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Enduring glucocorticoid-evoked exacerbation of synaptic plasticity disruption in male rats modelling early Alzheimer's disease amyloidosis

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Synaptic dysfunction is a likely proximate cause of subtle cognitive impairment in early Alzheimer's disease. Soluble oligomers are the most synaptotoxic forms of amyloid ß-protein (Aß) and mediate synaptic plasticity disruption in Alzheimer's disease amyloidosis. Because the presence and extent of cortisol excess in prodromal Alzheimer's disease predicts the onset of cognitive symptoms we hypothesised that corticosteroids would exacerbate the inhibition of hippocampal synaptic long-term potentiation in a rat model of Alzheimer's disease amyloidosis. In a longitudinal experimental design using freely behaving pre-plaque McGill-R-Thy1-APP male rats, three injections of corticosterone or the glucocorticoid methylprednisolone profoundly disrupted long-term potentiation induced by strong conditioning stimulation for at least 2 months. The same treatments had a transient or no detectible detrimental effect on synaptic plasticity in wild-type littermates. Moreover, corticosterone-mediated cognitive dysfunction, as assessed in a novel object recognition test, was more persistent in the transgenic animals. Evidence for the involvement of pro-inflammatory mechanisms was provided by the ability of the selective the NOD-leucine rich repeat and pyrin containing protein 3 (NLRP3) inflammasome inhibitor Mcc950 to reverse the synaptic plasticity deficit in corticosterone-treated transgenic animals. The marked prolongation of the synaptic plasticity disrupting effects of brief corticosteroid excess substantiates a causal role for hypothalamic-pituitary-adrenal axis dysregulation in early Alzheimer's disease.

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INTRODUCTION

Because Alzheimer's disease is predominantly sporadic many nongenetic, potentially modifiable, factors have been posited to play a key role in its pathogenesis. In particular, aberrant activation of the hypothalamic-pituitary-adrenal axis is considered to make a significant contribution, even at the early stages [1-4]. Indeed, the extent of cortisol excess provides a means to accurately predict the onset of clinical symptoms [5]. In most Alzheimer's disease cases it is difficult to determine what is driving the abnormal endocrine state, although it is generally assumed to be caused by excess psychological or cellular stress [4]. For example, Alzheimer's disease patients often display symptoms of major depressive disorder, a disease which doubles the risk for Alzheimer's disease [6] and it seems likely that Alzheimer's disease and depression lie on a continuum, sharing common glucocorticoid-mediated mechanisms [7]. Furthermore, the widespread clinical use of glucocorticoids is well known to cause mood and cognitive adverse effects [8, 9].

Exogenous administration of stress levels of corticosteroids over several weeks is used to model depressive syndromes in adult rodents [10]. Apart from emotional symptoms, such long-term exposure triggers age-related cognitive problems [10–12]. Synaptic pathology in brain areas such as the hippocampus is a likely proximate cause of impairment of learning and memory, with connections within and between certain networks being particularly vulnerable [13–15]. Of particular note, tau pathogenic mechanisms have been strongly implicated in mediating such synaptic and cognitive impairment [7].

In addition to triggering synaptic and tau pathology chronic administration of glucocorticoids exacerbates amyloid pathology, with accelerated amyloid deposition and cognitive deficits in transgenic mice overexpressing amyloid precursor protein (APP) with familial Alzheimer's disease mutations [16]. Indeed blocking glucocorticoid receptors can not only abrogate memory and synaptic deficits but also reduces amyloid load in APP transgenic mice [16–19]. Soluble amyloid ß (Aß) oligomers are particularly implicated in mediating disruption of synaptic plasticity [20] and recently the acute inhibition of long-term potentiation (LTP) by Aß in hippocampal slices was reported to depend on glucocorticoid receptor activation [21].

Although Aß-induced synaptic deficits are regulated by glucocorticoid status, and prolonged corticosteroid exposure can exacerbate amyloidosis-related synaptic "loss", it is not known if, and for how long, synaptic function in the pre-plaque stage is more susceptible to disruption by corticosteroid elevation. Here, we assess if even relatively brief elevations in corticosterone (CORT), the rodent equivalent of cortisol, exacerbate hippocampal synaptic plasticity dysfunction in young APP transgenic rats, prior

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to amyloid plaque deposition. These animals have an agedependent Aß oligomer-mediated inhibition of LTP which necessitates microglial NOD-leucine rich repeat and pyrin containing protein 3 (NLRP3) inflammasome activation [22, 23]. Since CORT can also activate this pro-inflammatory mechanism in the hippocampus [24], we hypothesised that inhibition of the inflammasome would reverse any persistent synaptic dysfunction.

MATERIALS AND METHODS

Animals

Male transgenic rats (4–5 months old at start) expressing human APP751 with Swedish and Indiana mutations under the control of the murine Thy1.2 promoter (McGill-R-Thy1-APP) [25] and their wild-type littermates were genotyped commercially by Transnetyx (Cordova, TN, USA) using real time polymerase chain reaction. McGill-R-Thy1-APP rats have an age-dependent accumulation of A β , first intracellularly, and then in extracellular neuritic plaques. Cognitive function is impaired before the formation of plaques, as early as 3 months of age [25]. We chose to inject animals at an age (4–5 month) when LTP is impaired by the build-up of A β oligomers but prior to the deposition of plaques [26, 27]. All experiments were carried out in accordance with the approval of the Health Products Regulatory Authority, Ireland, using methods similar to those described previously [22]. Animals had free access to food and water and a 12-h lights on/off cycle.

In vivo surgery and electrophysiology

For the chronic recording in the longitudinal studies the implantation procedure was carried out under anaesthesia using a mixture of ketamine and xylazine (80 and 8 mg/kg, respectively, i.p.) according to methods similar to those described previously [22]. For the longitudinal recovery experiments the rats were allowed at least 14 days after surgery before recordings began [22]. These rats were housed individually in their home cages post-surgery between recording sessions.

Both recording and stimulating electrodes were constructed from Teflon coated tungsten wires. The recording electrodes were single-strand wires (75 μ m inner core diameter, 112 μ m external diameter) targeted at 3.8 mm posterior to bregma and 2.5 mm lateral to midline, whereas the stimulating electrodes were twisted-pair wires (75 μ m inner core diameter, 112 μ m external diameter) targeted at 4.6 mm posterior to bregma and 3.8 mm lateral to midline. Field excitatory postsynaptic potentials (EPSPs) were recorded from the stratum radiatum in the CA1 area of the dorsal hippocampus in response to stimulation of the ipsilateral Schaffer collateral/commissural pathway. The final placement of electrodes was optimised by using electrophysiological criteria and confirmed via postmortem analysis [22].

Test stimuli were delivered to the Schaffer-collateral/commissural pathway every 30 s to evoke field EPSPs that were 45–60% maximum amplitude. To induce LTP we used a strong high frequency stimulation (sHFS) protocol consisting of 3 sets of 10 trains of 20 stimuli at high intensity (75% maximum) at 400 Hz with an inter-train interval of 2 s and an inter-set interval of 5 min [22]. The form of LTP induced by sHFS is both NMDA receptor-dependent and voltage-gated Ca2+ channel-dependent [26, 28]. For non-recovery experiments, only used in the Mcc950 study, the implantation procedure was comparable [22] but the rats were anaesthetised with urethane (1.5 g/kg, i.p.). Core body temperature was maintained at 37.5 \pm 0.5 °C.

Recovery animal experiments were carried out in a well-lit room. The recording compartment consisted of the base of the home cage, including normal bedding and food/water, but the sides were replaced with a translucent Perspex plastic box $(27 \times 22 \times 30 \text{ cm})$ with an open roof. The rats had access to food and water throughout the whole recording session from the same position as in the home cage. All animals were first habituated to the recording procedure over the post-surgery recovery period [22].

Behavioural tasks

Each animal was handled for 5 min per day for a week prior to testing. For the open field the rat was placed in the centre of an open square white apparatus ($L \times W \times H$, $60 \times 60 \times 40$ cm) and allowed to explore it for 5 min. Two parameters were measured: number of lines (marked on the floor) crossed and time spent in the central area (35×35 cm). Twenty four hour after the open field, novel object recognition (NOR) test was performed in the same apparatus, using previously described methods [29]. During the sample phase two identical objects were presented to the animal for 5 min. In the retention phase 3 h later, one of the familiar objects was replaced with a novel one and the animal was allowed to explore them for a further 5 min. The NOR task was repeated again in either 14 or 20 days using another set of objects completely novel to the animals. The discrimination index was calculated as the duration of novel object interaction/ duration of total interaction with both objects.

Drugs

CORT, Sigma was administered using polyethylene glycol 400 (PEG, Sigma, 1 ml/kg s.c.) as the vehicle. We chose a repeated dosing schedule over 3 days based on both previously published data reporting that 20 mg/kg is the minimum chronic dose necessary to trigger 'depression-like' symptoms in wild-type rats [10] and pilot experiments in which we found that 2 h after a single injection of CORT (10 mg/kg s.c.) LTP appeared normal in all animals tested regardless of genotype (WT, 156 \pm 9%, n = 5 and TG, 152 \pm 12%, n = 6). Both 6a-methylprednisolone 21-hemisuccinate sodium salt (MPL, Sigma) and Mcc950 sodium salt [30] (also known as CRID3; MedChemtronica, Sweden) were dissolved in H₂O vehicle (1 ml/kg s.c. and i.p., respectively). Although by no means receptor specific, MPL has been reported to be at least 100-fold more active than CORT at glucocorticoid over mineralocorticoid receptors in vitro [31, 32]. Based on published work reporting the pharmacokinetics/pharmacodynamics of MPL [33-35], we chose a dose of 50 mg/kg. Previously, a sub-chronic dose 30 mg/kg was found not to affect NMDA receptor-dependent LTP in wildtype rats [36]. Choice of the s.c., as opposed to the oral or i.p., route of administration for CORT and MPL avoids first pass metabolism [37] and promotes a gradual increase in concentration in the systemic circulation and hence the brain [38]. The dose schedule for Mcc950 was based on one found to reverse the impairment of NMDA receptor-dependent LTP in McGill-R-Thy1-APP rats [23].

Statistical analysis

Synaptic plasticity varies greatly depending on the stage of oestrous cycle (e.g. [39, 40]). Consistent with this literature, in pilot experiments we found that hippocampal LTP induced by either 200 or 400 Hz protocols was extremely variable in both wild type and transgenic female rats. In order to minimise animal numbers we restricted the present study to males. Animal numbers were also reduced by using a longitudinal design in the majority of experiments, requiring each animal to be recorded from multiple times and a repeated measures statistical evaluation.

The strength of synaptic transmission is expressed as a percentage of the EPSP amplitude recorded over a 30-60 min baseline recording period. The magnitude of LTP was measured at 3 h post-HFS and expressed as the mean ± SEM % baseline. No data were excluded. Control experiments were interleaved randomly throughout experimental sets. The choice of sample size was based on a priori testing to ensure adequate statistical power comparable to previously published papers (https://clincalc.com/ stats/samplesize.aspx). Experimental data passed Shapiro-Wilk and Kolmogorov-Smirnov normality tests. For statistical analysis and graphical display EPSP amplitude measurements were grouped into 10 min epochs. In longitudinal studies one-way ANOVA with repeated measures, followed by Bonferroni's multiple comparison test, was used to compare multiple time points in the same animals. In the cross-sectional Mcc950 study, oneway ANOVA, again followed by Bonferroni's multiple comparison test, was used to compare the magnitude of LTP between multiple groups. Paired and unpaired Student's t tests were used to compare between two time points within one group and between two groups, respectively. Onesample t test was used to analyse the bias towards the novel object [41]. A p value of <0.05 was considered statistically significant. Additional statistical information is available in Supplementary Tables 1 and 2.

RESULTS

Repeated strong conditioning stimulation reliably triggers robust LTP in freely behaving McGill-R-Thy1-APP transgenic rats and their wild-type littermates

Previously, we reported that LTP induced by 200 Hz, but not sHFS repeated high intensity 400 Hz trains, is inhibited in an agedependent manner in pre-plaque transgenic rats [22]. In this study we confirmed that there was no difference in the magnitude of sHFS-induced LTP between genotypes at 4–5 months of age



Fig. 1 Corticosterone transiently disrupts synaptic plasticity in wild-type rats. A In a longitudinal study design, wild-type (WT) rats received single daily s.c. injections of either CORT (10 mg/kg, open circles) or vehicle (PEG, closed circles) for 3 days. Strong high frequency conditioning stimulation (sHFS, arrowhead) was applied repeatedly in the hippocampus of freely behaving rats over a 2.5 month period. The time course of sHFS-induced potentiation from the same animals is displayed before (**B**, 0 month), 2 h after (**C**, 0.5 month), 1 (**D**, 1.5 month) and 2 (**E**, 2.5 month) months after the last injection, starting 4 months of age. Red and blue colours indicate CORT 10 or PEG treatments, respectively (online version only). Insets show representative field EPSP traces at the times indicated. Calibration bars: vertical, 1 mV; horizontal, 10 ms. The magnitude of potentiation 3 h post-sHFS (last 10 min) at the different recording sessions is plotted for WT CORT 10 (**F**) and WT PEG (**G**) rats, for individuals (left hand panel) and groups (right hand panel). The \ddagger symbol stands for a statistical comparison between groups. The magnitude of LTP was significantly different between recording sessions in WT PEG group (p > 0.05; one-way ANOVA with repeated measures). In contrast, LTP was significantly inhibited at 0.5 month in WT CORT 10 group. One symbol, p < 0.05; two symbols, p < 0.01. Values are mean \pm S.E.M. % pre-HFS baseline EPSP amplitude. Additional statistical information for this and the other figures is available in Supplementary Table 1.

(WT 161 ± 5% pre-sHFS baseline EPSP amplitude, n = 24 vs. TG 153 ± 4%, n = 26, p > 0.05, unpaired t test, data not shown). Therefore, we utilised the strong conditioning protocol to evaluate the effects of CORT in both wild-type and transgenic rats. Recordings were taken repeatedly in freely behaving chronically implanted animals. Because we used a longitudinal experimental design it was necessary to assess if the application of sHFS triggered similar magnitude LTP over the 2.5 month study period. Indeed, repeated application of the same strong protocol induced similar magnitude LTP at all recording sessions in both wild-type (157 ± 9% for session 0 month and 157 ± 12% for session 2.5 month) and transgenic (153 ± 6% for session 0 month and 153 ± 8% for session 2.5 month) animals (Supplementary Fig. 1).

Sub-chronic CORT-evoked transient inhibition of LTP in wildtype rats

We first examined the effect of CORT on sHFS-induced LTP in wildtype rats. Sub-chronic treatment (10 mg/kg s.c. daily for 3 days) impaired LTP induced 2 h after the last injection (119±6%, p < 0.05 compared with 160±10% of pre-treatment LTP, one-way ANOVA with repeated measures, Fig. 1B, C, F). The later phase of LTP appeared especially vulnerable to disruption. Importantly, the impairment of LTP by sub-chronic CORT in wild-type rats was transient. Thus, LTP was no longer inhibited when tested in the same animals 1 month after the last injection of CORT (152±10%, p > 0.05 compared with pre-treatment LTP, Fig. 1D, F). By contrast, similar magnitude LTP was triggered at all time points in vehicletreated wild-type rats (Fig. 1).

Sub-chronic CORT-evoked persistent inhibition of LTP in APP transgenic rats

Next, we examined the ability of sub-chronic administration of the same dose (10 mg/kg/day for 3 days) of CORT to disrupt LTP in freely behaving transgenic rats. We found that this brief CORT treatment caused a very long-lasting inhibition of LTP (Fig. 2). Similar to wild-type rats, LTP was strongly inhibited in transgenic animals at 2 h after the last CORT injection (110 \pm 7%, p < 0.05 compared with 151 \pm 10% of pre-treatment LTP, Fig. 2B, C, F). However, in contrast to wild-type rats, application of sHFS either 1 or 2 months after CORT treatment failed to trigger LTP in these transgenic rats (123 \pm 10%, p < 0.01 and 102 \pm 10%, p < 0.001, respectively, compared with pre-treatment LTP, Fig. 2D–F).

To determine if the 10 mg/kg dose was at or near the threshold to cause such a persistent disruption of synaptic plasticity in transgenic rats we also tested half this dose. We found that similar magnitude LTP was triggered at all recording sessions in transgenic rats receiving three injections of 5 mg/kg CORT (Fig. 2).

Sub-chronic CORT-evoked persistent impairment of NOR performance in APP transgenic rats

The possibility that the same sub-chronic CORT treatment protocol would also cause a persistent disruption of cognition was next tested using a relatively simple NOR test [42]. First, we confirmed that transgenic rats were not impaired at this preplaque age. Both untreated transgenic rats and their wild-type littermates had a similar small but significant bias towards the novel object when tested 3 h after the training session (p < 0.001 and p < 0.05, respectively, compared with the hypothetical no-bias value of 0.5, one sample *t* test, Supplementary Fig. 2, see also Fig. 3B, D).

In wild-type rats, consistent with the LTP studies (see Fig. 1), sub-chronic CORT reversibly disrupted NOR performance (Fig. 3B). Thus, in these animals on day 1 post-injection there was no preference for the novel object (p > 0.05), whereas on day 21 there was a significant bias (p < 0.01, Fig. 3E). In contrast, in transgenic rats, and again similar to the LTP studies (see Fig. 2), this sub-chronic CORT regime had a more persistent deleterious effect,

impairing performance of the NOR task both on days 1 and 21 (p > 0.05 on both days, Fig. 3C, F).

Of note, the performance of the same rats in an open field test 2 h after the last CORT injection, just prior to the NOR task, was statistically indistinguishable. Animals of both genotypes travelled a similar distance (lines crossed, mean \pm S.E.M., 90 \pm 7 in WT, n = 24, vs. 78 \pm 7 in TG, n = 23; p = 0.21) and spent comparable amount of time in the centre (mean \pm S.E.M., 14 \pm 3 s in WT, n = 24 vs. 8 \pm 2 s in TG, n = 23; p = 0.08) (data not shown).

The glucocorticoid methylprednisolone also persistently inhibits LTP in APP transgenic rats

The disruption of LTP by sub-chronic CORT is likely mediated via glucocorticoid receptor activation [13, 43]. Therefore, we predicted that sub-chronic treatment with another agonist of these receptors, MPL, would be sufficient to mimic this action of CORT. To our surprise, repeated treatment with a relatively high dose (50 mg/kg per day for 3 days) of MPL had no significant effect on sHFS-induced LTP magnitude at any time point in wild-type rats (148 \pm 9% for session 0 month and 149 \pm 14% for session 2.5 month, Supplementary Fig. 3).

In contrast to wild-type rats, the same MPL treatment protocol had a delayed persistent inhibitory effect in transgenic animals (Fig. 4). Although there was no observable change 2 h after the third injection, LTP was inhibited 1 month later ($107 \pm 8\%$, p < 0.05compared with $147 \pm 9\%$ of pre-treatment LTP, Fig. 4B, D, F), consistent with a role for glucocorticoid receptors in the prolonged disruptive action of CORT. Importantly, the lower dose of MPL (30 mg/kg) had no effect on LTP in transgenic animals (Fig. 4).

An NLRP3 inflammasome inhibitor reverses the persistent inhibition of LTP in CORT-treated APP transgenic rats

Previously we reported that sub-chronic treatment with Mcc950 reversed the disruption of the weaker 200 Hz HFS-induced LTP in pre-plague transgenic rats [23]. Here, we tested the efficacy of the same treatment protocol in 6-month-old CORT-pretreated transgenic rats to abrogate the persistent impairment of sHFS (400 Hz)-induced LTP. In these experiments we used a crosssectional study design and recorded LTP under anaesthesia. Consistent with the findings in freely behaving rats (Figs. 1 and 2), 1 month after repeated treatment with CORT the application of sHFS induced robust LTP in wild-type $(152 \pm 13\%)$ rats but only a weak potentiation in transgenic ($108 \pm 2\%$) animals (Fig. 5). Remarkably, i.p. injection with Mcc950 in the last 6 days (16 mg/kg/day) prior to recording reversed the deficit in transgenic rats. Indeed, LTP induced by sHFS in these rats was similar in magnitude to that recorded in CORT-pretreated wildtype animals $(144 \pm 9\%, p > 0.05)$.

Although there was a trend to increased number of Iba1-positive cells in the CA1 area 1 month after the last CORT injection in TG animals (Supplementary Fig. 4 and Supplementary Methods), a more sensitive technique would be needed to detect any CORT-induced, NLRP3-dependent microglia activation.

DISCUSSION

Recovery from the synaptic plasticity disrupting action of CORT was greatly delayed in an animal model of early Alzheimer's disease amyloidosis. Whereas in wild-type rats LTP inhibition after brief treatment with CORT was transient, the deficit in LTP lasted greater than 2 month in pre-plaque McGill APP rats. Enduring deficits in NOR memory were also triggered by CORT only in the transgenic animals. Moreover, the glucocorticoid MPL mimicked, and the selective NLRP3 inflammasome inhibitor McC950 reversed, the persistent impairment of LTP in the transgenic rats. These findings indicate that early amyloidosis increases the vulnerability of hippocampal synaptic circuits to persistent deleterious actions



Fig. 2 Corticosterone persistently disrupts synaptic plasticity in APP transgenic rats. A In a longitudinal study design, transgenic (TG) rats received single daily s.c. injections of CORT (either 10 mg/kg, closed circles, or 5 mg/kg, open circles) for 3 days. Strong high frequency conditioning stimulation (sHFS, arrowhead) was applied repeatedly in the hippocampus of freely behaving rats over a 2.5 month period. The time course of sHFS-induced potentiation from the same animals is displayed before (**B**, 0 month), 2 h after (**C**, 0.5 month), 1 (**D**, 1.5 month) and 2 (**E**, 2.5 month) months after the last injection, starting at 4 months. Red and blue colours indicate CORT 10 or CORT 5 treatments, respectively (online version only). Insets show representative field EPSP traces at the times indicated. Calibration bars: vertical, 1 mV; horizontal, 10 ms. The magnitude of potentiation 3 h post-sHFS (last 10 min) at the different recording sessions is plotted for 10 mg/kg (**F**) and 5 mg/kg (**G**) CORT-treated rats, for individuals (left hand panel) and groups (right hand panel). The $\frac{4}{3}$ symbol stands for a statistical comparison between groups. LTP was significantly inhibited at 0.5, 1.5, and 2.5 month in the TG CORT 10 group (one-way ANOVA with repeated measures). In contrast, the magnitude of LTP was not significantly different between recording sessions in the TG CORT 5 group. One symbol, p < 0.05; two symbols, p < 0.01; three symbols, p < 0.001. Values are mean \pm S.E.M. % pre-HFS baseline EPSP amplitude.



Fig. 3 Corticosterone persistently disrupts cognition in APP transgenic rats. In a longitudinal study design (**A**, **D**), wild-type (WT) and transgenic (TG) rats received single daily s.c. injections of CORT (10 mg/kg) for 3 days. In first group of animals, performance of WT (**B**) and TG (**C**) rats was tested in a relatively simple novel object recognition task on Day-14 before CORT treatment and 1 Day after the last CORT injection. To avoid potential confounds of multiple repeated behavioural testing in the NOR task, another group of animals was tested on Days 1 and 21. After initial memory disruption on Day 1, WT animals (**E**) significantly improved DI on Day 21. In contrast, TG rats (**F**) didn't show any recovery of performance. Summary of discrimination index (DI) for individuals and groups are in left and right hand panels, respectively. The # symbol stands for a statistical comparison of DI with the hypothetical no-bias value of 0.5 on the same day (one sample *t* test) whereas an * indicates a comparison between different days (paired *t* test). One symbol, *p* < 0.05; two symbols, *p* < 0.01; three symbols, *p* < 0.001. Values are mean \pm S.E.M. % of DI.

of glucocorticoids. The inflammasome inhibitor-mediated reversal of this persistent synaptoxicity lends support to further testing this therapeutic strategy in early Alzheimer's disease.

Corticosteroids and Aß can strongly interfere with similar hippocampal synaptic plasticity mechanisms and consequently have considerable potential to modulate each other's disruptive effects on LTP [13, 20, 21]. Here, we focused on the endurance of hippocampal synaptic plasticity disruption by corticosteroids both in healthy controls and a well-established transgenic rat model of Alzheimer's disease amyloidosis. We found that short-term highdose CORT was sufficient to transiently impair sHFS-induced LTP in wild-type rats. This finding is consistent with previous studies on the acute action of CORT on LTP induced using generally milder conditioning protocols [13, 43]. The latter protocols induce NMDA receptor-dependent LTP whereas sHFS-induced LTP is also voltage-gated Ca²⁺ channel-dependent [26, 28]. Here, short-term CORT acutely impaired a relatively late (3 h) phase of LTP while an earlier phase of LTP (<1 h) appeared unscathed. The apparent lack of change at the earlier phase may indicate that induction mechanisms triggered by the sHFS are relatively resistant to acute CORT.

In vitro, bath-application of CORT has been found to primarily inhibit NMDA receptor-dependent LTP in area CA1 [44–46]. We previously reported that in vivo, both these forms of LTP can be acutely impaired by CORT-elevating acute inescapable stress in wild-type rats [28], but see [44, 47]. Depending on the severity, escapability and duration of the acute stress, recovery of hippocampal NMDA receptor-dependent LTP has been reported to take minutes to over 4 weeks in wild-type animals [28, 48-54]. Although there is a paucity of information available regarding recovery from chronic corticosteroid-mediated inhibition of LTP [12], it is known that hippocampal area CA1 structural synaptic damage caused by high dose long-term CORT exposure reverts to normal between 3 weeks and a month of ceasing treatment in adult wild-type animals [55-57]. Three months injection of CORT in mid-aged (1 year old) wild-type rats caused a deficit in synaptic plasticity that extended to 1 month after ceasing treatment [12]. Thus, our finding that three days of CORT treatment in young preplaque transgenic rats impaired LTP for more than 2 months is remarkable. That the disruption of plasticity lasted so long indicates that the disease course was accelerated or aggravated by CORT. It will be important to determine if such brief CORT treatment persistently increases soluble Aß concentration, as has been reported for chronic glucocorticoid exposure in transgenic mice [58]. Recently, prolonged multi-modal 'modern life-like' acute stress was reported to rapidly increase Aß oligomers via upregulation of BACE and exacerbate hippocampal CA3 spine loss in transgenic mice [59]. We know that Alzheimer's disease



Fig. 4 Persistent disruption of synaptic plasticity in APP transgenic rats by the glucocorticoid methylprednisolone. A Study design: Transgenic rats (TG) received single daily s.c. injections of methylprednisolone (MPL either 50 mg/kg, closed circles, or 30 mg/kg, open circles) for 3 days. Strong high frequency conditioning stimulation (sHFS, arrow) was applied repeatedly in the hippocampus of freely behaving rats over a 2.5-month period. The time course of sHFS-induced potentiation from the same animals is displayed before (**B**, 0 month), 2 h after (**C**, 0.5 month), 1 (**D**, 1.5 month) and 2 (**E**, 2.5 month) months after the last injection, starting at 5 months. Red and blue colours indicate MPL 50 or MPL 30 treatments, respectively (online version only). Insets show representative field EPSP traces at the times indicated. Calibration bars: vertical, 1 mV; horizontal, 10 ms. The magnitude of potentiation 3 h post-sHFS (last 10 min) at the different recording sessions is plotted for TG MPL 30 (**G**) rats, for individuals (left hand panel) and groups (right hand panel). The \ddagger symbol stands for a statistical comparison between pre- and 3 h post-HFS values at each recording session within one group (paired *t* test) whereas an * indicates a comparison between pre- and 3 h post-tHFS values at each recording session within one group (paired *t* test) whereas an * indicates a comparison between groups. The magnitude of LTP was significantly inhibited at 1.5 and 2.5 month in TG MPL 30 group (p > 0.05; one-way ANOVA with repeated measures). In contrast, LTP was significantly inhibited at 1.5 and 2.5 month in TG MPL 50 group. One symbol, p < 0.05; two symbols, p < 0.01. Values are mean \pm S.E.M. % pre-HFS baseline EPSP amplitude.



120 150 180

Α

B

EPSP (%)

-60 -30 ò 30 60 90

TE CORT IN NCOSO Fig. 5 Reversal of the corticosterone-evoked persistent disruption of synaptic plasticity in anaesthetised transgenic rats by the inflammasome inhibitor Mcc950. A Schematic diagram of treatment regimen, starting at 5 months of age (online version in colour). B Whereas sHFS triggered robust LTP in wild-type rats pretreated 1 month previously with CORT (WT CORT 10), the same pretreatment in transgenic rats strongly inhibited LTP (TG CORT 10). Six-day i.p. treatment with Mcc950 (16 mg/kg/day) reversed the LTP deficit in transgenic rats (TG CORT 10 MCC950). An arrowhead indicates the time point of sHFS application. Inserts show representative EPSP traces at the times indicated. Calibration bars: Vertical, 1 mV, horizontal 10 ms. LTP magnitude was determined during the last 10 min. Statistical summary is in the right hand panel. Number of animals per group is shown on the bar. *p < 0.05 (one-way ANOVA followed by Bonferroni's multiple comparison test); p < 0.05, p < 0.05, p < 0.050.01 compared with pre-sHFS baseline (paired t test). Values are the mean ± S.E.M. % pre-HFS baseline EPSP amplitude.

Time (min)

brain-derived Aß oligomers can strongly inhibit sHFS-induced LTP [60]. In pilot studies we found that sHFS-induced LTP appeared unaffected in 9-10 months old control transgenic rats. Future studies need to investigate how long the CORT-triggered synaptic plasticity disruption lasts and at what age, if any, transgenic rats develop similar deficits in sHFS-induced LTP.

The ability of sub-chronic CORT to persistently inhibit LTP in the transgenic animals was not discernible when half the active dose was given. The requirement for a relatively high dose is consistent with the likely involvement of glucocorticoid receptors. Indeed similar brief administration of the glucocorticoid MPL also strongly inhibited LTP 2 months after the last injection, again in an apparently dose-dependent manner. Somewhat surprisingly, unlike CORT, brief MPL treatment did not significantly impair sHFS-induced LTP when tested 2 h after the last injection in either wild-type or transgenic rats. This difference between the time course of MPL and CORT action may at least partly lie in their different kinetics [61, 62], in particular the rate of penetration into the brain. Unlike CORT [63, 64], MPL has been found to enter the brain slowly by a saturable mechanism [65] and is a good substrate of P-glycoprotein, a major efflux pump of the bloodbrain barrier [66]. Consistent with this suggestion the application of sHFS 6 h after the last MPL injection (50 mg/kg, i.p. daily for 3 days) failed to trigger significant LTP in either WT or TG ure than e-an esthetized rats (n = 4 per group; unpublished

Sub-chronic CORT treatment also had more persistent disruptive effects on cognition in transgenic rats as assessed using a relatively simple NOR test. These transgenic rats have been reported not to be impaired on the version used here up to 1 year of age [42], again indicating that the disease course was accelerated or aggravated by CORT. Sub-chronic high-dose MPL is currently used clinically for disorders including certain forms of multiple sclerosis, and has been reported to cause a persistent, but reversible, impairment of memory function in these patients [70, 71]. The possibility of brief glucocorticoid-induced longlasting deficits in other cognitively vulnerable populations, including early Alzheimer's disease, needs further research [9, 72].

The NLRP3 inflammasome is a molecular complex central to the production of pro-inflammatory cytokines such as interleukin-1ß [73]. Evidence of its involvement in early Alzheimer's disease [74] includes enhanced cerebral expression of the key NLRP3 component, active caspase-1 [75]. The NLRP3 inflammasome has been reported to mediate microglial response to either Aß [76] or CORT [24]. Inhibition of the NLRP3 inflammasome reduces Aß burden and enhances cognition in plaque-bearing transgenic mice [75, 77], and reverses the LTP deficit in pre-plague transgenic rats [23]. The present finding that Mcc950 can also abrogate CORT-exacerbated LTP inhibition supports future evaluation of its efficacy against the CORT-exacerbated behavioural deficits. The finding also strongly supports a requirement for the NLRP3 inflammasome in the persistent deleterious synaptic actions of brief exposure to excess CORT on a background of Alzheimer's disease amyloidosis.

The present animal model findings on the synaptotoxic interaction between CORT and Aß, taken together with recent prospective longitudinal preclinical Alzheimer's disease human studies [5, 78], lend support to developing lifestyle changes to mitigate against exposure to glucocorticoid excess and underlie the potential benefit of therapies targeting shared mechanisms such as inflammasome activation early in the disease.

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REFERENCES

- 1. Sapolsky RM, Krey LC, McEwen BS. The neuroendocrinology of stress and aging: the glucocorticoid cascade hypothesis. Endocr Rev. 1986;7:284-301.
- 2. Rothman SM, Mattson MP. Adverse stress, hippocampal networks, and Alzheimer's disease. Neuromol Med. 2010;12:56-70.
- 3. Kalafatakis K, Russell GM, Zarros A, Lightman SL. Temporal control of glucocorticoid neurodynamics and its relevance for brain homeostasis, neuropathology and glucocorticoid-based therapeutics. Neurosci Biobehav Rev. 2016;61:12-25.
- 4. Ouanes S, Popp J. High cortisol and the risk of dementia and Alzheimer's Disease: a review of the literature. Front Aging Neurosci. 2019;11:43.
- 5. Ennis GE, An Y, Resnick SM, Ferrucci L, O'Brien RJ, Moffat SD. Long-term cortisol measures predict Alzheimer disease risk. Neurology 2017;88:371-78.
- 6. Cherbuin N, Kim S, Anstey KJ. Dementia risk estimates associated with measures of depression: a systematic review and meta-analysis. BMJ Open. 2015;5:e008853.

- Sotiropoulos I, Silva JM, Gomes P, Sousa N, Almeida OFX. Stress and the etiopathogenesis of Alzheimer's disease and depression. Adv Exp Med Biol. 2019;1184:241–57.
- 8. Brown ES. Effects of glucocorticoids on mood, memory, and the hippocampus. Treatment and preventive therapy. Ann N. Y Acad Sci. 2009;1179:41–55.
- Judd LL, Schettler PJ, Brown ES, Wolkowitz OM, Sternberg EM, Bender BG, et al. Adverse consequences of glucocorticoid medication: psychological, cognitive, and behavioral effects. Am J Psychiatry. 2014;171:1045–51.
- Sterner EY, Kalynchuk LE. Behavioral and neurobiological consequences of prolonged glucocorticoid exposure in rats: relevance to depression. Prog Neuropsychopharmacol Biol Psychiatry. 2010;34:777–90.
- 11. Dachir S, Kadar T, Robinzon B, Levy A. Cognitive deficits induced in young rats by long-term corticosterone administration. Behav Neural Biol. 1993;60:103–9.
- Bodnoff SR, Humphreys AG, Lehman JC, Diamond DM, Rose GM, Meaney MJ. Enduring effects of chronic corticosterone treatment on spatial learning, synaptic plasticity, and hippocampal neuropathology in young and mid-aged rats. J Neurosci. 1995;15:61–9.
- Joels M, Sarabdjitsingh RA, Karst H. Unraveling the time domains of corticosteroid hormone influences on brain activity: rapid, slow, and chronic modes. Pharm Rev. 2012;64:901–38.
- Chattarji S, Tomar A, Suvrathan A, Ghosh S, Rahman MM. Neighborhood matters: divergent patterns of stress-induced plasticity across the brain. Nat Neurosci. 2015;18:1364–75.
- Xiong H, Krugers HJ. Tuning hippocampal synapses by stress-hormones: relevance for emotional memory formation. Brain Res. 2015;1621:114–20.
- Green KN, Billings LM, Roozendaal B, McGaugh JL, LaFerla FM. Glucocorticoids increase amyloid-beta and tau pathology in a mouse model of Alzheimer's disease. J Neurosci. 2006;26:9047–56.
- Baglietto-Vargas D, Medeiros R, Martinez-Coria H, LaFerla FM, Green KN. Mifepristone alters amyloid precursor protein processing to preclude amyloid beta and also reduces tau pathology. Biol Psychiatry. 2013;74:357–66.
- Lante F, Chafai M, Raymond EF, Pereira AR, Mouska X, Kootar S, et al. Subchronic glucocorticoid receptor inhibition rescues early episodic memory and synaptic plasticity deficits in a mouse model of Alzheimer's disease. Neuropsychopharmacology 2015;40:1772–81.
- Pedrazzoli M, Losurdo M, Paolone G, Medelin M, Jaupaj L, Cisterna B, et al. Glucocorticoid receptors modulate dendritic spine plasticity and microglia activity in an animal model of Alzheimer's disease. Neurobiol Dis. 2019;132:104568.
- Li S, Selkoe DJ. A mechanistic hypothesis for the impairment of synaptic plasticity by soluble Abeta oligomers from Alzheimer's brain. J Neurochem. 2020;154:583–97.
- Kootar S, Frandemiche ML, Dhib G, Mouska X, Lorivel T, Poupon-Silvestre G, et al. Identification of an acute functional cross-talk between amyloid-beta and glucocorticoid receptors at hippocampal excitatory synapses. Neurobiol Dis. 2018;118:117–28.
- 22. Qi Y, Klyubin I, Harney SC, Hu N, Cullen WK, Grant MK, et al. Longitudinal testing of hippocampal plasticity reveals the onset and maintenance of endogenous human Aß-induced synaptic dysfunction in individual freely behaving pre-plaque transgenic rats: rapid reversal by anti-Aß agents. Acta Neuropathol Commun. 2014;2:175.
- Qi Y, Klyubin I, Cuello AC, Rowan MJ. NLRP3-dependent synaptic plasticity deficit in an Alzheimer's disease amyloidosis model in vivo. Neurobiol Dis. 2018;114: 24–30.
- Frank MG, Hershman SA, Weber MD, Watkins LR, Maier SF. Chronic exposure to exogenous glucocorticoids primes microglia to pro-inflammatory stimuli and induces NLRP3 mRNA in the hippocampus. Psychoneuroendocrinology 2014;40:191–200.
- Leon WC, Canneva F, Partridge V, Allard S, Ferretti MT, DeWilde A, et al. A novel transgenic rat model with a full Alzheimer's-like amyloid pathology displays preplaque intracellular amyloid-beta-associated cognitive impairment. J Alzheimers Dis. 2010;20:113–26.
- Qi Y, Klyubin I, Hu NW, Ondrejcak T, Rowan MJ. Pre-plaque Aß-mediated impairment of synaptic depotentiation in a transgenic rat model of Alzheimer's Disease Amyloidosis. Front Neurosci. 2019;13:861.
- Zhang D, Qi Y, Klyubin I, Ondrejcak T, Sarell CJ, Cuello AC, et al. Targeting glutamatergic and cellular prion protein mechanisms of amyloid beta-mediated persistent synaptic plasticity disruption: longitudinal studies. Neuropharmacology 2017;121:231–46.
- Ryan BK, Vollmayr B, Klyubin I, Gass P, Rowan MJ. Persistent inhibition of hippocampal long-term potentiation in vivo by learned helplessness stress. Hippocampus 2010;20:758–67.
- 29. Bevins RA, Besheer J. Object recognition in rats and mice: a one-trial nonmatching-to-sample learning task to study 'recognition memory'. Nat Protoc. 2006;1:1306–11.

- Coll RC, Robertson AA, Chae JJ, Higgins SC, Munoz-Planillo R, Inserra MC, et al. A small-molecule inhibitor of the NLRP3 inflammasome for the treatment of inflammatory diseases. Nat Med. 2015;21:248–55.
- De Kloet E, Ortiz Zacarias, NV and Meijer, OC. Manipulating the brain corticosteroid receptor balance: focus on ligands and modulators. in Neuroendocrinology and neurobiology handbook of stress vol 2. Fink G, editor. AMSTERDAM: Academic Press; 2017. p. 367–84.
- 32. Grossmann C, Scholz T, Rochel M, Bumke-Vogt C, Oelkers W, Pfeiffer AF, et al. Transactivation via the human glucocorticoid and mineralocorticoid receptor by therapeutically used steroids in CV-1 cells: a comparison of their glucocorticoid and mineralocorticoid properties. Eur J Endocrinol. 2004;151:397–406.
- Slivka AP, Murphy EJ. High-dose methylprednisolone treatment in experimental focal cerebral ischemia. Exp Neurol. 2001;167:166–72.
- Yao Z, DuBois DC, Almon RR, Jusko WJ. Pharmacokinetic/pharmacodynamic modeling of corticosterone suppression and lymphocytopenia by methylprednisolone in rats. J Pharm Sci. 2008;97:2820–32.
- Viscomi MT, Florenzano F, Latini L, Amantea D, Bernardi G, Molinari M. Methylprednisolone treatment delays remote cell death after focal brain lesion. Neuroscience 2008;154:1267–82.
- 36. Zhang B, Chen X, Lin Y, Tan T, Yang Z, Dayao C, et al. Impairment of synaptic plasticity in hippocampus is exacerbated by methylprednisolone in a rat model of traumatic brain injury. Brain Res. 2011;1382:165–72.
- 37. Lukas G, Brindle SD, Greengard P. The route of absorption of intraperitoneally administered compounds. J Pharm Exp Ther. 1971;178:562–4.
- Durk MR, Deshmukh G, Valle N, Ding X, Liederer BM, Liu X. Use of subcutaneous and intraperitoneal administration methods to facilitate cassette dosing in microdialysis studies in rats. Drug Metab Dispos. 2018;46:964–69.
- Warren SG, Humphreys AG, Juraska JM, Greenough WT. LTP varies across the estrous cycle: enhanced synaptic plasticity in proestrus rats. Brain Res. 1995;703:26–30.
- Wang W, Le AA, Hou B, Lauterborn JC, Cox CD, Levin ER, et al. Memory-related synaptic plasticity is sexually dimorphic in rodent hippocampus. J Neurosci. 2018;38:7935–51.
- Prado Lima MG, Schimidt HL, Garcia A, Dare LR, Carpes FP, Izquierdo I, et al. Environmental enrichment and exercise are better than social enrichment to reduce memory deficits in amyloid beta neurotoxicity. Proc Natl Acad Sci USA. 2018;115:E2403–E09.
- 42. Galeano P, Martino Adami PV, Do Carmo S, Blanco E, Rotondaro C, Capani F, et al. Longitudinal analysis of the behavioral phenotype in a novel transgenic rat model of early stages of Alzheimer's disease. Front Behav Neurosci. 2014;8:321.
- Xu L, Holscher C, Anwyl R, Rowan MJ. Glucocorticoid receptor and protein/RNA synthesis-dependent mechanisms underlie the control of synaptic plasticity by stress. Proc Natl Acad Sci USA. 1998;95:3204–8.
- Alfarez DN, Wiegert O, Joels M, Krugers HJ. Corticosterone and stress reduce synaptic potentiation in mouse hippocampal slices with mild stimulation. Neuroscience 2002;115:1119–26.
- Krugers HJ, Alfarez DN, Karst H, Parashkouhi K, van Gemert N, Joels M. Corticosterone shifts different forms of synaptic potentiation in opposite directions. Hippocampus 2005;15:697–703.
- Wiegert O, Pu Z, Shor S, Joels M, Krugers H. Glucocorticoid receptor activation selectively hampers N-methyl-D-aspartate receptor dependent hippocampal synaptic plasticity in vitro. Neuroscience 2005;135:403–11.
- Mesches MH, Fleshner M, Heman KL, Rose GM, Diamond DM. Exposing rats to a predator blocks primed burst potentiation in the hippocampus in vitro. J Neurosci. 1999;19:RC18.
- Foy MR, Stanton ME, Levine S, Thompson RF. Behavioral stress impairs long-term potentiation in rodent hippocampus. Behav Neural Biol. 1987;48:138–49.
- Shors TJ, Seib TB, Levine S, Thompson RF. Inescapable versus escapable shock modulates long-term potentiation in the rat hippocampus. Science 1989;244:224–6.
- 50. Xu L, Anwyl R, Rowan MJ. Behavioural stress facilitates the induction of long-term depression in the hippocampus. Nature 1997;387:497–500.
- Garcia R, Musleh W, Tocco G, Thompson RF, Baudry M. Time-dependent blockade of STP and LTP in hippocampal slices following acute stress in mice. Neurosci Lett. 1997;233:41–4.
- Sacchetti B, Lorenzini CA, Baldi E, Bucherelli C, Roberto M, Tassoni G, et al. Timedependent inhibition of hippocampal LTP in vitro following contextual fear conditioning in the rat. Eur J Neurosci. 2002;15:143–50.
- 53. Shors TJ, Gallegos RA, Breindl A. Transient and persistent consequences of acute stress on long-term potentiation (LTP), synaptic efficacy, theta rhythms and bursts in area CA1 of the hippocampus. Synapse 1997;26:209–17.
- Kohda K, Harada K, Kato K, Hoshino A, Motohashi J, Yamaji T, et al. Glucocorticoid receptor activation is involved in producing abnormal phenotypes of singleprolonged stress rats: a putative post-traumatic stress disorder model. Neuroscience 2007;148:22–33.

- Sousa N, Lukoyanov NV, Madeira MD, Almeida OF, Paula-Barbosa MM. Reorganization of the morphology of hippocampal neurites and synapses after stress-induced damage correlates with behavioral improvement. Neuroscience 2000;97:253–66.
- Shapiro LP, Omar MH, Koleske AJ, Gourley SL. Corticosteroid-induced dendrite loss and behavioral deficiencies can be blocked by activation of Abl2/Arg kinase. Mol Cell Neurosci. 2017;85:226–34.
- 57. Ortiz JB, Conrad CD. The impact from the aftermath of chronic stress on hippocampal structure and function: Is there a recovery? Front Neuroendocrinol. 2018;49:114–23.
- Joshi YB, Chu J, Pratico D. Stress hormone leads to memory deficits and altered tau phosphorylation in a model of Alzheimer's disease. J Alzheimers Dis. 2012; 31:167–76.
- Baglietto-Vargas D, Chen Y, Suh D, Ager RR, Rodriguez-Ortiz CJ, Medeiros R, et al. Short-term modern life-like stress exacerbates Abeta-pathology and synapse loss in 3xTg-AD mice. J Neurochem. 2015;134:915–26.
- 60. Klyubin I, Ondrejcak T, Hayes J, Cullen WK, Mably AJ, Walsh DM, et al. Neurotransmitter receptor and time dependence of the synaptic plasticity disrupting actions of Alzheimer's disease Abeta in vivo. Philos Trans R Soc Lond B Biol Sci. 2014;369:20130147.
- Sainio EL, Lehtola T, Roininen P. Radioimmunoassay of total and free corticosterone in rat plasma: measurement of the effect of different doses of corticosterone. Steroids 1988;51:609–22.
- 62. Hall ED. The neuroprotective pharmacology of methylprednisolone. J Neurosurg. 1992;76:13–22.
- Mason BL, Pariante CM, Thomas SA. A revised role for P-glycoprotein in the brain distribution of dexamethasone, cortisol, and corticosterone in wild-type and ABCB1A/B-deficient mice. Endocrinology 2008;149:5244–53.
- Mason BL, Pariante CM, Jamel S, Thomas SA. Central nervous system (CNS) delivery of glucocorticoids is fine-tuned by saturable transporters at the blood-CNS barriers and nonbarrier regions. Endocrinology 2010;151:5294–305.
- Chen TC, Mackic JB, McComb JG, Giannotta SL, Weiss MH, Zlokovic BV. Cellular uptake and transport of methylprednisolone at the blood-brain barrier. Neurosurgery 1996;38:348–54.
- Hashimoto N, Nakamichi N, Yamazaki E, Oikawa M, Masuo Y, Schinkel AH, et al. P-Glycoprotein in skin contributes to transdermal absorption of topical corticosteroids. Int J Pharm. 2017;521:365–73.
- Yang CH, Huang CC, Hsu KS. Behavioral stress modifies hippocampal synaptic plasticity through corticosterone-induced sustained extracellular signalregulated kinase/mitogen-activated protein kinase activation. J Neurosci. 2004; 24:11029–34.
- Gartner U, Holzer M, Arendt T. Elevated expression of p21ras is an early event in Alzheimer's disease and precedes neurofibrillary degeneration. Neuroscience 1999;91:1–5.
- 69. McShea A, Lee HG, Petersen RB, Casadesus G, Vincent I, Linford NJ, et al. Neuronal cell cycle re-entry mediates Alzheimer disease-type changes. Biochim Biophys Acta. 2007;1772:467–72.
- 70. Oliveri RL, Sibilia G, Valentino P, Russo C, Romeo N, Quattrone A. Pulsed methylprednisolone induces a reversible impairment of memory in patients with relapsing-remitting multiple sclerosis. Acta Neurol Scand. 1998;97:366–9.
- Uttner I, Muller S, Zinser C, Maier M, Sussmuth S, Claus A, et al. Reversible impaired memory induced by pulsed methylprednisolone in patients with MS. Neurology 2005;64:1971–3.
- Cheung V, Hoshide R, Bansal V, Kasper E, Chen CC. Methylprednisolone in the management of spinal cord injuries: Lessons from randomized, controlled trials. Surg Neurol Int. 2015;6:142.
- Rathinam VA, Fitzgerald KA. Inflammasome complexes: emerging mechanisms and effector functions. Cell 2016;165:792–800.
- Milner MT, Maddugoda M, Gotz J, Burgener SS, Schroder K. The NLRP3 inflammasome triggers sterile neuroinflammation and Alzheimer's disease. Curr Opin Immunol. 2020;68:116–24.

- Heneka MT, Kummer MP, Stutz A, Delekate A, Schwartz S, Vieira-Saecker A, et al. NLRP3 is activated in Alzheimer's disease and contributes to pathology in APP/ PS1 mice. Nature 2013;493:674–8.
- Halle A, Hornung V, Petzold GC, Stewart CR, Monks BG, Reinheckel T, et al. The NALP3 inflammasome is involved in the innate immune response to amyloidbeta. Nat Immunol. 2008;9:857–65.
- Dempsey C, Rubio Araiz A, Bryson KJ, Finucane O, Larkin C, Mills EL, et al. Inhibiting the NLRP3 inflammasome with MCC950 promotes non-phlogistic clearance of amyloid-beta and cognitive function in APP/PS1 mice. Brain Behav Immun. 2017;61:306–16.
- Pietrzak RH, Laws SM, Lim YY, Bender SJ, Porter T, Doecke J, et al. Plasma cortisol, brain amyloid-beta, and cognitive decline in preclinical Alzheimer's disease: a 6-year Prospective Cohort Study. Biol Psychiatry Cogn Neurosci Neuroimaging. 2017;2:45–52.

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AUTHOR CONTRIBUTIONS

Y.Q. and I.K. performed the experiments and analysed the data. I.K. and M.R. wrote the paper. All the authors contributed to study design, and read and approved the final paper.

ADDITIONAL INFORMATION

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