



Recurrent *FOS* rearrangement in proliferative fasciitis/proliferative myositis

Naohiro Makise¹ · Taisuke Mori^{2,3} · Toru Motoi⁴ · Junji Shibahara⁵ · Tetsuo Ushiku¹ · Akihiko Yoshida^{2,6}

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Abstract

Proliferative fasciitis (PF) and proliferative myositis (PM) are rare benign soft tissue lesions, usually affecting the extremities of middle-aged or older adults. Presenting as poorly circumscribed masses, they histologically show bland spindle cell proliferation in a myxoid to fibrous background and a hallmark component of large epithelioid “ganglion-like” cells in various numbers, which may lead to their misdiagnosis as sarcoma. PF/PM has been long considered as reactive, akin to nodular fasciitis; however, its pathogenesis has remained unknown. In this study, we analyzed the *FOS* status in 6 PF/PMs (5 PFs and 1 PM). Five PF/PMs occurred in adults, all showing diffuse strong expression of c-*FOS* primarily in the epithelioid cells, whereas spindle cell components were largely negative. Using fluorescence in situ hybridization (FISH), all 5 c-*FOS*-immunopositive tumors showed evidence of *FOS* gene rearrangement in the epithelioid cells. RNA sequencing in 1 case detected a *FOS-VIM* fusion transcript, which was subsequently validated by reverse transcriptase-polymerase chain reaction, Sanger sequencing, and *VIM* FISH. The one pediatric PF case lacked c-*FOS* expression and *FOS* rearrangement. c-*FOS* immunohistochemistry was negative in 45 cases of selected mesenchymal tumor types with epithelioid components that may histologically mimic PF/PM, including pleomorphic sarcoma with epithelioid features and epithelioid sarcoma. Recurrent *FOS* rearrangement and c-*FOS* overexpression in PF/PM suggested these lesions to be neoplastic. *FOS* abnormality was largely restricted to the epithelioid cell population, clarifying the histological composition of at least 2 different cell types. c-*FOS* immunohistochemistry may serve as a useful adjunct to accurately distinguish PF/PM from mimics.

Introduction

Proliferative fasciitis (PF) and proliferative myositis (PM) are rare benign soft tissue lesions. PM was first described formally by Kern in 1960 [1] and further characterized by Enzinger and Dulcey in 1967 [2]. Chung and Enzinger subsequently reported, in 1975, a subcutaneous counterpart of PM, known as PF [3]. PF and PM usually affect middle-aged and elderly adults with a roughly equal sex distribution [4]. Pediatric cases, though much rarer, have also been reported [5]. These tumors most commonly occur in the extremities, although trunk wall, head, and neck may also be involved. The tumors often present as rapidly growing masses, usually <5 cm in size, and may be associated with tenderness or pain. Microscopically, PF and PM are poorly circumscribed masses, composed of bland spindle cell proliferation in a myxoid to fibrous background and a hallmark component of large epithelioid “ganglion-like” cells in various numbers. Owing to the rapid growth, poor circumscription, and the presence of large epithelioid cells, PF/PM may be misdiagnosed as sarcoma [2, 3].

These authors jointly supervised this work: Tetsuo Ushiku, Akihiko Yoshida

✉ Akihiko Yoshida
akyoshid@ncc.go.jp

- ¹ Department of Pathology, Graduate School of Medicine, The University of Tokyo, Tokyo, Japan
- ² Department of Diagnostic Pathology, National Cancer Center Hospital, Tokyo, Japan
- ³ Division of Molecular Pathology, National Cancer Center Research Institute, Tokyo, Japan
- ⁴ Department of Pathology, Tokyo Metropolitan Cancer and Infectious Diseases Center Komagome Hospital, Tokyo, Japan
- ⁵ Department of Pathology, Kyorin University School of Medicine, Tokyo, Japan
- ⁶ Rare Cancer Center, National Cancer Center Hospital, Tokyo, Japan

Table 1 Summary of the primary antibodies and staining condition.

Antibody	Clone	Dilution	Pretreatment	Solution	Source
c-FOS	Polyclonal (ABE457)	1:2000	Water bath	Targeted Retrieval Solution, pH9 (Dako, Glostrup, Denmark)	Millipore, Billerica, MA, USA
FOSB	5G4	1:400	Autoclaving	Citrate buffer	Cell Signaling Technology, Danvers, MA, USA
ERG	EPR3864	1:2000	Autoclaving	Citrate buffer	Abcam, Cambridge, UK
SATB2	SATBA4B10	1:1000	Water bath	Targeted Retrieval Solution, pH9 (Dako)	Santa Cruz Biotechnology, Dallas, TX, USA

The pathogenesis of PF/PM remains unknown till date. Nonetheless, the lesions are widely considered as reactive, since they show self-limiting growth, scar-like histology, and a rare incidence of spontaneous involution [6]. Approximately 20% of PF/PM cases are associated with previous trauma [3]. These characteristics are so similar to those in the much more common nodular fasciitis that these lesions are often discussed together, or even considered as related entities. However, nodular fasciitis is now known to be a clonal disease, with *USP6* gene rearrangement in up to 92% of cases (most commonly *MYH9-USP6* fusion) [7, 8]; all PF/PMs tested thus far have lacked *USP6* rearrangement [8–10], challenging the association. Reported cytogenetic abnormalities of PF/PM, including trisomy 2 in 2 cases [11, 12] and t(6;14)(q23;q32) in 1 case [13] have not been validated or characterized further.

Recently, we noted diffuse strong immunohistochemical expression of c-FOS in a case of PF. Guided by this unexpected finding, we analyzed archival PF/PM cases to determine whether FOS abnormality is recurrent in these lesions.

Materials and methods

Case selection

This study was approved by the Institutional Review Boards. Samples from 6 PF/PM cases identified in the institutional pathology archives (National Cancer Center Hospital, University of Tokyo Hospital, Komagome Hospital, and Kyorin University Hospital, all in Tokyo, Japan) were available for additional immunohistochemical and molecular analyses. All tumors were histologically reviewed by a soft-tissue pathologist (AY) and the diagnosis was confirmed. Clinical data were retrieved from medical records.

We also retrieved soft-tissue tumors with epithelioid cells that can enter the histological differential diagnosis of PF/PM. These included 15 pleomorphic sarcomas with epithelioid component (9 myxofibrosarcomas and 6 undifferentiated pleomorphic sarcomas), 10 epithelioid sarcomas (all deficient in SMARCB1 expression), 5 epithelioid heman-gioendotheliomas (all immunopositive for CAMTA1), 5

pseudomyogenic hemangioendotheliomas (all immunopositive for FOSB), 5 embryonal rhabdomyosarcomas, and 5 tumors with ganglion cell component (2 ganglioneuromas and 3 ganglioneuroblastomas), although the latter 2 tumors types represent more historical than practical differentials. In addition, we collected 10 cases of nodular fasciitis (9 were tested positive for *USP6* rearrangement).

Immunohistochemistry

Sections (4 μm-thick) from the formalin-fixed, paraffin-embedded (FFPE) block of each specimen were routinely deparaffinized. The sections were exposed to 3% hydrogen peroxide to block endogenous peroxidase activity. Heat-induced epitope retrieval was performed. The primary antibodies and staining conditions are summarized in Table 1. The slides were incubated for 1 h at room temperature, and subsequently labeled with the EnVision system (Dako, Glostrup, Denmark). c-FOS was stained in all cases, whereas FOSB, ERG, and SATB2 were stained for a subset of PF/PM cases when needed in the study process. Appropriate controls were used. For PF/PMs and other tumors with epithelioid cells, strong nuclear immunoreactivity in >50% of epithelioid cells was considered positive. For nodular fasciitis, the staining results were recorded descriptively, since it lacked epithelioid cells except a few scattered osteoclast-like giant cells.

Fluorescence in situ hybridization (FISH)

FISH was performed using 4-μm-thick FFPE sections from PF/PM specimens. We used break-apart probes to detect gene rearrangement of *FOS*, *VIM*, and *FOSB* (all obtained from GSP Lab, Inc., Kobe, Japan). FISH images were captured using the Metafer Slide Scanning Platform (MetaSystems, Altlusheim, Germany). Samples in which more than 20% of the epithelioid cells showed break-apart signals were considered positive for rearrangement. Particular care was taken to differentiate epithelioid cell nuclei from surrounding spindle cell or endothelial nuclei, with a close correlation with H&E and c-FOS-immunostained slides. Relevant positive and negative control cases were used for each probe.

Table 2 Clinicopathological and molecular genetic summary of PF/PM.

Case	Age/Sex	Site	Diagnosis	c-FOS IHC	FOS FISH	VIM FISH	Sequencing
1	72/M	Thigh	PF	+	+	+	FOS-VIM
2	68/M	Forearm	PF	+	+	–	NA
3	1/M	Shoulder	PF	–	–	NA	NA
4	78/M	Groin	PF	+	+	–	NA
5	70/M	Arm	PM	+	+	–	NA
6	60/M	Arm	PF	+	+	–	NA

M male, *PF* proliferative fasciitis, *PM* proliferative myositis, *IHC* immunohistochemistry, *FISH* fluorescence in situ hybridization, *NA* data not available.

RNA sequencing

FFPE sample sections from case 1 were deparaffinized and subjected to RNA extraction using an RNeasy FFPE kit (Qiagen, Hilden, Germany). RNA sequencing was performed using the TruSight Pan-Cancer panel (Illumina, San Diego, CA, USA), which targets 1385 cancer-related genes, following the manufacturer's instruction. Sequencing was performed on a MiSeq instrument (Illumina) using MiSeq Reagent Kit v3 (Illumina) with 150 cycles. Fusion gene identification was performed using the RNA-Seq alignment applications STAR and Top-Hat2 (Illumina).

Reverse transcriptase-polymerase chain reaction (RT-PCR) and Sanger sequencing

Total RNA was extracted from the FFPE sample of case 1 using RecoverAll Total Nucleic Acid Isolation Kit (Invitrogen, Carlsbad, CA, USA). cDNA was synthesized from total RNA using Superscript III (Invitrogen). Tumor cDNA was subjected to PCR using the primer pairs 5'-GCTTCCCTTGATCTGACTGG-3' and 5'-TCTCCAAAGGCTG CAGAAGT-3' (predicted product size of 130 bp) to specifically amplify the fusion transcript. The PCR product was cleaned up using IllustraMicroSpin Columns (GE Healthcare, Chalfont St. Giles, UK). The isolated PCR product was then subjected to Sanger sequencing reaction using the BigDye Terminator version 3.1 Cycle Sequencing Kit (Applied Biosystems, Foster City, CA, USA) and analyzed on an ABI PRISM 3130xl Genetic Analyzer (Thermo Fisher Scientific, Waltham, MA, USA). The sequence chromatogram files were examined using FinchTV (version 1.4.0; Digital World Biology, Seattle, WA, USA).

Results

Clinicopathological findings

Clinicopathological findings of PF/PM cases are summarized in Table 2. All 6 patients were men. Five patients were adults (60–78 years old), and 1 was a child (1 year old). The lesions

were located in the arm ($n = 2$), forearm ($n = 1$), shoulder ($n = 1$), thigh ($n = 1$), and groin ($n = 1$). Five tumors were subcutaneous (PF), whereas 1 was intramuscular (PM). The lesions measured 7–30 mm (median, 20 mm). The tumor was resected in 4 cases, and expectantly observed after needle biopsy in 2 cases. Follow-up was available for 4 patients, and none had evidence of disease over 8 to 51 months. One tumor (case 4), which received needle biopsy only, involuted and was undetectable at 8 months. Histologically, all adult tumors were poorly circumscribed with variable infiltration into the surrounding adipose tissue (Fig. 1) or the skeletal muscle. The lesions consisted of fascicular proliferation of non-atypical long spindle cells in a fibro-edematous background, in which characteristic large epithelioid cells were admixed (Figs. 2a, c, e, g, 4a, 5a). The latter harbored abundant amphophilic cytoplasm, vesicular nuclei, and prominent nucleoli, similar to ganglion cells, and demonstrated no or focally mild nuclear pleomorphism. The number and distribution of epithelioid cells were variable, ranging from isolated and scattered to dense in adult cases. In resected specimens, the epithelioid cells were abundant in the center of the lesions, while they were scattered amongst predominant spindle cells in the periphery. In the only pediatric case, numerous epithelioid cells formed diffuse sheets, without accompanying spindle cell proliferation, in a well-circumscribed tumor (Fig. 7a), which was typical for this age group [5]. Overall, all 6 cases showed classic clinicopathological findings of PF/PM.

c-FOS overexpression and FOS rearrangement in adult PF/PM

The results are summarized in Table 2. Immunohistochemically, all 5 PF/PM tumors in adults showed c-FOS expression (Figs. 2b, d, f, h, 4b, 5b). The staining diffusely decorated the large epithelioid cells and the reactivity was so intense that, in addition to nuclear labeling, the cytoplasm also exhibited weaker staining. Similarly labeled smaller epithelioid cells, small round cells, and spindle cells were also observed in the tumor tissues in a much smaller number, some of which likely represented the truncated large epithelioid cells. In contrast, spindle cell components were mostly negative for c-FOS. Three of these cases (cases 1, 2,

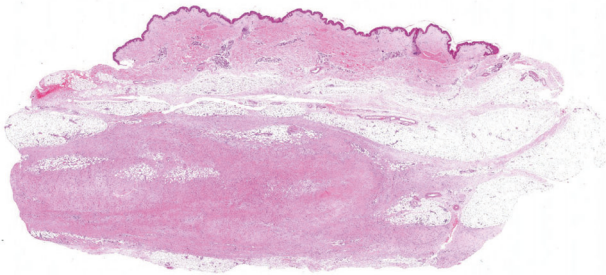


Fig. 1 A scanning view of proliferative fasciitis. The lesion shows a poorly circumscribed discoid fibrous mass in the subcutis, irregularly entrapping adipose tissue (case 1).

and 4) were also stained with FOSB antibody and showed negative to weak focal staining in spindle cells only.

Based on FISH, all the 5 c-FOS-immunopositive PF/PMs showed evidence of *FOS* gene rearrangement in the epithelioid cells, whereas spindle cell component was largely negative for *FOS* rearrangement (Figs. 3, 4c, 5c). A low-level *FOS* copy number increase was observed in some epithelioid cells with gene rearrangement.

One *FOS*-rearranged PF (case 1) had adequate material for targeted RNA sequencing, which detected a *FOS-VIM* fusion transcript. The fusion was validated using RT-PCR and Sanger sequencing. The breakpoint in *FOS* was located at c.724 in exon 4, which was fused to the reverse complementary strand of exon 1 in *VIM* (Fig. 6a). The predicted protein lost 138 amino acids from the C-terminus of wild-type c-FOS protein and gained additional 10 amino acids at the C-terminal end before the stop codon, maintaining the leucine zipper domain but losing the transactivation domain of c-FOS protein (Fig. 6b). The fusion was further validated using FISH with *VIM* break-apart probe (Fig. 6c). *VIM* break-apart FISH was performed for the remaining 4 *FOS*-rearranged PF/PM tumors; however, none showed evidence of *VIM* rearrangement.

Lack of c-FOS overexpression and *FOS* rearrangement in the pediatric PF case

The only pediatric case of PF studied (case 3) lacked both c-FOS immunoreexpression and *FOS* rearrangement (Fig. 7b, c). Both FOSB immunohistochemistry and *FOSB* FISH were also negative.

No evidence of endothelial or osteoblastic differentiation in PF/PM

To examine whether *FOS* rearrangement in PF/PM might result in similar lines of differentiation to epithelioid hemangioma or osteoid osteoma/osteoblastoma, we stained 4 *FOS*-rearranged PF/PM tumors (cases 1, 2, 4, and 5) using ERG and SATB2 antibodies, which are sensitive vascular

endothelial and osteoblastic markers, respectively. All 4 PF/PMs tested were negative for ERG. SATB2 was weakly and focally expressed in epithelioid cells in 2 of the 4 cases, while the remaining 2 showed negative results.

Utility of c-FOS immunohistochemistry in differential diagnosis

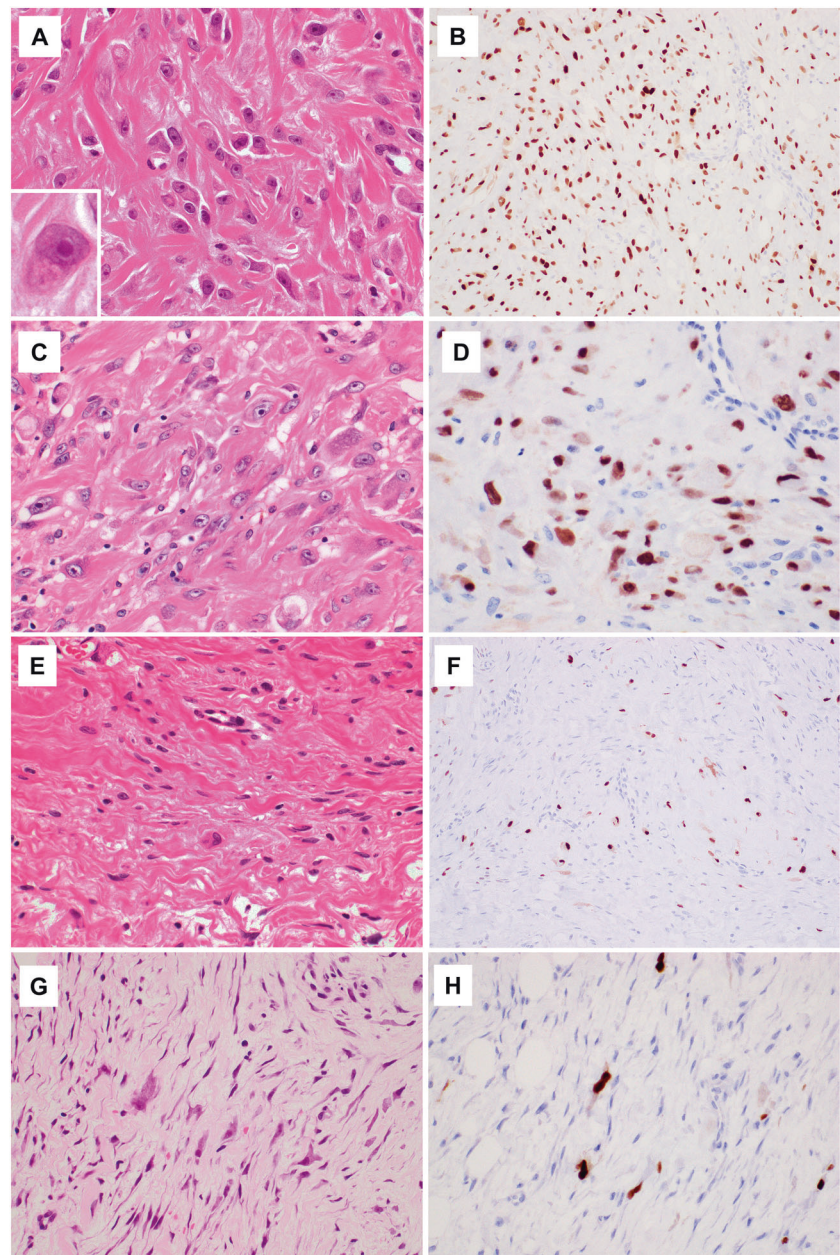
We performed c-FOS immunohistochemistry in histological mimics of PF/PM to test its diagnostic utility. The results are presented in Table 3 and illustrated in Fig. 8. None of the 45 mesenchymal tumors with epithelioid components showed strong nuclear staining in >50% of cells, although heterogeneous weak to strong immunoreactive cells were focally observed in many cases. All 10 cases of nodular fasciitis contained a small number of spindle or small rounded cells that were weakly to strongly positive for c-FOS in up to 10% of cells.

Discussion

PF and PM are rare benign soft tissue lesions with distinct clinical and histological features, and yet unknown pathogenesis. In this study, we identified recurrent *FOS* rearrangement in archival PF/PM cases with classic clinicopathological features. Among a total of 6 PF/PMs, 5 adult tumors showed *FOS* rearrangement as per FISH, along with diffuse and strong c-FOS positivity by immunohistochemistry. One of these cases, analyzed by RNA sequencing, showed a *FOS-VIM* fusion transcript, which was validated by RT-PCR, Sanger sequencing, and *VIM* FISH. Although the karyotype t(6;14)(q23;q32) was previously identified in a case of PM through conventional G-banding¹³, the *FOS* locus 14q24 was not involved in it.

FOS gene encodes a leucine zipper protein c-FOS, which dimerizes with the JUN family protein and forms the transcription factor complex AP-1 [14]. *FOS* physiologically plays important roles in cell growth, differentiation, and survival. Oncogenic c-FOS activation via *FOS* gene rearrangement has been reported in epithelioid hemangioma [15, 16] and osteoid osteoma/osteoblastoma [17], in which *FOS* fuses with diverse partners, including *LMNA*, *VIM*, *MBNL1*, and lincRNA (*RP11-326N17.1*) in epithelioid hemangioma [15, 16], and *ANKH*, *KIAA1199*, *MYO1B*, and intergenic non-coding regions in osteoid osteoma/osteoblastoma [17]. In the reported cases, *FOS* gene is invariably rearranged within exon 4 and fused to various protein-coding or non-coding regions, forming a stop codon at or immediately after the breakpoint. Truncated c-FOS protein maintains the leucine zipper domain while losing the transactivating domain and C-terminal regulatory sequences, thus hampering normal rapid mRNA

Fig. 2 Histological and c-FOS immunohistochemical findings of proliferative fasciitis. **a, c** Characteristic large epithelioid cells densely proliferate, particularly in the center of the lesion. These cells harbor vesicular chromatin, prominent nucleoli, and amphophilic cytoplasm, similar to ganglion cells (A, case 1; C, case 2). **Inset** in (a) shows a magnified view of a ganglion-like cell. **b, d** c-FOS immunohistochemistry (in the areas depicted in (a) and (c), respectively) shows diffuse strong expression in epithelioid cells. **e, g** In areas, particularly at the periphery of the lesion, fascicular proliferation of bland spindle cells predominates, with only a few scattered epithelioid cells admixed within (E, case 1, G, case 4). **f, h** c-FOS immunohistochemistry in these areas (depicted in (e) and (g), respectively) identifies a few scattered epithelioid cells, whereas spindle cell component is virtually negative for c-FOS expression.



and protein degradation and leading to abnormally stabilized mutant c-FOS expression [17, 18]. The causal relationship between *FOS* rearrangement and epithelioid hemangioma was supported by an in vitro experiment in which ectopic expression of truncated c-FOS stimulated endothelial sprouting via upregulation of matrix metalloproteinases and components of the Notch signaling pathway [18]. The fusion structure of *FOS-VIM* in our case 1 was similar to that reported in epithelioid hemangioma [16], which, along with diffuse strong c-FOS immunorexpression, suggests a similar c-FOS activating mechanism in PF and PM. However, further studies are required to understand the detailed pathogenesis.

PF and PM have long been considered as reactive, owing to self-limiting growth, scar-like microscopic appearance, association with trauma in a subset, and rare spontaneous involution [1–3, 6]. The demonstration of recurrent *FOS* rearrangement in PF/PM challenges this notion and suggested a neoplastic nature of this disorder, adding to the growing list of “pseudotumors” that have been re-interpreted as neoplasms after the discovery of recurrent genetic abnormalities (most notably, the frequent *USP6* rearrangement in nodular fasciitis [7, 8] and myositis ossificans [19]). Spontaneous regression, which is rarely documented and observed in one of our cases after needle biopsy, may indicate PF/PM to possibly represent another

Fig. 3 *FOS* break-apart FISH in cases of proliferative fasciitis. The assay shows evidence of *FOS* rearrangement in epithelioid cell nuclei (**a** case 2; **b** case 1; arrows indicate split 3' and 5' probes). Notice background spindle cells in (**a**) lack *FOS* gene rearrangement.

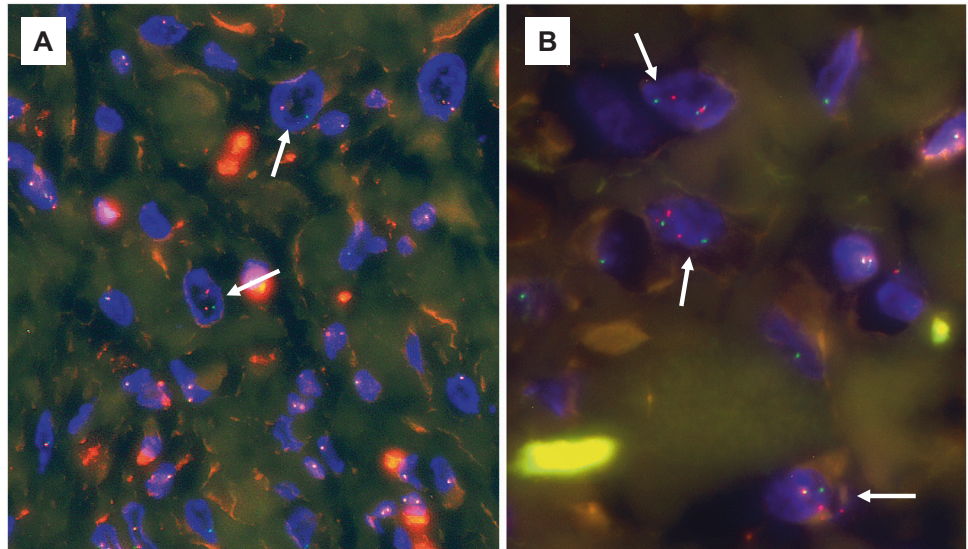


Fig. 4 Additional case of proliferative fasciitis (case 6). **a** The tumor consists of large epithelioid cells with mild pleomorphism and spindle cell proliferation. **b** The epithelioid cells are immunohistochemically positive for c-FOS, whereas spindle cells are negative. **c** The epithelioid cells harbor *FOS* gene rearrangement as detected by FISH (arrows indicate split 3' and 5' probes), as well as a low-level copy number increase.

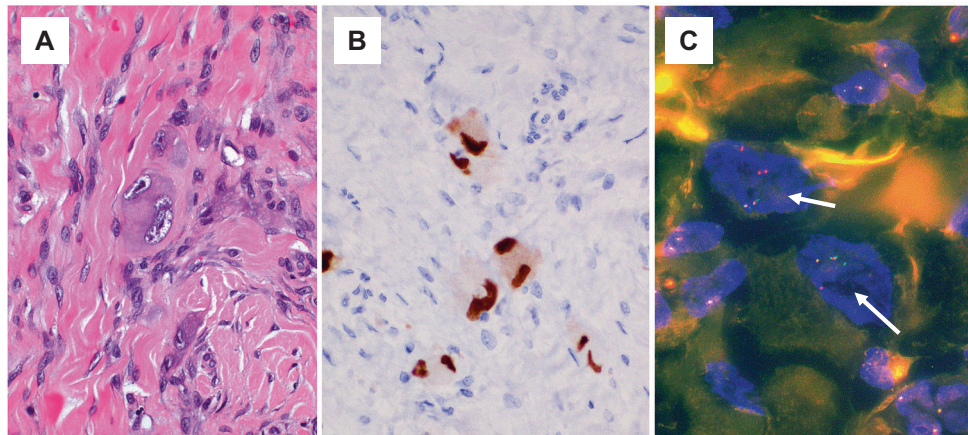
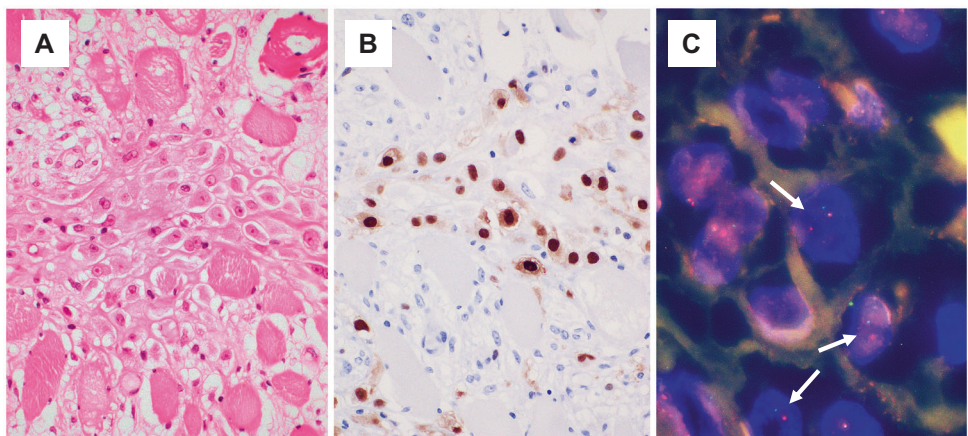


Fig. 5 A case of proliferative myositis (case 5). **a** Entrapping skeletal muscle fibers, large epithelioid cells are observed. **b** The epithelioid cells are intensely positive for c-FOS immunostaining. **c** The epithelioid cells harbor *FOS* gene rearrangements as detected by FISH (arrows indicate split 3' and 5' probes).



example of “transient neoplasia”, a concept originally proposed for nodular fasciitis [7].

c-FOS immunoreactivity and *FOS* rearrangement were primarily observed in epithelioid cells, a hallmark of PF/

PM, but they were mostly absent in the accompanying spindle cells. The nature of epithelioid cells in PF/PM has been a subject of debate, with proposed hypotheses including myogenic, fibroblastic, myofibroblastic, and

histiocytic lineages [20]. Electron microscopic evidence have suggested these cells to be modified fibroblasts, since they harbor abundant rough endoplasmic reticulum, thin filament of actin type, along with intracytoplasmic collagen production [3, 20, 21]. Immunohistochemically, epithelioid cells have been positive for vimentin, and negative or focally positive for smooth muscle actin [21]. Enzinger et al. had speculated epithelioid and spindle cells to possibly form a spectrum, since they observed an intermediate form

between these cells [2, 3]. However, the existence of intermediate cells was not confirmed by a subsequent ultrastructural study by Lundgren et al [21]. Our study suggested the epithelioid and spindle cells to be distinct populations, with the only former likely representing a clonal element, admixed with reactive spindle cells. Nonetheless, a much smaller number of small round cells and spindle cells that expressed c-FOS were also admixed, and their nature and relationship to large epithelioid cells remain unclear.

The heterogeneous histological composition of PF/PM is reminiscent of osteoid osteoma, in which *FOS* fusion is restricted to the osteoblastic cells that line immature bone and osteoid within conspicuous granulation-like reactive tissue. PF/PM is reportedly associated with bone formation in rare instances [1, 2, 13], leading to an early hypothesis that it might represent a precursor to myositis ossificans [1]. Later studies had also repeatedly suggested epithelioid cells in PF/PM to show some resemblance with differentiating osteoblasts, both at the light and electron microscopic levels [2, 3, 21]. However, none of the epithelioid cells in the 4 *FOS*-rearranged PF/PMs tested in this study showed diffuse strong SATB2 expression, unlike osteoid osteoma/osteoblastoma [22], thus not supporting the immediate phenotypic link between these distinctive diseases in soft tissues and bone.

Both *FOS* rearrangement and c-FOS immunoreactivity were negative in the only pediatric PF sample examined in this study. Pediatric PF/PM is an exceedingly rare lesion, with some recognized differences from adult counterparts, including good circumscription, lobulation, diffuse sheet of epithelioid cells, and lack of spindle cell component [5]. This pediatric tumor also lacked FOSB expression or gene rearrangement, although *FOSB* fusion has been reported as an alternative oncogenic mechanism in epithelioid hemangioma or osteoblastoma that lack *FOS* alterations [17, 23]. Interestingly, a previous study identified diffuse strong FOSB immunorepression in a small subset of PF [24]. As

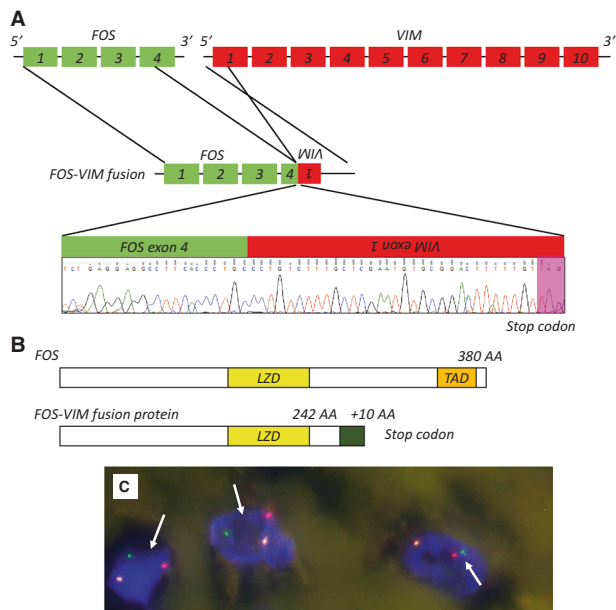


Fig. 6 *FOS-VIM* fusion in case 1. **a** RNA sequencing detected a *FOS-VIM* fusion transcript, which was validated by RT-PCR and Sanger sequencing. The breakpoint in *FOS* was located at c.724 in the middle of exon 4, which was fused to the reverse complementary strand of exon 1 of *VIM*. **b** The predicted protein lost 138 C-terminal amino acids (AA) of wild-type c-FOS protein and gained additional 10 AA at C-terminal end before the stop codon, which maintains the leucine zipper domain (LZD) but loses the transactivation domain (TAD) of c-FOS protein. **c** The presence of *FOS-VIM* fusion was also validated by *VIM* rearrangement as detected by a *VIM* break-apart FISH assay (arrows indicate split 3' and 5' probes).

Fig. 7 A pediatric case of proliferative fasciitis (case 3). **a** The tumor consists of dense proliferation of large epithelioid cells. **b** c-FOS immunohistochemistry is negative. **c** FISH shows no evidence of *FOS* rearrangement (only fused yellow signals are observed).

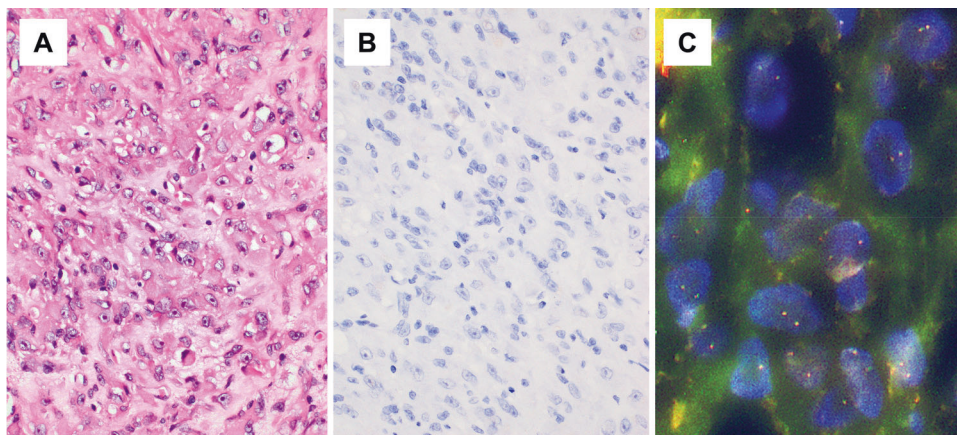


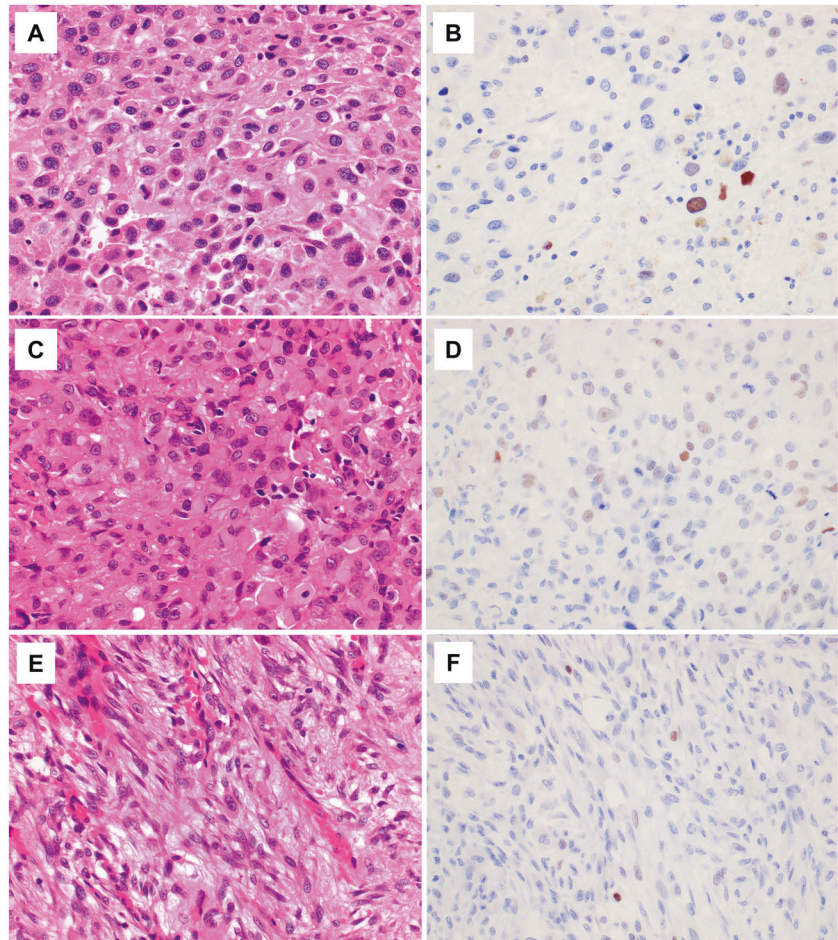
Table 3 c-FOS immunohistochemistry in PF/PM and histological mimics.

Tumor types	N	Positive [#]	Comments
Proliferative fasciitis/myositis	6	5	Strong staining in virtually all epithelioid cells in 5/6 cases.
Pleomorphic sarcoma with epithelioid features	15	0	Weak to strong staining in <5% of cells in 10/15 cases.
Epithelioid sarcoma	10	0	Weak to moderate staining in <1 to 40% of cells in 8/10 cases.
Epithelioid hemangioendothelioma	5	0	Strong staining in 5% of cells in 1/5 cases.
Pseudomyogenic hemangioendothelioma	5	0	Moderate staining in <1% of cells in 1/5 cases.
Embryonal rhabdomyosarcoma	5	0	Weak to strong staining in <5% in 4/5 cases.
Ganglioneuroma/Ganglioneuroblastoma	5	0	Weak to strong staining in ≤5% of ganglion cells in 2/5 cases.
Nodular fasciitis	10	–	Weak to strong staining in <1 to 10% of spindle cells and small cells in 10/10 cases.

[#]Immunopositivity was defined by strong nuclear staining in >50% of epithelioid cells. This definition was not applicable to nodular fasciitis.

Fig. 8 c-FOS immunohistochemistry in histological mimics of proliferative fasciitis/myositis.

a, b An undifferentiated pleomorphic sarcoma with epithelioid features focally shows heterogeneous weak to strong c-FOS immunoreactivity. **c, d** An epithelioid sarcoma focally shows heterogeneous weak to moderate c-FOS immunoreactivity. **e, f** A case of nodular fasciitis containing a small number of spindled or small rounded cells that weakly to moderately expressed c-FOS.



the present study is limited by the small number of cases, future studies on a larger cohort are necessary to establish the prevalence of *FOS* rearrangement, and to determine whether alternative *FOSB* rearrangement may exist in a subset and whether pediatric PF/PM shares pathogenesis with adult cases.

c-FOS immunohistochemistry can be diagnostically helpful. Unlike PF/PM, none of the 45 mimicking tumors

with epithelioid component showed diffuse (>50%) strong c-FOS expression. In bone, c-FOS staining has been proposed as an ancillary tool for distinguishing osteoid osteoma/osteoblastoma from osteosarcoma [25, 26]. Nodular fasciitis invariably contained a small number of scattered spindled or small rounded cells that were weakly to strongly stained for c-FOS. Although this low-power appearance may superficially resemble that of PF/PM, the

c-FOS-positive cells in nodular fasciitis do not possess large epithelioid cytology, and the staining is heterogeneous unlike uniform strong labeling in PF/PM.

In summary, we demonstrated that PF/PM harbors recurrent *FOS* rearrangement and c-FOS overexpression, thus suggesting these lesions as neoplasms. *FOS* abnormality was largely restricted to the epithelioid cell population, clarifying the histological composition of at least 2 different cell types. c-FOS immunohistochemistry may serve as a useful adjunct to accurately distinguish PF/PM from mesenchymal mimics with epithelioid elements.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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