ARTICLE

Open Access

Hippocampal glutamate and hippocampus subfield volumes in antipsychotic-naive first episode psychosis subjects and relationships to duration of untreated psychosis

Frédéric Briend¹, Eric A. Nelson², Omar Maximo¹, William P. Armstrong¹, Nina V. Kraguljac¹ and Adrienne C. Lahti¹

Abstract

Evidence points toward a relationship between longer duration of untreated psychosis (DUP) and worse long-term outcomes in patients with first episode psychosis (FEP), but the underlying neurobiology remains poorly understood. Proton magnetic resonance spectroscopy studies have reported altered hippocampus glutamatergic neurotransmission, and structural MRI as reported hippocampal atrophy that may be associated with memory impairment in schizophrenia. Here, we quantify left hippocampus glutamate (Glx) and left hippocampus subfield volumes in 54 antipsychotic-naive FEP and 41 healthy controls (HC), matched on age, sex, and parental occupation. While there were no significant group difference in Glx levels, hippocampal Glx levels were significantly higher in those who underwent a long DUP (>12 months) compared to those with a short DUP, and compared to HC. Compared to HC, FEP had significantly reduced whole hippocampus volume, as well as of CA1, CA4, granule cell layer, subiculum, and presubiculum subfields. Smaller whole hippocampal volume, as well as CA1, molecular layer, subiculum, presubiculum, and hippocampal tail volumes were significantly associated with longer DUP. However, we found no significant association between hippocampal Glx levels and hippocampal volume or subfields, suggesting that these alterations are not related, or their relationship does not follow a linear pattern. However, our results strongly suggest that one or several pathophysiological processes underlie the DUP. Importantly, our data highlight the critical need for reducing the DUP and for early pharmacological intervention with the hope to prevent structural deficits and, hopefully, improve clinical outcomes.

Introduction

Meta-analyses have consistently identified an association between longer duration of untreated psychosis (DUP), the duration between the onset of positive symptoms and treatment, and worse clinical outcome¹. Longer DUP is associated with worse negative symptoms at first treatment contact, and with poorer symptomatic

Correspondence: Adrienne C. Lahti (alahti@uabmc.edu)

¹Department of Psychiatry and Behavioral Neurobiology, University of Alabama at Birmingham, Birmingham, AL, USA

© The Author(s) 2020

and functional recovery from the first psychotic episode¹⁻⁴. The relationship between DUP and outcome is found across various lengths of follow-up periods, suggesting DUP influences the long-term course of the illness⁴. In addition, there is evidence that the association of DUP with outcome is more pronounced for longer DUP (>12 months) compared to shorter DUP (ref. ⁴). A deleterious effect of psychosis has been hypothesized; later it was proposed that hypofunction of *N*-methyl-D-aspartate receptors on gamma-aminobutyric acid (GABA) inhibitory neurons results in a hyperactivation of glutamate

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

²Department of Psychology, University of Alabama at Birmingham, Birmingham, AL, USA

neurons, leading to excess glutamate release, and, if the disruption is sustained, neuronal insult 5,6 .

The hippocampus plays a key role in memory processes, especially in long-term episodic memory, which has been shown to be impaired in schizophrenia (SZ)⁸. At the cytoarchitectonical level, hippocampal subdivisions include the subiculum complex, cornu ammonis (CA) 1-4, and the dentate gyrus $(DG)^9$. At the neuronal level, the pyramidal layers of the hippocampus are tightly packed with glutamatergic neurons that have a low firing threshold, ensuring a high level of neuroplasticity in the region¹⁰. The glutamatergic pyramidal neurons account for ~90% of hippocampal neurons, a much higher percentage than in other parts of the cortex¹¹. The remaining 10% of hippocampal neurons consist of inhibitory GABAergic interneurons tasked with regulation of the easily excited glutamatergic neurons^{10,12}. Postmortem studies in SZ have indicated normal hippocampal pyramidal neuronal density and number^{13,14}, but reduced number of GABAergic interneurons¹⁵. In addition, the GABA-synthesizing enzymes glutamic acid decarboxylase (GAD)65 and GAD67 are decreased in the hippocampus in SZ (ref. ¹⁶).

Converging lines of evidence from memory assessments⁸, molecular studies^{17,18} to multimodal brain imaging^{19–21} points to hippocampal alterations in SZ. In addition, hippocampal dysfunction is at the root of some prominent neurobiological models of SZ (refs. ^{10,22,23}). Hippocampal alterations have been identified in family members²⁴, and those at risk to develop psychosis²⁵. More generally, hippocampal dysfunction has also been described in psychotic disorders^{2,21,25–30}, suggesting this dysfunction plays a key role in etiology of the psychotic disorders.

Consistent with a deleterious effect of psychosis on brain structure, a recent study reported that, in FEP, longer DUP was associated with accelerated hippocampal atrophy over the initial 8 weeks of antipsychotic treatment, suggesting a persistent effect of DUP on brain structure³¹. In addition, we previously reported elevated hippocampal glutamate + glutamine (Glx) levels in a group of unmedicated patients with SZ (refs. 32-34), and an association between decreased hippocampal volume and increased Glx (ref. 32). Furthermore, clinical outcomes in individuals at clinical high risk for psychosis may be associated with an increase in baseline hippocampus Glx levels³⁵. Taken together, these studies suggest that hippocampal dysfunction plays a critical role in the onset of psychosis; however, the biologic basis for this dysfunction and its relationship to the DUP have not been identified³⁶.

In this study, we obtained measurements of hippocampal Glx and hippocampal volume segmentation in a group of antipsychotic-naive FEP and matched healthy controls (HC). We systematically assessed the DUP based on information provided by the patient and their caregivers during screening and, at any time during the 32week follow-up. Based on the existing literature, we hypothesized that we would observe higher Glx levels in FEP compared to controls. We further hypothesized that, in FEP, longer DUP would be associated with elevated Glx levels, and with smaller whole hippocampal and subfield volumes. Finally, we hypothesized that in FEP higher Glx levels would be associated with smaller hippocampal volumes, and mediate the association between DUP and smaller whole hippocampus and subfield volumes. Finally, we also conducted exploratory analyses to investigate associations with clinical and behavioral memory measures.

Material and methods

Participants

Sixty-six antipsychotic-naive FEP subjects were recruited from the emergency department, inpatient units, and outpatient psychiatry clinics at the University of Alabama at Birmingham (UAB). Seven patients dropped out for the following reasons: they withdrew consent before scan (n = 2), they did not tolerate the scan environment (n = 3) or for unknown reason (n = 2).

Diagnoses were established according to DSM-V criteria by review of medical records and consensus of two board-certified psychiatrists (A.C.L. and N.V.K.). Fortyone HC, matched on age, gender, and parental socioeconomic status were recruited by advertisements. Exclusion criteria were major neurological or medical conditions, history of significant head trauma, substance use disorders (excluding nicotine and cannabis) within 1 month of imaging, >5 days of lifetime antipsychotic exposure, pregnancy or breastfeeding, and MRI contraindications. HC with a personal or family history in a firstdegree relative of a psychiatric disorder were excluded. The UAB Institutional Review Board gave approval for this study, and written informed consent was obtained prior to enrollment and after subjects were deemed to have capacity to provide consent³⁷.

Duration of untreated psychosis

DUP was defined as the duration between the first onset of discernable positive symptoms to the time of initial treatment contact (the time at which the first antipsychotic prescription was written, which also coincided with enrollment in the study) and is reported in months. Two experienced psychiatrists systematically assessed the DUP based on information provided by the patient and their caregivers during screening, and at any time during the follow-up period. Moreover, we dichotomized DUP into short (<12 months) and long (>12 months) based on a number of studies that used this cutoff to define long and short DUP (refs. ^{4,38–41}), including those predicting treatment outcomes in early psychosis⁴² and poorer outcomes after the first hospitalization⁴³.

Clinical and behavioral scales

The brief psychiatric rating scale (BPRS) was used for assessments of symptom severity⁴⁴. The repeatable battery for the assessment of neuropsychological status (RBANS) was used to characterize memory cognitive function of our subject, via their total, immediate, and delayed memory subscales⁴⁵.

Image acquisition of the proton magnetic resonance spectroscopy

Data were collected from a voxel in the left hippocampus, such that the amount of gray matter was maximized while avoiding major vessels $(27 \times 15 \times 10 \text{ mm}^3)$; Fig. 1). Following automatic and manual shimming to optimize field homogeneity across the voxel, chemical shift selective pulses were used to suppress the water signal. Then, spectra were obtained using a point-resolved spectroscopy sequence (TR/TE = 2000/80 ms, flip angle = 90°, vector size 1024, 192 averages; the echo time was chosen in order to resolve and separate the C4 resonance of Glx from other *j*coupling metabolites^{46,47}). Moreover, eight averages of unsuppressed water scans with the same acquisition parameters were acquired for quantify metabolite according to the water peak.

Proton magnetic resonance spectroscopy data processing

All spectra were analyzed in jMRUI version 6.0 using the AMARES algorithm⁴⁸. Prior knowledge derived from in vitro and in vivo spectra was included in the model. A phantom solution of 20 mM glutamate in buffer (30 mM sodium hydrogen carbonate and 30 mM sodium carbonate; pH, 7.1) was imaged using the same acquisition parameters from the in vivo study. The model consisted of peaks for *N*-acetyl aspartate, choline, creatine (Cr + Cr2), and Glx was modeled as a triplet (large peak with two small outer wings), as previously described³². After removing the residual water peak using the Hankel-Lanczos singular values decomposition filter, the amplitude of the center Glx peak was estimated, and Glx levels were calculated relative to the unsuppressed voxel water and expressed in institutional units⁴⁹. Metabolite levels were corrected for partial volume effects according to Gasparovic and colleagues^{50,51}; the fraction of cerebrospinal fluid, gray, and white matter were calculated by segmentation of the T1-weighted images in SPM 8 (see in Supplementary Data).

Exclusion criteria for Glx were failure of the fitting algorithm, signal to noise ratio < 3, full-width at half-maximum > 0.1 ppm (ref. 52), and Cramer–Rao lower





bounds > 20%. Five FEP subjects were excluded from the analyses based on those criteria.

Hippocampus subregions segmentation

Each participant's high-resolution T1- and T2-weighted images were preprocessed using the FreeSurfer 6.0 (ref. ⁵³). All structural images underwent skull stripping via FSL (ref. ⁵⁴) before submission to the recon-all preprocessing pipeline⁵³. Following this step, we used Freesurfers' hippocampus subfield segmentation module to calculate each participants left and right subregions volumes⁵⁵. Freesurfer 6.0 has a significantly improved segmentation algorithm that uses Bayesian inference combined with a manually delineated hippocampal atlas⁵⁵. The estimated total intracranial volume (ICV) was also obtained via FreeSurfer 6.0 (ref. ⁵⁶; Table 1).

We assessed the overall hippocampal volume, and considered eight subregions across allocortical regions of the hippocampal formation²⁹: the hippocampal tail (comprised of portions of CA and DG), CA1, CA3 (CA2 and CA3 are combined in the Freesurfer atlas), CA4, molecular layer (ML) of the subiculum sub volumes and the DG, the granule cell layer of DG (GC-ML-DG: a combination of the granular cell layer, the ML, and the DG), and finally subiculum subfields (subiculum and presubiculum). These regions have been found to be altered in SZ (refs. ^{21,27,30}). Smaller area, as the fimbria, the hippocampal fissure and the parasubiculum were not included in this study due to reliability concerns in their segmentations^{21,57}.

Data quality was assessed using the Qoala-T-tool⁵⁸ that uses a supervised-learning model to assess accuracy of manual quality control of automated segmentation. Each participant's subfield segmentation was assessed by visual inspection (E.N. and O.M.), no hippocampal segmentation failures were reported.

Statistical analysis

Statistical analyses were performed in R (http://cran.rproject.org).

To compare Glx and hippocampus volumes between groups, we used a two-sample *t*-test (FEP vs HC) and a one-way ANCOVA to compared HC, long and short DUP, controlling for age, sex, and smoking status (packday). To assess the relationship between Glx, hippocampus volume measures, and clinical variables, we used partial correlations with age, sex, and smoking status included as covariates. For any analyses, including hippocampus volume, we included ICV as an additional covariate.

To complement analyses, we also calculated Cohen's d effect sizes. All analyses were controlled for multiple comparisons using the false detection rate (FDR) correction method by Benjamini and Hochberg⁵⁹.

Results

FEP and HC were well matched in terms of age, gender, and socioeconomic status. However, they differ in smoking status (Table 1, P < 0.01).

Glx levels in HC, FEP, and relationship with DUP

There was no significant group difference in Glx levels in the left hippocampus (Fig. 1c, Cohen's d = -0.17; t(94) = -0.86; P = 0.39).

Although Glx levels were not significantly correlated with DUP in FEP (r = 0.22, P = 0.11), when patients were dichotomized into those with long and short DUP based on a cutoff point of 12 months, the ANOVA revealed a significant group effect (F(2,89) = 4.92; P < 0.01). Those with long DUP (n = 17) had significantly higher Glx levels than those with shorter DUP (n = 37; Fig. 2, Cohen's d = -0.82; t(89) = 3.14; P < 0.01) and than HC (Cohen's d = -0.74; t(89) = 2.67; P = 0.03).

Hippocampus volume in HC, FEP, and relationship with DUP

Left whole hippocampus volume was significantly reduced in FEP compared to HC (Cohen's d = 0.59; t(85.7) = 2.87; P < 0.01). We also found lower volumes in the following hippocampus subfields: subiculum (Cohen's d = 0.53; t(85.7) = 2.53; P = 0.03), presubiculum (Cohen's d = 0.71; t(85.7) = 3.41; P < 0.01), CA1 (Cohen's d = 0.62; t(85.7) = 3.04; P = 0.01), GC-ML-DG (Cohen's d = 0.48; t(85.7) = 2.35; P = 0.04), and CA4 (Cohen's d = 0.45; t(85.7) = 2.21; P = 0.04; P < 0.05 FDR corrected; Table 2). Similar group differences were obtained for the right hippocampus (Supplementary Table 1). Although left and right whole hippocampus volumes, and all the subregions are reduced in long DUP compared to short DUP, these did not survive multiple comparisons (Table 2, Supplementary Table 1).

In FEP subjects, longer DUP was significantly associated with smaller whole hippocampus volume (Fig. 3, Table 3, r = -0.37, P < 0.01). This relationship was also seen in the following subfields: hippocampus tail (r = -0.32, P = 0.04), CA1 (r = -0.31, P = 0.04), ML (r = -0.30, P = 0.04), subiculum (r = -0.41, P = 0.02), and presubiculum (r = -0.33, P = 0.04) volumes.

Relationship between Glx and hippocampal volumes

In both FEP and HC, there were no significant relationship between Glx levels and hippocampal volumes (Table 3).

Clinical and behavioral relationships

Memory function: relationship with DUP, volume, and Glx

There were no significant relationship between the memory subscales of the RBANS and DUP, hippocampal volumes or Glx levels for the HC, the FEP, or the short and long DUP subgroups (Supplementary Tables 2 and 3).

Table 1 Demographic and clinica	l information of the sa	ample.				
	НС	FEP all	FEP subgroup $n = 54$		t-Test (HC vs FEP)	
			Long DUP	Short DUP		t-Test (long vs short)
	n = 41	n = 54	<i>n</i> = 17	n = 37		
Age (years)	24.75 (6.35)	23.88 (6.16)	24.58 (6.63)	23.56 (6.00)	P = 0.50; d = 0.14	P = 0.57; d = 0.16
Gender (% males)	60.97	64.81	58.82	67.56	Chi ² : 0.86	Chi ² : 0.07
Socioeconomic status (SES)	4.21 (4.20)	6.03 (5.05)	5.62 (5.28)	6.22 (5.01)	P = 0.09; d = -0.38	P = 0.70; d = -0.12
Smoking status (pack-day)	0.015 (0.07)	0.21 (0.34)	0.18 (0.34)	0.22 (0.34)	*P < 0.01; d = -0.74	P = 0.66; d = -0.13
Duration of untreated psychosis (weeks)		19.93 (36.60)	55.76 (49.18)	3.47 (3.58)		* $P < 0.01; d = 1.90$
Intracranial volume (mm^3)	10,000,907 (118056.2)	985,471.4 (126406.7)	1,002,273 (133221.7)	987,526.5 (117958.4)	P = 0.54; d = 0.12	P = 0.62; d = 0.12
Brief psychiatric rating scale (BPRS)						
Positive		11.50 (3.31)	10.41 (3.26)	12 (3.25)		P = 0.11; d = -0.48
Negative		5.92 (3.22)	5.11 (2.49)	6.29 (3.47)		P = 0.16; d = -0.36
Total		49.90 (11.03)	46.88 (10.24)	51.29 (11.23)		P = 0.16; d = -0.40
RBANS						
Total	94.06 (10.29)	75.69 (15.93)	81.07 (18.83)	73.24 (14.05)	* <i>P</i> < 0.01; <i>d</i> = 1.31	P = 0.16; d = 0.50
Immediate memory	102.87 (15.57)	83.14 (19.03)	84.53 (19.83)	82.51 (18.94)	*P < 0.01; d = 1.11	P = 0.74; d = 0.10
Delayed memory	92.58 (8.72)	77.41 (16.34)	83.40 (16.89)	74.70 (15.60)	*P < 0.01; d = 1.09	P = 0.10; d = 0.54
HC healthy controls, FEP first episode psychosi	s's patients, DUP duration of t	untreated psychosis (long >	12 months > short), SES pare	ntal socioeconomic ranks de	termined from diagnostic in	terview for genetic studies

HC healthy controls, *FEP* first episode psychosis's patients, *DUP* duration of untreated psychosis (long > 12 months > short), *SES* parenta (1–18 scale), *BPRS* brief psychological status, *d* Cohen's *d*.



Symptoms: relationship with DUP, volume, and Glx

Across the entire FEP sample, after multiple comparison correction, lower severity of positive symptoms were associated with longer DUP (r = -0.34, P = 0.03), but not severity of negative (r = -0.09, P = 0.52) or general symptoms (r = -0.22, P = 0.16). Across the DUP subgroups, no significant correlations were found (Supplementary Tables 3 and 4).

There were no significant associations between positive or negative symptoms, and volume (Supplementary Table 4) or Glx levels for the entire FEP sample or for DUP groups (Supplementary Table 3).

Discussion

To our knowledge, were are the first to simultaneously evaluate hippocampal glutamate levels and volumes, including subfields, in antipsychotic-naive FEP in order to shed light into the neurobiological underpinnings of the relationship between longer DUP and poorer clinical outcomes. In medication-naive FEP, we found that hippocampal Glx levels were significantly higher in those who underwent a long DUP (>12 months) compared to those with a short DUP, and compared to HC. In addition, we found that smaller hippocampal volumes, including CA1, ML, tail, subiculum, and presubiculum, were significantly associated with longer DUP. However, we found no significant association between hippocampal Glx levels, and hippocampal volume or subfields. Our results strongly suggest that one or several pathophysiological processes underlie the DUP.

Glutamate levels in left hippocampus

Contrary to our hypothesis, we found no significant differences in levels of Glx between medication-naive FEP and HC in left hippocampus. This is in agreement with another study in a small sample of medication-naive FEP (n = 15) where no group difference in Glx levels was identified in the medical temporal cortex⁶⁰. This is in contrast to our prior study, where elevated hippocampus Glx levels was found in a group of unmedicated patients with SZ (refs. $^{32-34}$). Illness chronicity or prior exposure antipsychotic medication may affect hippocampus Glx. In FEP, Glx levels showed greater variance compared to controls, suggesting heterogeneity in glutamatergic metabolism in FEP. It is possible that only a subset of patients have developed a hyperglutamatergic state^{61,62}. Our findings underscore the importance of considering heterogeneity when characterizing psychosis spectrum patients in the early illness stages.

Association between left hippocampus glutamate level and DUP

Because there is evidence that the association of DUP with outcome is more pronounced for longer DUP (>12 months) compared to shorter DUP (ref. ⁴), we dichotomized our FEP patients based on that cutoff value. We found increased hippocampus Glx levels in those with a DUP > 12 months, compared to those with a DUP < 12 months, and compared to HC. The two DUP subgroups were not significantly different based on demographic characteristics or symptom burden, indicating difference in Glx levels was not related to these variables (Table 1). Our results are in line with the report that higher hippocampus glutamate levels in clinical high-risk individuals for psychosis were associated with a poor functional outcome³⁵.

Left hippocampus volume and subregions in FEP compared to HC

Here, we report significantly lower whole left hippocampus volumes, as well as CA1, CA4, GC-ML-DG, subiculum, and presubiculum in antipsychotic-naive FEP patients compared to HC. While volume deficits of the hippocampus in SZ have been consistently reported [21, 26, 28, 29, 56, 57], only two prior studies measured hippocampal volumes in antipsychotic-naive FEP (refs. ^{31,63}). Consistent with our findings, one of them³¹ observed decreased hippocampal volumetric integrity in a cohort of 71 FEP, but they did not measure subfield volumes. The other⁶³, reported no significant change in total hippocampal volumes and significantly greater subfield volumes

H (n = 41) FE (n = 54) M6.37) P a FD6 0.05 Color et whele lippocampal at hppocampal at hppocampal at hppocampal at hppocampal at a 2314 (3457) 23714 (3455) 23714 (3453) 2371 2470 2401<		טווא טו ובור וווףףטכמוווףמו אמטוב				
Left whole hippocampairs 343.34 (36.5) 32/14 (34.5) 28/1 $(-0)^+$ <th< th=""><th></th><th>HC (<i>n</i> = 41)</th><th>FEP (<i>n</i> = 54)</th><th>t(85.7)</th><th><i>P</i> at FDR 0.05</th><th>Cohen's <i>d</i></th></th<>		HC (<i>n</i> = 41)	FEP (<i>n</i> = 54)	t(85.7)	<i>P</i> at FDR 0.05	Cohen's <i>d</i>
left hippocampal fall S81.5 (7.60) S33.6 (5.55) 1.89 0.06 0.1 left sbiculum 40.43 (5.17) 13.38 (4735) 2.33 0.07* 0.03* 0.03 left sbiculum 40.43 (5.17) 13.38 (4735) 2.881 (5.6.4.5) 2.881 (5.6.4.5) 0.07* 0.03* left corrun ammonis 1 31.46 (5.06) 2.881 (5.6.4.5) 2.881 (5.6.4.5) 0.07* 0.04 left molecula layer 52.00 (56180) 2.881 (56.4) 2.94 0.07* 0.07* 0.04 left molecula layer 52.00 (56180) 2.894 (5630) 2.94 0.06* 0.06* 0.44 left molecula layer 52.350 (5535) 2.994 (504) 2.34 0.06* 0.44 left molecula layer 2350 (5550) 2.944 (500) 2.34 0.06* 0.44 left corru ammonis 3 2.355 (560) 2.944 (500) 2.34 0.06* 0.44 left corru ammonis 3 2.355 (560) 2.944 (500) 2.31 0.06* 0.44 left corru ammonis 3 1.443 (560) 2.31	Left whole hippocampus	3483.54 (366.9)	3271.4 (346.5)	2.87	<0.01*	0.59
left abliculum 40.43 (6.1) 41.38 (4.73) 233 003* 033 left corrun ammonis 1 64.70 (79.45) 58.62 (8.28) 3.41 $0.01*$ 0.01 left corrun ammonis 1 64.70 (79.45) 58.62 (8.28) 3.41 $0.01*$ 0.01 left presubliculum 31.46 (59.06) 28.81 (56.13) 2.04 $0.01*$ 0.01 left molecular/jeer 57.01 (96.180) 28.81 (56.63) 2.04 $0.05*$ $0.02*$ left corrun ammonis 3 21.289 (29.70) $203-30$ (58.91) 2.397 (50.61) $0.05*$ $0.05*$ left corrun ammonis 3 21.289 (29.70) $299-36$ (50.91) 2.31 $0.05*$ $0.05*$ left corrun ammonis 4 573.520 (59.56) $299-4$ (50.04) 1.45 $0.05*$ $0.05*$ left corrun ammonis 1 61.70 0.71 $0.05*$ $0.05*$ $0.05*$ left presubliculum 582.64 (50.81) $0.317.44$ (20.60) 1.45 $0.05*$ $0.05*$ left molecular (10) 582.64 (50.81) 0.3	Left hippocampal tail	581.15 (76.80)	553.36 (63.65)	1.89	0.06	0.4
left conta anmonic 1 647.20 (79.45) 596.2 (82.81) 3.04 001* 021 left precidiculum 314.6 (80.06) 28811 (65.14) 3.41 0.00* 0.01 left precidiculum 314.6 (80.06) 28811 (65.14) 3.41 0.00* 0.01 left precidiculum 374.3 (56.76) 289.1 (65.14) 2.33 0.04 0.01 left conductular layer 3.07.43 (56.76) 289.1 (56.14) 2.33 0.04* 0.04 left conturations 3 2.12.89 (29.70) 2.930 (28.91) 1.91 0.04* 0.04 left conturations 3 2.33.53 (29.25) 2.939 (30.04) 2.21 0.04* 0.04 left conturations 3 2.33.53 (29.55) 2.939 (30.04) 1.91 0.04* 0.04 left conturations 4 1.17.1 (n=37) (n=37) (n=37) 0.04* 0.04* left conturations 1 0.71.3 (66.0) 1.43.4 (32.06) 1.43 0.04* 0.04* left conturations 1 0.71.3 (66.0) 1.43.4 (32.06) 1.43 0.04* 0.04* <td>Left subiculum</td> <td>440.43 (56.17)</td> <td>413.38 (47.35)</td> <td>2.53</td> <td>0.03*</td> <td>0.53</td>	Left subiculum	440.43 (56.17)	413.38 (47.35)	2.53	0.03*	0.53
Let peculiculum 3146 (59.06) 38.11 (56.14) 314 $< 001^{+}$ 01^{-1} Let molecular layer 32.01 (961.86) 46.92 (56.83) 2.04 0.06 0.42 Let GC-ML-DG 307.33 (56.76) 2.033 (25.93) 2.039 (261.61) 1.91 0.06 0.33 Let GC-ML-DG 307.33 (56.76) 2.033 (25.93) 2.039 (261.61) 2.13 0.06* 0.34 Let GC-ML-DG 373.50 (295.53) 2.394 (30.04) 2.01 0.06* 0.34 Let GC-ML-DG 373.50 (295.53) 2.394 (30.04) 2.21 0.06* 0.34 Let GC-ML-DG 317.41 (30.60) 1.45 0.06* 0.34 0.34 Let Whole hiptocampus 317.41 (30.60) 1.45 0.04* 0.05 0.34 Let Whole hiptocampus 317.41 (30.60) 1.45 0.145 0.04* 0.35 Let whole hiptocampus 317.41 (30.60) 1.45 0.45 0.45 0.45 Let conu ammonis 1 2.32.4 (30.61) 1.45 1.45 0.45 0.42 <	Left cornu ammonis 1	647.20 (79.45)	596.62 (82.81)	3.04	0.01*	0.62
Lett molecular layer 520.10 (661.66) 64.92 (56.83) 2.04 006 0.04 Lett Gc-Mi-DS 307.43 (56.76) 28.97 (66.16) 2.35 0.04* 0.06 0.04 Lett Gc-Mi-DS 307.43 (56.76) 2013 (28.91) 1.91 0.06* 0.64 Lett Gc-Mi-DS 307.43 (56.76) 2013 (28.91) 212.98 0.04* 0.64 0.64 Lett Grow ammonis 4 233.50 (29.55) 2399.4 (60.04) 2.21 0.04* 0.64 0.64 Lett Grow ammonis 4 233.50 (29.55) 2399.4 (60.04) 7.21 0.04* 0.64 0.64 Lett Grow ammonis 4 17.17 A 4.41 0.64 0.64 0.64 Lett Mole Hippocampus 317.12 (88.6) 3317.41 (32.60) 1.45 0.16 0.64 Lett Mole Hippocampus 317.12 (88.6) 317.41 (32.60) 1.45 0.16 0.65 Lett Mole Hippocampus 317.12 (88.6) 317.14 (32.60) 1.45 0.16 0.65 Lett Mole Hippocampus 355.64 (66.8) 317.41 (32.60) <td>Left presubiculum</td> <td>314.66 (39.06)</td> <td>288.11 (36.14)</td> <td>3.41</td> <td><0.01*</td> <td>0.71</td>	Left presubiculum	314.66 (39.06)	288.11 (36.14)	3.41	<0.01*	0.71
Lett G-Mi-DG $30'43(36.76)$ $28979(36.16)$ 233 0.04° 0.64 Lett Groun ammoris 3 $212.89(29.70)$ $20130(28.91)$ 191 0.06° 0.39 Lett Groun ammoris 3 $212.89(29.70)$ $20130(28.91)$ $20130(28.91)$ 0.06° 0.39 Lett Groun ammoris 4 $23330(29.55)$ $2039(400)$ 221 0.04° 0.64° FFB subgroup Integ DU Short DU 1734 12642 Pat FDR 0.05 0.04° Lett whole hippocampus $31714(2306)$ 145 0.145 0.04° 0.35 Lett whole hippocampus $31714(2306)$ 145 0.167 0.16 0.02° Lett whole hippocampus $31714(2306)$ $142(306)$ 145 0.166 0.03° Lett whole hippocampus $31714(2306)$ 0.145 0.36 0.04° 0.03° Lett whole hippocampus $31714(2306)$ $1423(206)$ 145 0.46 0.03° Lett whole hippocampus $3374(3206)$	Left molecular layer	520.10 (961.86)	464.92 (56.83)	2.04	0.06	0.42
	Left GC-ML-DG	307.43 (36.76)	289.79 (36.16)	2.35	0.04*	0.48
	Left cornu ammonis 3	212.89 (29.70)	201.30 (28.91)	1.91	0.06	0.39
FEP subgroup FEP subgroup Cohen Long DUP Short DUP (15.42) P at FDR 0.05 Cohen Long DUP Short DUP (1 = 37) (1 = 37) - 0.02 - 0.02 Left hippocampus 317.14 (38.6) 313.74 (32.06) 1.45 0.18 - 0.02 Left hippocampul tail 538.24 (% 0.3) 56.30 (55.70) - 1.05 0.46 - 0.35 Left subliculum 355.6 (46.8) 4.21.13 (46.04) - 1.87 0.36 - 0.35 Left subliculum 355.6 (46.8) 2.93.6 (35.20) - 1.07 0.46 - 0.35 Left molecular layer 275.98 (39.01) 606.58 (79.06) - 1.25 0.46 - 0.35 Left molecular layer 275.98 (33.83) 293.48 (35.24) - 1.17 0.46 - 0.35 Left molecular layer 275.98 (33.83) 293.48 (79.06) - 1.17 0.46 - 0.35 Left corru ammonis 1 275.98 (33.83) 293.48 (79.06) - 1.17 0.46 - 0.35 Left corru ammonis 3 197.35 (31.15) 201.42 (53.25)	Left cornu ammonis 4	253.50 (29.55)	239.94 (30.04)	2.21	0.04*	0.45
Imaguu (n=17) Imaguu (n=37) Imaguu (FEP subgroup				
Left whole hippocampus 317142 (388.6) 331741 (320.60) 1.45 0.18 -0.42 Left whole hippocampul tail 538.24 (780.3) 560.30 (57.70) -1.05 0.46 -0.35 Left subiculum 395.56 (46.68) 421.13 (46.04) -1.87 0.36 -0.36 Left corru ammonis 1 574.93 (89.01) 606.58 (79.06) -1.25 0.46 -0.35 Left corru ammonis 1 275.98 (33.33) 293.68 (36.24) -1.26 0.36 -0.36 Left presubiculum 275.98 (33.33) 501.42 (53.25) -1.74 0.36 -0.37 Left molecular layer 480.73 (63.31) 501.42 (53.25) -1.17 0.46 -0.37 Left docular layer 282.96 (43.38) 201.42 (53.25) -1.17 0.46 -0.27 Left corru ammonis 3 197.35 (31.15) 203.12 (28.08) -0.65 0.65 -0.65 Left corru ammonis 3 197.35 (31.15) 203.12 (28.08) -0.65 0.76 -0.22 Left corru ammonis 4 234.06 (35.81) 242.64 (77.11) -0.68 0.76 -0.22		Long DUP $(n=17)$	Short DUP $(n = 37)$	t(26.42)	P at FDR 0.05	Cohen's <i>d</i>
Left hippocampal tail538.24 (78.03)560.30 (55.70)-1.050.46-0.35Left subiculum395.56 (46.68)421.13 (46.04)-1.870.36-0.36Left subiculum574.93 (89.01)606.58 (79.06)-1.250.46-0.36Left cornu ammonis 1574.93 (89.01)606.58 (79.06)-1.250.46-0.37Left presubiculum275.98 (33.83)293.68 (36.24)-1.740.36-0.37Left molecular layer480.73 (6.3.1)501.42 (53.25)-1.170.46-0.37Left molecular layer282.95 (43.38)292.94 (32.50)-1.170.46-0.37Left cornu ammonis 3197.35 (31.15)203.12 (28.08)-0.650.46-0.27Left cornu ammonis 4234.06 (35.81)242.64 (27.11)-0.680.46-0.22	Left whole hippocampus	3171.42 (388.6)	3317.41 (320.60)	1.45	0.18	-0.42
Left subiculum395.56 (46.68)421.13 (46.04)-1.870.36-0.55Left corru ammonis 1574.93 (89.01)606.58 (79.06)-1.250.46-0.38Left roru ammonis 1275.98 (33.33)293.68 (36.24)-1.740.36-0.51Left molecular layer480.73 (63.31)501.42 (53.25)-1.170.46-0.37Left molecular layer480.73 (63.31)501.42 (53.25)-1.170.46-0.37Left conular layer282.95 (43.38)292.94 (32.50)-0.870.46-0.27Left corru ammonis 3197.35 (31.15)203.12 (28.08)-0.650.52-0.21Left corru ammonis 4234.06 (35.81)242.64 (27.11)-0.880.46-0.28	Left hippocampal tail	538.24 (78.03)	560.30 (55.70)	-1.05	0.46	-0.35
Left cornu ammonis 1574.93 (89.01)606.58 (79.06)-1.250.46-0.38Left presubiculum275.98 (33.33)293.68 (36.24)-1.740.36-0.51Left molecular layer480.73 (6.31)501.42 (53.25)-1.170.46-0.37Left molecular layer282.95 (43.38)292.94 (32.50)-1.170.46-0.27Left cornu ammonis 3197.35 (31.15)203.12 (28.08)-0.650.46-0.21Left cornu ammonis 4234.06 (35.81)242.64 (27.11)-0.880.46-0.28	Left subiculum	395.56 (46.68)	421.13 (46.04)	-1.87	0.36	-0.55
Left presubiculum 275.98 (3.3.83) 293.68 (36.24) -1.74 0.36 -0.51 Left molecular layer 480.73 (6.3.1) 501.42 (53.25) -1.17 0.46 -0.37 Left molecular layer 282.95 (43.38) 501.42 (53.25) -1.17 0.46 -0.37 Left GC-ML-DG 282.95 (43.38) 292.94 (32.50) -0.87 0.46 -0.27 Left corru ammonis 3 197.35 (31.15) 203.12 (28.08) -0.65 0.52 -0.21 Left corru ammonis 4 234.06 (35.81) 242.64 (27.11) -0.88 0.46 -0.28	Left cornu ammonis 1	574.93 (89.01)	606.58 (79.06)	-1.25	0.46	-0.38
Left molecular layer 480.73 (5.3.1) 501.42 (53.25) -1.17 0.46 -0.37 Left GC-ML-DG 282.95 (43.38) 292.94 (32.50) -0.87 0.46 -0.27 Left GC-ML-DG 197.35 (31.15) 292.91 (32.50) -0.65 0.46 -0.21 Left cornu ammonis 3 197.35 (31.15) 203.12 (28.08) -0.65 0.52 -0.21 Left cornu ammonis 4 234.06 (35.81) 242.64 (27.11) -0.88 0.46 -0.28	Left presubiculum	275.98 (33.83)	293.68 (36.24)	-1.74	0.36	-0.51
Left GC-ML-DG 282.95 (43.38) 292.94 (32.50) -0.87 0.46 -0.27 Left cornu ammonis 3 197.35 (31.15) 203.12 (28.08) -0.65 0.52 -0.21 Left cornu ammonis 4 234.06 (35.81) 242.64 (27.11) -0.88 0.46 -0.28	Left molecular layer	480.73 (63.31)	501.42 (53.25)	-1.17	0.46	-0.37
Left corru ammonis 3 197.35 (31.15) 203.12 (28.08) -0.65 0.52 -0.21 Left corru ammonis 4 234.06 (35.81) 242.64 (27.11) -0.88 0.46 -0.28	Left GC-ML-DG	282.95 (43.38)	292.94 (32.50)	-0.87	0.46	-0.27
Left cornu ammonis 4 234.06 (35.81) 242.64 (27.11) -0.88 0.46 -0.28	Left cornu ammonis 3	197.35 (31.15)	203.12 (28.08)	-0.65	0.52	-0.21
	Left cornu ammonis 4	234.06 (35.81)	242.64 (27.11)	-0.88	0.46	-0.28

(mm³) in FEP and HC. subfield volumes ¢ C of left hinn .i. Į ē Statistical

the dentate gyrus, ML molecular layer, FEP: n = 54.



in the ML, granular layer, and CA4 in a cohort of 41 FEP. Interestingly, Ho and colleagues³⁰ measured hippocampal volumes and subfields in a large group of patients with SZ in the early stages of illness (mean duration of illness of 7 years), and found deficits in hippocampal whole volume and CA1, but not in other subfields. In addition, they measured hippocampal subfields in an older cohort (mean duration of illness of 18 years) of patients and report deficits in all subfields. It is not clear why this study detected deficits limited to the CA1 region in the early stage psychosis cohort, while we observed more extensive deficits in medication-naive FEP. This cohort was collected in Singapore, while ours was collected from the southeast of the US. It is possible that differences in DUP, stress, or other environmental factors could explain those differences. Another possibility is their hippocampal segmentation was obtained using T1-weighted image, and not T1- and T2-weighted images like we did, which has been shown to improve segmentation⁵⁵. More consistent with our results are the report of deficits in CA1, CA2/3, and CA4/DG in ultra-high-risk subjects compared to HC, deficits that were not as extensive as in a group of SZ patients²¹. However, because $\sim 2/3$ of the high-risk population will not convert to psychosis, the interpretation of high-risk studies is complicated. In high-risk individuals who were followed until conversion to psychosis, elevated cerebral blood volume in CA1 region, thought to represent increased metabolism secondary to elevated neuronal activity, predicted subsequent

conversion⁶⁴. The progression of atrophy from CA1 to other hippocampal subfields could originate from the particularly vulnerability of CA1 to dysregulation of glutamatergic neurotransmission and excitotoxic injury^{30,65,66}.

In summary, our results support that hippocampal abnormalities are already present at psychosis onset but indicate a more extensive reduction in hippocampal subfield volumes in early psychosis than previously thought.

Relationship between left hippocampus subregions and DUP

We found significant associations between longer DUP and smaller whole hippocampus volumes, as well as CA1, ML, subiculum, presubiculum, and tail. Our findings are consistent with those of Goff et al.³¹ who reported that, in FEP, longer DUP was associated with accelerated hippocampal atrophy over just 8 weeks of antipsychotic treatment, suggesting a persistent effect of DUP on brain structure. These results are consistent with evidence of a damaging effect of DUP on other brain structures^{31,67}. Although we did not find an association between hippocampal volumes and memory performances, another study did find associations between hippocampus subregions and cognitive function in high risk and SZ (ref.²¹). Taken together these data could potentially explain how longer DUP acting through deficits in hippocampus subfields is linked to worse outcome in FEP patients^{1,4}.

	нс	FEP all	FEP subgroup				
			Long DUP	Short DUP			
	<i>n</i> = 41	<i>n</i> = 54	<i>n</i> = 17	n = 37	<i>t</i> -Test (long vs short)		
Hippocampus subfield volum	es and DUP						
Left whole hippocampus	—	r = −0.37, P < 0.01*	r = -0.05, P = 0.86	$r = -0.38, P = 0.02^*$	P = 0.18; d = -0.42		
Left hippocampal tail	—	$r = -0.32, P = 0.04^*$	r = -0.26, P = 0.96	r = -0.37, P = 0.13	P = 0.46; d = -0.35		
Left subiculum	—	$r = -0.41, P = 0.02^*$	r = 0.16, P = 0.96	r = -0.42, P = 0.10	P = 0.36; d = -0.55		
Left cornu ammonis 1	—	$r = -0.31, P = 0.04^*$	r = -0.04, P = 0.96	r = -0.32, P = 0.16	P = 0.46; d = -0.38		
Left presubiculum	—	$r = -0.33, P = 0.04^*$	r = 0.26, P = 0.96	r = -0.26, P = 0.26	P = 0.36; d = -0.50		
Left molecular layer	—	$r = -0.30, P = 0.04^*$	r = -0.14, P = 0.96	r = -0.24, P = 0.32	P = 0.46; d = -0.36		
Left GC-ML-DG	—	r = -0.20, P = 0.17	r = 0.02, P = 0.96	r = -0.19, P = 0.32	P = 0.46; d = -0.28		
Left cornu ammonis 3	—	r = -0.18, P = 0.21	r = -0.12, P = 0.96	r = -0.18, P = 0.32	P = 0.52; d = -0.20		
Left cornu ammonis 4	—	r = -0.21, P = 0.17	r = -0.02, P = 0.96	r = -0.18, P = 0.32	P = 0.46; d = -0.28		
Hippocampus subfield volum	es and Glx level						
Left whole hippocampus	r = -0.12, P = 0.47	r = 0.14, P = 0.32	r = 0.47, P = 0.10	r = 0.03, P = 0.88	—		
Left hippocampal tail	r = 0.04, P = 0.80	r = 0.22, P = 0.69	r = 0.29, P = 0.43	r = 0.25, P = 0.67	—		
Left subiculum	r = -0.20, P = 0.80	r = 0.06, P = 0.69	r = 0.29, P = 0.43	r = 0.06, P = 0.88	—		
Left cornu ammonis 1	r = -0.08, P = 0.89	r = 0.06, P = 0.69	r = 0.22, P = 0.47	r = 0.03, P = 0.88	—		
Left presubiculum	r = -0.09, P = 0.80	r = 0.10, P = 0.69	r = 0.44, P = 0.26	r = 0.05, P = 0.88	—		
Left molecular layer	r = -0.22, P = 0.80	r = 0.13, P = 0.69	r = 0.51, P = 0.26	r = 0.04, P = 0.88	—		
Left GC-ML-DG	r = -0.08, P = 0.80	r = 0.08, P = 0.69	r = 0.49, P = 0.26	r = -0.22, P = 0.67	—		
Left cornu ammonis 3	r = -0.06, P = 0.80	r = 0.07, P = 0.69	r = 0.26, P = 0.43	r = -0.03, P = 0.88	_		
Left cornu ammonis 4	r = -0.05, P = 0.80	r = 0.06, P = 0.69	r = 0.45, P = 0.26	r = -0.20, P = 0.67	_		

Table 3	Correlations	between l	eft hippocan	pus subfield	volumes a	and DUP,	Glx level.
---------	--------------	-----------	--------------	--------------	-----------	----------	------------

Pearson partial correlation coefficients were used to assess relationships between the left hippocampus subfields and DUP or glutamate (with controlling variables, for DUP: age, gender, and ICV; for glutamate level: age, gender, tobacco taking (pack-day), and ICV); all statistical analyses were controlled for multiple comparisons by false detection rate correction.

FEP first episode psychosis's patients, DUP duration of untreated psychosis (long > 12 months > short), BPRS brief psychiatric rating scale, GC-ML-DG granular cell of the dentate gyrus, d Cohen's d.

Glutamate as a candidate to explain the hippocampus atrophy

We did not report association between whole hippocampus or subfield volumes and Glx levels. Our prior study in unmedicated SZ suggested that alterations in hippocampus glutamatergic neurotransmission may play a role in hippocampus deficits³². The lack of association may be the consequence of a nonoptimal overlap between the MRS voxel and the hippocampus volumes. It is also possible that there is a threshold value for a toxic effect of psychosis, rather than a linear relationship between DUP and a neurotoxic effect³⁶. Other mechanism, such as increased dopaminergic activity⁶⁸, or persistent catecholaminergic and hypothalamic pituitary adrenal axis activity⁶⁹ have been proposed as causal factors for the association of DUP with poor outcome.

Limitations

Our findings need to be placed in the context of several strengths and limitations. One of the major strengths here is that all FEP were enrolled at the time of first treatment contact that allowed us to mitigate possible confounds of antipsychotic medication exposure. We also recruited a control group that was carefully matched on key demographic variables, and we controlled for several possible confounding variables (e.g., total ICV) during statistical analyses. We determined DUP with a clinical interview. A quantitative review comparing methods of assessing DUP found that clinical interviews are no less reliable than standardized assessment tools⁷⁰. While our MRS acquisition sequence does have some drawbacks such as J-modulation and T2 relaxation effects on the spectrum, as well as a water signal that is highly T2-weighted and sensitive to cerebrospinal fluid contamination⁵², a significant advantage of these acquisition parameters is that it allows us put findings in context of a number of our previous studies for which we used the same acquisition parameters^{32,71,72}. Regarding the hippocampus segmentation, the previous version of computerized segmentation of the hippocampus by Freesurfer was controversial⁷³, but the new version, by using highresolution T2-weighted MR imaging, addresses these criticisms⁵⁵.

Summary

Here, we demonstrate evidence of lower hippocampal subfield volumes and altered glutamatergic metabolism in those with longer DUP, suggesting possible biological mechanisms underlying the clinical observation that patients with longer DUP suffer from worse overall outcomes. Our data highlight the critical need for reducing the DUP and for early pharmacological intervention, with the hope to prevent structural deficits and improve clinical outcomes.

Acknowledgements

This work was supported by NIMH grants R01 MH102951, MH113800 (A.C.L.), and K23MH106683 (N.V.K).

Conflict of interest

The authors declare that they have no conflict of interest.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Supplementary Information accompanies this paper at (https://doi.org/ 10.1038/s41398-020-0812-z).

Received: 30 October 2019 Revised: 14 April 2020 Accepted: 21 April 2020 Published online: 12 May 2020

References

- Perkins, D. O. Review: longer duration of untreated psychosis is associated with worse outcome in people with first episode psychosis. *Evid. Based Ment. Health* 9, 36 (2006).
- Fusar-Poli, P. et al. Neurofunctional correlates of vulnerability to psychosis: a systematic review and meta-analysis. *Neurosci. Biobehav. Rev.* **31**, 465–484 (2007).
- Perkins, D. O., Gu, H., Boteva, K. & Lieberman, J. A. Relationship between duration of untreated psychosis and outcome in first-episode schizophrenia: a critical review and meta-analysis. *Am. J. Psychiatry* 162, 1785–1804 (2005).
- Marshall, M. et al. Association Between duration of untreated psychosis and outcome in cohorts of first-episode patients: a systematic review. Arch. Gen. Psychiatry 62, 975–983 (2005).
- Olney, J. W. & Farber, N. B. Glutamate receptor dysfunction and schizophrenia. Arch. Gen. Psychiatry 52, 998–1007 (1995).
- Olney, J. W. et al. Drug-induced apoptotic neurodegeneration in the developing brain. Brain Pathol. 12, 488–498 (2002).
- Bird, C. M. & Burgess, N. The hippocampus and memory: insights from spatial processing. *Nat. Rev. Neurosci.* 9, 182–194 (2008).
- Aleman, A., Hijman, R., de Haan, E. H. & Kahn, R. S. Memory impairment in schizophrenia: a meta-analysis. *Am. J. Psychiatry* **156**, 1358–1366 (1999).

- Wael, R. V. et al. Anatomical and microstructural determinants of hippocampal subfield functional connectome embedding. *Proc. Natl Acad. Sci. USA* **115**, 10154–10159 (2018).
- Tamminga, C. A., Stan, A. D. & Wagner, A. D. The hippocampal formation in schizophrenia. *Am. J. Psychiatry* 167, 1178–1193 (2010).
- Olbrich, H.-G. & Braak, H. Ratio of pyramidal cells versus non-pyramidal cells in sector CA1 of the human Ammon's horn. *Anat. Embryol. (Berl.)* 173, 105–110 (1985).
- Freund, T. F. & Buzsáki, G. Interneurons of the hippocampus. *Hippocampus* 6, 347–470 (1996).
- Heckers, S., Heinsen, H., Geiger, B. & Beckmann, H. Hippocampal neuron number in schizophrenia. A stereological study. Arch. Gen. Psychiatry 48, 1002–1008 (1991).
- Schmitt, A. et al. Stereologic investigation of the posterior part of the hippocampus in schizophrenia. Acta Neuropathol. (Berl.) 117, 395–407 (2009).
- Konradi, C. et al. Hippocampal interneurons are abnormal in schizophrenia. Schizophr. Res. 131, 165–173 (2011).
- Heckers, S. et al. Differential hippocampal expression of glutamic acid decarboxylase 65 and 67 messenger RNA in bipolar disorder and schizophrenia. *Arch. Gen. Psychiatry* 59, 521–529 (2002).
- Fujihara, K. C. et al. Glutamate decarboxylase 67 deficiency in a subset of GABAergic neurons induces schizophrenia-related phenotypes. *Neuropsychopharmacology* 40, 2475–2486 (2015).
- Benes, F. M. et al. Regulation of the GABA cell phenotype in hippocampus of schizophrenics and bipolars. Proc. Natl Acad. Sci. USA 104, 10164–10169 (2007).
- van Erp, T. G. M. et al. Subcortical brain volume abnormalities in 2028 individuals with schizophrenia and 2540 healthy controls via the ENIGMA consortium. *Mol. Psychiatry* 21, 547–553 (2016).
- Vita, A. & De Peri, L. New meta- and mega-analyses of magnetic resonance imaging findings in schizophrenia: do they really increase our knowledge about the nature of the disease process? *Biol. Psychiatry* 85, e33 (2019).
- Vargas, T. et al. Hippocampal subregions across the psychosis spectrum. Schizophr. Bull. 44, 1091–1099 (2018).
- Lodge, D. J. & Grace, A. A. Aberrant hippocampal activity underlies the dopamine dysregulation in an animal model of schizophrenia. *J. Neurosci. J.* Soc. Neurosci. 27, 11424–11430 (2007).
- Lieberman, J. A. et al. Hippocampal dysfunction in the pathophysiology of schizophrenia: a selective review and hypothesis for early detection and intervention. *Mol. Psychiatry* 23, 1764–1772 (2018).
- Xiao, Y. et al. Altered cortical thickness related to clinical severity but not the untreated disease duration in schizophrenia. *Schizophr. Bull.* **41**, 201–210 (2015).
- Harrisberger, F. et al. Impact of polygenic schizophrenia-related risk and hippocampal volumes on the onset of psychosis. *Transl. Psychiatry* 6, e868 (2016).
- 26. Watson, D. R. et al. Structural changes in the hippocampus and amygdala at first episode of psychosis. *Brain Imaging Behav.* **6**, 49–60 (2012).
- Baglivo, V. et al. Hippocampal subfield volumes in patients with first-episode psychosis. Schizophr. Bull. 44, 552–559 (2018).
- Wang, A. Y. et al. Bipolar disorder type 1 and schizophrenia are accompanied by decreased density of parvalbumin- and somatostatin-positive interneurons in the parahippocampal region. *Acta Neuropathol. (Berl.)* **122**, 615–626 (2011).
- McHugo, M. et al. Regionally specific volume deficits along the hippocampal long axis in early and chronic psychosis. *NeuroImage Clin.* 20, 1106–1114 (2018).
- Ho, N. F. et al. Progression from selective to general involvement of hippocampal subfields in schizophrenia. *Mol. Psychiatry* 22, 142–152 (2017).
- Goff, D. C. et al. Association of hippocampal atrophy with duration of untreated psychosis and molecular biomarkers during initial antipsychotic treatment of first-episode psychosis. JAMA Psychiatry 75, 370–378 (2018).
- Kraguljac, N. V., White, D. M., Reid, M. A. & Lahti, A. C. Increased hippocampal glutamate and volumetric deficits in unmedicated patients with schizophrenia. *JAMA Psychiatry* **70**, 1294–1302 (2013).
- Kraguljac, N. V., White, D. M., Hadley, J., Reid, M. A. & Lahti, A. C. Hippocampalparietal dysconnectivity and glutamate abnormalities in unmedicated patients with schizophrenia. *Hippocampus* 24, 1524–1532 (2014).
- Kraguljac, N. V. et al. A longitudinal magnetic resonance spectroscopy study investigating effects of risperidone in the anterior cingulate cortex and hippocampus in schizophrenia. *Schizophr. Res.* **210**, 239–244 (2019).
- Bossong, M. G. et al. Association of hippocampal glutamate levels with adverse outcomes in individuals at clinical high risk for psychosis. JAMA Psychiatry 76, 199–207 (2018).

- Rund, B. R. Does active psychosis cause neurobiological pathology? A critical review of the neurotoxicity hypothesis. *Psychol. Med.* 44, 1577–1590 (2014).
- Carpenter, W. T. et al. Decisional capacity for informed consent in schizophrenia research. Arch. Gen. Psychiatry 57, 533–538 (2000).
- Haas, G. L., Garratt, L. S. & Sweeney, J. A. Delay to first antipsychotic medication in schizophrenia: impact on symptomatology and clinical course of illness. J. Psychiatr. Res. 32, 151–159 (1998).
- Altamura, A. C., Bassetti, R., Bignotti, S., Pioli, R. & Mundo, E. Clinical variables related to suicide attempts in schizophrenic patients: a retrospective study. *Schizophr. Res.* 60, 47–55 (2003).
- Primavera, D. et al. Does duration of untreated psychosis predict very long term outcome of schizophrenic disorders? Results of a retrospective study. *Ann. Gen. Psychiatry* **11**, 21 (2012).
- Allott, K et al. Duration of untreated psychosis and neurocognitive functioning in first-episode psychosis: a systematic review and meta-analysis. *Psychol. Med.* 48, 1592–1607 (2018).
- 42. Black, K. et al. Duration of untreated psychosis predicts treatment outcome in an early psychosis program. *Schizophr. Res.* **47**, 215–222 (2001).
- Bottlender, R. et al. The impact of duration of untreated psychosis and premorbid functioning on outcome of first inpatient treatment in schizophrenic and schizoaffective patients. *Eur. Arch. Psychiatry Clin. Neurosci.* 252, 226–231 (2002).
- Overall, J. E. & Gorham, D. R. The brief psychiatric rating scale. *Psychol. Rep.* 10, 799–812 (1962).
- Randolph, C., Tierney, M. C., Mohr, E. & Chase, T. N. The Repeatable Battery for the Assessment of Neuropsychological Status (RBANS): preliminary clinical validity. J. Clin. Exp. Neuropsychol. 20, 310–319 (1998).
- Mullins, P. G., Chen, H., Xu, J., Caprihan, A. & Gasparovic, C. Comparative reliability of proton spectroscopy techniques designed to improve detection of J-coupled metabolites. *Magn. Reson. Med.* **60**, 964–969 (2008).
- Schubert, F., Gallinat, J., Seifert, F. & Rinneberg, H. Glutamate concentrations in human brain using single voxel proton magnetic resonance spectroscopy at 3 Tesla. *NeuroImage* 21, 1762–1771 (2004).
- Vanhamme, L, van den Boogaart, A. & Van Huffel, S. Improved method for accurate and efficient quantification of mrs data with use of prior knowledge. J. Magn. Reson. 129, 35–43 (1997).
- Scheidegger, O. et al. Optimized quantitative magnetic resonance spectroscopy for clinical routine. *Magn. Reson. Med.* **70**, 25–32 (2013).
- Gasparovic, C. et al. Use of tissue water as a concentration reference for proton spectroscopic imaging. *Magn. Reson. Med.* 55, 1219–1226 (2006).
- Gussew, A., Erdtel, M., Hiepe, P., Rzanny, R. & Reichenbach, J. R. Absolute quantitation of brain metabolites with respect to heterogeneous tissue compositions in (1)H-MR spectroscopic volumes. *MAGMA* 25, 321–333 (2012).
- Wilson, M. et al. Methodological consensus on clinical proton MRS of the brain: Review and recommendations. *Magn. Reson. Med.* 82, 527–550 (2019).
- Dale, A. M., Fischl, B. & Sereno, M. I. Cortical surface-based analysis. I. Segmentation and surface reconstruction. *NeuroImage* 9, 179–194 (1999).
- 54. Smith, S. M. et al. Advances in functional and structural MR image analysis and implementation as FSL. *NeuroImage* **23**, S208–S219 (2004).
- Iglesias, J. E. et al. A computational atlas of the hippocampal formation using ex vivo, ultra-high resolution MRI: Application to adaptive segmentation of in vivo MRI. *NeuroImage* **115**, 117–137 (2015).

- 56. Buckner, R. L. et al. A unified approach for morphometric and functional data analysis in young, old, and demented adults using automated atlas-based head size normalization: reliability and validation against manual measurement of total intracranial volume. *NeuroImage* 23, 724–738 (2004).
- Van Leemput, K. et al. Automated segmentation of hippocampal subfields from ultra-high resolution in vivo MRI. *Hippocampus* 19, 549–557 (2009).
- Klapwijk, E. T., van de Kamp, F., van der Meulen, M., Peters, S. & Wierenga, L. M. Qoala-T: a supervised-learning tool for quality control of FreeSurfer segmented MRI data. *NeuroImage* 189, 116–129 (2019).
- Benjamini, Y. & Hochberg, Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. J. R. Stat. Soc. Ser. B Methodol. 57, 289–300 (1995).
- Wood, S. J. et al. A 1H-MRS investigation of the medial temporal lobe in antipsychotic-naïve and early-treated first episode psychosis. *Schizophr. Res.* 102, 163–170 (2008).
- Szeszko, P. R. et al. Smaller anterior hippocampal formation volume in antipsychotic-naive patients with first-episode schizophrenia. *Am. J. Psychiatry* 160, 2190–2197 (2003).
- Haukvik, U. K. et al. In vivo hippocampal subfield volumes in schizophrenia and bipolar disorder. *Biol. Psychiatry* 77, 581–588 (2015).
- Li, W. et al. Volume alteration of hippocampal subfields in first-episode antipsychotic-naïve schizophrenia patients before and after acute antipsychotic treatment. *NeuroImage Clin.* 20, 169–176 (2018).
- Schobel, S. A. et al. Differential targeting of the CA1 subfield of the hippocampal formation by schizophrenia and related psychotic disorders. *Arch. Gen. Psychiatry* 66, 938–946 (2009).
- Schobel, S. A. et al. Imaging patients with psychosis and a mouse model establishes a spreading pattern of hippocampal dysfunction and implicates glutamate as a driver. *Neuron* 78, 81–93 (2013).
- Small, S. A., Schobel, S. A., Buxton, R. B., Witter, M. P. & Barnes, C. A. A pathophysiological framework of hippocampal dysfunction in ageing and disease. *Nat. Rev. Neurosci.* 12, 585–601 (2011).
- Anderson, K. K. et al. Minimal evidence that untreated psychosis damages brain structures: a systematic review. *Schizophr. Res.* **162**, 222–233 (2015).
- Simantov, R. et al. Dopamine-induced apoptosis in human neuronal cells: inhibition by nucleic acids antisense to the dopamine transporter. *Neuroscience* 74, 39–50 (1996).
- Keshavan, M. S. et al. Superior temporal gyrus and the course of early schizophrenia: progressive, static, or reversible? J. Psychiatr. Res. 32, 161–167 (1998).
- Register-Brown, K. & Hong, L. E. Reliability and validity of methods for measuring the duration of untreated psychosis: a quantitative review and metaanalysis. *Schizophr. Res.* 160, 20–26 (2014).
- Hutcheson, N. L. et al. Multimodal analysis of the hippocampus in schizophrenia using proton magnetic resonance spectroscopy and functional magnetic resonance imaging. *Schizophr. Res.* **140**, 136–142 (2012).
- Kraguljac, N. V. et al. Ketamine modulates hippocampal neurochemistry and functional connectivity: a combined magnetic resonance spectroscopy and resting-state fMRI study in healthy volunteers. *Mol. Psychiatry* 22, 562–569 (2017).
- Wisse L. E. M., Biessels G. J., Geerlings M. I. A critical appraisal of the hippocampal subfield segmentation package in FreeSurfer. *Front. Aging Neurosci.* 6, 261 https://doi.org/10.3389/fnagi.2014.00261 (2014).