



ARTICLE

An integrated PK/PD model investigating the impact of tumor size and systemic safety on animal survival in SW1990 pancreatic cancer xenograft

Qing-yu Yao^{1,2}, Jun Zhou³, Ye Yao¹, Jun-sheng Xue¹, Yu-chen Guo¹, Wei-zhe Jian¹, Ren-wei Zhang¹, Xiao-yan Qiu² and Tian-yan Zhou¹

Survival is one of the most important endpoints in cancer therapy, and parametric survival analysis could comprehensively reveal the overall result of disease progression, drug efficacy, toxicity as well as their interactions. In this study we investigated the efficacy and toxicity of dexamethasone (DEX) combined with gemcitabine (GEM) in pancreatic cancer xenograft. Nude mice bearing SW1990 pancreatic cancer cells derived tumor were treated with DEX (4 mg/kg, i.g.) and GEM (15 mg/kg, i.v.) alone or in combination repeatedly (QD, Q3D, Q7D) until the death of animal or the end of study. Tumor volumes and net body weight (NBW) were assessed every other day. Taking NBW as a systemic safety indicator, an integrated pharmacokinetic/pharmacodynamic (PK/PD) model was developed to quantitatively describe the impact of tumor size and systemic safety on animal survival. The PK/PD models with time course data for tumor size and NBW were established, respectively, in a sequential manner; a parametric time-to-event (TTE) model was also developed based on the longitudinal PK/PD models to describe the survival results of the SW1990 tumor-bearing mice. These models were evaluated and externally validated. Only the mice with good tumor growth inhibition and relatively stable NBW had an improved survival result after DEX and GEM combination therapy, and the simulations based on the parametric TTE model showed that NBW played more important role in animals' survival compared with tumor size. The established model in this study demonstrates that tumor size was not always the most important reason for cancer-related death, and parametric survival analysis together with safety issues was also important in the evaluation of oncology therapies in preclinical studies.

Keywords: gemcitabine; pharmacokinetic/pharmacodynamic model; parametric survival model; tumor size; net body weight; pancreatic cancer

Acta Pharmacologica Sinica (2023) 44:465–474; <https://doi.org/10.1038/s41401-022-00960-0>

INTRODUCTION

Cancer is considered a systemic disease with not only tumor lesions but also metabolic disorders, inflammation and other morbidities, which may lead to unexpected loss of body weight, aggravated toxicity and complications of anti-cancer therapies [1]. However, a lot of preclinical cancer studies absolutely choose tumor growth inhibition (TGI) derived from tumor size as a primary indicator to evaluate the in-vivo anti-cancer efficacy in animal xenografts, which might neglect other potential information related to safety issues and lead to a bias evaluation of anti-cancer therapies [2, 3]. Survival has been one of the most important endpoints in clinical cancer therapy, which may comprehensively reflect the overall result of disease progression, drug efficacy, toxicity, and their interactions [4]. Survival analysis in animal xenograft model that takes both drug effect and safety issues into consideration makes better use of experimental data and could provide more insights for specific therapy [5].

In addition to the commonly used techniques in survival analysis including nonparametric and semiparametric methods,

such as Kaplan-Meier plot, log-rank test and Cox proportional hazard model [6, 7], parametric time-to-event (TTE) model is raising more and more interest, for its potential in associating time-course of important biomarkers with survival-related endpoints as well as its better ability in simulation [8, 9]. Combination of longitudinal tumor size model and TTE model is increasingly applied to facilitate decision making in oncology clinical trials and predicting the survival of individuals [10]. Nevertheless, except for tumor size, time-courses of other factors that may also relate to the survival are less considered. For example, nearly 80% of patients with advanced cancer experience progressive loss of body weight due to cachexia [11]. In preclinical studies, body weight of animals is also generally taken for a biomarker for systemic safety, individuals with significant change in body weight after being treated are considered to be not tolerated and not benefit from therapy [12]. However, there is little research investigating on the longitudinal body weight and its quantitative relationship with anti-tumor efficacy and survival.

¹Beijing Key Laboratory of Molecular Pharmaceutics and New Drug Delivery System, Department of Pharmaceutics, School of Pharmaceutical Sciences, Peking University, Beijing 100191, China; ²Department of Immunology, School of Basic Medical Sciences, Peking University, Beijing 100191, China and ³Department of Gastrointestinal Oncology, Key Laboratory of Carcinogenesis and Translational Research (Ministry of Education/Beijing), Peking University Cancer Hospital and Institute, Beijing 100142, China
Correspondence: Xiao-yan Qiu (qiuxy@bjmu.edu.cn) or Tian-yan Zhou (tianyanzhou@bjmu.edu.cn)

These authors contributed equally: Qing-yu Yao, Jun Zhou

Received: 21 March 2022 Accepted: 13 July 2022

Published online: 11 August 2022

Pancreatic cancer is a malignant tumor with 80%–90% risk of weight loss due to cachexia and an extremely high mortality [13]. In our previous study, dexamethasone (DEX) was found to be efficacious in inhibiting the growth and migration of pancreatic cancer in vitro and in vivo, where the blockade of NF-κB phosphorylation, reversion of EMT and down-regulation of IL-6 and VEGF were involved [14]. However, long-term and high dose of DEX may lead to muscle atrophy and aggravate the weight loss related with cancer cachexia [15–17].

In the current study, the efficacy and toxicity of DEX combined with gemcitabine (GEM), one of the first-line treatments for pancreatic cancer, was investigated in pancreatic cancer xenograft. Taking body weight as a systemic safety indicator, this study aimed to develop an integrated pharmacokinetic/pharmacodynamic (PK/PD) model to quantitatively demonstrating the impact of tumor size and systemic safety on animal survival.

MATERIALS AND METHODS

Chemicals and reagents

DEX and GEM were purchased from Meilunbio (Dalian, China). The DMEM medium was supplied by Maccgene Biotech Co., Ltd (Beijing, China), fetal bovine serum (FBS) was provided by PAN-Biotech (Bavaria, Germany) and penicillin/streptomycin was provided by Gibco (CA, USA).

Cell culture

Human pancreatic cancer cell line SW1990 was supplied by American Type Culture Collection. Cells were cultured in DMEM medium with 10% FBS and 1% penicillin/streptomycin at 37 °C in a humidified 5% CO₂ atmosphere.

Animals

The animal experiments were approved by the Institutional Animal Care and Use Committee of Peking University Health Science Center. Female BALB/c nude mice (5 weeks old) were provided by Department of Laboratory Animal Science, Peking University Health Science Center. The mice were maintained in individual ventilated cages with standard pathogen-free condition at 22–24 °C, 50%–60% humidity, and a 12 h/12 h light/dark cycles. Every effort was made to minimize the stress and the number of the animals used in the experiments.

PD study

In vivo study was carried out to investigate on the efficacy, toxicity as well as the impact on survival of DEX combined with GEM. SW1990 cells (3×10^6) were injected subcutaneously into BALB/c nude mice to establish the SW1990 xenograft model. The mice were randomly assigned into 8 groups and the treatment started when the tumor volumes reached 50–100 mm³. The dosage for DEX and GEM was selected according to our previous researches [14, 18, 19]. Considering the side-effect caused by long-term and high dose of DEX [16], DEX with longer dosing interval and lower total dose were also investigated in this study. Group A was set as vehicle control, Group B–D were treated with DEX 4 mg/kg by gavage every day/3 days/7 days respectively. Group E was given GEM 15 mg/kg every 3 days by tail vein injection, while Group F–H were given GEM 15 mg/kg every 3 days combined with DEX 4 mg/kg every day/3 days/7 days, respectively. DEX was dissolved in 10% hydroxypropyl-β-cyclodextrin (HP-β-CD) and GEM was in saline.

The two agents were given repeatedly until the death of animal or the end of study. In order to exclude the potential influence of solvent and administration on body weight or tumor size, 10% HP-β-CD was administered orally every day and saline was injected every 3 days for Group A. The length and width of the tumors were measured with caliper, and the body weight of the animals were recorded every other day. The tumor volumes were calculated as: tumor volume

(mm³) = length × width² × 0.5 [20]. Net body weight (NBW) was derived by subtracting tumor weight from animal's body weight assuming that tumor has a density of 1 g/cm³, in order to exclude the numeric influence from the rapid-progressing tumor on the weight of other non-tumor parts of the animal. Finally, the survival of every mouse was recorded in a 60 days period, individuals that survived more than 60 days were considered as right-censored, which were euthanized at the end of the study.

PK/PD model

The PK/PD models describing the time-course of both tumor size and NBW were developed in a sequential manner, respectively. Figure 1 showed the global scheme of the integrated model. Firstly, the PK model of DEX was described by a two-compartment model with first-order absorption rate, which was developed in our previous study [21]. Differential equations for the PK model are shown as follows:

$$\frac{dX_{a,DEX}}{dt} = -k_{a,DEX} \cdot X_{a,DEX}, X_{a,DEX}(0) = X_{0,DEX} \quad (1)$$

$$\frac{dX_{C,DEX}}{dt} = k_{a,DEX} \cdot X_{a,DEX} \cdot F_{DEX} - Q_{DEX} \cdot C_{C,DEX} + Q_{DEX} \cdot C_{P,DEX} - CL_{DEX} \cdot C_{C,DEX}, X_{C,DEX}(0) = 0 \quad (2)$$

$$\frac{dX_{P,DEX}}{dt} = Q_{DEX} \cdot C_{C,DEX} - Q_{DEX} \cdot C_{P,DEX}, X_{P,DEX}(0) = 0 \quad (3)$$

$$C_{C,DEX} = \frac{X_{C,DEX}}{V_{C,DEX}}, C_{C,DEX}(0) = 0 \quad (4)$$

$$C_{P,DEX} = \frac{X_{P,DEX}}{V_{P,DEX}}, C_{P,DEX}(0) = 0 \quad (5)$$

where $X_{a,DEX}$, $X_{C,DEX}$, $X_{P,DEX}$ represent the amount of DEX in absorption, central and peripheral compartment, respectively. $V_{C,DEX}$ and $V_{P,DEX}$ stand for apparent volume of distribution in central and peripheral compartment, while $C_{C,DEX}$ and $C_{P,DEX}$ are the concentrations of DEX in the relative compartment. $k_{a,DEX}$ is the first order absorption rate constant, CL_{DEX} is the clearance of DEX and Q_{DEX} is the intercompartmental clearance. F_{DEX} represents the bioavailability of DEX.

In addition, the two-compartment model of GEM's PK was also obtained from publication [19]:

$$\frac{dX_{C,GEM}}{dt} = Q_{GEM} \cdot C_{P,GEM} - Q_{GEM} \cdot C_{C,GEM} - CL_{GEM} \cdot C_{C,GEM}, \quad (6)$$

$$X_{C,GEM}(0) = X_{0,GEM}$$

$$\frac{dX_{P,GEM}}{dt} = Q_{GEM} \cdot C_{C,GEM} - Q_{GEM} \cdot C_{P,GEM}, X_{P,GEM}(0) = 0 \quad (7)$$

$$C_{C,GEM} = \frac{X_{C,GEM}}{V_{C,GEM}}, C_{C,GEM}(0) = 0 \quad (8)$$

$$C_{P,GEM} = \frac{X_{P,GEM}}{V_{P,GEM}}, C_{P,GEM}(0) = 0 \quad (9)$$

where $X_{C,GEM}$, $X_{P,GEM}$ represent the amount of GEM in central and peripheral compartment, $V_{C,GEM}$ and $V_{P,GEM}$ stand for apparent volume of distribution in the relative compartment. CL_{GEM} is the clearance of GEM and Q_{GEM} is the intercompartmental clearance. $C_{C,GEM}$ and $C_{P,GEM}$ are the concentrations of GEM in central and peripheral compartment, respectively.

The tumor natural growth model was characterized by Koch et al. [22], where the tumor growth was described by an exponential rate λ_0 followed by a linear growth rate λ_1 with no

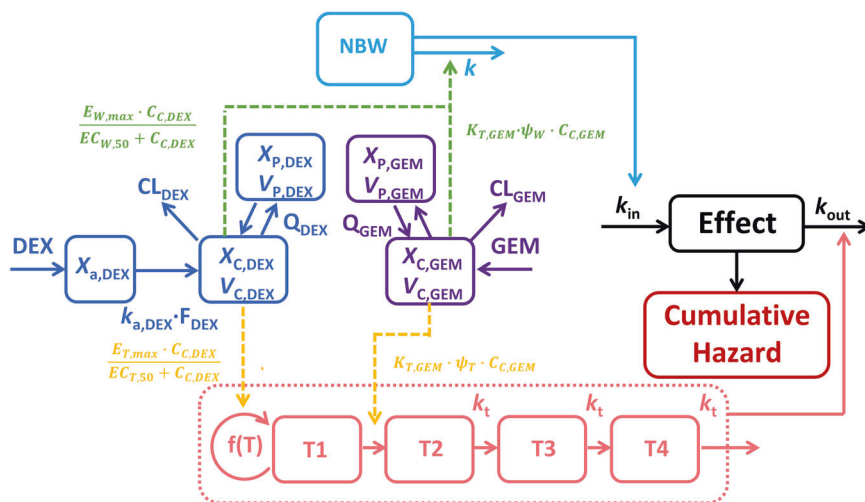


Fig. 1 The schematic structure of the integrated PK/PD model for the combination use of DEX and GEM. The schematic structure of the integrated PK/PD model for the combination use of DEX and GEM. $X_{a,DEX}$, $X_{c,DEX}$, $X_{p,DEX}$: amount of DEX in absorption, central and peripheral compartment; $V_{c,DEX}$, $V_{p,DEX}$: apparent volume of distribution in central and peripheral compartment; $C_{c,DEX}$, $C_{p,DEX}$: concentrations of DEX in central and peripheral compartment; $k_{a,DEX}$: first order absorption rate constant of DEX; CL_{DEX} : clearance of DEX; Q_{DEX} : intercompartmental clearance of DEX; F_{DEX} : bioavailability of DEX; $X_{c,GEM}$, $X_{p,GEM}$: amount of GEM in central and peripheral compartment; $V_{c,GEM}$, $V_{p,GEM}$: apparent volume of distribution in central and peripheral compartment; CL_{GEM} : clearance of GEM; Q_{GEM} : intercompartmental clearance of GEM; $C_{c,GEM}$, $C_{p,GEM}$: concentrations of GEM in central and peripheral compartment; CL_{GEM} : clearance of GEM; Q_{GEM} : intercompartmental clearance of GEM; $C_{c,GEM}$, $C_{p,GEM}$: concentrations of GEM in central and peripheral compartment; $E_{T,max}$: maximum effect of DEX in tumor growth inhibiting; $EC_{T,50}$: DEX's concentration to achieve half of the maximum effect; $K_{T,GEM}$: linear potency factor of GEM on damaging tumor cells; k_t : transition rate between the tumor compartments; ψ_T : the combination index of DEX and GEM in tumor inhibition; k : first order rate of NBW decreasing; $E_{W,max}$: maximum effect of DEX on NBW; $EC_{W,50}$: concentration of DEX to achieve half of the maximum effect on NBW; $K_{W,GEM}$: linear potency factor of GEM on NBW; ψ_W : combination index of DEX and GEM in NBW; k_{in} : turnover rate of the Effect compartment ($k_{in} = k_{out}$).

threshold point between two phases. The equation is shown as follows:

$$\frac{dT}{dt} = \frac{2 \cdot \lambda_0 \cdot \lambda_1 \cdot T}{\lambda_1 + 2 \cdot \lambda_0 \cdot T}, T(0) = T_0 \tag{10}$$

T indicates the tumor volume and T_0 is the initial tumor volume at the beginning of treatment.

The PK/PD models describing the anti-tumor effect of DEX and GEM were developed separately and the typical values of the drug efficacy parameters were fixed when the model for their combination was established. DEX showed its effect by inhibiting the proliferation of tumor cells, which was similar with our previous study [23], and a Hill's function was employed to describe it. GEM was assumed to convert the proliferating cells into non-proliferating cells, and the tumor cells were gradually brought to death through a series of transit compartments, describing the time delay between GEM exposure and tumor shrinkage [19]. Equations showing effect of DEX and GEM on the tumor size are shown as follows:

$$\frac{dT1}{dt} = \frac{2\lambda_0\lambda_1 \cdot T1^2}{(\lambda_1 + 2\lambda_0T1) \cdot T} \cdot \left(1 - \frac{E_{T,max} \cdot C_{C,DEX}}{EC_{T,50} + C_{C,DEX}}\right) - K_{T,GEM} \cdot \psi_T \cdot C_{C,GEM} \cdot T1, T1(t = 0) = T_0 \tag{11}$$

$$\frac{dT2}{dt} = K_{T,GEM} \cdot C_{C,GEM} \cdot T1 - k_t \cdot T2, T2(t = 0) = 0 \tag{12}$$

$$\frac{dT3}{dt} = k_t \cdot (T2 - T3), T3(t = 0) = 0 \tag{13}$$

$$\frac{dT4}{dt} = k_t \cdot (T3 - T4), T4(t = 0) = 0 \tag{14}$$

$$T = T1 + T2 + T3 + T4 \tag{15}$$

where $T1$ stands for the proliferating tumor cells and $T2, T3, T4$ stand for the non-proliferating cells, and T is the whole tumor including all these cells. $E_{T,max}$ is the maximum effect of DEX in tumor growth inhibiting with a theoretical upper limitation of 1, and $EC_{T,50}$ is DEX's concentration to achieve half of the maximum effect. $K_{T,GEM}$ is the linear potency factor of GEM on damaging tumor cells, and k_t denotes the transition rate between the tumor compartments. ψ_T is the combination index of DEX and GEM in tumor inhibition, it is assumed that the two drugs showed synergistic effect if ψ_T was greater than 1 while they antagonize with each other if ψ_T was less than 1, and no interaction exists in PD level if ψ_T equal to 1.

The NBW of tumor-bearing mice was progressively decreasing since the tumor's malignant proliferation required increasing nutrients from the host [24]. The unperturbed NBW decreasing was described with a first order rate as follows:

$$\frac{dNBW}{dt} = -k \cdot NBW, NBW(t = 0) = NBW_0 \tag{16}$$

where k is the first order rate of NBW decreasing, and NBW_0 is the initial NBW at the start of the treatment.

The models of DEX and GEM's effect on the time-course of NBW were developed separately, too. E_{max} function was used to describe DEX's effect on NBW, and GEM influenced the NBW in a linear way. The typical values of the drug-specific parameters were fixed in their combination model. The equation is shown as follows:

$$\frac{dNBW}{dt} = k \cdot \left(1 + \frac{E_{W,max} \cdot C_{C,DEX}}{EC_{W,50} + C_{C,DEX}} + K_{W,GEM} \cdot \psi_W \cdot C_{C,GEM}\right) \cdot NBW, NBW(t = 0) = NBW_0 \tag{17}$$

where $E_{W,max}$ is the maximum effect of DEX on NBW, while $EC_{W,50}$ is the concentration of DEX to achieve half of the maximum effect. $K_{W,GEM}$ is the linear potency factor of GEM on NBW, and ψ_W is the combination index of DEX and GEM in NBW.

Survival model

A parametric TTE model was developed to further explore the relationship between tumor size, NBW, and survival in the tumor-bearing mice. The hazard, cumulative hazard, and survival function are shown as follows:

$$h(t) = h_0 \quad (18)$$

$$H(t) = \int_0^t h(t) \quad (19)$$

$$S(t) = e^{-H(t)} \quad (20)$$

where h_0 is the baseline hazard, and $H(t)$ is the integration of $h(t)$. $S(t)$ stands for the probability of survival. In addition, probability density of observing death event was defined as the production of $h(t)$ and $S(t)$. Multiple probability density functions were tried for the baseline hazard including exponential, Weibull, log-logistic, and Gompertz distribution.

To investigate on the relationship between tumor size, NBW and survival, empirical Bayesian estimates were derived from the developed PK/PD models for tumor size and NBW, respectively, and longitudinal tumor size and NBW were considered as time-varying predictors in the TTE model. An indirect-response (IDR) model was used to fit the time-lag of drug effect better, which is assumed as a "Effect" compartment:

$$\frac{dEffect}{dt} = k_{in} - k_{out} \cdot Effect, Effect(t = 0) = 1 \quad (21)$$

where k_{in} and k_{out} were equal, for the homeostasis was maintained when the initial $Effect = 1$ at time = 0 and NBW and tumor size remained stable. This homeostasis was interrupted by the implantation of the xenograft tumor and the treatment of DEX and GEM. DEX and GEM were assumed to influence the tumor size and NBW simultaneously and further affect the *Effect* compartment to work on the cumulative hazard, which derived survival:

$$\frac{dEffect}{dt} = k_{in} \cdot \frac{NBW(t)}{NBW_0} - k_{out} \cdot (1 + POW \cdot \log_{10}(T(t)/T_0)) \cdot Effect, \quad t > 0, Effect(t = 0) = 1 \quad (22)$$

where $NBW(t)/NBW_0$ represent the relative change of NBW at time = t versus the initial NBW. $\log_{10}(T(t)/T_0)$ stands for the change of tumor size at time = 0, and POW is its potency factor on k_{out} . Since the tumor size increased and NBW decreased with time, the value of *Effect* would gradually decrease from the initial value 1, and therefore $h(t)$ increased until the death event occurred or censored.

Model development, evaluation, and external validation

The modeling and simulation in this study were conducted using NONMEM software (version 7.4.0, ICON Development Solutions, Ellicott City, MD, USA) and PsN (version 4.9.0, Uppsala University, Uppsala, Sweden). For the PK/PD models, first-order conditional estimation with interaction (FOCEI) method was used, and the inter-individual variability (IIV) was estimated in an exponential model. The residual variability was tested with additive, proportional, and mixture model. As for the parametric TTE model, Laplacian estimation method was used for parameter estimation. Since the variability of baseline hazard could not be estimated based on the survival data, all the individuals were assumed to have the same baseline hazard and therefore no IIV was estimated [5].

Models were evaluated and selected based on the rationality of the estimates, change in objective function value (OFV), and diagnostic plots. Relative standard error (RSE) was shown to evaluate the precision of the estimates and bootstrap based on 1000 runs was performed to access the parameter uncertainty of

the tumor size model as well as the NBW model. On the other hand, sampling importance resampling (SIR) was applied to evaluate the robustness and parameter uncertainty of the TTE model. Besides, visual predictive check (VPC) based on 1000 times of simulation was executed for the PK/PD models and 100 times for the TTE model.

In order to avoid misleading exploration due to inaccurate estimation for hazard distribution and further improve the reliability of the parametric TTE model, additional two groups of tumor-bearing mice treated with different dosing schedules were employed as external validation set for the final model. Group I was treated with Schedule A (DEX 4 mg/kg at day 1, GEM 15 mg/kg at day 4, 7 days/cycle), and Group J was treated with Schedule B (DEX 1.33 mg/kg at day 1–3, GEM 15 mg/kg at day 4, 7 days/cycle). Tumor volume, NBW, and survival were recorded as mentioned above.

Simulations

The survival of the tumor-bearing mice under different NBW changes and tumor sizes changes were simulated to illustrate their impact on the survival of animals.

RESULTS

DEX and GEM's effect on tumor growth, NBW, and survival in SW1990 xenograft

The effects of DEX, GEM as well as their combination on tumor growth, NBW and the survival of SW1990 tumor-bearing mice were shown in Fig. 2. The number of mice in Group A (control) is 6 while other groups were 5, except a mouse died by accident in Group D. DEX inhibited tumor growth in a dose-dependent manner, while GEM significantly inhibited tumor growth both alone and in combination with DEX (Fig. 2a). NBW decreased with time for all the animals, and the NBW of mice treated with GEM decreased rapidly in spite of a rather slow tumor progression, except for Group H treated with DEX 4 mg/kg every 7 days combined with GEM 15 mg/kg every 3 days that showed a relatively stable NBW time-course after an obvious weight loss at the beginning of the treatment, and only Group H had significantly improved animals' survival compared with control group (Fig. 2b, c), suggesting that the tumor sizes can not necessarily reflect survival results of tumor-bearing mice, and it would be more comprehensive to investigate on the survival as well as systemic safety biomarkers such as NBW of the animal xenografts.

PK/PD models for tumor size and NBW

The PK/PD models were developed sequentially and the parameter estimates, as well as the bootstrap 90% confidence intervals, are shown in Table 1. Some of the IIV in the final models were fixed to zero due to the limitation of data, resulting in an overestimation in the IIVs of the other parameters such as λ_0 and $K_{T,GEM}$, which were over 100%. For the tumor size model, the combination index ψ_T is 1.14 (90% CI is 0.78–1.42), indicating there was no obvious synergistic between DEX and GEM. On the other hand, the combination index in the NBW model ψ_W is 0.247, revealing the strong antagonism in the body weight loss between them. As the GOF plots shown (Supplementary Fig. S1), most of the population predictions and the individual predictions symmetrically distributed around the diagonal line, and the conditional weighted residuals (CWRES) were mostly within -4 and 4 .

Individual fitting plots (Supplementary Fig. S2) indicated that the individual predictions basically agreed with the observations. The VPC results based on 1000 simulations were also performed (Supplementary Fig. S3). The medians of observations and simulations generally matched well, while the 10% and 90% percentiles of the simulations were wider than that of the

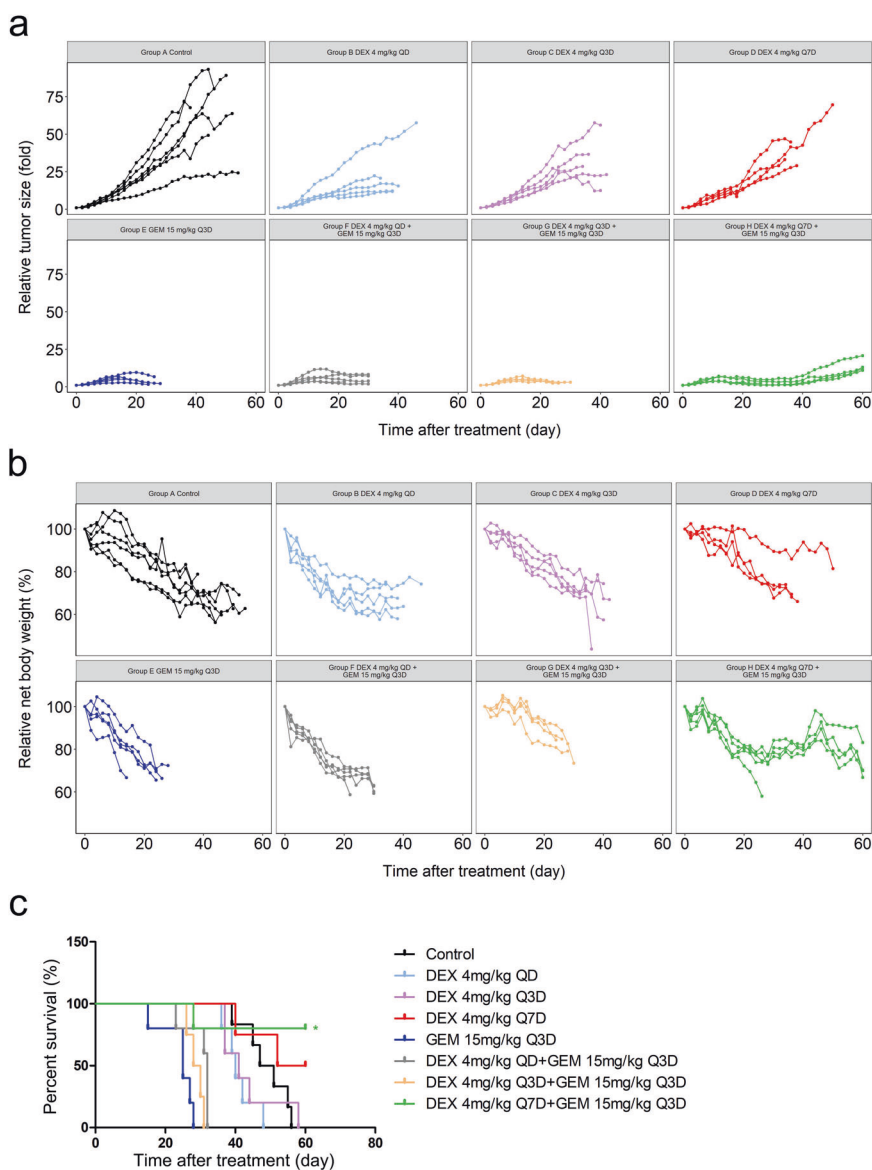


Fig. 2 The effect of DEX, GEM and their combination on SW1990 xenograft. **a** Tumor growth, **b** NBW, and **c** the survival of SW1990 tumor-bearing mice. The statistical results are shown as $*P < 0.05$.

observations, indicating the uncertainty of the model possibly due to the limited sample size. Overall, the PK/PD models were evaluated to be qualified to be integrated with the survival model. The main process for establishing the PK/PD models is summarized in Supplementary Table S1.

Parametric TTE model for survival

Empirical Bayesian estimates were derived from the aforementioned PK/PD models and the parametric TTE model for the survival of SW1990 tumor-bearing mice was further developed, the parameter estimates are shown in Table 2 and the main process for establishing the TTE model was summarized in Supplementary Table S1. The RSE of all the parameter estimates were under 30%, and the SIR results suggested that the parameters were estimated with reasonable uncertainty. The potency factor POW, representing the effect of tumor size on the assumed Effect compartment, was estimated to be rather small (9.4×10^{-7}), suggesting that tumor size contributed less to the final survival results compared with NBW changes. The VPC results in Fig. 3 showed that most of the observed events were

within the predicted 95% confidential interval, indicating that the model was with good ability to predict survival results in SW1990 tumor-bearing mice with the treatment of DEX and GEM, even though some misfits could still be found in Group D-H, which may be attributed to the limited sample size for model developing.

External validation

To further validate the predicting ability of the models, two groups of SW1990 xenografts with different dosing schedules were set as the external validations, where Group I contained 5 mice and Group J contained 4. For the PK/PD models, all the observed data were within the 95% prediction intervals (Fig. 4a, b). Post hoc Bayesian estimation was also performed for the external validation data set. The mean relative error (MRE) as well as root mean square error (RMSE) were calculated as 9.0% and 247.96 mm^3 for the tumor size model, and 4.9% and 0.70 g for the NBW model, respectively. For the TTE model, the death events also fell into the prediction intervals (Fig. 4c). Basically, the external validation data sets met the model predictions well.

Simulations

The survival of animals with different NBW reduction and tumor size progression was simulated based on the developed model. NBW reduction played an important role in animals' survival in the current model, where individuals with less NBW reduction might

have a much longer survival under the assumption that tumor size remains stable (Fig. 5). On the other hand, changes in tumor size made little difference in survival when the NBW were set to be stable, where the individuals with the same NBW reduction overlapped in spite of the different tumor ratio up to 50-fold change (Supplementary Fig. S4). These results suggested that there was a relatively weak connection between tumor size and survival in SW1990 xenografts, and NBW could possibly be a good predictor for the survival of tumor-bearing mice in preclinical studies of SW1990 xenograft model.

Table 1. Parameter estimates of the PK/PD models for tumor size and NBW.

Parameters	Estimates (RSE%)	Bootstrap results	
		Median	Bootstrap 90% CI
Tumor size model			
T_0 (mm ³)	110 (7.5)	110.5	97.3–125.5
λ_0 (1/day)	0.167 FIX		
λ_1 (mm ³ /day)	196 FIX		
$E_{T,max}$	0.599 FIX		
$EC_{T,50}$ (μg/mL)	0.999 FIX		
$K_{T,GEM}$ (mL/μg)	1.51 FIX		
ψ_T	1.14 (15.6)	1.14	0.78–1.42
k_t (1/day)	0.171 FIX		
IIV T_0 (%)	46.6 (13.6)	45.2	35.0–55.4
IIV λ_0 (%)	102 (11.9)	102.0	83.0–125.7
IIV λ_1 (%)	54.1 (13.3)	54.9	41.3–66.5
IIV $K_{T,GEM}$ (%)	112.7 (21.7)	110.1	77.4–169.8
Proportional error (%)	21.7 (14.9)	21.6	16.3–26.5
Net body weight (NBW) model			
NBW ₀ (g)	14.1 (1.6)	14.1	13.7–14.4
k (1/day)	0.00937 FIX		
$E_{W,max}$	0.369 FIX		
$EC_{W,50}$ (μg/mL)	0.673 FIX		
$K_{W,GEM}$ (mL/μg)	11.6 FIX		
ψ_W	0.247 (1.7)	0.314	0.062–0.720
IIV NBW ₀ (%)	9.8 (15.8)	9.4	7.1–11.8
IIV k (%)	41.4 (14.5)	39.1	27.5–48.6
IIV $K_{W,GEM}$ (%)	54.8 (76.2)	54.1	23.9–89.2
IIV ψ_W (%)	62.2 (64.5)	60.5	26.1–117.3
Additive error (g)	0.59 (0.1)	0.59	0.54–0.64

Definitions: T_0 : initial tumor volume; λ_0 : exponential tumor growth rate; λ_1 : linear tumor growth rate; $E_{T,max}$: maximum effect of DEX in tumor growth inhibiting; $EC_{T,50}$: DEX's concentration to achieve half of the maximum effect; $K_{T,GEM}$: linear potency factor of GEM on damaging tumor cells; k_t : transition rate between the tumor compartments; ψ_T : the combination index of DEX and GEM in tumor inhibition; NBW_0 : initial NBW; k : first order rate of NBW decreasing; $E_{W,max}$: maximum effect of DEX on NBW; $EC_{W,50}$: concentration of DEX to achieve half of the maximum effect on NBW; $K_{W,GEM}$: linear potency factor of GEM on NBW; ψ_W : combination index of DEX and GEM in NBW.

DISCUSSION

Survival is accepted as a gold standard in oncology clinical trials for it could show the most clinically meaningful improvements from therapies [25]. Tumor inhibition has also been an important therapeutic target in both clinical and preclinical oncology studies. At preclinical stage, mice xenografts bearing cells or patient-derived tumors were commonly developed for researches in anti-tumor efficacy and TGI based on tumor size is often calculated to evaluate the efficacy of specific cancer therapy in mice xenografts [26]. However, it could be biased to only consider tumor size in oncology studies. It is reported that nutritional and psychological supportive care could significantly improve the prognosis for cancer patients [27], indicating that tumor size is not the only reason for cancer-related death. Similarly, it could be observed in preclinical studies that therapies significantly inhibiting tumor growth may also possibly came up with major safety issues and even lethal toxicity [28]. In this study, we developed an integrated PK/PD model considering both tumor size and systemic safety, in order to quantitatively investigate their impact on animal survival in pancreatic cancer xenograft.

Progressive weight loss is an important symptom of cancer cachexia which is often accompanied with advanced carcinoma, it has been reported that over 10% losses in body weight could be a significant risk factor for the survival of pancreatic cancer patients [29]. Besides, body weight of animals in preclinical studies is also considered as an indicator of cachexia as well as a systemic safety biomarker for certain therapy [30]. With the tumor size increasing, it would require more and more nutrition supplies from the host animals and also release more and more TGF-β, lactic acid and myostatin, further leading to systemic inflammation, liver glycogen metabolism promotion and inhibiting skeletal muscle formation [24]. Although tumor weights are neglectable compared with human body weights, sometimes it could make a difference in animal xenografts where the increasing tumor weights may cover the body weight loss of the tumor-bearing mice, and therefore resulted in an underestimation of the systemic safety issues. In this study, NBW that derived from the body weight and tumor weight was selected as the systemic safety indicator to separate the tumor from the non-tumor part of the body weight, and the time course of NBW exhibited the weight loss caused by tumor progression together with anti-tumor therapies in a more direct manner.

Table 2. Parameter estimates of the survival model.

Parameters	Estimates (RSE%)	SIR results	
		Median (RSE%)	95% confidence interval
λ	3129 (15.3)	3149 (15.2)	2172–3977
k_{in}	9.4×10^{-7} (5.4)	9.4×10^{-7} (6.5)	8.2×10^{-7} – 1.0×10^{-6}
POW	9.99×10^{-8} (0.1)	9.99×10^{-8} (0.2)	9.96×10^{-8} – 1.01×10^{-7}

Definitions: λ : scale factor of the exponential hazard distribution; k_{in} : turnover rate of the Effect compartment ($k_{in} = k_{out}$); POW: potency factor on k_{out} .

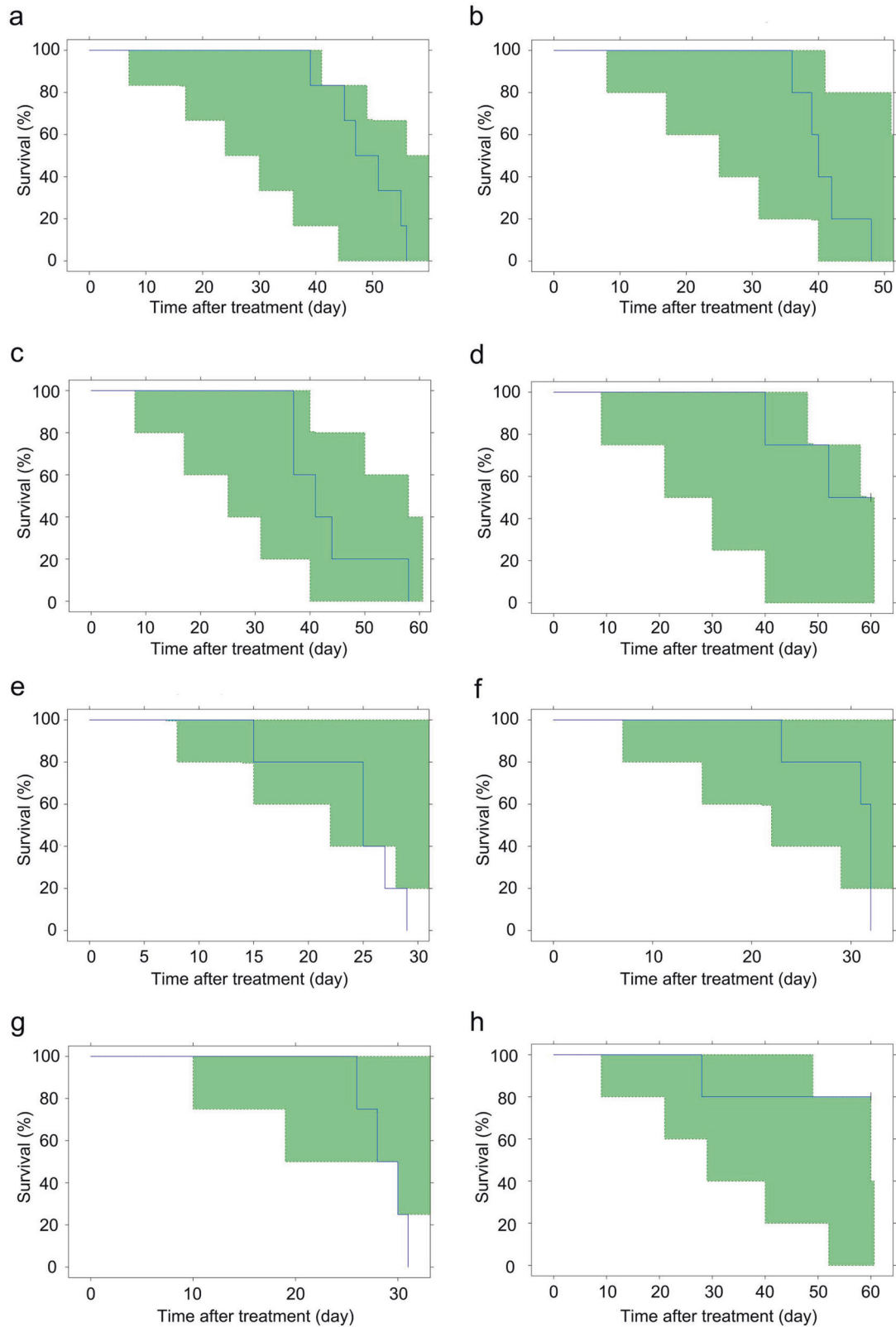


Fig. 3 Visual predictive check (VPC) for survival model for DEX combined with GEM in SW1990 xenograft. **a–h** Stand for related treatment groups. Group **a** was set as vehicle control; Group **b–d** were treated with DEX 4 mg/kg by gavage every day/3 days/7 days respectively. Group **e** was given GEM 15 mg/kg every 3 days by tail vein injection, while Group **f–h** were given GEM 15 mg/kg every 3 days combined with DEX 4 mg/kg every day/3 days/7 days, respectively. The blue solid line is the observations, the green shaded area stands for its 95% prediction interval.

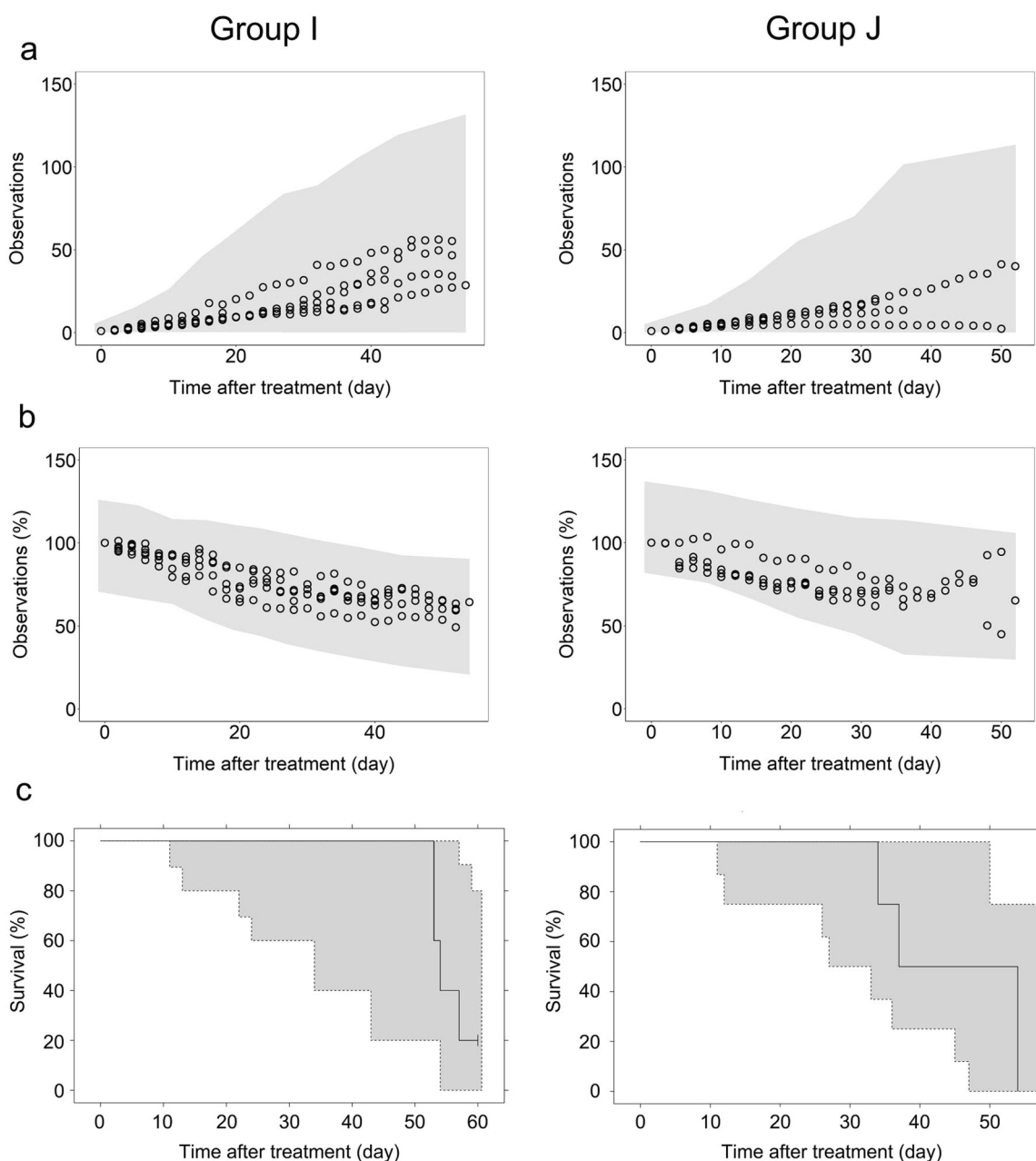


Fig. 4 External validation of the PK/PD models for the relative changes. Tumor size (a), NBW (b) and the TTE model (c) by Group I (left panel) and Group J (right panel). The shaded area represented the prediction interval, while the dots in **a** and **b** as well as lines in **c** represented the observations.

In the PD study, DEX showed a dose-dependent tumor growth inhibition in SW1990 xenograft, which was consistent with our previous results, and therefore a Hill's function was used to describing DEX's inhibition on the proliferation of tumor cells based on the GR-related mechanism [14]. Linear function was used for GEM's effect because there was only one dose level in the experimental data set. No PK interaction was assumed in the combination PK/PD model for the reason that DEX is mainly metabolized by CYP3A while GEM and its metabolites are excreted via renal, and it has been reported that no PK interaction was found between them in human [31]. In contrast to the synergistic effect in breast cancer xenograft [19], the combination index for tumor size model in SW1990 xenograft is slightly greater than 1, and the 90% confidence interval based on bootstrap indicated no significant synergistic effect was observed, which might be

attributed to the relative lower expression of GR in SW1990 cells, and further resulting in a worse response to DEX therapy either alone or in combination with GEM [14].

Body weight was usually taken as a result of blood glucose changes in previous PK/PD modelling studies. For example, Iwasaki et al. established a semi-mechanism PK/PD model where exenatide influenced the body weight of mice via impacting their food intake [32]. However, the time course of body weight in tumor-bearing mice was seldomly quantitatively investigated. In this study, an empirical model for the longitudinal NBW data was developed by using a first order rate constant k to describe the NBW reduction caused by the increasing tumor size, which is simplified and fit-for-purpose. GEM sharply increased the NBW reduction rate with a potency factor $K_{W,GEM}$ of 11.6 mL/ μ g, while DEX slightly decreased the NBW by a $E_{W,max}$ of 0.369. The

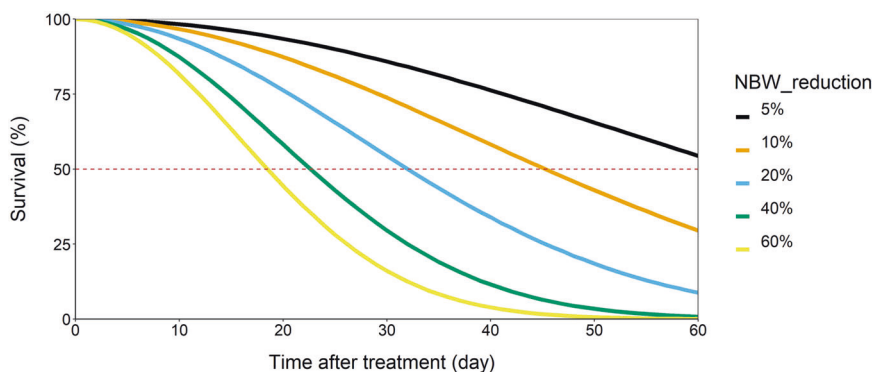


Fig. 5 Simulated survival of SW1990 tumor bearing mice in different net body weight reduction.

combination index ψ_W was 0.247, suggesting that the safety issue of weight loss could be improved when DEX and GEM were used in combination.

During the development of TTE mode, exponential, Weibull, log-logistic and Gompertz distributions for the baseline hazard were tested, and exponential distribution was selected in the final model for better fitness and lowest AIC (Supplementary Table S1). To integrate tumor size and NBW into the survival model, we firstly tried a proportional hazard style as follow:

$$h(t) = h_0 \times e^{\beta_1 \times X_1 + \beta_2 \times X_2 + \dots + \beta_n \times X_n} \quad (23)$$

where h_0 is the baseline hazard, X_i is risk factor such as tumor size, NBW and their derived metrics, and β_i is the associated potency factor. However, it did not provide a satisfying fitness. Considering the time lag between drug administration, tumor/NBW response and the cumulative hazard, the IDR model with an “Effect” compartment was utilized [5]. A difference with the model developed by Xie et al. was that the “Effect” compartment of IDR model was assumed to be a function to drug concentration in peritoneum and blood in that publication, which was considered to be related with the antitumor-efficacy and toxicity respectively [5]. In the current model, we assumed that both DEX and GEM showed their effect on tumor size and NBW, which were not only observed in the treatment groups but also in control group, so we tried this IDR model firstly in the control group and then in all the groups, rather than followed the sequence of “control group – single agent group – combination group” that we used in the development of tumor size model and NBW model.

Many different attempts were made to add tumor size into the survival model and the model was successfully minimized when the tumor size was added in a $\log_{10}(T(t)/T_0)$ manner with an extremely small potency factor POW near to zero and little improvement on fitness. The extremely small potency factor digitalized this neglectable influence of tumor size on the survival of tumor-bearing mice in this case, and therefore it was reserved in the final model to fit the purpose of the study better. Finally, the longitudinal relative changes of NBW and tumor size were integrated into the survival model as covariates. It should be emphasized that this structural model was limited to tumor-bearing mice, because it assumed that the hazard function was zero at hemostasis where the NBW and tumor size remained stable, which may not be extrapolated to other disease models. Besides, the hazard function $h(t)$ should be limited to non-negative so as to avoid Lazarus distribution [33]. Since all the NBW predictions after treatment were less than the initial NBW and the tumor sizes were also greater than the initial volumes, no additional assumption was made to restrict $h(t)$ to non-negative.

The simulation results showed that NBW played much more important role in the survival of the animals compared with tumor size. The reduction of NBW not only came from drug effect but also the increasing demand for nutrition of rapid-progressing

tumor with not any treatment [34], the relationship between NBW and the increasing tumor size was actually neglected when an empirical first order rate was used to capture the data. However, a more complicated model could be unstable due to the limitation of our data, as a result it was not taken into consideration in the current study. On the other hand, NBW could be more directly associated with the survival of tumor-bearing mice than tumor sizes, suggesting that tumor-growth inhibition is not the only thing should be cared when evaluating the therapeutic efficacy in preclinical studies.

One of the limitations of this study is that the results were obtained from immunodeficient mice bearing SW1990 cells derived tumors, while other covariates except NBW might be more important to the survival results in other animal models and tumors, such as the immune status of the host, which required further investigation in more different xenografts. In addition, the PK models for DEX as well as GEM were adopted from previous publications [19, 21], and the individual PK for the mice was not considered in the PK/PD model, which may lead to over-estimation for the variability of the parameters in the PK/PD models. Furthermore, it would be a more complicated situation when it comes to clinical cancer therapy, and even more risk factors that may associate with the clinical survival endpoints need to be taken into consideration. Last but not least, the small sample size for model development and external validation was a significant limitation in the current study, and there were still some misfits in the model possibly due to the limited data, which may bring some uncertainties and keep us modest with the conclusions.

Nevertheless, the current study has revealed the importance of body weight monitoring and survival analysis in preclinical oncology studies, which was often left in the basket in the past but actually worth considering in future preclinical studies, and the proposed model structure with DEX and GEM could provide some reference to other modeling and simulation studies in preclinical stages. For other anti-tumor drugs with different mechanisms, tumor size and NBW might not possibly be the only factors connecting drug efficacy, toxicity and survival outcomes in preclinical studies. In future studies, the applicability of the model would be tested in various situations including different anti-cancer drugs and different cancer xenograft models, and the workflow of parametric survival analysis in preclinical studies could be further explored and optimized.

CONCLUSION

In summary, a parametric TTE model of pancreatic cancer SW1990 xenografts’ survival integrated with longitudinal PK/PD models based on tumor sizes and NBW was developed and validated. NBW showed a more direct and important relation with the survival results of SW1990 tumor-bearing mice compared with tumor sizes, suggesting that it could be biased to evaluate the

anti-cancer therapeutic efficacy with tumor growth inhibition alone, and survival analysis together with systemic safety biomarkers such as body weight might deserve more attention in preclinical oncology studies.

ACKNOWLEDGEMENTS

The study was supported by Peking University Medicine Seed Fund for Interdisciplinary Research (Approval No. BMU2021MX003).

AUTHOR CONTRIBUTIONS

TYZ and QYY designed research; QYY, YCG, WZJ and RWZ performed research; QYY, JZ, YY and JSX analyzed the data; QYY, JZ, TYZ and XYQ wrote and revised the manuscript.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41401-022-00960-0>.

Competing interests: The authors declare no competing interests.

REFERENCES

1. Baracos VE, Martin L, Korc M, Guttridge DC, Fearon KCH. Cancer-associated cachexia. *Nat Rev Dis Prim.* 2018;4:17105.
2. Jung J, Seol HS, Chang S. The generation and application of patient-derived Xenograft model for cancer research. *Cancer Res Treat.* 2018;50:1–10.
3. Al-Huniti N, Feng Y, Yu JJ, Lu Z, Nagase M, Zhou D, et al. Tumor growth dynamic modeling in oncology drug development and regulatory approval: past, present, and future opportunities. *CPT Pharmacometrics Syst Pharmacol.* 2020;9:419–27.
4. Kamat AM, Sylvester RJ, Böhle A, Palou J, Lamm DL, Brausi M, et al. Definitions, end points, and clinical trial designs for non-muscle-invasive bladder cancer: recommendations from the International Bladder Cancer Group. *J Clin Oncol: Off J Am Soc Clin Oncol.* 2016;34:1935–44.
5. Xie F, De Clercq K, Vervaet C, Van Bocxlaer J, Colin P, Vermeulen A. Model-based analysis of treatment effects of paclitaxel microspheres in a microscopic peritoneal carcinomatosis model in mice. *Pharmacol Res.* 2019;36:127.
6. George B, Seals S, Aban I. Survival analysis and regression models. *J Nucl Cardiol.* 2014;21:686–94.
7. Schober P, Vetter TR. Survival analysis and interpretation of time-to-event data: the tortoise and the hare. *Anesth Analg.* 2018;127:792–8.
8. Brilleman SL, Crowther MJ, Moreno-Betancur M, Buros Novik J, Duniyakh J, Al-Huniti N, et al. Joint longitudinal and time-to-event models for multilevel hierarchical data. *Stat Methods Med Res.* 2019;28:3502–15.
9. Hansson EK, Amantea MA, Westwood P, Milligan PA, Houk BE, French J, et al. PKPD modeling of VEGF, sVEGFR-2, sVEGFR-3, and sKIT as predictors of tumor dynamics and overall survival following sunitinib treatment in GIST. *CPT Pharmacometrics Syst Pharmacol.* 2013;2:e84.
10. Krishnan SM, Friberg LE, Bruno R, Beyer U, Jin JY, Karlsson MO. Multistate model for pharmacometric analyses of overall survival in HER2-negative breast cancer patients treated with docetaxel. *CPT Pharmacometrics Syst Pharmacol.* 2021; 10:1255–66.
11. Vagnildhaug OM, Blum D, Wilcock A, Fayers P, Strasser F, Baracos VE, et al. The applicability of a weight loss grading system in cancer cachexia: a longitudinal analysis. *J Cachexia Sarcopenia Muscle.* 2017;8:789–97.
12. Tang J, Zhang J, Liu Y, Liao Q, Huang J, Geng Z, et al. Lung squamous cell carcinoma cells express non-canonically glycosylated IgG that activates integrin-FAK signaling. *Cancer Lett.* 2018;430:148–59.
13. Biswas AK, Acharyya S. Understanding cachexia in the context of metastatic progression. *Nat Rev Cancer.* 2020;20:274–84.

14. Yao Y, Yao QY, Xue JS, Tian XY, An QM, Cui LX, et al. Dexamethasone inhibits pancreatic tumor growth in preclinical models: Involvement of activating glucocorticoid receptor. *Toxicol Appl Pharmacol.* 2020;401:115118.
15. Braun TP, Grossberg AJ, Krasnow SM, Levasseur PR, Szumowski M, Zhu XX, et al. Cancer- and endotoxin-induced cachexia require intact glucocorticoid signaling in skeletal muscle. *FASEB J.* 2013;27:3572–82.
16. Schakman O, Gilson H, Thissen JP. Mechanisms of glucocorticoid-induced myopathy. *J Endocrinol.* 2008;197:1–10.
17. Siddiqui JA, Pothuraju R, Jain M, Batra SK, Nasser MW. Advances in cancer cachexia: Intersection between affected organs, mediators, and pharmacological interventions. *Biochim Biophys Acta Rev Cancer.* 2020;1873:188359.
18. Wang LJ, Li J, Hao FR, Yuan Y, Li JY, Lu W, et al. Dexamethasone suppresses the growth of human non-small cell lung cancer via inducing estrogen sulfotransferase and inactivating estrogen. *Acta Pharmacol Sin.* 2016;37:845–56.
19. Yuan Y, Zhou X, Ren Y, Zhou S, Wang L, Ji S, et al. Semi-mechanism-based pharmacokinetic/pharmacodynamic model for the combination use of dexamethasone and gemcitabine in breast cancer. *J Pharm Sci.* 2015;104:4399–408.
20. Tomayko MM, Reynolds CP. Determination of subcutaneous tumor size in athymic (nude) mice. *Cancer Chemother Pharmacol.* 1989;24:148–54.
21. Yuan Y, Zhou X, Li J, Ye S, Ji X, Li L, et al. Development and validation of a highly sensitive LC-MS/MS method for the determination of dexamethasone in nude mice plasma and its application to a pharmacokinetic study. *Biomed Chromatogr.* 2015;29:578–83.
22. Koch G, Walz A, Lahu G, Schropp J. Modeling of tumor growth and anticancer effects of combination therapy. *J Pharmacokinet Pharmacodyn.* 2009;36:179–97.
23. Yao Y, Yao Q, Fu Y, Tian X, An Q, Yang L, et al. Pharmacokinetic/Pharmacodynamic modeling of the anti-cancer effect of dexamethasone in pancreatic cancer xenografts and anticipation of human efficacious doses. *J Pharm Sci.* 2020;109:1169–77.
24. Schmidt SF, Rohm M, Herzog S, Berriel, Diaz M. Cancer Cachexia: more than skeletal muscle wasting. *Trends Cancer.* 2018;4:849–60.
25. Wilson MK, Karakasis K, Oza AM. Outcomes and endpoints in trials of cancer treatment: the past, present, and future. *Lancet Oncol.* 2015;16:e32–42.
26. Zhu AZ. Quantitative translational modeling to facilitate preclinical to clinical efficacy & toxicity translation in oncology. *Future Sci OA.* 2018;4:Fso306.
27. Lu Z, Fang Y, Liu C, Zhang X, Xin X, He Y, et al. Early Interdisciplinary supportive care in patients with previously untreated metastatic esophagogastric cancer: a Phase III randomized controlled trial. *J Clin Oncol.* 2021;39:748–56.
28. Christodoulou I, Ho WJ, Marple A, Ravich JW, Tam A, Rahnama R, et al. Engineering CAR-NK cells to secrete IL-15 sustains their anti-AML functionality but is associated with systemic toxicities. *J Immunother Cancer.* 2021;9:e003894.
29. Dalal S, Hui D, Bidaut L, Lem K, Del Fabbro E, Crane C, et al. Relationships among body mass index, longitudinal body composition alterations, and survival in patients with locally advanced pancreatic cancer receiving chemoradiation: a pilot study. *J Pain Symptom Manag.* 2012;44:181–91.
30. Talbert EE, Cuitiño MC, Ladner KJ, Rajasekera PV, Siebert M, Shakya R, et al. Modeling human cancer-induced Cachexia. *Cell Rep.* 2019;28:1612–1622.e1614.
31. Leggas M, Kuo KL, Robert F, Cloud G, deShazo M, Zhang R, et al. Intensive anti-inflammatory therapy with dexamethasone in patients with non-small cell lung cancer: effect on chemotherapy toxicity and efficacy. *Cancer Chemother Pharmacol.* 2009;63:731–43.
32. Iwasaki S, Hamada T, Chisaki I, Andou T, Sano N, Furuta A, et al. Mechanism-based pharmacokinetic/pharmacodynamic modeling of the glucagon-like peptide-1 receptor agonist exenatide to characterize its antiobesity effects in diet-induced obese mice. *J Pharmacol Exp Ther.* 2017;362:441–9.
33. Holford N. A time to event tutorial for pharmacometricians. *CPT Pharmacometrics Syst Pharmacol.* 2013;2:e43.
34. Bose S, Le A. Glucose metabolism in cancer. *Adv Exp Med Biol.* 2018;1063:3–12.

Springer Nature or its licensor holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.