomes among patients with ECOG 2–4, R-ISS 3, EMD, last pre- apheresis treatment with BCMA-targeted therapy, alkylator-based BT, and lack of CR by 3 months. Stepwise Cox regression revealed a significant association of BT with worse PFS (p = 0.027), particularly with alkylator use ((hazard ratio (HR) = 2.54, 95% CI (1.39–4.62); p = 0.002). A similar trend, though not statistically significant, was noted in Selinexor and PI combos. Response less than CR [(PR/VGPR (very good PR): HR = 2.05, p = 0.019; stable/progression disease (SD/PD): HR = 31.32; p < 0.0001)] was associated with inferior PFS. For OS, responses less than CR (HR = 3.91; p = 0.0002) and high-risk cytogenetics (HR = 5.77; p = 0.008) were associated with poore outcomes.

Our analysis highlighted alkylators as linked to worse PFS despite no initial differences in tumor burden or inflammatory markers among BT subgroups. Therefore, we examined alkylator types to gauge their impact. In alkylator BT, 92% received Cy, others had oral melphalan or melphalan flufena-mide; bendamustine was not used. Among Cy-treated patients, 43% had intensified/infusional (hyperfractionated Cy-based (hyperCy), DCEP, PACE), and 57% had weekly doses (e.g., CyBorD, KCD, etc.). Intensified/infusional vs. weekly Cy showed no significant difference in median PFS (4.61 vs. 8.49 months, p = 0.089) or OS (10 vs. 15.5 months, p = 0.11). Comparison of weekly Cy with no-alkylator group showed no significant differences in median PFS (p = 0.3) or OS (p = 0.3).

However, intensified/infusional resulted in poorer outcomes compared to no-alkylator group (median PFS: 4.6 vs. 12.0 months, p = 0.002; median OS: 10 vs. NR months, p = 0.006) (Fig. 1E, F, supplementary 4). In a recent analysis [7] of CAR T-cell therapies for RRMM, grouping BTs by Cy usage (solely hyperCy-based, weekly Cy, no Cy/alkylator) demonstrated comparable CRS, ICANS rates, and PFS. Notably, hyperCy group patients experienced prolonged platelet recovery and lower OS, possibly due to disease aggressiveness.

Note that, in retrospective studies, assessing tumor burden and baseline markers offers only a snapshot, potentially missing full disease progression or patient status. Earlier studies suggest chemotherapy's lasting impact on T cells, potentially affecting their function [8, 9]. While specific pre-apheresis T cell quality data is lacking and no observed PFS difference based on preapheresis alkylator exposure, pre-collection T cell quality might affect outcomes. Moreover, alkylator use in aggressive disease contexts may hinder sustained treatment response, affecting outcomes.

Among 170 BT patients, 12% responded to BT (n = 21; 1 CR, 8 VGPR, 12 PR); response rates didn't significantly differ among subgroups (p = 0.503). BT response showed no significant association with CRS, ICANS rates, or 3-month post CAR-T response (p = 1.00, p = 1.00, p = 0.425, respectively). Median PFS didn't significantly differ based on BT response (6.51 months in \ge PR vs. 8.48 months in SD/PD; p = 0.6). Despite an 88% BT rate in the KarMMa trial, the response remained low at 5% (n = 5) [2]. This implies that BT may not confer superiority or necessity for stable patients. Further evaluation through prospective studies or extensive registry analysis with larger cohorts is essential.

In our study, only 4 patients received BCMA-directed antibody-drug conjugate as BT. They were excluded from subgroup analysis due to limited cases and significantly inferior PFS. No patients received bispecific T-cell engagers as BT.

Limited by retrospective design and lacking BCMA data, reduced PFS might relate to decreased tumor BCMA expression [10]. Our previous data showed lower PFS post BCMA-targeted therapy [11], and hinted at compromised ide-cel efficacy within six months of such treatment [12].

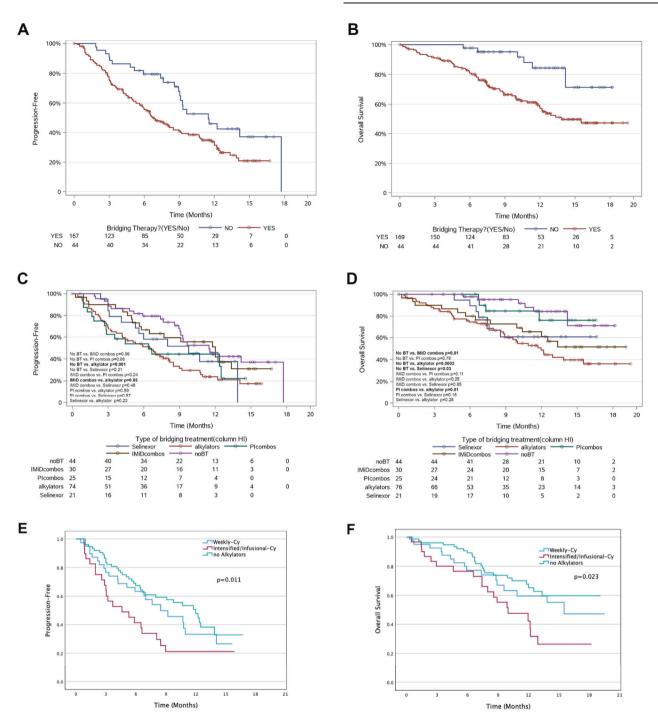


Fig. 1 Survival based on bridging therapy strategy. Kaplan-Meier plots estimates of PFS A and OS B in RRMM patients treated with idecabtagene vicleucel, between those with and without bridging therapy. Kaplan-Meier estimates of PFS C and OS D categorized by bridging therapy type (no-BT, Selinexor, alkylator, PI combos, IMiD±mAb combos). Kaplan-Meier estimates of PFS E and OS F categorized based on cyclophosphamide dosing and exposure as bridging therapy.

The retrospective nature of our study introduces potential selection biases. Additionally, smaller sample sizes within certain treatment regimens limit drawing definitive conclusions, alongside the probable influence of disease aggressiveness on BT choice.

In summary, patients with no-BT before ide-cel showed prolonged PFS and OS, possibly indicating less aggressive disease. Conversely, alkylator-based BT resulted in inferior PFS and OS compared to other BT types or no-BT approaches. This trend might relate more to refractory myeloma than directly to alkylators, proposing the potential benefit of early CAR T-cell therapy before standard treatment resistance. Tailoring BT based on patient history, toxicity risks, and disease traits is crucial. While reconsidering the necessity of BT in stable disease, considering the risk of toxicity due to inadequate cytoreduction remains crucial. However, caution with intensified/infusional Cy, if feasible, is advisable. Personalized assessments are pivotal in selecting bridging therapy. Aimaz Afrough ^{1,16 ⊠}, Hamza Hashmi^{2,16}, Doris K. Hansen ³, Surbhi Sidana ⁴, Chul Ahn⁵, Lauren C. Peres ³, Danai Dima ⁶ Ciara L. Freeman³, Omar Castaneda Puglianini 🔞 Mehmet H. Kocoglu⁷, Shebli Atrash⁸, Peter M. Voorhees⁸ Leyla Shune⁹, Joseph P. McGuirk¹⁰⁹, Gary Simmons¹⁰, Douglas W. Sborov¹¹, James A. Davis², Gurbakhash Kaur¹⁰, Aishwarya Sannareddy¹, Christopher J. Ferreri¹², Mahmoud R. Gaballa¹², Scott Goldsmith¹³, Omar Nadeem¹⁴, Shonali Midha¹⁴, Charlotte B. Wagner ^[11], Frederick L. Locke³, Krina K. Patel ^[12], Jack Khouri^{6,17}, Larry D. Anderson Jr. ^[117] and Yi Lin 15,17 ¹Myeloma, Waldenstrom's, and Amyloidosis Program. Hematoloaic Malianancies and Cellular Therapy Program, Simmons Comprehensive Cancer Center, UT Southwestern Medical Center, Dallas, TX, USA. ²Medical University of South Carolina, Charleston, SC, USA. ³H. Lee Moffitt Cancer Center & Research Institute, Tampa, FL, USA. ⁴Stanford University School of Medicine, Stanford, CA, USA. ⁵Peter O'Donnell Jr. School of Public Health, UT Southwestern Medical Center, Dallas, TX, USA. ⁶Cleveland Clinic Taussig Cancer Center, Cleveland, OH, USA. ⁷University of Maryland Marlene and Stewart Greenebaum Comprehensive Cancer Center, Baltimore, MD, USA. ⁸Levine Cancer Institute, Charlotte, NC, USA. ⁹The University of Kansas Medical Center, Kansas City, KS, USA. ¹⁰Virginia Commonwealth University Massey Cancer Center, Richmond, VA, USA. ¹¹The University of Utah Huntsman Cancer Institute, Salt Lake City, UT, USA. ¹²The University of Texas MD Anderson Cancer Center, Houston, TX, USA. ¹³City of Hope National Medical Center, Duarte, CA, USA. ¹⁴Dana-Farber Cancer Institute, Boston, MA, USA. ¹⁵Mayo Clinic, Rochester, MN, USA.¹⁶These authors contributed eaually: Aimaz Afrough, Hamza Hashmi. ¹⁷These authors jointly supervised this work: Jack Khouri, Larry D. Anderson Jr., Yi Lin. [™]email: Aimaz.afrough@utsouthwestern.edu; larry.anderson@utsouthwestern.edu

DATA AVAILABILITY

The data that support the findings of this study are available from the corresponding author upon reasonable request.

REFERENCES

- Bhaskar ST, Dholaria BR, Sengsayadeth SM, Savani BN, Oluwole OO. Role of bridging therapy during chimeric antigen receptor T cell therapy. EJHaem. 2022;3:39–45.
- Munshi NC, Anderson LD Jr, Shah N, Madduri D, Berdeja J, Lonial S, et al. Idecabtagene vicleucel in relapsed and refractory multiple myeloma. N Engl J Med. 2021;384:705–16.
- Lee DW, Santomasso BD, Locke FL, Ghobadi A, Turtle CJ, Brudno JN, et al. ASTCT consensus grading for cytokine release syndrome and neurologic toxicity associated with immune effector cells. Biol Blood Marrow Transplant. 2019;25:625–38.
- Common Terminology Criteria for Adverse Events (CTCAE) Version 5.0: U.S. Department of Health and Human Services; November 2017 [Available from: https://ctep.cancer.gov/protocoldevelopment/electronic_applications/docs/ ctcae_v5_quick_reference_5x7.pdf.
- Kumar S, Paiva B, Anderson KC, Durie B, Landgren O, Moreau P, et al. International Myeloma Working Group consensus criteria for response and minimal residual disease assessment in multiple myeloma. Lancet Oncol. 2016;17:e328–e46.
- Gust J, Hay KA, Hanafi LA, Li D, Myerson D, Gonzalez-Cuyar LF, et al. Endothelial activation and blood-brain barrier disruption in neurotoxicity after adoptive immunotherapy with CD19 CAR-T cells. Cancer Discov. 2017;7:1404–19.
- Zafar A, Huang CY, Lo M, Arora S, Chung A, Wong SW, et al. Intensity of cyclophosphamide-based bridging therapy before chimeric antigen receptor T cell therapy in myeloma. Transplant Cell Ther. 2023;29:504.e1–504.e7.
- 8. Das RK, O'Connor RS, Grupp SA, Barrett DM. Lingering effects of chemotherapy on mature T cells impair proliferation. Blood Adv. 2020;4:4653–64.
- 9. Das RK, Vernau L, Grupp SA, Barrett DM. Naïve T-cell deficits at diagnosis and after chemotherapy impair cell therapy potential in pediatric cancers. Cancer Discov. 2019;9:492–9.
- Samur MK, Fulciniti M, Aktas Samur A, Bazarbachi AH, Tai Y-T, Prabhala R, et al. Biallelic loss of BCMA as a resistance mechanism to CAR T cell therapy in a patient with multiple myeloma. Nat Commun. 2021;12:868.

- Hansen DK, Sidana S, Peres LC, Leitzinger CC, Shune L, Shrewsbury A, et al. Idecabtagene Vicleucel for Relapsed/Refractory Multiple Myeloma: Real-World Experience From the Myeloma CAR T Consortium. J Clin Oncol. 2023;41:2087–97.
- Ferreri CJ, Hildebrandt MAT, Hashmi H, Shune LO, McGuirk JP, Sborov DW, et al. Real-world experience of patients with multiple myeloma receiving ide-cel after a prior BCMA-targeted therapy. Blood Cancer J. 2023;13:117.

AUTHOR CONTRIBUTIONS

AA, HH, JK, LDA Jr, YL contributed to study design. All authors contributed to data acquisition. AA, CA, HH, JK, LDA Jr, YL contributed to data analysis and interpretation. AA wrote the original draft of the manuscript and incorporated the comments by the co-authors in all subsequent drafts. All authors provided review and edits, and approved the final version of the manuscript. AA and HH contributed equally as co-first authors. JK, LDA Jr, YL contributed equally as co-senior authors.

COMPETING INTERESTS

A.A. reports research funding from Abbvi, Adaptive Biotech, and advisory role for Bristol-Myers Squibb, Karvopharm Therapeutics, H.H. reports consulting or advisory role for Janssen, Bristol-Myers Squibb, Sanofi; Speakers' bureau for Sanofi, GlaxoSmithKline, and Karvopharm, D.K.H. reports research funding from Bristol-Myers Squibb, Janssen, Karyopharm, International Myeloma Society Young Investigator Award, and the Pentecost Family Myeloma Research Center and Adaptive Biotech and consulting or advisory role for Bristol-Myers Squibb, Janssen, Karyopharm and Pfizer, and member of the Bristol-Myers Squibb IMW Ide-Cel Academic Advisory Board, Bristol-Myers Squibb Multiple Myeloma ASH Steering Committee, and Multiple Myeloma Pfizer Advisory Board, and received net honoraria from OncLive and Survivorship. S.S. reports consulting or advisory role for Janssen, Bristol-Myers Squibb, Magenta Therapeutics, Sanofi, Takeda, Pfizer and Legend Biotech; Research funding from Janssen, Magenta Therapeutics, Allogene Therapeutics, Bristol-Myers Squibb and Novartis. C.A report advisory role for Psomagen, and consulting for PPD Global, LSK Global, IAVIA, Advarra, Syneos Health. L.C.P. reports research funding from Bristol-Myers Squibb and Karyopharm. C.L.F. reports honoraria/consulting BMS, Seattle Genetics, Celgene, Abbvie, Sanofi, Incyte, Amgen, and ONK therapeutics & Janssen; and has research funding from BMS, Janssen and Roche/Genentech. O.C.P. reports payment on speaker bureaus from Adaptive Biotechnologies. S.A. reports honoraria from Janssen; Research funding from GlaxoSmithKline, Amgen, Karyopharm Therapeutics, Janssen, Bristol-Myers Squibb; Honoraria from Janssen, P.M.V. reports consulting or advisory role for Oncopeptides, Abbvie/Genentech, Karyopharm Therapeutics, Bristol-Myers Squibb, Secura Bio, Pfizer, Sanofi, Janssen, GlaxoSmithKline; Research funding from Abbvie, Janssen, GlaxoSmithKline, and TeneoBio; Travel, accommodations, and expenses from Sanofi, and involvement on a Data Safety and Monitoring Committee for Sanofi. L.S reports consulting or advisory role for Janssen. J.P.M. reports consultancy fees and honoraria from Magenta Therapeutics, Novartis, Bristol Myers Squibb, Juno Therapeutics, Kite, CRISPR Therapeutics, Nektar, and Allovir, reports honoraria from Sana, reports payment on speaker bureaus from Bristol Myers Squibb, Kite, and Allovir, and research funding from Magenta Therapeutics, Juno Therapeutics, Kite, Orca Bio, and Allovir. G.S. reports speaker bureau payment from Kite/Gilead. D.W.S. reports consulting or advisory role for Sanofi, GlaxoSmithKline, Janssen, Pfizer, Abbvie, Arcellx and BiolineRx: Research funding from Janssen, BioLineRx, Sanofi, Bristol-Myers Squibb, Amgen, Pfizer, Arcellx and Gilead Sciences. G.K. reports consulting or advisory committees for Bristol Myers Squibb, Cellectar, Sanofi, Janssen, and Arcellx, and research funding from Bristol Myers Squibb, Janssen, and Arcellx. C.J.F. reports payment from participation on an advisory board from Sanofi, and currently holds equity in a publicly traded company, Affimed. M.R.G. reports consulting for Boxer Capital, LLC. S.G. reports consultancy fees from BMS, Janssen, Sanofi, Meditope Biosciences, and research funding from BMS, Ipsen, Adaptive Biothechnology, and speaker bureau payment from Janssen, Adaptive Biotech, O.N reports membership on an entity's Board of Directors or advisory committees from Bristol Myers Squibb, Karvopharm, GPCR Therapeutics, Adaptive Biotechnologies, GSK, and Janssen, and reports research funding from Takeda and Janssen. F.L.L reports a scientific advisory role for A2, Allogene, Amgen, Bluebird Bio, BMS/Celgene, Calibr, Cellular Biomedicine Group, GammaDelta Therapeutics, lovance, Kite Pharma, Janssen, Legend Biotech, Novartis, Sana, Takeda, Wugen, Umoja; research funding from Kite Pharma (Institutional), Allogene (Institutional), Novartis (Institutional), Blue-Bird Bio (Institutional), CERo Therapeutics (Institutional), and BMS (Institutional); patents, royalties, and other intellectual property including several patents held by the institution in his name (unlicensed) in the field of cellular immunotherapy; consulting roles for Cowen, EcoR1, Emerging Therapy Solutions, and Gerson Lehrman Group (GLG); and education or editorial activity for Aptitude Health, ASH, BioPharma Communications CARE Education, Clinical Care Options Oncology, Imedex, and Society of Immunotherapy of Cancer. K.K.P reports consulting or advisory role for Bristol-Myers

Squibb, Janssen, Pfizer, Arcellx, and Karyopharm Therapeutics; Research funding from Bristol-Myers Squibb, Poseida Therapeutics, Takeda, Janssen, Cellectis, Nektar, Abbvie/Genentech, Precision Biosciences, and Allogene Therapeutics; Travel, accommodations, and expenses from Bristol-Myers Squibb. J.K. reports honoraria from OncLive. L.A.D. Jr reports honoraria and membership on an entity's Board of Directors or advisory committees for Bristol Myers Squibb, Celgene, GSK, AbbVie, Pharmacyclics, Karyopharm, Janssen, Prothena, Sanofi, Beigene, Cellectar, and Amgen. Y.L. reports consultancy fees from Janssen, Juno, Vineti, Kite/Gilead, Novartis, Legend, Sorrento, Gamida Cell, Celgene, Bluebird Bio, and research funding from Janssen, Kite/Gilead, Merck, Celgene, Bluebird Bio, and Takeda. Remaining authors with no potential conflicts of interest.

ETHICS APPROVAL

This multi-center study was approved by the respective institutions' Institutional Review Board.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at https://doi.org/10.1038/s41408-024-00993-0.

Correspondence and requests for materials should be addressed to Aimaz Afrough or Larry D. Anderson Jr.

Reprints and permission information is available at http://www.nature.com/ reprints

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http:// creativecommons.org/licenses/by/4.0/.

© The Author(s) 2024