



REVIEW ARTICLE



# Evolution of prefrontal cortex

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Subdivisions of the prefrontal cortex (PFC) evolved at different times. Agranular parts of the PFC emerged in early mammals, and rodents, primates, and other modern mammals share them by inheritance. These are limbic areas and include the agranular orbital cortex and agranular medial frontal cortex (areas 24, 32, and 25). Rodent research provides valuable insights into the structure, functions, and development of these shared areas, but it contributes less to parts of the PFC that are specific to primates, namely, the granular, isocortical PFC that dominates the frontal lobe in humans. The first granular PFC areas evolved either in early primates or in the last common ancestor of primates and tree shrews. Additional granular PFC areas emerged in the primate stem lineage, as represented by modern strepsirrhines. Other granular PFC areas evolved in simians, the group that includes apes, humans, and monkeys. In general, PFC accreted new areas along a roughly posterior to anterior trajectory during primate evolution. A major expansion of the granular PFC occurred in humans in concert with other association areas, with modifications of corticocortical connectivity and gene expression, although current evidence does not support the addition of a large number of new, human-specific PFC areas.

*Neuropsychopharmacology* (2022) 47:3–19; <https://doi.org/10.1038/s41386-021-01076-5>

## INTRODUCTION

Many neuroscientists assume that cortical organization is highly conserved among mammals—the cortex might differ in size or number of neurons, but its basic elements and pattern of organization are shared by all. The current concentration of research on a few “model” species reinforces this view. In addition to the cerebral cortex of humans, neuroscience devotes the vast majority of its effort to study the favored four: rats, mice, and rhesus monkeys, with common marmosets recently augmenting this limited roster. Because translational neuroscience depends on features of organization shared among species, presumably including humans, there is a tendency to neglect the diversity of mammalian cortex (for discussions, see [1–6]).

If one adopts a broader, comparative perspective, however, it becomes difficult to sustain this view. Of course, some features of the mammalian cortex are widely shared among species by virtue of common ancestry. Nevertheless, the cortex also exhibits the same high level of diversity characteristic of other aspects of mammalian biology [7], different lineages having evolved a wide variety of cortical specializations, just as they evolved numerous specializations of behavior, skeletal anatomy, physiology, macromolecules, and genome.

The same principles apply to the topic of this special issue: the prefrontal cortex (PFC). In this paper, we discuss features of PFC organization that appear to be widely shared among mammals, along with evidence that primates possess a set of PFC areas that most or all other mammals lack: namely, the granular PFC, the part of the PFC that dominates the human frontal lobe.

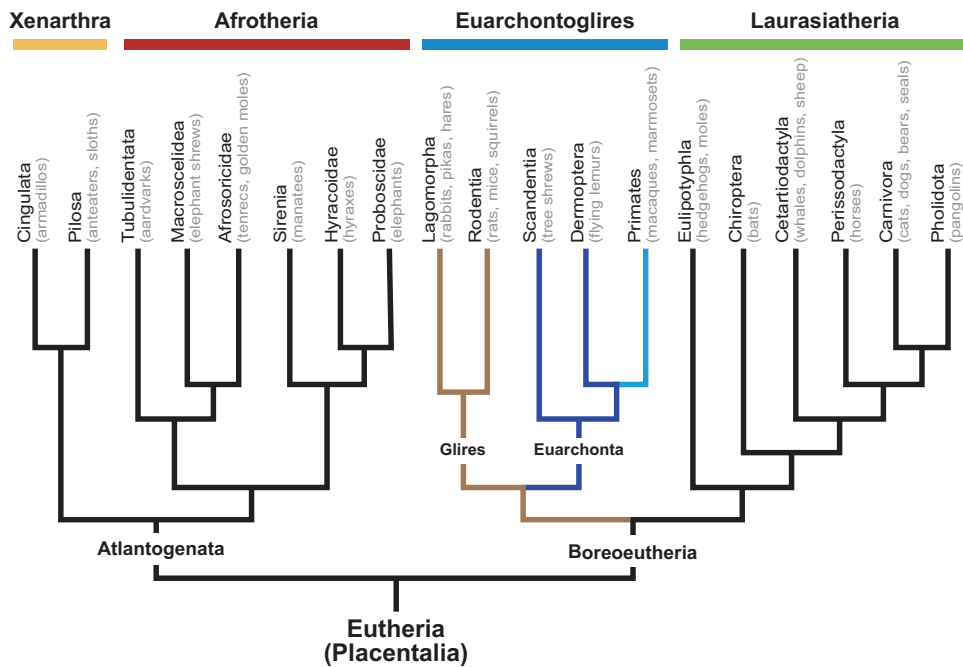
## SOME EVOLUTIONARY FUNDAMENTALS

In order to reconstruct the evolution of the PFC, we need an accurate picture of who is related to whom among mammals. In addition, we need a terminology for designating shared features of brain organization that reflects those relationships.

### Who's related to whom?

Prior to the development of efficient DNA sequencing techniques in the 1990s [8], accounts of the relationships of mammals came mainly from anatomy and have been fraught with uncertainties. For example, at one time bats and certain insectivores were thought to be closely related to primates [9], a view no longer supported [10]. Those inadequacies not only obscured our understanding of which mammals are most closely related to primates but also how primate brains differ from those of our closest relatives. With the newfound ability to sequence large blocks of DNA, the relationship of primates to other mammals have largely been resolved (Fig. 1). Now, we can be confident that the closest relatives of primates are tree shrews and colugos (flying lemurs), although which of those two groups is most closely related to primates remains uncertain, and they could be equally closely related [11, 12]. Together with primates, these animals constitute a group called Euarchonta. Molecular phylogenies also show that rodents are closely related to rabbits (settling another long-running debate), and together this group, called Glires, is the lineage most closely related to Euarchonta. Euarchonta and Glires constitute an even higher-order group, the Euarchontoglires (Fig. 1). And so it goes, with increasingly distantly related branches coalescing in deeper and deeper nodes of the evolutionary tree.

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**Fig. 1 A phylogenetic tree of placental mammals, based on molecular phylogenomics.** The best-established supraordinal clades are labeled at the top. Adapted from Murphy et al. [12, 327].

### Homology and analogy

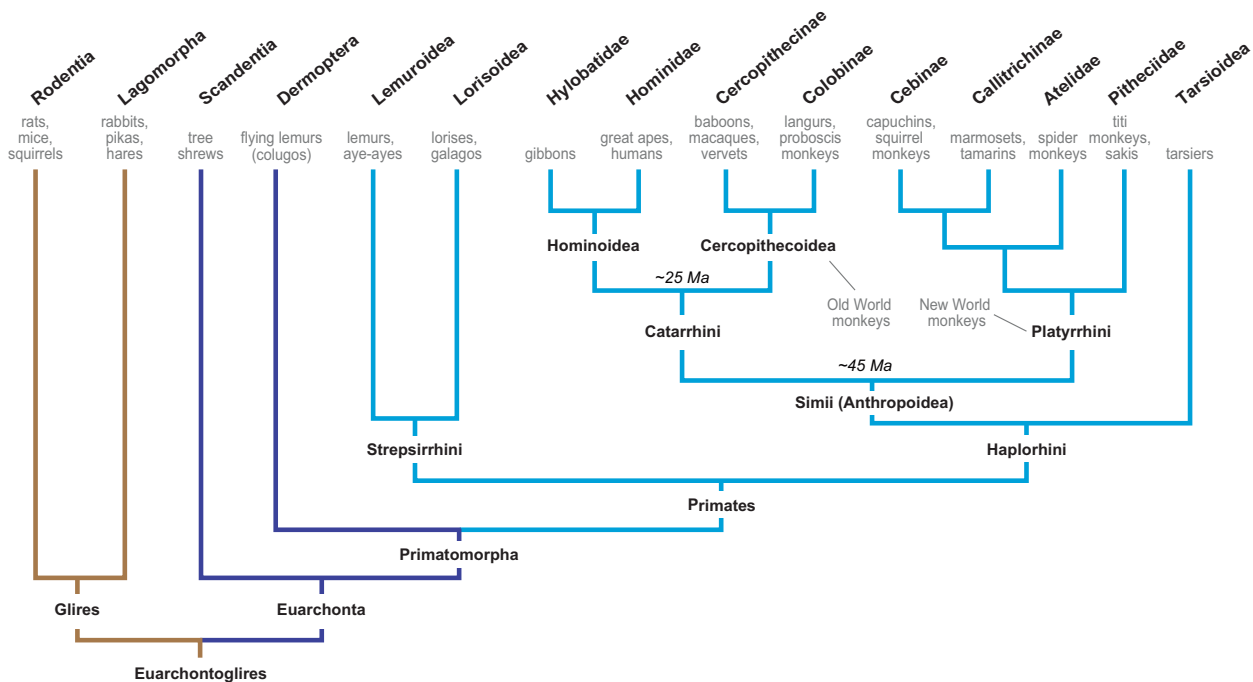
Among the most important concepts in biology are homology and analogy, which provide the framework for understanding similarities and differences among organisms [13, 14]. The concept of homology refers to the “same” feature of an organism present in different species, where sameness implies descent from a common ancestor. A feature can be anything a lineage reproduces across generations. While homology suggests that a feature present in two species shares some attributes, it does not require that they be identical in all respects or even that they have a high degree of similarity. Analogy, in common English, refers to similarity, but in biology it indicates a particular kind of similarity, one that results from independent evolution (also known as convergent or parallel evolution), the feature in question having not been present in the common ancestor of the taxa involved. Because homology and analogy are defined in terms of their relationship to ancestry, they are mutually exclusive. Neuroscientists have sometimes been reluctant to use the term “homology” when comparing the cortex of different species, even when it is clear from the context that they are making a claim of homology, as that concept is currently understood. Even worse, they may imply homology by applying the same term to regions that are similar in some respects, although the balance of evidence indicates they are likely not homologous. For example, we believe the evidence adduced in this review indicates that rodents possess homologs of the agranular medial frontal (MF) and agranular orbital areas of primates but lack homologs of the granular cortex that makes up the largest part of PFC in most primate species. To label the rodent agranular areas as “PFC” and to generalize results in rodents to primates (including humans) without reference to both rodent–primate homologs and primate specializations can only create confusion.

Recognizing homologs and analogs is thus every bit as important in neuroscience as in other branches of biology, and there are some compelling examples. We have good reasons to conclude that all mammals possess homologs of the primary visual (V1), auditory (A1), and somatosensory (S1) areas [15, 16]. For one, in all mammals studied, they occur in the same relative locations

within the cerebral cortex: V1 at the posterior end of the cortex; A1 laterally; and S1 anteriorly. For another, in all mammals studied, V1 receives inputs from the retina via the thalamus; A1 gets inputs from the cochlea via a different thalamic nucleus; and S1 has thalamic inputs that relay signals from cutaneous mechanoreceptors. While the location of an area within the cortical mantle and its connections with the thalamus are commonly employed as indicators of homology for cortical areas, any feature is grist for the mill: other features of connectivity, topographic organization (especially for the sensorimotor areas), architectonics (cyto-, myelo-, and chemoarchitecture), neurophysiological properties, behavioral functions, and so forth. In comparative anatomy, the location of a structure within the body plan has long been considered a critical clue to homology, on the assumption that bodypart locations tend to be stable in evolution. Functions, by contrast, can be quite changeable: for example, homologs of Broca’s and Wernicke’s areas exist in nonhuman primates [17], but only humans have language.

Comparative cortical neuroscience is not all about homology. Convergent evolution is also important. For example, primates and carnivores both possess a large number of extrastriate visual areas [18], and in cats and at least some other carnivores, area V1 includes prominent “blobs” in their upper layers, similar in their enrichment with cytochrome oxidase and certain features of connectivity to those of primates [19]. Because mammals more closely related to primates than carnivores, specifically rodents and tree shrews (Fig. 1), lack these features, we can be confident they evolved convergently, and are thus analogous [19, 20].

Identifying homologous cortical areas or regions in different mammalian groups is complicated by the fact that the number of areas differs across mammals [15, 21]. Moreover, areas that are located in close proximity to each other often share many features, especially of connectivity and function, such as the multiple extrastriate visual areas of primates. It is not enough, then, to simply identify a set of similarities between areas to declare them homologous. Rather, we need to identify sets of diagnostic features that distinguish areas from each other.



**Fig. 2 Relationships among Euarchontoglires [12, 28].** Estimated times for the catarrhine–platyrrhine and the hominoid–cercopithecoidean divergences appear in italics. Ma million years ago.

### Scale thinking versus tree thinking

Today, the accepted metaphor for evolution is, for most purposes, a branching tree, not a phylogenetic scale [13, 14, 22–24]. While one sometimes still hears neuroscientists speak of primates as “higher mammals” and rodents as “lower mammals,” you are unlikely to hear that from an evolutionary biologist. To reject the older metaphor of the phylogenetic scale is not to deny that primates evolved distinctive specializations, but rather to acknowledge that primates, rodents, and other mammalian orders each evolved distinctive features since their divergence in the Mesozoic Era (Fig. 1) [12]. The turn to tree thinking also applies to the Order Primates. In the past, primate evolution was commonly seen as an ascending scale, with tree shrews (which are no longer considered primates) at the bottom, then progressing through a ranked series of lemurs, tarsiers, monkeys, and apes, culminating in the highest rank: humans [25, 26]. Modern views of primate evolution emphasize diversification rather than ascent, with hundreds of species organized in multiple, nested lineages [27, 28]. Figure 2 presents an evolutionary tree of primates in the context of other Euarchontoglires.

### What, if anything, is a monkey?

This change in perspective has important consequences for how neuroscientists should think about primates. For example, one thing that is clear from Fig. 2 is that there is no such thing as “the monkey” from an evolutionary viewpoint. A “thing” from that perspective is a monophyletic group, also known as a natural group or a clade. A monophyletic group comprises the complete set of species that descended from a common ancestor and only those species [13, 22].

New World and Old World monkeys are not a natural group, but rather two distinct lineages that are not even each others’ closest relatives. The closest relatives of the Old World monkeys (the Cercopithecoidea) are members of the ape–human clade (the Hominoidea). Collectively, these two groups compose the Catarrhini. The closest relatives, collectively, of the Catarrhini are the New World monkeys (the Platyrrhini). Thus, references to “the monkey” are problematic; it is important to specify which

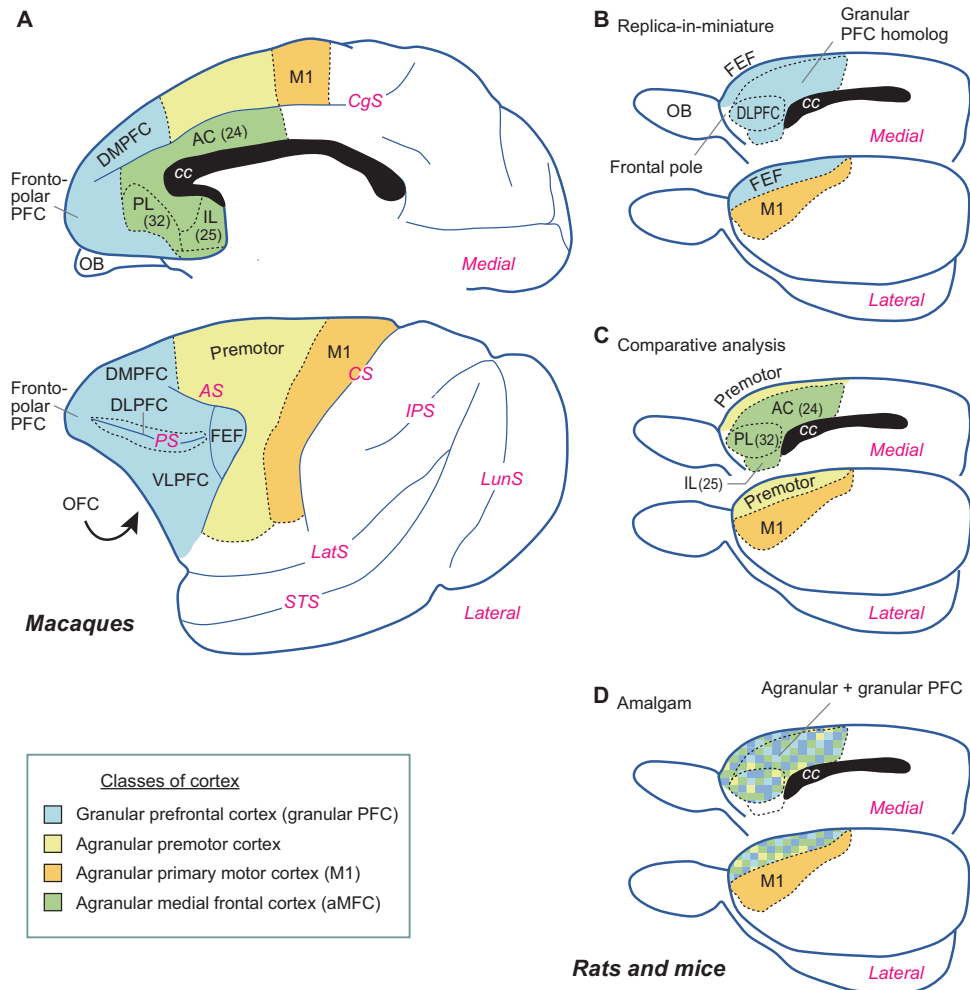
monkeys are under consideration. Similarly, the term “prosimian”—consisting of lemurs, bushbabies, and tarsiers—does not refer to a natural group because the descendants of their last common ancestor include the simians (the catarrhines and platyrrhines), which are not prosimians. Because tarsiers are more closely related to simians than to the lemur–loris–bushbaby group, primates are now usually considered to be comprised of two main clades, the Strepsirrhini (lemurs, lorises, and bushbabies) and the Haplorhini (tarsiers plus simians) (Fig. 2).

If “the monkey” is problematic, references to “the primate” are even more so, unless it is clear from the context exactly which of the myriad primate species are under discussion (Figs. 1 and 2). Brain organization varies among primates, so it is dangerous to assume that what is true of one primate species is true of all. Similar considerations apply to “the mammal” and “the rodent.” However, given that most rodent studies are carried out in rats and mice, we will use “rodent” as convenient shorthand for those taxa. A somewhat wider array of primates has been studied, so we will usually specify from which primate data were obtained.

Although it is desirable to obtain data from a broad range of species when reconstructing evolutionary change, the general paucity of species studied by neuroscientists means that analyses of brain structures usually depend on a regrettably small number of species (e.g., [1, 2, 6, 19, 29–32]). Even in primates, almost all our information about the frontal cortex of strepsirrhines comes from studies of bushbabies (also known as galagos), and then from just two species of the genus *Otolemur*. (These two species were formerly classified as members of the genus *Galago* but are now recognized as a separate genus in the galagid family, all of which are termed “galagos.”) Similarly, almost all our knowledge of Old World monkeys comes from studies of rhesus macaques (*Macaca mulatta*) and a few other macaque species.

### CORTICAL ORGANIZATION

Having established the evolutionary rules of the road, we now turn to the organization and evolution of cerebral cortex, with emphasis on the frontal cortex. As has long been understood, the



**Fig. 3 Proposed homologies among frontal areas in primates and rodents.** Areas shaded with the same color have been advanced as homologs. **A** Macaque brains serve as representative simians for comparison with rodents. The curved arrow indicates the location of OFC on the hidden, ventral surface of the macaque frontal lobe. **B** An example of the idea that rodents have some or all of the granular PFC areas observed in simians, albeit in miniature form. **C** The view advocated here, which is based on comparative neuroanatomy. **D** The amalgam theory, in which alternating colored voxels indicate the intermixing of areas. AC anterior cingulate cortex (area 24 in primates), aMFC agranular medial frontal cortex, AS arcuate sulcus, cc corpus callosum, CgS cingulate sulcus, CS central sulcus, DLPFC dorsolateral PFC (also known as the periprincipal PFC), DMPFC dorsomedial PFC, FEF frontal eye field, IL infralimbic cortex (area 25 in primates), IPS intraparietal sulcus, LatS lateral sulcus, LunS lunate sulcus, M1 primary motor cortex, OB olfactory bulb, PFC prefrontal cortex, OFC orbitofrontal cortex, VLPFC ventrolateral PFC, PL prelimbic cortex (area 32 in primates), PS principal sulcus, STS superior temporal sulcus. Adapted from Preuss and Robert [6].

gray matter of the cortex is a mantle or sheet of tissue, however much it might be folded in large-brained mammals. The central region (or core) of the sheet is occupied by isocortex, which contains most of the sensorimotor and association areas. The isocortex is surrounded by three rings of cortex [33–36]. The outermost ring, which forms the rim of the cortical mantle, is the three-layered cortex called allocortex. This consists of the hippocampus and the primary olfactory (piriform) cortex, along with some smaller olfactory structures. It develops in a different way than the isocortex and the other rings, all of which emerge in an “inside-out” manner, meaning that neurons born earlier take positions in the deeper layers.

Two additional rings of cortical tissue lie between the core isocortex and the allocortex: the periallocortex and proisocortex (sometimes collectively referred to as “mesocortex”). The periallocortex, which borders the allocortex, includes the entorhinal cortex, subiculum, para- and presubiculum, part of the insular cortex, and, in the frontal lobe, the posterior-most orbital cortex, contiguous with the insular cortex. The frontal proisocortex, which is sandwiched between the core isocortex and the periallocortex,

is comprised of the agranular MF cortex (aMFC), consisting of area 24 (the anterior cingulate area, AC), area 32 (the prelimbic area, PL), and area 25 (the infralimbic area, IL), as well as parts of the orbital and insular cortex adjacent to isocortex. The aMFC corresponds to the anterior part of Brodmann’s cingulate region. Primates, but not rodents, have subdivisions of area 32 that are dysgranular as well as agranular [37], but it is convenient to refer to this cingulate region collectively as aMFC. Similarly, primate orbitofrontal cortex (OFC) is a component of the PFC that includes posterior agranular and dysgranular components, as well as anterior, granular divisions, whereas rodent OFC is exclusively agranular. Posterior proisocortex includes the retrosplenial and parahippocampal cortex. Histologically, many of the proisocortical areas resemble isocortex, but may lack one or more layers characteristic of it.

The three rings surrounding the isocortex—proisocortex, periallocortex, and allocortex—have long been considered parts of the limbic system, owing not only to their location along the margin (limbus) of the cortical mantle but also to their close functional relationship with the autonomic nervous system [38–46]. The

distinction between isocortex and proisocortex, in particular, is significant for understanding homologies relevant to PFC evolution, because agranular parts of the orbital and MF cortex, much of which is proisocortex, are usually (but not always) classified as part of the PFC. This inconsistency raises a deceptively simple question: What is the PFC?

## WHAT IS THE PFC?

### Historical perspective

For the past 60–70 years, neuroscientists have commonly defined the PFC as the cortical territory targeted by projections from the mediodorsal nucleus (MD) of the thalamus [47, 48]. Brodmann [49] recognized a large *Regio frontalis* in multiple primate species, including areas occupying the anterior-most lateral, dorsal, medial, and orbital surfaces of the hemisphere. This region became known as the “PFC,” “granular frontal cortex,” “frontal association cortex,” or some variant of those terms. We will call it the granular PFC. Brodmann indicated it consists of isocortex with a “compact inner granular layer” (layer 4). In fact, this is not true for the entirety of his frontal region, because layer 4 granule cells progressively diminish from anterior to posterior in the OFC, which thus consists of granular, dysgranular, and agranular territories [50–55], and only the granular areas are definitely isocortical. Brodmann also concluded that carnivores and ungulates have a single, small region of granular PFC, whereas rodents and rabbits have none.

Brodman’s conclusions have been controversial because they imply that the granular PFC, and presumably the higher-level cognitive functions it supports, is absent in the most widely used neuroscience models: rodents. By the middle of the 20th century, however, it seemed difficult to reconcile Brodmann’s views with evidence about thalamocortical connectivity. Based on studies of retrograde degeneration in the thalamus following cortical lesions, anatomists generated a parcellation of the cortex based on its thalamic afferents (e.g., [56–58]), illustrations of which can still be seen in modern neuroanatomy textbooks. According to this schema, MD projects to the granular PFC, whereas other parts of the frontal lobe receive projections from other nuclei—the cingulate gyrus (including the aMFC) from the anterior thalamic nuclei, and the primary and nonprimary motor areas from the ventral tier nuclei.

On the assumption that MD projections are diagnostic of the granular PFC in primates, and the fact that MD is a readily identifiable in all the commonly studied mammalian brains, a solution to Brodmann’s dilemma presented itself: to identify the granular PFC homolog in nonprimate mammals, even if it is not granular, one need but find the MD-projection cortex. Rose and Woolsey pursued this approach in their lesion-degeneration studies of sheep (cetartiodactyls) and cats (carnivores) [59], and in both, the MD-projection cortex was localized to anterior parts of the frontal lobe. They concluded: “... a cortical field equivalent to the frontal granular cortex of primates is present in all the animals studied.” Subsequently, Akert [60] went further, proposing homologs in cats of the primate OFC, dorsolateral PFC (DLPFC), and the frontal eye field (FEF; granular area 8), based on the medial-to-lateral distribution of degeneration in MD after cortical lesions, which matches the topography of connections between MD and the granular PFC in nonhuman primates.

One vexing problem remained: where is the MD-projection cortex in rodents? Definitive resolution of this issue awaited the development of improved axonal fiber-tracing techniques—first, stains to identify anterogradely degenerating fibers, then injectable tracers. In 1969, in studies of rats, Leonard reported tracing fibers to the orbital cortex along the anterior end of the rhinal sulcus after lesions of the medial-most MD, to the medial wall of the hemisphere superior and anterior to the genu of the corpus callosum after lesions more laterally in MD, and dorsomedially, along the “shoulder” of the hemisphere (where the medial and

lateral surfaces meet) after lesions of the lateral-most MD ([61–63], see also [64, 65]). Subsequent studies using injectable tracers in rats confirmed the existence of reciprocal connections between MD and the sulcal cortex (i.e., the agranular orbito-insular cortex) and the aMFC, as well as the “shoulder” cortex, although it is significant for claims about homology that they also revealed reciprocal connections with the anterior thalamic nuclei, as well as the ventral, intralaminar, and midline thalamic nuclei [66–81]. Research in other rodents [82–84] and in rabbits [68, 85] yielded similar results.

Leonard’s [61] initial interpretation of homologies was similar to that of Akert [60], as illustrated in Fig. 3B. On that view, rat orbital cortex corresponds to the most posterior parts of primate OFC and the shoulder cortex to the primate FEF. As for the cortex of the medial wall (the aMFC), Leonard argued, “tentatively and largely by exclusion,” for homology with the granular PFC in primates. Interestingly, Leonard explicitly acknowledged that while the granular PFC in primates is isocortical, rat aMFC is a “primitive, relatively undifferentiated type of cortex”—that is, *not* isocortex [61]. Although they did not address the histological classification of the aMFC, Krettek and Price [86] reached a similar conclusion, stating: “... the functional significance of the cortical areas rostral to the level of the genu of the corpus callosum in the rat should be considered in terms of their relationship with the MD nucleus and their possible correspondence to PFC of primates rather than on their traditional association with the cingulate gyrus, which was based largely on topographical considerations.”

Collectively, these studies were instrumental in establishing the idea that rodents possess cortical fields corresponding to those that comprise primate PFC including, significantly, the granular PFC [87–90]. Typically, the proposed homolog or counterpart of the primate granular PFC is localized mainly to the rat’s aMFC (areas 24, 32, and 25). That raises a serious problem, however: if the aMFC of rodents is homologous to primate granular PFC, then where is the rodent homolog of primate aMFC?

### Comparing the PFC in primates and rodents

At the same time that new fiber-tracing techniques were being employed in rodents, they were also being applied in primates. The results threw a monkey wrench, as it were, into the revised interpretation of rodent frontal cortex (reviewed by Preuss [48]; see also [91–93]).

These studies in Old World macaques, New World owl monkeys and marmosets, and strepsirrhine galagos confirmed some of the results of the older lesion-degeneration studies in macaques and humans, specifically that nucleus MD projects to the dorsal, lateral, and medial granular PFC and to the OFC. Significantly, however, they also demonstrated that MD connections reach a much greater expanse of cortex than Brodmann’s *Regio frontalis*, including the aMFC (areas 24, 32, and 25), the agranular insula, and the temporal pole [55, 94–102]. MD also projects to primary and premotor areas (e.g., [96, 97, 103–106]), although these are not as numerous as the projections from the ventral nuclei. There are also weak connections with parahippocampal cortex and temporal and parietal isocortex, and possibly the posterior insular cortex, although these may be mainly corticothalamic projections only [107–114]. Moreover, primate PFC, including granular PFC, agranular MFC, and agranular OFC, is connected not only with MD but also with additional thalamic nuclei including the anterior, ventral, midline, intralaminar, and medial pulvinar nuclei (see the citations earlier in this paragraph, plus [115–118]), although not every part of PFC is connected with all these nuclei.

These findings show that MD projects to areas outside the PFC, and they highlight two problems. First, to sustain a practical definition of the primate PFC in terms of MD projections, one would have to restrict the PFC to those regions that receive a majority or plurality of their thalamic inputs from MD, which would likely yield something corresponding closely to

Brodmann's *Regio frontalis*, with the addition of agranular insular cortex. But quantification of axons or terminals in the requisite way remains more aspirational than practical. An alternative, simpler approach would be to define PFC as all the frontal cortex exclusive of motor and premotor cortex [91]. Other definitions have been suggested, such as the cortical territory with MD projections plus certain additional attributes (e.g., [119–121]). A recent data-driven approach in mice based on thalamic and cortical connectivity yielded a PFC “module” consisting of the agranular OFC and aMFC [122, 123]. Second, if primate PFC includes both the granular PFC and the aMFC, then the idea that the rodent aMFC is homologous to the former rather than the latter, as illustrated in Fig. 3B, is severely challenged.

There is additional evidence undermining claims of homology of the rodent aMFC with the primate granular PFC but supporting its homology with the primate aMFC. For one thing, primate aMFC and rodent aMFC share the same location in the cortical mantle: on the medial wall superior, anterior, and ventral to the genu of the corpus callosum. Both consist of agranular proisocortex. Both receive projections from nucleus MD, and detailed analysis of the topography of MD projections to aMFC cortex in rats and macaques highlights their similarity [124, 125]. Significantly, both have connections with the anterior thalamic nuclei—classically regarded as the hallmark of the aMFC. Moreover, both have efferent projections to nucleus accumbens, hypothalamus, and periaqueductal gray, reciprocal connections with the amygdala, and inputs from the hippocampus [43, 45, 94, 102, 126–135], indicating that they are elements of the limbic system. The homology of these rodent and primate regions is now commonly acknowledged (e.g., [37, 43, 44, 47, 91, 131, 132, 136–140]).

If the rodent aMFC is homologous to the primate aMFC (Fig. 3C), and the rodent sulcal cortex to the caudal-most, agranular parts of the primate OFC (the latter not being disputed), then there simply is no good candidate for a granular PFC homolog in rodents. Evidence for such a homolog would require the identification of features that are: (1) characteristic of both the granular PFC of primates and its proposed homolog in rodents and (2) absent from the aMFC or agranular OFC of primates. MD projections are not diagnostic of granular PFC because they fail this test and because, as explained above, they also target areas outside of the PFC by any definition. Other similarities between granular PFC and rodent aMFC have been cited, notably the existence of strong projections from dopaminergic neurons [71], and involvement in spatial-delay tasks, such as the delayed alternation task (e.g., [87, 88, 141, 142]), the latter being typified by a delay period prior to a choice between two spatial locations. Neither of these is diagnostic of the primate granular PFC, either. The aMFC and OFC of primates receive dense dopaminergic innervation, as do some (but not all) of the granular PFC areas [143–151]. However, the primary motor and premotor areas are also strongly innervated by dopaminergic neurons in macaques and humans, but not in rats [152]. Likewise, although lesions of the periprincipal region of granular PFC (the cortex within and surrounding the principal sulcus) produce impairments on spatial-delay tasks in macaques, modest impairments also follow lesions of the aMFC in macaques [153, 154]. There is also evidence of delay-period single-neuron activity [155, 156] and functional-imaging activations [157] in the macaque aMFC during performance of these tasks. In humans, a distributed network of areas that includes the aMFC and the DLPFC is involved in spatial-delay tasks [158]. Thus, spatial-delay-period activity and lesion effects are not diagnostic of the periprincipal PFC. What is more, if the analysis of area homologies presented below is correct, homologs of periprincipal cortex are not even present in all primates, being a specialization of simians.

Studies of corticostriatal projections provide additional support for the conclusion that rodents and primates share homologs of the agranular OFC and MFC areas, and that rodents lack a homolog of the granular PFC. Using conventional tract-tracing

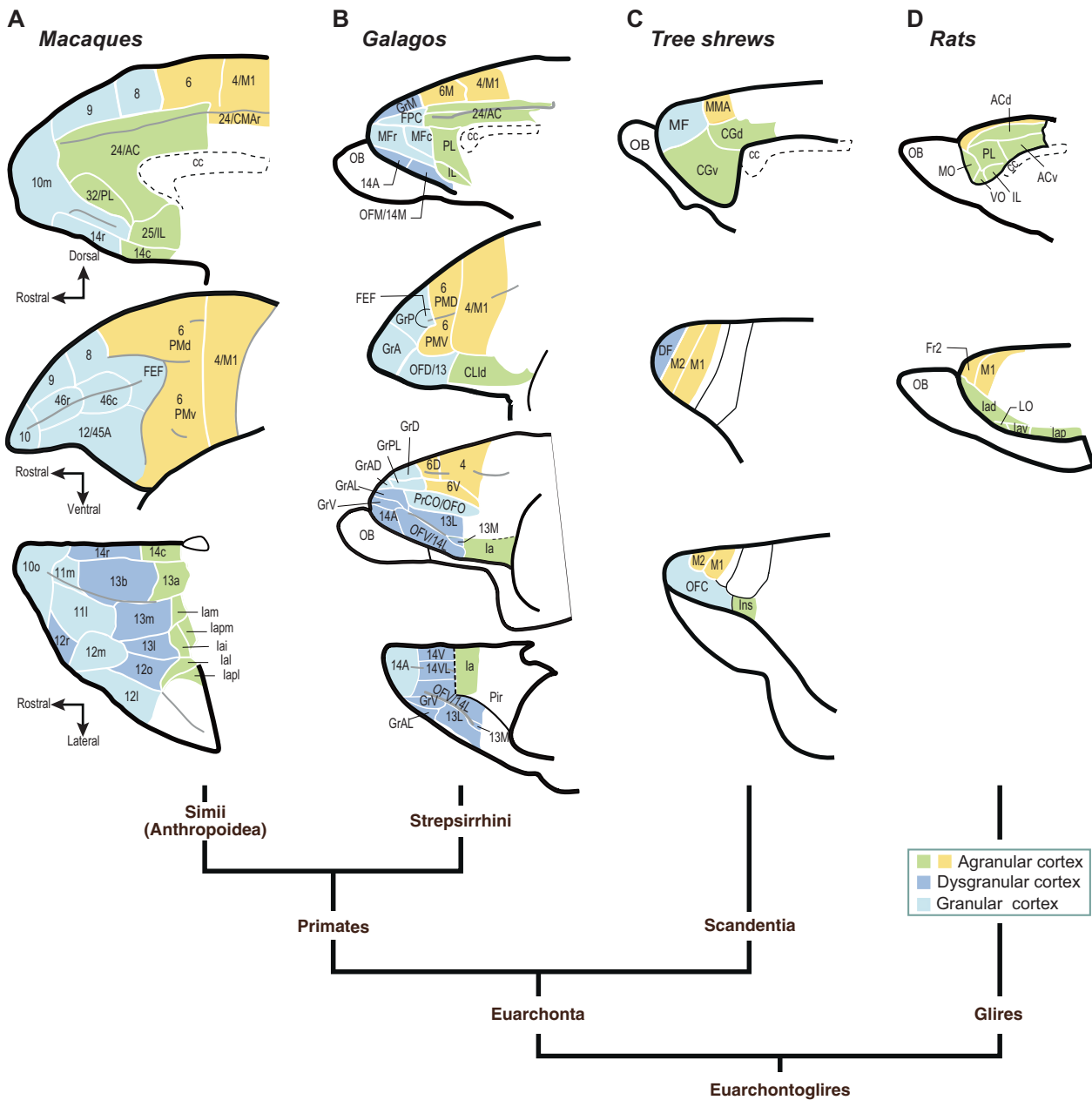
techniques to compare rats and macaques, Heilbronner et al. [137] found that projections from the aMFC and OFC of rats and macaques occupy topographically corresponding domains within the striatum. Using resting-state functional magnetic resonance imaging (MRI) signals in mice, macaques, and humans, Balsters et al. [159] examined signal covariance that depended on corticostriatal projections. They then analyzed the total pattern of corticostriatal projections from each voxel of the imaged brains, creating a “connectional fingerprint” for different areas. Balsters et al. concluded that anterior parts of the granular PFC in macaques and humans have no counterparts in mouse brains.

There is also the matter of rodent shoulder cortex and the proposal that it is homologous to the primate FEF, as depicted in Fig. 3B. Support for this idea came from reports that eye movements can be evoked from this area with intracortical microstimulation [160–162]. More recent results, however, suggest that this region represents movements of the vibrissae [161, 163–168] and may be part of M1 [166, 168]. In addition, there is evidence that in rodents, eye movements are represented in the aMFC rather than in the shoulder cortex [163]. Notably, there is evidence for an oculomotor representation in the macaque aMFC [169] as well as in multiple premotor areas of macaques and other simians, in addition to the arcuate FEF [169–173]. Thus, an oculomotor representation is not diagnostic of the FEF or any other part of granular PFC.

Figure 3D illustrates another idea about homologies, which is that the rodent aMFC, and perhaps the “shoulder” cortex, contain an amalgam of the aMFC and granular PFC areas of primates [62, 174]. This idea attempts to resolve the problem of the missing aMFC in rodents by positing that the granular PFC and aMFC are undifferentiated, and the region in question mixes their features. The proposal has little to recommend it, as rodent aMFC does not appear to have properties of the primate granular PFC that are not also properties of primate aMFC, such as afferents from the anterior thalamic nucleus and involvement in spatial-delay tasks. If the granular PFC is a primate specialization, moreover, we should expect it to have properties that rodents and other mammals lack. There are indeed such properties, both structural and functional.

Perhaps the most important primate specialization from a comparative perspective is the system of connections of the granular PFC. This is best understood in the broader context of primate cortical organization. All primates that have been examined possess regions that correspond to the classical higher-order association territories: the granular PFC, the posterior parietal cortex, and large portions of the temporal cortex. Each of these regions includes multiple areas, and those areas are linked in multiple transcortical networks, which are themselves linked with limbic cortical areas (e.g., [102, 175–182]). No other mammal studied to date has a comparable system of transcortical networks. Moreover, in macaques and marmosets, in which the connections have been studied adequately, these association and limbic areas receive inputs from the medial pulvinar [101, 112, 115, 117, 183–188], a thalamic nucleus present in all primates studied that has no apparent homolog in nonprimate mammals ([189, 190], and see [191]). Thus, the granular PFC is part of a larger system of association areas with features unique to primates among mammals that have been studied.

The granular PFC also has functional properties that the aMFC lacks. Although involvement in spatial-delay tasks per se is not diagnostic of the periprincipal granular PFC, aMFC lesions in rats do result in mild and temporary impairments on such tasks. However, these effects contrast with the severe and permanent effects seen after lesions of the periprincipal cortex in macaques (reviewed in [48, 92]). In addition, macaque granular PFC neurons encode associations between acoustic stimuli and abstract behavior-guiding rules [192] and between color–shape stimuli and abstract problem-solving strategies [193], some of which



**Fig. 4 Types of cortical areas in the frontal cortex of Euarchontoglires.** **A** Macaques (*Macaca*) as representative simians. Adapted from Carmichael and Price [328]. **B** Galagos (*Otolemur*) as representative strepsirrhines. This is an amalgamation of the interpretations of Preuss and Goldman-Rakic [203] and Wong and Kaas [205]. **C** Tree shrews (*Tupaia*) as representative non-primate Euarchontans. Adapted from Wong and Kaas [198]. **D** Rats (*Rattus*) as representative Glires. Adapted from Palomero-Gallagher and Zilles [329]. Labeling has been retained from the originals to the extent possible. a or A anterior or a subdivision of an area (versus b or B), AC anterior cingulate cortex, c caudal, cc corpus callosum, CG cingulate-gyrus cortex, CLI claustral isocortex, CMA cingulate motor area, d or D dorsal, DF dorsal frontal area, FPC frontopolar cortex, Fr2 second frontal area, also known as the medial agranular area, Gr granular, i inferior, la agranular insular cortex, IL infralimbic cortex, Ins insular cortex, l or L lateral, LO lateral orbitofrontal cortex, m or M medial, M1 primary motor cortex, M2, in this case, another part of M1 or a premotor area (not homologous with M2 in simians or rodents, which, in turn, are not homologous with each other), MF medial frontal area, MMA medial motor area, MO medial orbitofrontal cortex, o or O orbital, OB olfactory bulb, OF orbitofrontal cortex, OFO opercular orbitofrontal cortex, p posterior, Pir piriform cortex, PL prefrontal cortex, PM premotor cortex, PrCO opercular preopercular cortex, r rostral, v or V ventral, VO ventral orbitofrontal cortex.

correlate with correct or incorrect task performance [194]. No evidence for such properties has been reported for the aMFC of rodents. Furthermore, lesions of specific parts of the granular PFC in macaques cause profound impairments in rapid learning of arbitrary associations between color–shape stimuli and behavioral goals [195], whereas lesions of the aMFC cause no impairment in rats performing a similar task [196], and even facilitate early stages of learning these associations [197].

Recall that the original intention of the Rose–Woolsey–Akert project was to identify homologs of the primate granular PFC in nonprimate mammals [48]. While studies in rodents initially seemed to indicate that their aMFC (Fig. 3B) filled that role, the balance of evidence indicates that this part of the rodent cortex is homologous to the aMFC of primates (Fig. 3A, C), and that those animals lack a homolog of the primate granular PFC (Figs. 3C and 4). Indeed, the effect of defining the PFC as MD-projection cortex

has not been so much to find the rodent PFC [63], as was the original intent, as to rebrand the aMFC as part of the PFC [6]. We see nothing inherently wrong with including the rodent aMFC and the agranular OFC or orbito-insular cortex in the PFC, as long as the correct homologies among PFC regions are recognized [6, 48, 91]. In our view, however, attempts to interpret data from rodent aMFC or agranular OFC based on homology or similarity to the granular PFC of humans or nonhuman primates are unwarranted.

### Comparing the PFC of primates and tree shrews

Although it now seems likely that the granular PFC is absent in rodents, rodents are not the mammals most closely related to primates, and it is possible that some or all the areas and attributes found in the primate granular PFC are present in mammals that are more closely related to primates, specifically, in colugos (“flying lemurs”) and/or tree shrews (Fig. 2). Colugos have rarely been studied. In tree shrews, however, there is a small area, labeled DF (dorsal frontal cortex) in Fig. 4C, that is perhaps best described as dysgranular based on Wong and Kaas [198]. However, they describe both an anterior part of the OFC and a medial frontal area (MF) as having a “well-developed layer 4.” It is unknown whether either MF or DF is homologous with parts of the granular PFC in primates. More likely is the possibility that the granular OFC of tree shrews is homologous with small parts of the granular OFC in galagos and macaques. These proposed homologies, however, lack support from data about connectivity and other attributes. The connections of tree shrew areas DF and MF are also largely unknown, but we know something about connections they do not have. In contrast to the parietal visual areas of galagos, macaques, and humans, which have extensive interconnections with dorsolateral and dorsomedial PFC (DLPFC and DMPFC in Fig. 3A), the visual areas of tree shrews confine their projections to motor areas [199]. This suggests that the network connecting posterior parietal and granular prefrontal areas, which is such a prominent feature of primate cortical organization, is absent in tree shrews.

### Changes in shared mammalian areas

In focusing on the frontal cortex at the level of its broad regional organization, we have perhaps created the impression that the evolution of primate PFC simply involves the addition of the granular PFC to the aMFC and agranular OFC of rodents and other mammals. Evolution is not that boring: within the aMFC, macaques and humans appear to have more subdivisions of area 24 than do rodents, and there are differences in the connectivity, functions, and receptor distribution of rodents and primates in the areas they share (e.g., [37, 43, 140, 152]). There are also differences in the aMFC among rodent species (e.g., [37, 200]) and among primates, as discussed below.

### PFC IN PRIMATE EVOLUTION

Primates are a diverse group of mammals and understanding how the PFC evolved requires acknowledging that diversity. Comparative neuroanatomy indicates that much of the action in primate PFC evolution involved the granular PFC.

### Shared primate areas

All modern primates share certain granular PFC areas, while additional areas evolved later, during simian evolution. Comparing galago (strepsirrhines) and New World and Old World simians indicates that these animals share at least two parts of the granular PFC: a region located posteriorly, adjacent to the premotor cortex along the anterior bank of the arcuate sulcus in macaques, that includes the FEF (part of area 8 of macaques), and a region that includes the granular, and possibly dysgranular, components of the OFC.

The evidence for homologous FEFs in strepsirrhine and simian primates is very strong (for strepsirrhines (galagos) see, e.g., [97, 180, 181, 201–206]; for platyrrhines: [173, 207–217]; for catarrhine monkeys: [209, 212, 218–225]; and for apes and humans, see [226]). The area in question is located on the lateral surface immediately anterior to the junction of the dorsal and ventral premotor areas, and it is a strongly myelinated isocortical field, has major connections with the most lateral part of nucleus MD and with visual areas of both the dorsal and ventral streams, and projects to the superficial and intermediate layers of the superior colliculus. Intracortical microstimulation of this region with very low currents elicits eye movements, although as mentioned earlier, eye movements are represented in the premotor cortex and aMFC, as well.

What of the possibility that the FEF evolved prior to the divergence of primates and their euarchontan relatives? Two comprehensive mapping studies of the frontal cortex in tree shrews failed to find any evidence for an FEF [227, 228]. It is possible that FEF was present in an ancestral lineage but lost secondarily in tree shrews, although this seems unlikely because these animals have a very well-developed visual system [229].

The OFC has been well-studied only in simians, but the architectonics and corticocortical connections of this region in galagos closely resemble those of simians [180, 181, 203, 205, 230]. In platyrrhines and catarrhines, the OFC has direct connections with the agranular insular cortex, which represents conjunctions of olfactory, gustatory, somatosensory, and visceral inputs. In addition, the OFC is interconnected with the aMFC and lateral granular PFC, with the temporopolar cortex, and with components of the ventral visual stream, including both inferior temporal cortex and the perirhinal cortex [52, 231–238]. The OFC also projects to the nucleus accumbens, ventral striatum, and hypothalamus, and has reciprocal connections with the amygdala [102, 126, 127, 239–248].

### Simian-specific areas

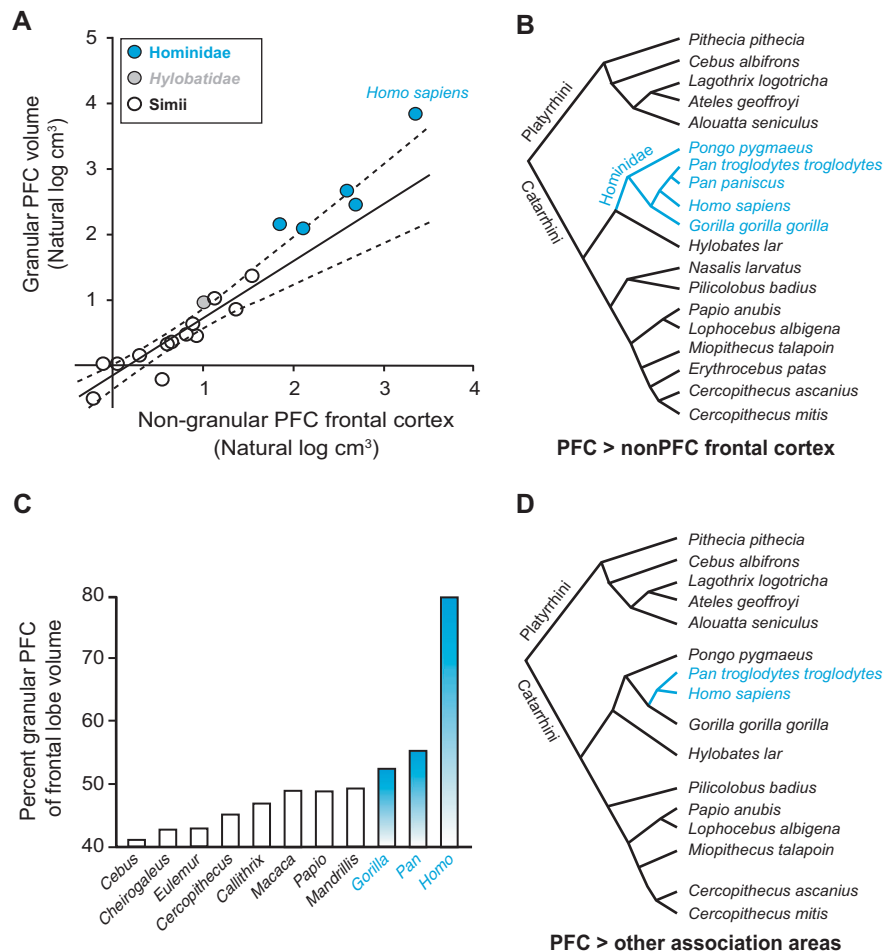
Comparison of galago and simian brains (Fig. 4A, B) reveals that the granular PFC of the latter possesses a number of additional areas, located mainly anterior to the FEF. Some of these areas are less heavily myelinated than are the granular PFC areas shared by galagos and simians, such as the FEF [203, 208, 236, 249]. Figure 4A, B depicts cytoarchitectonics by different shading, as indicated by the key. Based on these observations, and features of corticocortical connectivity, Preuss and Goldman-Rakic [48, 180, 203] concluded that most, if not all, of these more anterior granular PFC areas evolved in haplorhines or simians after their divergence from strepsirrhines. Unfortunately, there is little information about tarsiers, although their frontal lobes are tiny and appear to have little cortex anterior to the precentral region [250, 251], which suggests that the additional areas evolved in simians. The presumably simian-specific areas correspond to the DLPFC (Fig. 3A; also known as the periprincipal cortex and as areas 46 and 9/46 in macaques), ventrolateral PFC (areas 12 and 45, along with area 47 in some species), DMPFC (area 9), and (with somewhat less confidence) frontopolar PFC (area 10).

To summarize the comparative evidence, the FEF appears to be an evolutionary innovation of primates, while most of the anterior, lateral, and medial components of the granular PFC—including the periprincipal cortex—are innovations of simian primates. Most of the granular, and probably dysgranular, OFC is also a primate specialization, although a small part might predate the divergence of tree shrews and primates.

### Diversity among simians

Comparative studies of differences in relative brain size (or encephalization—i.e., brain volume scaled for body mass) reveal that simians almost always have larger brains than strepsirrhines [252–255]. What is more, studies of fossil brain casts reveal that





**Fig. 5 Enlargement of the granular PFC in human evolution.** **A** Granular PFC size relative to the remainder of the frontal lobe. Solid line: regression; dashed lines: confidence limits. **B** Phylogenetic statistical tests reveal that the increase in hominids (blue) is significant. **C** Percentage of granular PFC within the frontal lobe. **D** Phylogenetic statistics show that the clade including chimpanzees and humans underwent a significantly greater increase in relative granular PFC volume contrasted with other homotypical association areas, namely those in the parietal and temporal lobes. **A, B, D** Reproduced, with permission (RightsLink license 5035380571023, license date March 24, 2021) from Smaers et al. [282]. **C** Plotted from data in Elston et al. [330]. PFC prefrontal cortex.

relative brain size increased independently in many primate lineages, with early members of the strepsirrhine, platyrrhine, and catarrhine lineages usually having smaller brains (and smaller frontal lobes) than most of their extant counterparts [256–260]. Given these differences, we might expect that those lineages have different complements of granular PFC areas. Alternatively, all the simian-specific parts of the granular PFC could have evolved in the last common ancestor of simians, and subsequently expanded independently in platyrrhines and catarrhines. Unfortunately, we do not have contemporary, high-quality frontal-lobe maps for many of the numerous platyrrhine species to compare to the well-studied macaque frontal lobe, apart from capuchins (*Cebus*) and marmosets (*Callithrix*). The results are interesting, nonetheless, as the large-brained *Cebus* is reported to have a very similar complement of prefrontal areas as macaques [236], which it closely resembles in its convergently acquired sulcal morphology. By contrast, the small-brained marmosets evidently have a somewhat simplified areal organization, especially in the mid-frontal granular PFC (DLPFC), with a relatively small area 46 and possibly fewer subdivisions than *Cebus* or *Macaca* [208, 233].

The New World callitrichid monkeys (marmosets and tamarins) are especially interesting from the standpoint of size. They are the smallest of the platyrrhines, similar in size to the small strepsirrhines, probably as a result of evolutionary dwarfism [261–263]. It is

unclear whether callitrichids underwent a corresponding reduction in relative brain size in evolution, but their brains are among the smallest in absolute size of all simians [264]. This is potentially functionally significant, given the evidence that absolute brain size is a better predictor of cognitive ability than relative brain size across primate species [265, 266].

#### PFC in human evolution

Recent years have seen a surge of new research on human brain evolution (see, e.g., [267–271]). The evolutionary specializations of the human PFC and especially of the granular PFC have received particular attention.

**Size.** Since at least the time of Brodmann [49, 272], it has commonly been accepted that association cortex, including the granular PFC, underwent enormous expansion in human evolution, both absolutely and relative to the amount of primary sensorimotor cortex. This view has been challenged, however. Based on scaling studies, Barton and colleagues [273, 274] have argued that even though PFC is absolutely much larger in humans than in other primates, humans have the expected amount of PFC for a primate of our brain size. Semendeferi and colleagues [275, 276] have made a similar claim, arguing that the frontal lobe occupies about the same proportion of the cortex in humans as it does in the great apes. So,

those authors would argue that human PFC is not exceptionally large. Other authors, however, have maintained that the data support the traditional view [249, 277–282].

We also support the traditional view, based in part on recent phylogenetic regression analyses (Fig. 5) and studies of cranial endocasts in fossil humans [267]. But one can look at the issue a bit differently [268]. One would never expect a great ape to have a brain as large as ours. In body size, humans overlap the chimpanzees and other African great apes (our closest relatives), but our brains, at about 1450 cc on average, are 3–4 times larger than theirs. One would expect humans to have a brain size similar to that of a chimpanzee (~400 cc). What is the difference? Most human primary sensorimotor areas are only marginally larger than those of chimpanzees, but certain parts of the association cortex—including the dorsolateral, frontopolar, and anterior orbital parts of the granular PFC—are enormously larger [283]. Figure 5A–C shows, for example, that great apes and humans have significantly more granular PFC relative to other components of the frontal lobe. Thus, the increased size of the human brain reflects the nonuniform enlargement of specific cortical areas, especially in the higher-order association cortex. This is the real crux of the matter: humans have far more neural machinery in our granular PFC and in certain other association regions than do great apes or other primates, presumably because natural selection favored enhancement of their functional capacities. Figure 5D indicates that, in humans and chimpanzees, the granular PFC expanded marginally but significantly more than the other association areas.

**Areas.** If granular PFC areas evolved in the earliest primates from ancestral brains that lacked them, and if additional granular PFC areas evolved in simians, it is reasonable to suppose that the complement of granular PFC areas differs between humans and other simians, given the enormous expansion of this region in human evolution, and the view that the addition of cortical areas is an important correlate or cause of brain-size enlargement and the acquisition of new functions (e.g., [20, 49]). While measuring brain size or cortical extent is seemingly straightforward, comparing complements of cortical areas across species is fraught with difficulties, as meaningful comparisons require the application of a common set of reliable techniques across species. Historically, the most widely used technique has been cytoarchitectonics, which involves microscopic inspection of Nissl-stained sections is now considered inadequate by itself. Today, it is widely accepted that parcellations are better when based on multiple techniques, including architectonics (especially observer-independent, quantitative architectonics), connectivity, physiological mapping, and roles in behavior [284, 285].

Naturally, we would especially like to know more about our own species, *Homo sapiens*. There have been a number of modern area-mapping studies comparing humans and macaques, and these suggest that both species share a similar complement of granular PFC areas (e.g., [286–289]). Sallet et al. [289] and Neubert et al. [288] have, however, highlighted some possible differences based on their structural-connectivity MRI parcellation of frontal areas. For example, they found that the cortex on the lateral aspect of the frontal pole in humans has no clear counterpart in macaques, an intriguing result given the involvement of that region in higher cognitive function, such as generalized relational reasoning (e.g., [290, 291]). The study of Balsters et al. [159], based on corticostriatal “fingerprints,” led to a similar conclusion. In their comparison of human and macaque brains, they found that the major difference involved the lateral frontopolar cortex and the territories to which it projects in the anterior caudate nucleus. In addition, there appear to be important differences in the aMFC of macaques and humans, with humans possessing additional dysgranular subdivisions of area 32 [37].

While findings of human–macaque differences such as these certainly bear on what we can learn about human PFC functions

from studying macaques, we cannot assume that areas humans possess but macaques lack are necessarily human specializations. For one thing, we have no comparable information about apes, so we do not know whether these are hominoid (i.e., ape–human) specializations or human specializations (for an example of the difference this makes, see [292]). Second, it is possible, if seemingly unlikely, that these features of PFC were present in the ancestors of macaques, but subsequently lost. Addressing this possibility requires studying additional species, with platyrrhines being especially useful for reconstructing the ancestral state of the catarrhine lineage. Nevertheless, the currently available—albeit limited—evidence suggests that the complement of granular PFC areas in macaques and humans is quite similar, which, given that human frontal lobes are much larger than those of macaques (and apes), implies that at least some of the areas shared by these primates are much larger in humans than in the other species.

**Histology, connectivity, and genomics.** Even though we currently lack the kinds of maps required to compare the complement of areas that make up the granular PFC in apes and humans, we do have tools that enable us to compare other features of PFC organization of humans to chimpanzees, our closest relatives, and to macaques. There are new comparative histological studies, employing Nissl and Golgi staining, and immunohistochemistry (e.g., [148, 293–298]). There are also MRI studies in all three species, providing information, for example, about myeloarchitecture [249] and hemispheric asymmetries [299]. In addition, diffusion-weighted MRI studies have demonstrated human specializations of connectivity, including modified connections of the arcuate fasciculus and other systems that interconnect temporal, parietal, and frontal association cortex [300–305].

There are now also abundant resources, and data, for comparing genes and gene expression in humans, chimpanzees, and other nonhuman primates. Space limitations preclude a review of this active area of research. Significantly, however, much of the gene-expression research has focused on the granular PFC (typically area 46 in the DLPFC), and numerous human specializations involving (if not necessarily limited to) this region have been identified (e.g., [283, 306–312]). These gene-expression changes are likely to have modified the cellular organization and physiology of human cortex, although we currently lack direct evidence of such modifications.

## PFC EVOLUTION IN MAMMALS OTHER THAN THE EUARCHONTOGLIRES

The account above has focused on the PFC in the Euarchontoglires, especially primates. There have been few studies of other mammalian groups, apart from carnivores, a group of placental mammals that, as indicated in Fig. 1, is quite distantly related to primates. Many different mammalian groups underwent enlargement of their frontal cortex during their evolution, including carnivores [14, 254, 313]. The predominant cortical connections of carnivore PFC are with limbic regions [314, 315], suggesting it underwent elaboration of the agranular OFC and/or aMFC. It is reasonable to assume that the PFC expanded independently in other mammalian lineages as well.

## FUTURE RESEARCH DIRECTIONS

There remain several outstanding issues regarding PFC evolution:

- (1) The status of dysgranular areas, especially those in the OFC, requires further attention: Are these separate cortical fields or are they transition zones? Some evidence from macaques points to the former [316], but more data are needed. Also unknown is whether there are homologs of the dysgranular OFC in rodents or other nonprimates.

- (2) Are there homologs of the granular PFC areas in the nonprimate members of the Euarchonta (i.e., tree shrews and flying lemurs)? We know that tree shrews have small granular PFC areas but not whether they are homologs of primate areas. We should be cautious about assessing “granularity” across species based on qualitative descriptions, however, as these accounts are often in poor agreement even among closely related species (see, for example, the contrasting descriptions of layer 4 in carnivore preoral cortex [49, 60, 317–319]). Such disparities emphasize the need to adopt multimethod parcellations, including observer-independent architectonics [320–322].
- (3) Given that platyrrhines and catarrhines underwent independent enlargement of the brain, did they also undergo independent addition of new granular PFC areas? If so, could it be that some of the areas assigned the same numbers and names in platyrrhines and catarrhines evolved independently?

## CONCLUSIONS

The PFC is evolutionarily dynamic and diverse, with new areas and new systems of connections evolving in primates. Despite deficiencies in the data, we can state with reasonable confidence that the rodent PFC consists of homologs of the primate aMFC (areas 24, 32, and 25) and primate posterior, agranular OFC. We can also state with reasonable confidence that the granular PFC is a specialization of primates or possibly of primates and their close euarchontan relatives. From comparisons among primates, we infer that early primates possessed a small set of granular PFC areas, while additional areas, including the DLPFC, evolved later in the simian branch of the primate tree. Our understanding of the evolution of the limbic, agranular PFC is comparatively poor, but given its involvement in social behavior (e.g., [174, 323, 324]) and given the diversity of social behavior among mammals, its organization was likely modified in many primate groups.

We are aware that our view regarding the uniqueness of the primate granular PFC has been unpopular among some neuroscientists who study rodents and other nonprimate species, although we note that in the 30 years since it was first articulated no data have been advanced that convincingly contradict it. We believe, moreover, that many of the ambiguities about the behavioral role of the PFC in rodents as compared to primates are resolved by the interpretation of homologies offered here. What is more, our view by no means negates the importance of rodents as biomedical models. For one thing, the parts of the PFC that rodents and primates do share—namely, the limbic parts—are unquestionably of functional importance, and in some respects of even greater clinical importance than the granular PFC. That is not to say that we should study the limbic PFC only in rodents—any nonhuman primate is more closely related to humans than any rodent and their behavioral phenotypes are more readily compared to those of humans [325]—but rather that our evolutionary analysis does not exclude an important role for rodent research. We note, too, that the caution that we have expressed about uncritically extrapolating findings from model animals to humans applies to catarrhine primates as well as to rodents and platyrrhines, although the much closer relationship of human to other catarrhines mitigates this problem to a considerable extent [326].

## FUNDING AND DISCLOSURE

TMP is supported by NIH ORIP/OD P51 OD011132. SPW has nothing to disclose.

## REFERENCES

1. Manger PR, Cort J, Ebrahim N, Goodman A, Henning J, Karolia M, et al. Is 21st century neuroscience too focussed on the rat/mouse model of brain function and dysfunction? *Front Neuroanat.* 2008;2:5.
2. Bolker J. Model organisms: there's more to life than rats and flies. *Nature.* 2012;491:31–3.
3. Logan CA. The altered rationale for the choice of a standard animal in experimental psychology: Henry H. Donaldson, Adolf Meyer and 'the' albino rat. *Hist Psychol.* 1999;2:3–24.
4. Murray EA, Wise SP, Graham KS. The evolution of memory systems: ancestors, anatomy, and adaptations. Oxford, UK: Oxford University Press; 2017.
5. Preuss TM. The discovery of cerebral diversity: an unwelcome scientific revolution. In: Falk D, Gibson KR, editors. *Evolutionary anatomy of the primate cerebral cortex.* Cambridge, UK: Cambridge University Press; 2001. p. 138–64.
6. Preuss TM, Robert JS. Animals models of the human brain: repairing the paradigm. In: Gazzaniga MS, Mangun GR, editors. *The cognitive neurosciences.* 5th ed. Cambridge, MA: MIT Press; 2014. p. 59–66.
7. Preuss TM. Reinventing primate neuroscience for the twenty-first century. In: Platt ML, Ghazanfar AA, editors. *Primate neuroethology.* Oxford, UK: Oxford University Press; 2010. p. 422–53.
8. Murphy WJ, Pevzner PA, O'Brien SJ. Mammalian phylogenomics comes of age. *Trends Genet.* 2004;20:631–9.
9. Gregory W. The orders of mammals. *Bull Am Mus Nat Hist.* 1910;27:1–524.
10. Cartmill M. New views on primate origins. *Evol Anthropol.* 2005;1:105–11.
11. Halliday TJ, Upchurch P, Goswami A. Resolving the relationships of paleocene placental mammals. *Biol Rev Camb Philos Soc.* 2017;92:521–50.
12. Murphy WJ, Foley NM, Bredemeyer KR, Gatesy J, Springer MS. Phylogenomics and the genetic architecture of the placental mammal radiation. *Annu Rev Anim Biosci.* 2021;9:29–53.
13. Baum DA, Smith SD. *Tree thinking: an introduction to phylogenetic biology.* Greenwood Village, CO: Roberts; 2012.
14. Striedter GF. *Principles of brain evolution.* Sunderland, MA: Sinauer Associates; 2005.
15. Kaas JH. The evolution of brains from early mammals to humans. *Wiley Interdiscip Rev Cogn Sci.* 2013;4:33–45.
16. Krubitzer LA, Seelke AM. Cortical evolution in mammals: the bane and beauty of phenotypic variability. *Proc Natl Acad Sci USA.* 2012;109:10647–54.
17. Rilling JK. Comparative primate neurobiology and the evolution of brain language systems. *Curr Opin Neurobiol.* 2014;28:10–4.
18. Lyon DC. The evolution of visual cortex and visual systems. In: Kaas JH, Krubitzer LA, editors. *Evolution of nervous systems, vol. 3: mammals.* London: Elsevier; 2007. p. 267–306.
19. Preuss TM. Taking the measure of diversity: comparative alternatives to the model-animal paradigm in cortical neuroscience. *Brain Behav Evol.* 2000;55:287–99.
20. Kaas JH. The organization and evolution of neocortex. In: Wise SP, editor. *Higher brain function: recent explorations of the brain's emergent properties.* New York: John Wiley; 1987. p. 347–78.
21. Krubitzer L, Huffman KJ. Arealization of the neocortex in mammals: genetic and epigenetic contributions to the phenotype. *Brain Behav Evol.* 2000;55:322–35.
22. Nunn CL. *The comparative approach in evolutionary anthropology and biology.* Chicago: The University of Chicago Press; 2011.
23. Smulders TV. The relevance of brain evolution for the biomedical sciences. *Biol Lett.* 2009;5:138–40.
24. Preuss TM. *What's human about the human brain? The new cognitive neurosciences.* 2nd ed. Cambridge, MA: MIT Press; 2000. p. 1219–34.
25. Le Gros Clark WE. *The antecedents of man.* Edinburgh: Edinburgh University Press; 1959.
26. Romer AS. *The vertebrate body.* Philadelphia: Saunders; 1970.
27. Fleagle JG. *Primate adaptation and evolution.* 3rd ed. Oxford: Academic; 2013.
28. Fleagle JG, Seiffert ER. The phylogeny of primates. In: Kaas JH, Krubitzer L, editors. *Evolution of nervous systems, second edition, vol. 3: primates.* Amsterdam/Boston: Elsevier/Academic; 2017. p. 1–34.
29. Brenowitz EA, Zakon HH. Emerging from the bottleneck: benefits of the comparative approach to modern neuroscience. *Trends Neurosci.* 2015;38:273–8.
30. Johnson ZV, Young LJ. Evolutionary diversity as a catalyst for biological discovery. *Integr Zool.* 2018;13:616–33.
31. Keifer J, Summers CH. Putting the “biology” back into “neurobiology”: the strength of diversity in animal model systems for neuroscience research. *Front Syst Neurosci.* 2016;10:69.
32. Yartsev MM. The emperor's new wardrobe: rebalancing diversity of animal models in neuroscience research. *Science.* 2017;358:466–9.
33. Braak H. *Architectonics of the human telencephalic cortex.* Berlin: Springer; 1980.

34. Mesulam MM. Principles of behavioral and cognitive neurology. Oxford: Oxford University Press; 2000.
35. Sanides F. Functional architecture of motor and sensory cortices in primates in the light of a new concept of neocortex evolution. *Adv Primatol.* 1970;1:137–201.
36. Stephan H. Evolutionary trends in limbic structures. *Neurosci Biobehav Rev.* 1983;7:367–74.
37. Vogt BA, Hof PR, Zilles K, Vogt LJ, Herold C, Palomero-Gallagher N. Cingulate area 32 homologues in mouse, rat, macaque and human: cytoarchitecture and receptor architecture. *J Comp Neurol.* 2013;521:4189–204.
38. Catani M, Dell'acqua F, Thiebaut De Schotten M. A revised limbic system model for memory, emotion and behaviour. *Neurosci Biobehav Rev.* 2013;37:1724–37.
39. Critchley HD. Neural mechanisms of autonomic, affective, and cognitive integration. *J Comp Neurol.* 2005;493:154–66.
40. Critchley HD, Mathias CJ, Josephs O, O'Doherty J, Zanini S, Dewar BK, et al. Human cingulate cortex and autonomic control: converging neuroimaging and clinical evidence. *Brain.* 2003;126:2139–52.
41. Heimer L, Van, Hoesen GW. The limbic lobe and its output channels: Implications for emotional functions and adaptive behavior. *Neurosci Biobehav Rev.* 2006;30:126–47.
42. Lew CH, Semendeferi K. Evolutionary specializations of the human limbic system. In: Kaas JH, Preuss TM, editors. *Evolution of nervous systems, vol. 4: the evolution of the human brain: apes and other ancestors (second edition).* Amsterdam/Boston: Elsevier/Academic; 2017. p. 277–91.
43. Öngür D, Price JL. The organization of networks within the orbital and medial prefrontal cortex of rats, monkeys and humans. *Cereb Cortex.* 2000;10:206–19.
44. Price JL. Definition of the orbital cortex in relation to specific connections with limbic and visceral structures and other cortical regions. *Ann N Y Acad Sci.* 2007;1121:54–71.
45. Vogt BA. Cingulate cortex in the three limbic subsystems. *Handb Clin Neurol.* 2019;166:39–51.
46. Westerhaus MJ, Loewy AD. Central representation of the sympathetic nervous system in the cerebral cortex. *Brain Res.* 2001;903:117–27.
47. Laubach M, Amarante LM, Swanson K, White SR. What, if anything, is rodent prefrontal cortex? *eNeuro.* 2018;5:1–14.
48. Preuss TM. Do rats have prefrontal cortex? The Rose-Woolsey-Akert program reconsidered. *J Cogn Neurosci.* 1995;7:1–24.
49. Brodmann K. *Vergleichende Lokalisationslehre der Grosshirnrinde.* Leipzig: Barth; 1909. (Reprinted as Brodmann's 'Localisation in the cerebral cortex,' translated and edited by L. J. Garey, London: Smith-Gordon, 1994).
50. Barbas H, Pandya DN. Architecture and intrinsic connections of the prefrontal cortex in the rhesus monkey. *J Comp Neurol.* 1989;286:353–75.
51. Beck E. A cytoarchitectural investigation into the boundaries of cortical areas 13 and 14 in the human brain. *J Anat.* 1949;83:147–57.
52. Carmichael ST, Price JL. Architectonic subdivision of the orbital and medial prefrontal cortex in the macaque monkey. *J Comp Neurol.* 1994;346:366–402.
53. Hof PR, Mufson EJ, Morrison JH. Human orbitofrontal cortex: cytoarchitecture and quantitative immunohistochemical parcellation. *J Comp Neurol.* 1995;359:48–68.
54. Mesulam MM, Mufson EJ. Insula of the Old World monkey. I. Architectonics in the insulo-orbito-temporal component of the paralimbic brain. *J Comp Neurol.* 1982;212:1–22.
55. Morecraft RJ, Geula C, Mesulam MM. Cytoarchitecture and neural afferents of orbitofrontal cortex in the brain of the monkey. *J Comp Neurol.* 1992;323:341–58.
56. McLardy T. Thalamic projection to frontal cortex in man. *J Neurol Neurosurg Psychiatry.* 1950;13:198–202.
57. Rose JE, Woolsey CN. Organization of the mammalian thalamus and its relationships to the cerebral cortex. *Electroencephalogr Clin Neurophysiol.* 1949;1:391–403.
58. Walker AE. *The primate thalamus.* Chicago, IL: The University of Chicago Press; 1938.
59. Rose JE, Woolsey CN. The orbitofrontal cortex and its connections with the mediodorsal nucleus in rabbit, sheep and cat. *Res Publ Assoc Res Nerv Ment Dis.* 1948;27:210–32.
60. Akert K. Comparative anatomy of frontal cortex and thalamofrontal connections. In: Warren JM, Akert K, editors. *The frontal granular cortex and behavior.* New York: McGraw-Hill; 1964. p. 372–96.
61. Leonard CM. The prefrontal cortex of the rat. I. Cortical projection of the mediodorsal nucleus. II. Efferent Connections. *Brain Res.* 1969;12:321–43.
62. Leonard CM. The connections of the dorsomedial nuclei. *Brain Behav Evol.* 1972;6:524–41.
63. Leonard CM. Finding prefrontal cortex in the rat. *Brain Res.* 2016;1645:1–3.
64. Domesick VB. Projections from the cingulate cortex in the rat. *Brain Res.* 1969;12:296–320.
65. Domesick VB. Thalamic relationships of the medial cortex in the rat. *Brain Behav Evol.* 1972;6:457–83.
66. Beckstead RM. Convergent thalamic and mesencephalic projections to the anterior medial cortex in the rat. *J Comp Neurol.* 1976;166:403–16.
67. Beckstead RM. An autoradiographic examination of corticocortical and sub-cortical projections of the mediodorsal-projection (prefrontal) cortex in the rat. *J Comp Neurol.* 1979;184:43–62.
68. Benjamin RM, Jackson JC, Golden GT. Cortical projections of the thalamic mediodorsal nucleus in the rabbit. *Brain Res.* 1978;141:251–65.
69. Berendse HW, Groenewegen HJ. Restricted cortical termination fields of the midline and intralaminar thalamic nuclei in the rat. *Neuroscience.* 1991;42:73–102.
70. Condé F, Audinat E, Maire-Lepoivre E, Crépel F. Afferent connections of the medial frontal cortex of the rat. A study using retrograde transport of fluorescent dyes. I. Thalamic afferents. *Brain Res Bull.* 1990;24:341–5.
71. Divac I, Bjorklund A, Lindvall O, Passingham RE. Converging projections from the mediodorsal thalamic nucleus and mesencephalic dopaminergic neurons to the neocortex in three species. *J Comp Neurol.* 1978;180:59–71.
72. Van Groen T, Wyss JM. Projections from the anterodorsal and anteroventral nucleus of the thalamus to the limbic cortex in the rat. *J Comp Neurol.* 1995;358:584–604.
73. Van Groen T, Kadish I, Wyss JM. Efferent connections of the anteromedial nucleus of the thalamus of the rat. *Brain Res Rev.* 1999;30:1–26.
74. Groenewegen HJ. Organization of the afferent connections of the mediodorsal thalamic nucleus in the rat, related to the mediodorsal-prefrontal topography. *Neuroscience.* 1988;24:379–431.
75. Heidebreder CA, Groenewegen HJ. The medial prefrontal cortex in the rat: Evidence for a dorso-ventral distinction based upon functional and anatomical characteristics. *Neurosci Biobehav Rev.* 2003;27:555–79.
76. Hoover WB, Vertes RP. Anatomical analysis of afferent projections to the medial prefrontal cortex in the rat. *Brain Struct Funct.* 2007;212:149–79.
77. Horikawa K, Kinjo N, Stanley LC, Powell EW. Topographic organization and collateralization of the projections of the anterior and laterodorsal thalamic nuclei to cingulate areas 24 and 29 in the rat. *Neurosci Res.* 1988;6:31–44.
78. Marini G, Pianca L, Tredici G. Thalamocortical projection from the parafascicular nucleus to layer v pyramidal cells in frontal and cingulate areas of the rat. *Neurosci Lett.* 1996;203:81–4.
79. Sarter M, Markowitsch HJ. Collateral innervation of the medial and lateral prefrontal cortex by amygdaloid, thalamic, and brain-stem neurons. *J Comp Neurol.* 1984;224:445–60.
80. Sesack SR, Deutch AY, Roth RH, Bunney BS. Topographical organization of the efferent projections of the medial prefrontal cortex in the rat: an anterograde tract-tracing study with *Phaseolus vulgaris* leucoagglutinin. *J Comp Neurol.* 1989;290:213–42.
81. Shibata H. Efferent projections from the anterior thalamic nuclei to the cingulate cortex in the rat. *J Comp Neurol.* 1993;330:533–42.
82. Fillinger C, Yalcin I, Barrot M, Veinante P. Afferents to anterior cingulate areas 24a and 24b and midcingulate areas 24a' and 24b' in the mouse. *Brain Struct Funct.* 2017;222:1509–32.
83. Guldin WO, Pritzel M, Markowitsch HJ. Prefrontal cortex of the mouse defined as cortical projection area of the thalamic mediodorsal nucleus. *Brain Behav Evol.* 1981;19:93–107.
84. Markowitsch HJ, Pritzel M. Prefrontal cortex of the guinea pig (*Cavia porcellus*) defined as cortical projection area of the thalamic mediodorsal nucleus. *Brain Behav Evol.* 1981;18:80–95.
85. Vogt LJ, Vogt BA, Sikes RW. Limbic thalamus in rabbit: architecture, projections to cingulate cortex and distribution of muscarinic acetylcholine, GABAA, and opioid receptors. *J Comp Neurol.* 1992;319:205–17.
86. Krettek JE, Price JL. The cortical projections of the mediodorsal nucleus and adjacent thalamic nuclei in the rat. *J Comp Neurol.* 1977;171:157–91.
87. Eichenbaum H, Clegg RA, Feeley A. Reexamination of functional subdivisions of the rodent prefrontal cortex. *Exp Neurol.* 1983;79:434–51.
88. Kesner RP. Subregional analysis of mnemonic functions of the prefrontal cortex in the rat. *Psychobiology.* 2000;28:219–28.
89. Kolb B, Nanneman AJ, Singh RK. Double dissociation of spatial impairments and perseveration following selective prefrontal lesions in rats. *J Comp Physiol Psychol.* 1974;87:772–80.
90. Markowitsch HJ, Pritzel M. Comparative analysis of prefrontal learning functions in rats, cats, and monkeys. *Psychol Bull.* 1977;84:817–37.
91. Passingham RE, Wise SP. *The neurobiology of the prefrontal cortex: anatomy, evolution, and the origin of insight.* Oxford, UK: Oxford University Press; 2012.
92. Wise SP. Forward frontal fields: phylogeny and fundamental function. *Trends Neurosci.* 2008;31:599–608.
93. Wise SP. The evolution of the prefrontal cortex in early primates and anthropoids. In: Kaas JH, Krubitzer LA, editors. *Evolution of nervous systems, vol. 3: the*

- nervous systems of non-human primates, second edition. Amsterdam/Boston: Elsevier/Academic; 2017. p. 387–422.
94. Baleyrier C, Maudguiere F. The duality of the cingulate gyrus in monkey. Neuroanatomical study and functional hypothesis. *Brain*. 1980;103:525–54.
  95. Barbas H, Henion TH, Dermon CR. Diverse thalamic projections to the prefrontal cortex in the rhesus monkey. *J Comp Neurol*. 1991;313:65–94.
  96. Burman KJ, Bakola S, Richardson KE, Yu HH, Reser DH, Rosa MG. Cortical and thalamic projections to cytoarchitectural areas 6va and 8c of the marmoset monkey: connectionally distinct subdivisions of the lateral premotor cortex. *J Comp Neurol*. 2015;523:1222–47.
  97. Fang PC, Stepniewska I, Kaas JH. The thalamic connections of motor, premotor, and prefrontal areas of cortex in a prosimian primate (*Otolemur garnetti*). *Neuroscience*. 2006;143:987–1020.
  98. Giguere M, Goldman-Rakic PS. Mediodorsal nucleus: areal, laminar, and tangential distribution of afferents and efferents in the frontal lobe of rhesus monkeys. *J Comp Neurol*. 1988;277:195–213.
  99. Goldman-Rakic PS, Porrino LJ. The primate mediodorsal (MD) nucleus and its projection to the frontal lobe. *J Comp Neurol*. 1985;242:535–60.
  100. Kievit J, Kuypers HG. Organization of the thalamo-cortical connexions to the frontal lobe in the rhesus monkey. *Exp Brain Res*. 1977;29:299–322.
  101. Mufson EJ, Mesulam MM. Thalamic connections of the insula in the rhesus monkey and comments on the paralimbic connectivity of the medial pulvinar nucleus. *J Comp Neurol*. 1984;227:109–20.
  102. Roberts AC, Tomic DL, Parkinson CH, Roeling TA, Cutter DJ, Robbins TW, et al. Forebrain connectivity of the prefrontal cortex in the marmoset monkey (*Callithrix jacchus*): an anterograde and retrograde tract-tracing study. *J Comp Neurol*. 2007;502:86–112.
  103. Ilinsky IA, Jouandet ML, Goldman-Rakic PS. Organization of the nigrothalamo-cortical system in the rhesus monkey. *J Comp Neurol*. 1985;236:315–30.
  104. Schell GR, Strick PL. The origin of thalamic inputs to the arcuate premotor and supplementary motor areas. *J Neurosci*. 1984;4:539–60.
  105. Stepniewska I, Preuss TM, Kaas JH. Thalamic connections of the primary motor cortex (M1) of owl monkeys. *J Comp Neurol*. 1994;349:558–82.
  106. Stepniewska I, Preuss TM, Kaas JH. Thalamic connections of the dorsal and ventral premotor areas in New World owl monkeys. *Neuroscience*. 2007;147:727–45.
  107. Bachevalier J, Meunier M, Lu MX, Ungerleider LG. Thalamic and temporal cortex input to medial prefrontal cortex in rhesus monkeys. *Exp Brain Res*. 1997;115:430–44.
  108. Cappe C, Morel A, Rouiller EM. Thalamocortical and the dual pattern of corticothalamic projections of the posterior parietal cortex in macaque monkeys. *Neuroscience*. 2007;146:1371–87.
  109. Divac I, Lavail JH, Rakic P, Winston KR. Heterogeneous afferents to the inferior parietal lobule of the rhesus monkey revealed by the retrograde transport method. *Brain Res*. 1977;123:197–207.
  110. Hardy SG, Lynch JC. The spatial distribution of pulvinar neurons that project to two subregions of the inferior parietal lobule in the macaque. *Cereb Cortex*. 1992;2:217–30.
  111. Russchen FT, Amaral DG, Price JL. The afferent input to the magnocellular division of the mediodorsal thalamic nucleus in the monkey, *Macaca fascicularis*. *J Comp Neurol*. 1987;256:175–210.
  112. Schmahmann JD, Pandya DN. Anatomical investigation of projections from thalamus to posterior parietal cortex in the rhesus monkey: a WGA-HRP and fluorescent tracer study. *J Comp Neurol*. 1990;295:299–326.
  113. Stanton GB, Cruce WLR, Goldberg ME, Robinson DL. Some ipsilateral projections to areas PF and PG of the inferior parietal lobule in monkeys. *Neurosci Lett*. 1977;6:243–50.
  114. Yeterian EH, Pandya DN. Corticothalamic connections of the posterior parietal cortex in the rhesus monkey. *J Comp Neurol*. 1985;237:408–26.
  115. Baleyrier C, Maudguiere F. Anatomical evidence for medial pulvinar connections with the posterior cingulate cortex, the retrosplenial area, and the posterior parahippocampal gyrus in monkeys. *J Comp Neurol*. 1985;232:219–28.
  116. Barbas H, Mesulam MM. Cortical afferent input to the principals region of the rhesus monkey. *Neuroscience*. 1985;15:619–37.
  117. Homman-Ludiyé J, Mundinano IC, Kwan WC, Bourne JA. Extensive connectivity between the medial pulvinar and the cortex revealed in the marmoset monkey. *Cereb Cortex*. 2020;30:1797–812.
  118. Romanski LM, Giguere M, Bates JF, Goldman-Rakic PS. Topographic organization of medial pulvinar connections with the prefrontal cortex in the rhesus monkey. *J Comp Neurol*. 1997;379:313–32.
  119. Kolb B. Do all mammals have a prefrontal cortex? In: Kaas JH, Krubitzer L, editors. *Evolution of nervous systems, vol. 3: mammals*. London: Elsevier; 2007. p. 443–50.
  120. Markowitsch HJ, Pritzel M. The prefrontal cortex: projection area of the thalamic mediodorsal nucleus? *Physiol Psychol*. 2013;7:1–6.
  121. Reep R. Relationship between prefrontal and limbic cortex: a comparative anatomical review. *Brain Behav Evol*. 1984;25:5–80.
  122. Harris JA, Mihalas S, Hirokawa KE, Whitesell JD, Choi H, Bernard A, et al. Hierarchical organization of cortical and thalamic connectivity. *Nature*. 2019;575:195–202.
  123. Le Merre P, Ahrlund-Richter S, Carlen M. The mouse prefrontal cortex: unity in diversity. *Neuron*. 2021;109:1925–44.
  124. Ray JP, Price JL. The organization of the thalamocortical connections of the mediodorsal thalamic nucleus in the rat, related to the ventral forebrain-prefrontal cortex topography. *J Comp Neurol*. 1992;323:167–97.
  125. Ray JP, Price JL. The organization of projections from the mediodorsal nucleus of the thalamus to orbital and medial prefrontal cortex in macaque monkeys. *J Comp Neurol*. 1993;337:1–31.
  126. Aggleton JP, Wright NF, Rosene DL, Saunders RC. Complementary patterns of direct amygdala and hippocampal projections to the macaque prefrontal cortex. *Cereb Cortex*. 2015;25:4351–73.
  127. Barbas H, Saha S, Rempel-Clower N, Ghoshghaei T. Serial pathways from primate prefrontal cortex to autonomic areas may influence emotional expression. *BMC Neurosci*. 2003;4:25.
  128. Condé F, Maire-Lepoivre E, Audinat E, Crepel F. Afferent connections of the medial frontal cortex of the rat. II. Cortical and subcortical afferents. *J Comp Neurol*. 1995;352:567–93.
  129. Devinsky O, Morrell MJ, Vogt BA. Contributions of anterior cingulate cortex to behaviour. *Brain*. 1995;118:279–306.
  130. Freedman LJ, Insel TR, Smith Y. Subcortical projections of area 25 (subgenual cortex) of the macaque monkey. *J Comp Neurol*. 2000;421:172–88.
  131. Neafsey EJ. Prefrontal cortical control of the autonomic nervous system: anatomical and physiological observations. *Prog Brain Res*. 1991;85:147–66.
  132. Rolls ET. The cingulate cortex and limbic systems for action, emotion, and memory. *Handb Clin Neurol*. 2019;166:23–37.
  133. Swanson LW. A direct projection from Ammon's horn to prefrontal cortex in the rat. *Brain Res*. 1981;217:150–4.
  134. Terreberry RR, Neafsey EJ. The rat medial frontal cortex projects directly to autonomic regions of the brainstem. *Brain Res Bull*. 1987;19:639–49.
  135. Wang J, John Y, Barbas H. Pathways for contextual memory: the primate hippocampal pathway to anterior cingulate cortex. *Cereb Cortex*. 2021;31:1807–26.
  136. Bicks LK, Koike H, Akbarian S, Morishita H. Prefrontal cortex and social cognition in mouse and man. *Front Psychol*. 2015;6:1805.
  137. Heilbronner SR, Rodriguez-Romaguera J, Quirk GJ, Groenewegen HJ, Haber SN. Circuit-based corticostriatal homologies between rat and primate. *Biol Psychiatry*. 2016;80:509–21.
  138. Heukelum SV, Mars RB, Guthrie M, Buitelaar JK, Beckmann CF, Tiesinga PHE, et al. Where is cingulate cortex? A cross-species view. *Trends Neurosci*. 2020;43:285–99.
  139. Ledoux J. Rethinking the emotional brain. *Neuron*. 2012;73:653–76.
  140. Vogt BA, Paxinos G. Cytoarchitecture of mouse and rat cingulate cortex with human homologies. *Brain Struct Funct*. 2014;219:185–92.
  141. Divac I. The prefrontal system: a smorgasbord. *Prog Brain Res*. 1994;100:169–75.
  142. Kolb B. Functions of the frontal cortex of the rat: a comparative review. *Brain Res*. 1984;320:65–98.
  143. Berger B, Trottier S, Verney C, Gaspar P, Alvarez C. Regional and laminar distribution of the dopamine and serotonin innervation in the macaque cerebral cortex: a radioautographic study. *J Comp Neurol*. 1988;273:99–119.
  144. Foote SL, Morrison JH. Development of the noradrenergic, serotonergic, and dopaminergic innervation of neocortex. *Curr Topics Dev Biol*. 1987;21:391–423.
  145. Gaspar P, Berger B, Febvret A, Vigny A, Henry JP. Catecholamine innervation of the human cerebral cortex as revealed by comparative immunohistochemistry of tyrosine hydroxylase and dopamine-beta-hydroxylase. *J Comp Neurol*. 1989;279:249–71.
  146. Gaspar P, Stepniewska I, Kaas JH. Topography and collateralization of the dopaminergic projections to motor and lateral prefrontal cortex in owl monkeys. *J Comp Neurol*. 1992;325:1–21.
  147. Lewis DA, Foote SL, Goldstein M, Morrison JH. The dopaminergic innervation of monkey prefrontal cortex: a tyrosine hydroxylase immunohistochemical study. *Brain Res*. 1988;449:225–43.
  148. Raghanti MA, Stimpson CD, Marcinkiewicz JL, Erwin JM, Hof PR, Sherwood CC. Cortical dopaminergic innervation among humans, chimpanzees, and macaque monkeys: a comparative study. *Neuroscience*. 2008;155:203–20.
  149. Rosenberg DR, Lewis DA. Postnatal maturation of the dopaminergic innervation of monkey prefrontal and motor cortices: a tyrosine hydroxylase immunohistochemical analysis. *J Comp Neurol*. 1995;358:383–400.
  150. Williams SM, Goldman-Rakic PS. Characterization of the dopaminergic innervation of the primate frontal cortex using a dopamine-specific antibody. *Cereb Cortex*. 1993;3:199–222.

151. Williams SM, Goldman-Rakic PS. Widespread origin of the primate mesofrontal dopamine system. *Cereb Cortex*. 1998;8:321–45.
152. Berger B, Gaspar P, Vernay C. Dopaminergic innervation of the cerebral cortex: unexpeded differences between rodents and primates. *Trends Neurosci*. 1991;14:21–7.
153. Bachevalier J, Mishkin M. Visual recognition impairment follows ventromedial but not dorsolateral prefrontal lesions in monkeys. *Behavioural Brain Res*. 1986;20:249–61.
154. Meunier M, Bachevalier J, Mishkin M. Effects of orbital frontal and anterior cingulate lesions on object and spatial memory in rhesus monkeys. *Neuropsychologia*. 1997;35:999–1015.
155. Fuster JM. Unit activity in prefrontal cortex during delayed-response performance: neuronal correlates of transient memory. *J Neurophysiol*. 1973;36:61–78.
156. Niki H, Watanabe M. Cingulate unit activity and delayed response. *Brain Res*. 1976;110:381–6.
157. Inoue M, Mikami A, Ando I, Tsukada H. Functional brain mapping of the macaque related to spatial working memory as revealed by pet. *Cereb Cortex*. 2004;14:106–19.
158. Liu J, Xia M, Dai Z, Wang X, Liao X, Bi Y, et al. Intrinsic brain hub connectivity underlies individual differences in spatial working memory. *Cereb Cortex*. 2017;27:5496–508.
159. Balsters JH, Zerbi V, Sallet J, Wenderoth N, Mars RB. Primate homologs of mouse cortico-striatal circuits. *Elife*. 2020;9. <https://doi.org/10.7554/eLife.53680>.
160. Hall RD, Lindholm EP. Organization of motor and somatosensory neocortex in albino rat. *Brain Res*. 1974;66:23–38.
161. Neafsey EJ, Bold EL, Haas G, Hurley-Gius KM, Quirk G, Sievert CF, et al. The organization of the rat motor cortex: a microstimulation mapping study. *Brain Res*. 1986;396:77–96.
162. Reep RL, Goodwin GS, Corwin JV. Topographic organization in the corticocortical connections of medial agranular cortex in rats. *J Comp Neurol*. 1990;294:262–80.
163. Brecht M, Krauss A, Muhammad S, Sinai-Esfahani L, Bellanca S, Margrie TW. Organization of rat vibrissa motor cortex and adjacent areas according to cytoarchitectonics, microstimulation, and intracellular stimulation of identified cells. *J Comp Neurol*. 2004;479:360–73.
164. Brecht M, Schneider M, Sakmann B, Margrie TW. Whisker movements evoked by stimulation of single pyramidal cells in rat motor cortex. *Nature*. 2004;427:704–10.
165. Colechio EM, Alloway KD. Differential topography of the bilateral cortical projections to the whisker and forepaw regions in rat motor cortex. *Brain Struct Funct*. 2009;213:423–39.
166. Halley AC, Baldwin MKL, Cooke DF, Englund M, Krubitzer L. Distributed motor control of limb movements in rat motor and somatosensory cortex: the sensorimotor amalgam revisited. *Cereb Cortex*. 2020;30:6296–312.
167. Smith JB, Alloway KD. Rat whisker motor cortex is subdivided into sensory-input and motor-output areas. *Front Neural Circuits*. 2013;7:4.
168. Tandon S, Kambi N, Jain N. Overlapping representations of the neck and whiskers in the rat motor cortex revealed by mapping at different anaesthetic depths. *Eur J Neurosci*. 2008;27:228–37.
169. Moschovakis AK, Gregoriou GG, Ugolini G, Doldan M, Graf W, Guldin W, et al. Oculomotor areas of the primate frontal lobes: a transneuronal transfer of rabies virus and [<sup>14</sup>C]-2-deoxyglucose functional imaging study. *J Neurosci*. 2004;24:5726–40.
170. Fujii N, Mushiaki H, Tanji J. Intracortical microstimulation of bilateral frontal eye field. *J Neurophysiol*. 1998;79:2240–4.
171. Fujii N, Mushiaki H, Tanji J. Distribution of eye- and arm-movement-related neuronal activity in the SEF and in the SMA and pre-SMA of monkeys. *J Neurophysiol*. 2002;87:2158–66.
172. Mitz AR, Wise SP. The somatotopic organization of the supplementary motor area: intracortical microstimulation mapping. *J Neurosci*. 1987;7:1010–21.
173. Preuss TM, Stepniewska I, Kaas JH. Movement representation in the dorsal and ventral premotor areas of owl monkeys: a microstimulation study. *J Comp Neurol*. 1996;371:649–76.
174. Uylings HB, Groenewegen HJ, Kolb B. Do rats have a prefrontal cortex? *Behav Brain Res*. 2003;146:3–17.
175. Cavada C, Goldman-Rakic PS. Posterior parietal cortex in rhesus monkey: II. Evidence for segregated corticocortical networks linking sensory and limbic areas with the frontal lobe. *J Comp Neurol*. 1989;287:422–45.
176. Cavada C, Goldman-Rakic PS. Posterior parietal cortex in rhesus monkey: I. Parcellation of areas based on distinctive limbic and sensory corticocortical connections. *J Comp Neurol*. 1989;287:393–421.
177. Goldman-Rakic PS. Topography of cognition: parallel distributed networks in primate association cortex. *Annu Rev Neurosci*. 1988;11:137–56.
178. Jones EG, Powell TP. An anatomical study of converging sensory pathways within the cerebral cortex of the monkey. *Brain*. 1970;93:793–820.
179. Pandya DN, Seltzer B. Association areas of the cerebral cortex. *Trends Neurosci*. 1982;5:386–90.
180. Preuss TM, Goldman-Rakic PS. Ipsilateral cortical connections of granular frontal cortex in the strepsirrhine primate *Galago*, with comparative comments on anthropoid primates. *J Comp Neurol*. 1991;310:507–49.
181. Stepniewska I, Cerkevich CM, Kaas JH. Cortical connections of the caudal portion of posterior parietal cortex in prosimian galagos. *Cereb Cortex*. 2016;26:2753–77.
182. Yeo BT, Krienen FM, Sepulcre J, Sabuncu MR, Lashkari D, Hollinshead M, et al. The organization of the human cerebral cortex estimated by intrinsic functional connectivity. *J Neurophysiol*. 2011;106:1125–65.
183. Asanuma C, Andersen RA, Cowan WM. The thalamic relations of the caudal inferior parietal lobule and the lateral prefrontal cortex in monkeys: divergent cortical projections from cell clusters in the medial pulvinar nucleus. *J Comp Neurol*. 1985;241:357–81.
184. Baleyrier C, Manguiere F. Network organization of the connectivity between parietal area 7, posterior cingulate cortex and medial pulvinar nucleus: a double fluorescent tracer study in monkey. *Exp Brain Res*. 1987;66:385–93.
185. Baleyrier C, Morel A. Segregated thalamocortical pathways to inferior parietal and inferotemporal cortex in macaque monkey. *Vis Neurosci*. 1992;8:391–405.
186. Burton H, Jones EG. The posterior thalamic region and its cortical projection in New World and Old World monkeys. *J Comp Neurol*. 1976;168:249–301.
187. Trojanowski JQ, Jacobson S. Areal and laminar distribution of some pulvinar cortical efferents in rhesus monkey. *J Comp Neurol*. 1976;169:371–92.
188. Yeterian EH, Pandya DN. Thalamic connections of the cortex of the superior temporal sulcus in the rhesus monkey. *J Comp Neurol*. 1989;282:80–97.
189. Preuss TM. Evolutionary specializations of primate brain systems. In: Ravosa MJ, Dagasto M, editors. *Primate origins: adaptations and evolution*. Boston: Springer; 2007. p. 625–75.
190. Preuss TM. Primate brain evolution in phylogenetic context. In: Kaas JH, Preuss TM, editors. *Evolution of nervous systems, vol. 4: primates*. London: Elsevier; 2007. p. 1–34.
191. Baldwin MKL, Krubitzer L. Architectonic characteristics of the visual thalamus and superior colliculus in titi monkeys. *J Comp Neurol*. 2018;526:1760–76.
192. Wallis JD, Miller EK. From rule to response: neuronal processes in the premotor and prefrontal cortex. *J Neurophysiol*. 2003;90:1790–806.
193. Genovesio A, Brasted PJ, Mitz AR, Wise SP. Prefrontal cortex activity related to abstract response strategies. *Neuron*. 2005;47:307–20.
194. Genovesio A, Tsujimoto S, Wise SP. Encoding problem-solving strategies in prefrontal cortex: activity during strategic errors. *Eur J Neurosci*. 2008;27:984–90.
195. Bussey TJ, Wise SP, Murray EA. The role of ventral and orbital prefrontal cortex in conditional visuomotor learning and strategy use in rhesus monkeys (*Macaca mulatta*). *Behav Neurosci*. 2001;115:971–82.
196. Bussey TJ, Muir JL, Everitt BJ, Robbins TW. Triple dissociation of anterior cingulate, posterior cingulate, and medial frontal cortices on visual discrimination tasks using a touchscreen testing procedure for the rat. *Behav Neurosci*. 1997;111:920–36.
197. Bussey TJ, Muir JL, Everitt BJ, Robbins TW. Dissociable effects of anterior and posterior cingulate cortex lesions on the acquisition of a conditional visual discrimination: facilitation of early learning vs. Impairment of late learning. *Behav Brain Res*. 1996;82:45–56.
198. Wong P, Kaas JH. Architectonic subdivisions of neocortex in the tree shrew (*Tupaia belangeri*). *Anat Rec*. 2009;292:994–1027.
199. Remple MS, Reed JL, Stepniewska I, Lyon DC, Kaas JH. The organization of frontoparietal cortex in the tree shrew (*Tupaia belangeri*): II. Connectional evidence for a frontal-posterior parietal network. *J Comp Neurol*. 2007;501:121–49.
200. Deacon RM, Penny C, Rawlins JN. Effects of medial prefrontal cortex cytotoxic lesions in mice. *Behav Brain Res*. 2003;139:139–55.
201. Baldwin MK, Kaas JH. Cortical projections to the superior colliculus in prosimian galagos (*Otolemur garnetti*). *J Comp Neurol*. 2012;520:2002–20.
202. Fang PC, Stepniewska I, Kaas JH. Ipsilateral cortical connections of motor, pre-motor, frontal eye, and posterior parietal fields in a prosimian primate, *Otolemur garnetti*. *J Comp Neurol*. 2005;490:305–33.
203. Preuss TM, Goldman-Rakic PS. Myelo- and cytoarchitecture of the granular frontal cortex and surrounding regions in the strepsirrhine primate *Galago* and the anthropoid primate *Macaca*. *J Comp Neurol*. 1991;310:429–74.
204. Stepniewska I, Pouget P, Kaas JH. Frontal eye field in prosimian galagos: intracortical microstimulation and tracing studies. *J Comp Neurol*. 2018;526:626–52.
205. Wong P, Kaas JH. Architectonic subdivisions of neocortex in the galago (*Otolemur garnetti*). *Anat Rec*. 2010;293:1033–69.
206. Wu CW, Bichot NP, Kaas JH. Converging evidence from microstimulation, architecture, and connections for multiple motor areas in the frontal and cingulate cortex of prosimian primates. *J Comp Neurol*. 2000;423:140–77.

207. Blum B, Kulikowski JJ, Carden D, Harwood D. Eye movements induced by electrical stimulation of the frontal eye fields of marmosets and squirrel monkeys. *Brain Behav Evol.* 1982;21:34–41.
208. Burman KJ, Palmer SM, Gamberini M, Rosa MG. Cytoarchitectonic subdivisions of the dorsolateral frontal cortex of the marmoset monkey (*Callithrix jacchus*), and their projections to dorsal visual areas. *J Comp Neurol.* 2006;495:149–72.
209. Cusick CG. Anatomical organization of the superior colliculus in monkeys: corticotectal pathways for visual and visuomotor functions. *Prog Brain Res.* 1988;75:15.
210. Collins CE, Lyon DC, Kaas JH. Distribution across cortical areas of neurons projecting to the superior colliculus in New World monkeys. *Anat Rec A Discov Mol Cell Evol Biol.* 2005;285:619–27.
211. Gould HJ 3rd, Cusick CG, Pons TP, Kaas JH. The relationship of corpus callosum connections to electrical stimulation maps of motor, supplementary motor, and the frontal eye fields in owl monkeys. *J Comp Neurol.* 1986;247:297–325.
212. Huerta MF, Krubitzer LA, Kaas JH. Frontal eye field as defined by intracortical microstimulation in squirrel monkeys, owl monkeys, and macaque monkeys: I. Subcortical connections. *J Comp Neurol.* 1986;253:415–39.
213. Huerta MF, Krubitzer LA, Kaas JH. Frontal eye field as defined by intracortical microstimulation in squirrel monkeys, owl monkeys, and macaque monkeys. II. Cortical connections. *J Comp Neurol.* 1987;265:332–61.
214. Leichnetz GR, Gonzalo-Ruiz A. Prearcuate cortex in the cebus monkey has cortical and subcortical connections like the macaque frontal eye field and projects to fastigial-recipient oculomotor-related brainstem nuclei. *Brain Res Bull.* 1996;41:1–29.
215. Tian JR, Lynch JC. Subcortical input to the smooth and saccadic eye movement subregions of the frontal eye field in cebus monkey. *J Neurosci.* 1997;17:9233–47.
216. Tian JR, Lynch JC. Corticocortical input to the smooth and saccadic eye movement subregions of the frontal eye field in cebus monkeys. *J Neurophysiol.* 1996;76:2754–71.
217. Tian JR, Lynch JC. Functionally defined smooth and saccadic eye movement subregions in the frontal eye field of cebus monkeys. *J Neurophysiol.* 1996;76:2740–53.
218. Bruce CJ, Goldberg ME. Primate frontal eye fields. I. Single neurons discharging before saccades. *J Neurophysiol.* 1985;53:603–35.
219. Bruce CJ, Goldberg ME, Bushnell MC, Stanton GB. Primate frontal eye fields. II. Physiological and anatomical correlates of electrically evoked eye movements. *J Neurophysiol.* 1985;54:714–34.
220. Cerkevich CM, Lyon DC, Balaram P, Kaas JH. Distribution of cortical neurons projecting to the superior colliculus in macaque monkeys. *Eye Brain.* 2014;2014:121–37.
221. Fries W. Cortical projections to the superior colliculus in the macaque monkey: a retrograde study using horseradish peroxidase. *J Comp Neurol.* 1984;230:55–76.
222. Robinson DA, Fuchs AF. Eye movements evoked by stimulation of frontal eye fields. *J Neurophysiol.* 1969;32:637–48.
223. Stanton GB, Bruce CJ, Goldberg ME. Topography of projections to posterior cortical areas from the macaque frontal eye fields. *J Comp Neurol.* 1995;353:291–305.
224. Stanton GB, Goldberg ME, Bruce CJ. Frontal eye field efferents in the macaque monkey: II. Topography of terminal fields in midbrain and pons. *J Comp Neurol.* 1988;271:493–506.
225. Stanton GB, Goldberg ME, Bruce CJ. Frontal eye field efferents in the macaque monkey: I. Subcortical pathways and topography of striatal and thalamic terminal fields. *J Comp Neurol.* 1988;271:473–92.
226. Schall JD, Zinke W, Cosman JD, Schall MS, Paré M, Pouget P. On the evolution of the frontal eye field: comparisons of monkeys, apes, and humans. In: Kaas JH, Preuss TM, editors. *Evolution of nervous systems, vol. 4: the evolution of the human brain: apes and other ancestors* (second edition). Amsterdam/Boston: Elsevier/Academic Press. p. 249–75.
227. Baldwin MK, Cooke DF, Krubitzer L. Intracortical microstimulation maps of motor, somatosensory, and posterior parietal cortex in tree shrews (*Tupaia belangeri*) reveal complex movement representations. *Cereb Cortex.* 2017;27:1439–56.
228. Remple MS, Reed JL, Stepniewska I, Kaas JH. Organization of frontoparietal cortex in the tree shrew (*Tupaia belangeri*). I. Architecture, microelectrode maps, and corticospinal connections. *J Comp Neurol.* 2006;497:133–54.
229. Lyon DC, Jain N, Kaas JH. Cortical connections of striate and extrastriate visual areas in tree shrews. *J Comp Neurol.* 1998;401:109–28.
230. Stepniewska I, Cerkevich CM, Fang PC, Kaas JH. Organization of the posterior parietal cortex in galagos: II. Ipsilateral cortical connections of physiologically identified zones within anterior sensorimotor region. *J Comp Neurol.* 2009;517:783–807.
231. Baylis LL, Rolls ET, Baylis GC. Afferent connections of the caudolateral orbito-frontal cortex taste area of the primate. *Neuroscience.* 1995;64:801–12.
232. Burman KJ, Reser DH, Yu HH, Rosa MG. Cortical input to the frontal pole of the marmoset monkey. *Cereb Cortex.* 2011;21:1712–37.
233. Burman KJ, Rosa MG. Architectural subdivisions of medial and orbital frontal cortices in the marmoset monkey (*Callithrix jacchus*). *J Comp Neurol.* 2009;514:11–29.
234. Carmichael ST, Clugnet MC, Price JL. Central olfactory connections in the macaque monkey. *J Comp Neurol.* 1994;346:403–34.
235. Carmichael ST, Price JL. Sensory and premotor connections of the orbital and medial prefrontal cortex of macaque monkeys. *J Comp Neurol.* 1995;363:642–64.
236. Cruz-Rizzolo RJ, De Lima MA, Ervolino E, De Oliveira JA, Casatti CA. Cyto-, myelo- and chemoarchitecture of the prefrontal cortex of the cebus monkey. *BMC Neurosci.* 2011;12:6.
237. Mesulam MM, Mufson EJ. Insula of the Old World monkey. III: efferent cortical output and comments on function. *J Comp Neurol.* 1982;212:38–52.
238. Rosa MGP, Soares JGM, Chaplin TA, Majka P, Bakola S, Phillips KA, et al. Cortical afferents of area 10 in cebus monkeys: implications for the evolution of the frontal pole. *Cereb Cortex.* 2019;29:1473–95.
239. Amaral DG, Price JL. Amygdalo-cortical projections in the monkey (*Macaca fascicularis*). *J Comp Neurol.* 1984;230:465–96.
240. Barbas H, De Olmos J. Projections from the amygdala to basoventral and mediadorsal prefrontal regions in the rhesus monkey. *J Comp Neurol.* 1990;300:549–71.
241. Barbas H. Connections underlying the synthesis of cognition, memory, and emotion in primate prefrontal cortices. *Brain Res Bull.* 2000;52:319–30.
242. Timbie C, Barbas H. Specialized pathways from the primate amygdala to posterior orbitofrontal cortex. *J Neurosci.* 2014;34:8106–18.
243. Carmichael ST, Price JL. Limbic connections of the orbital and medial prefrontal cortex in macaque monkeys. *J Comp Neurol.* 1995;363:615–41.
244. Porrino LJ, Crane AM, Goldman-Rakic PS. Direct and indirect pathways from the amygdala to the frontal lobe in rhesus monkeys. *J Comp Neurol.* 1981;198:121–36.
245. Ferry AT, Öngür D, An X, Price JL. Prefrontal cortical projections to the striatum in macaque monkeys: evidence for an organization related to prefrontal networks. *J Comp Neurol.* 2000;425:447–70.
246. Haber SN, Kim KS, Maily P, Calzavara R. Reward-related cortical inputs define a large striatal region in primates that interface with associative cortical connections, providing a substrate for incentive-based learning. *J Neurosci.* 2006;26:8368–76.
247. Haber SN, Kunishio K, Mizobuchi M, Lynd-Balta E. The orbital and medial prefrontal circuit through the primate basal ganglia. *J Neurosci.* 1995;15:4851–67.
248. Saleem KS, Kondo H, Price JL. Complementary circuits connecting the orbital and medial prefrontal networks with the temporal, insular, and opercular cortex in the macaque monkey. *J Comp Neurol.* 2008;506:659–93.
249. Glasser MF, Goyal MS, Preuss TM, Raichle ME, Van Essen DC. Trends and properties of human cerebral cortex: correlations with cortical myelin content. *Neuroimage.* 2014;93:165–75.
250. Wong P, Collins CE, Kaas JH. Overview of sensory systems of *Tarsius*. *Int J Primatol.* 2010;31:1002–31.
251. Woolard HH. The cortical lamination of *Tarsius*. *J Anat.* 1925;60:86–105.
252. Armstrong E. Relative brain size in monkeys and prosimians. *Am J Phys Anthropol.* 1985;66:263–73.
253. Boddy AM, McGowen MR, Sherwood CC, Grossman LI, Goodman M, Wildman DE. Comparative analysis of encephalization in mammals reveals relaxed constraints on anthropoid primate and cetacean brain scaling. *J Evol Biol.* 2012;25:981–94.
254. Jerison HJ. *Evolution of the brain and intelligence.* New York: Academic Press; 1973.
255. Jerison HJ. Brain, body and encephalization in early primates. *J Hum Evol.* 1979;8:615–35.
256. Bush EC, Simons EL, Allman JM. High-resolution computed tomography study of the cranium of a fossil anthropoid primate, *Parapithecus grangeri*: new insights into the evolutionary history of primate sensory systems. *Anat Rec A Discov Mol Cell Evol Biol* 2004;281:1083–7.
257. Gonzales LA, Benefit BR, Mccrossin ML, Spoor F. Cerebral complexity preceded enlarged brain size and reduced olfactory bulbs in Old World monkeys. *Nat Commun.* 2015;6:7580.
258. Gurche J. Early primate brain evolution. In: Armstrong E, Falk D, editors. *Primate brain evolution.* New York: Plenum Press; 1982. p. 227–46.
259. Ni X, Flynn JJ, Wyss AR, Zhang C. Cranial endocast of a stem platyrrhine primate and ancestral brain conditions in anthropoids. *Sci Adv.* 2019;5:eaav7913.
260. Radinsky L. Primate brain evolution. *Am Sci.* 1975;63:656–63.
261. Ford SM. Callitrichids as phyletic dwarfs, and the place of the callitrichidae in platyrrhini. *Primates.* 1980;21:31–43.
262. Martin RD. Goeldi and the dwarfs: the evolutionary biology of the small New World monkeys. *J Hum Evol.* 1992;22:367–93.

263. Montgomery SH, Mundy NI. Parallel episodes of phyletic dwarfism in callitrichid and cheirogaleid primates. *J Evol Biol.* 2013;26:810–9.
264. Isler K, Christoph Kirk E, Miller JM, Albrecht GA, Gelvin BR, Martin RD. Endocranial volumes of primate species: scaling analyses using a comprehensive and reliable data set. *J Hum Evol.* 2008;55:967–78.
265. Deaner RO, Isler K, Burkart J, Van Schaik C. Overall brain size, and not encephalization quotient, best predicts cognitive ability across non-human primates. *Brain Behav Evol.* 2007;70:115–24.
266. Gibson K, Rumbaugh D, Beran M. Bigger is better: primate brain size in relationship to cognition. In: Falk D, Gibson K, editors. *Evolutionary anatomy of primate cerebral cortex.* Cambridge, UK: Cambridge University Press; 2001. p. 79–97.
267. Ponce De Leon MS, Bienvenu T, Marom A, Engel S, Tafforeau P, Alatorre Warren JL, et al. The primitive brain of early *Homo*. *Science* 2021;372:165–71.
268. Preuss TM. The human brain. In: Tibayrenc M, Ayala FJ, editors. *On human nature.* Amsterdam/Boston: Elsevier/Academic Press; 2017. p. 125–49.
269. Kaas JH, Preuss TM, editors. *Evolution of nervous systems, volume 4: the evolution of the human brain: apes and other ancestors.* 2nd ed. Amsterdam/Boston: Elsevier/Academic Press; 2017.
270. Rilling JK. Comparative primate neuroimaging: Insights into human brain evolution. *Trends Cogn Sci.* 2014;18:46–55.
271. Sherwood CC, Bauernfeind AL, Bianchi S, Raghanti MA, Hof PR. Human brain evolution writ large and small. In: Hofman MA, Falk D, editors. *Evolution of the primate brain.* Amsterdam: Elsevier; 2012. p. 237–54.
272. Brodmann K. Neue ergebnisse über die vergleichende histologische lokalisation der grosshirnrinde mit besonderer berücksichtigung des stirnhirns. *Anat Anz.* 1912;41:157–216.
273. Barton RA, Venditti C. Human frontal lobes are not relatively large. *Proc Natl Acad Sci USA.* 2013;110:9001–6.
274. Barton RA, Montgomery SH. Proportional versus relative size as metrics in human brain evolution. *Proc Natl Acad Sci USA.* 2019;116:3–4.
275. Semendeferi K, Damasio H, Frank R, Van Hoesen GW. The evolution of the frontal lobes: a volumetric analysis based on three-dimensional reconstructions of magnetic resonance scans of human and ape brains. *J Hum Evol.* 1997;32:375–88.
276. Semendeferi K, Lu A, Schenker N, Damasio H. Humans and great apes share a large frontal cortex. *Nat Neurosci.* 2002;5:272–6.
277. Donahue CJ, Glasser MF, Preuss TM, Rilling JK, Van Essen DC. Quantitative assessment of prefrontal cortex in humans relative to nonhuman primates. *Proc Natl Acad Sci USA.* 2018;115:E5183–92.
278. Donahue CJ, Glasser MF, Preuss TM, Rilling JK, Van Essen DC. Reply to barton and montgomery: a case for preferential prefrontal cortical expansion. *Proc Natl Acad Sci USA.* 2019;116:5–6.
279. Passingham RE, Smaers JB. Is the prefrontal cortex especially enlarged in the human brain? Allometric relations and remapping factors. *Brain Behav Evol.* 2014;84:156–66.
280. Passingham RE, Smaers JB, Sherwood CC. Evolutionary specializations of the human prefrontal cortex. In: Kaas JH, Preuss TM, editors. *Evolution of nervous systems, vol. 4: the evolution of the human brain: apes and other ancestors, second edition.* Amsterdam/Boston: Elsevier/Academic Press. p. 207–26.
281. Smaers JB, Steele J, Case CR, Cowper A, Amunts K, Zilles K. Primate prefrontal cortex evolution: human brains are the extreme of a lateralized ape trend. *Brain Behav Evol.* 2011;77:67–78.
282. Smaers JB, Gomez-Robles A, Parks AN, Sherwood CC. Exceptional evolutionary expansion of prefrontal cortex in great apes and humans. *Curr Biol.* 2017;27:714–20.
283. Wei Y, De Lange SC, Scholtens LH, Watanabe K, Ardesch DJ, Jansen PR, et al. Genetic mapping and evolutionary analysis of human-expanded cognitive networks. *Nat Commun.* 2019;10:4839.
284. Glasser MF, Coalson TS, Robinson EC, Hacker CD, Harwell J, Yacoub E, et al. A multi-modal parcellation of human cerebral cortex. *Nature.* 2016;536:171–8.
285. Kaas JH. The organization of neocortex in mammals: implications for theories of brain function. *Annu Rev Psychol.* 1987;38:129–51.
286. Petrides M, Pandya DN. Comparative architectonic analysis of the human and the macaque frontal cortex. In: Boofer F, Grafman J, editors. *Handbook of neuropsychology.* Amsterdam: Elsevier; 1994. p. 17–58.
287. Rajkowska G, Goldman-Rakic PS. Cytoarchitectonic definition of prefrontal areas in the normal human cortex: I. Remapping of areas 9 and 46 using quantitative criteria. *Cereb Cortex.* 1995;5:307–22.
288. Neubert FX, Mars RB, Thomas AG, Sallet J, Rushworth MF. Comparison of human ventral frontal cortex areas for cognitive control and language with areas in monkey frontal cortex. *Neuron.* 2014;81:700–13.
289. Sallet J, Mars RB, Noonan MP, Neubert FX, Jbabdi S, O'reilly JX, et al. The organization of dorsal frontal cortex in humans and macaques. *J Neurosci.* 2013;33:12255–74.
290. Hartogsveld B, Bramson B, Vijayakumar S, Van Campen AD, Marques JP, Roelofs K, et al. Lateral frontal pole and relational processing: activation patterns and connectivity profile. *Behav Brain Res.* 2018;355:2–11.
291. Wendelken C, O'hare ED, Whitaker KJ, Ferrer E, Bunge SA. Increased functional selectivity over development in rostralateral prefrontal cortex. *J Neurosci.* 2011;31:17260–8.
292. Preuss TM, Coleman GQ. Human-specific organization of primary visual cortex: alternating compartments of dense cat-301 and calbindin immunoreactivity in layer 4a. *Cereb Cortex.* 2002;12:671–91.
293. Raghanti MA, Stimpson CD, Marcinkiewicz JL, Erwin JM, Hof PR, Sherwood CC. Cholinergic innervation of the frontal cortex: differences among humans, chimpanzees, and macaque monkeys. *J Comp Neurol.* 2008;506:409–24.
294. Raghanti MA, Stimpson CD, Marcinkiewicz JL, Erwin JM, Hof PR, Sherwood CC. Differences in cortical serotonergic innervation among humans, chimpanzees, and macaque monkeys: a comparative study. *Cereb Cortex.* 2008;18:584–97.
295. Allman JM, Tetreault NA, Hakeem AY, Manaye KF, Semendeferi K, Erwin JM, et al. The von Economo neurons in the fronto-insular and anterior cingulate cortex. *Ann N Y Acad Sci.* 2011;1225:59–71.
296. Bianchi S, Stimpson CD, Bauernfeind AL, Schapiro SJ, Baze WB, McArthur MJ, et al. Dendritic morphology of pyramidal neurons in the chimpanzee neocortex: regional specializations and comparison to humans. *Cereb Cortex.* 2013;23:2429–36.
297. Schenker NM, Buxhoeveden DP, Blackmon WL, Amunts K, Zilles K, Semendeferi K. A comparative quantitative analysis of cytoarchitecture and minicolumnar organization in broca's area in humans and great apes. *J Comp Neurol.* 2008;510:117–28.
298. Semendeferi K, Teffer K, Buxhoeveden DP, Park MS, Bludau S, Amunts K, et al. Spatial organization of neurons in the frontal pole sets humans apart from great apes. *Cereb Cortex.* 2011;21:1485–97.
299. Xiang L, Crow TJ, Hopkins WD, Roberts N. Comparison of surface area and cortical thickness asymmetry in the human and chimpanzee brain. *Cereb Cortex.* 2020. Online ahead of print.
300. Bryant KL, Li L, Eichert N, Mars RB. A comprehensive atlas of white matter tracts in the chimpanzee. *PLoS Biol.* 2020;18:e3000971.
301. Eichert N, Robinson EC, Bryant KL, Jbabdi S, Jenkinson M, Li L, et al. Cross-species cortical alignment identifies different types of anatomical reorganization in the primate temporal lobe. *Elife.* 2020;9:e53232.
302. Hecht EE, Gutman DA, Bradley BA, Preuss TM, Stout D. Virtual dissection and comparative connectivity of the superior longitudinal fasciculus in chimpanzees and humans. *Neuroimage.* 2015;108:124–37.
303. Hecht EE, Gutman DA, Preuss TM, Sanchez MM, Parr LA, Rilling JK. Process versus product in social learning: comparative diffusion tensor imaging of neural systems for action execution-observation matching in macaques, chimpanzees, and humans. *Cereb Cortex.* 2013;23:1014–24.
304. Rilling JK, Glasser MF, Preuss TM, Ma X, Zhao T, Hu X, et al. The evolution of the arcuate fasciculus revealed with comparative DTI. *Nat Neurosci.* 2008;11:426–8.
305. Rilling JK, Glasser MF, Jbabdi S, Andersson J, Preuss TM. Continuity, divergence, and the evolution of brain language pathways. *Front Evol Neurosci.* 2011;3:1–6.
306. Enard W, Khaitovich P, Klose J, Zollner S, Heissig F, Giavalisco P, et al. Intra- and interspecific variation in primate gene expression patterns. *Science.* 2002;296:340–3.
307. Khaitovich P, Muetzel B, She X, Lachmann M, Hellmann I, Dietzsch J, et al. Regional patterns of gene expression in human and chimpanzee brains. *Genome Res.* 2004;14:1462–73.
308. Somel M, Franz H, Yan Z, Lorenc A, Guo S, Giger T, et al. Transcriptional neoteny in the human brain. *Proc Natl Acad Sci USA.* 2009;106:5743–8.
309. Cáceres M, Lachuer J, Zapala MA, Redmond JC, Kudo L, Geschwind DH, et al. Elevated gene expression levels distinguish human from non-human primate brains. *Proc Natl Acad Sci USA.* 2003;100:13030–5.
310. Cáceres M, Suwyn C, Maddox M, Thomas JW, Preuss TM. Increased cortical expression of two synaptogenic thrombospondins in human brain evolution. *Cereb Cortex.* 2007;17:2312–21.
311. Berto S, Mendizabal I, Usui N, Toriumi K, Chatterjee P, Douglas C, et al. Accelerated evolution of oligodendrocytes in the human brain. *Proc Natl Acad Sci USA.* 2019;116:24334–42.
312. Konopka G, Friedrich T, Davis-Turak J, Winden K, Oldham MC, Gao F, et al. Human-specific transcriptional networks in the brain. *Neuron.* 2012;75:601–17.
313. Swanson EM, Holekamp KE, Lundrigan BL, Arsznov BM, Sakai ST. Multiple determinants of whole and regional brain volume among terrestrial carnivores. *PLoS ONE.* 2012;7:e38447.
314. Cavada C, Reinoso-Suarez F. Topographical organization of the cortical afferent connections of the prefrontal cortex in the cat. *J Comp Neurol.* 1985;242:293–324.
315. Markow-Rajkowska G, Kosmal A. Organization of cortical afferents to the frontal association cortex in dogs. *Acta Neurobiol Exp.* 1987;47:137–61.



316. Murray EA, Moylan EJ, Saleem KS, Basile BM, Turchi J. Specialized areas for value updating and goal selection in the primate orbitofrontal cortex. *Elife*. 2015;4:e11695.
317. Duque A, McCormick DA. Circuit-based localization of ferret prefrontal cortex. *Cereb Cortex*. 2010;20:1020–36.
318. Rajkowska G, Kosmal A. Intrinsic connections and cytoarchitectonic data of the frontal association cortex in the dog. *Acta Neurobiol Exp*. 1988;48:169–92.
319. Salazar I, Ruiz Pesini P, Fernandez Troconiz P, Gonzalez Soriano J, Fernandez Alvarez P. The neocortex of the dog. 1. A classical cytoarchitectonic map. *Anat Histol Embryol*. 1988;17:169–87.
320. Mackey S, Petrides M. Architecture and morphology of the human ventromedial prefrontal cortex. *Eur J Neurosci*. 2014;40:2777–96.
321. Schleicher A, Amunts K, Geyer S, Morosan P, Zilles K. Observer-independent method for microstructural parcellation of cerebral cortex: a quantitative approach to cytoarchitectonics. *Neuroimage*. 1999;9:165–77.
322. Semendeferi K, Armstrong E, Schleicher A, Zilles K, Van, Hoesen GW. Prefrontal cortex in humans and apes: a comparative study of area 10. *Am J Phys Anthropol*. 2001;114:224–41.
323. Apps MA, Rushworth MF, Chang SW. The anterior cingulate gyrus and social cognition: tracking the motivation of others. *Neuron*. 2016;90:692–707.
324. Kolb B, Pellis S, Robinson TE. Plasticity and functions of the orbital frontal cortex. *Brain Cogn*. 2004;55:104–15.
325. Rudebeck PH, Rich EL, Mayberg HS. From bed to bench side: reverse translation to optimize neuromodulation for mood disorders. *Proc Natl Acad Sci USA*. 2019;116:26288–96.
326. Phillips KA, Bales KL, Capitanio JP, Conley A, Czoty PW, T Hart BA, et al. Why primate models matter. *Am J Primatol*. 2014;76:801–27.
327. Murphy WJ, Eizirik E, O'Brien SJ, Madsen O, Scally M, Douady CJ, et al. Resolution of the early placental mammal radiation using bayesian phylogenetics. *Science*. 2001;294:2348–51.
328. Carmichael ST, Price JL. Connectional networks within the orbital and medial prefrontal cortex of macaque monkeys. *J Comp Neurol*. 1996;371:179–207.
329. Palomero-Gallagher N, Zilles K. Isocortex. In: Paxinos G, editor. *The rat nervous system*. San Diego, CA: Elsevier Academic Press; 2004. p. 729–57.
330. Elston GN, Benavides-Piccione R, Elston A, Zietsch B, Defelipe J, Manger P, et al. Specializations of the granular prefrontal cortex of primates: implications for cognitive processing. *Anat Rec A Discov Mol Cell Evol Biol*. 2006;288:26–35.

## ACKNOWLEDGEMENTS

The authors thank the editors for inviting this contribution.

## AUTHOR CONTRIBUTIONS

Both authors wrote the manuscript.

## COMPETING INTERESTS

The authors declare no competing interests.

## ADDITIONAL INFORMATION

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