BRIEF COMMUNICATION



Compound heterozygous mutations of *SH3TC2* in Charcot–Marie–Tooth disease type 4C patients

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Abstract

Charcot-Marie-Tooth disease type 4C (CMT4C) is an autosomal recessive neuropathy caused by *SH3TC2* mutations, characterized by spine deformities and cranial nerve involvement. This study identified four CMT4C families with compound heterozygous *SH3TC2* mutations from 504 Korean demyelinating or intermediate CMT patients. The frequency of the CMT4C was calculated as 0.79% in demyelinating and intermediate patients (n = 504), but it was calculated as 2.02% in patients without *PMP22* duplication (n = 198). The CMT4C frequency was similar to patients in Japan, but it was relatively low compared to those patients in other populations. The symptom was less severe and slowly progressed compared to the other AR-CMT. A patient harboring an intermediate neuropathy showed cranial nerve involvement but did not have scoliosis. This study will be helpful in making molecular diagnoses of demyelinating or intermediate CMT due to *SH3TC2* mutations.

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Introduction

Mutations in *SH3TC2* cause autosomal recessive (AR) demyelinating or intermediate Charcot–Marie–Tooth disease type 4C (CMT4C) characterized by spine deformities and cranial nerve impairment [1–7]. *SH3TC2* encodes Src homology-3 (SH3) domains and tetratricopeptide repeat (TPR) motifs which are localized in the plasma membrane of the Schwann cells of peripheral nerves [8, 9].

The frequencies of CMT4C vary according to countries in AR demyelinating patients without *PMP22* duplication. CMT4C is the most frequent form of AR-CMT in several countries: >20% in Italy [4] and Czech [10]. However, CMT4C is relatively rare in Germany (5.2%) [5] and Japan (1.76%) [6]. This study examined *SH3TC2* mutations in Korean demyelinating or intermediate CMT patients.

Materials and methods

Subjects and clinical examinations

We collected 504 unrelated demyelinating or intermediate CMT patients from the Korean CMT cohort study group, of which 306 patients were proven to be positive for *PMP22* duplication. This study performed exome sequencing or

 Table 1 Compound heterozygous SH3TC2 mutations in four CMT4C patients

Family ID	Mutations		dbSNP no	Mutant allele frequencies				GERP	In silico analysis ^b			
	Nucleotide ^a	Amino acid		1000G	ESP	ExAC	KRGDB		PROVEAN	PP2	MUpro	Fathmm
FC523, FC657	c.929G>A	p.G310E	NR	NR	NR	0.00001	NR	4.88	-6.84*	1.00*	-0.29*	-0.17*
	c.2831A>G	p.E944G	NR	NR	NR	NR	0.00091	6.02	-6.65*	1.00*	-1.00*	-1.21*
FC703, FC1080	c.929G>A	p.G310E	NR	NR	NR	0.00001	NR	4.88	-6.84*	1.00*	-0.29*	-0.17*
	c.3272G>T	p.G1091V	NR	NR	NR	0.00001	NR	5.76	-8.27*	1.00*	-1.00*	-1.73*

1000G 1000 Genomes project (http://www.1000genomes.org/), CMT4C Charcot-Marie-Tooth disease type 4C, ESP exome sequencing project (http://exs.gs.washington.edu/EVS/), ExAC exome aggregation consortium (http://exac.broadinstitute.org/), GERP genomic evolutionary rate profiling score (http://mendel.stanford.edu/SidowLab/downloads/gerp/index.html), KRGDB Korean reference genome database (http://152.99.75. 168/KRGDB/menuPages/introKor.jsp), NR not reported

^aReference DNA sequence of SH3TC2: NM_024577.3

^bScores of PROVEAN (http://provean.jcvi.org/seq_submit.php) <-2.5, PolyPhen-2 (PP2, http://genetics.bwh.harvard.edu/pph2/) ~1, MUpro (http://www.ics.uci.edu/~baldig/mutation) <0, and Fathmm (http://fathmm.biocompute.org.uk/) \leq -1.5 indicate a prediction of pathogenicity (asterisk denotes a "pathogenic" prediction)

targeted sequencing for CMT-related genes in 198 patients without *PMP22* duplication.

Physical disability was determined by the functional disability scale (FDS) and the CMT neuropathy score (CMTNS). Nerve conduction values and MRIs were obtained by previous methods [11]. Distal sural nerve was biopsied from two patients (FC657 and FC703). This study was approved by the Institutional Review Board of Sung-kyunkwan University, Samsung Medical Center.

Molecular genetic studies

Exome sequencing and targeted sequencing of the CMTrelated genes were performed using the genomic DNAs isolated from blood [12, 13]. The exome was captured using the SureSelect Human All Exon 50 M Kit (Agilent Technologies, Santa Clara, CA, USA), and the subsequent sequencing was performed using the HiSeq 2000 Genome Analyzer (Illumina, San Diego, CA, USA).

RESULTS

Identification of *SH3TC2* compound heterozygous mutations

From the 198 CMT families without *PMP22* duplication, we identified four patients having likely pathogenic compound heterozygous mutations in *SH3TC2*: c.929G>A (p.G310E) and c.2831A>G (p.E944G) in two families (FC523 and FC667) and c.929G>A (p.G310E), and c.3272G>T (p.G1091V) in two families (FC703 and FC1080) (Table 1, Fig. 1a, b). Thus far, these compound heterozygous combinations have not been reported in either unaffected controls or CMT patients. The mutations are located at the highly conserved regions (Fig. 1c, d), and

several in silico analyses suggested a pathogenic prediction of the *SH3TC2* mutations.

Several other rare heterozygous variants were found in *SH3TC2* (Supplementary Table 1). They were excluded from the genetic causes because of the deviation from the AR mode. However, if these variants exist in a homozygous or compound heterozygous state with other variants, their pathogenicity could not be ruled out.

Clinical manifestations

The clinical features of the four patients are summarized in Table 2. Foot deformities, pes cavus, and walking difficulties were present in all the patients. No one showed lower limb proximal weakness. Areflexia was noticed in the early stages of the disease, but pathologic reflexes were not found. Mild scoliosis, cranial nerve involvement, and hearing loss were present in three patients. Disease severity according to the CMTNS varied from mild to moderate.

Nerve conduction velocities (NCVs) indicated demyelinating or intermediate type neuropathies. Median motor NCVs in the FC657 patient were 36.2–38.0 m/s, and the sural nerve biopsy findings revealed both axonal and demyelinating neuropathies, which indicate the intermediate type. Compound muscle action potentials of the lower limb muscles were considerably decreased compared to the upper limb muscles.

T1-weighted coronal MRIs of a patient (FC703) showed a fatty infiltration and atrophy in the distal leg muscles (Fig. 1e–j). Although the axial MRIs showed normal images at the hip and thigh levels (Fig. 1g, h), whole compartment muscles in the calf revealed fatty replacement (Fig. 1e, f). The T1 signal intensity was increased in the anterior muscle compartment of the lower leg which was accompanied by fatty infiltration. Hypertrophy was observed in both sciatic nerves, and these findings are appropriate for



Fig. 1 Charcot–Marie–Tooth disease type 4C (CMT4C) families with *SH3TC2* mutations. **a** Four Pedigrees of CMT4C families. Genotypes of the *SH3TC2* mutations are indicated at the bottom of all the examined family members. Filled and open symbols represent affected and unaffected individuals, respectively. **b** Sequencing chromatograms of the *SH3TC2* mutations. **c** Domain structure of the SH3TC2 protein. The three observed mutations are indicated by arrows (SH3: Src homology-3 domain, TPR: tetratricopeptide repeat motif). **d** Conservation of the amino acids at the mutation sites among several vertebrate species (*H. sapiens*: NP_078853.2, *M. musculus*: NP_766216.2, and *B. taurus*: XP_015327803.1, *C. lupus*:

XP_022273537.1, *G. gallus*: XP_025010675.1, and *X. topicalis*: XP_002939154.2). **e-j** Hip, thigh, and calf MRIs of the FC703 patient (16 years old). **e, f** Coronal images of the thigh (**e**) and calf muscles (**f**). T1-weighted coronal leg MRIs of the lower extremities showed fatty replacement and muscle atrophies in the calf. Axial images of the hip (**g**) and thigh (**h**) muscles revealed no fatty replacements. Axial images at the upper third (**i**) and lower third calf (**j**) showed fatty streaks which were involved in the whole compartment calf muscles. Those findings were consistent with the patterns of muscle involvements in length dependent axonal degeneration

neuromuscular disease which may be related to neuromuscular junction changes in the CMT4C mouse model [14]. Although they had cranial nerve involvement, the brain MRI did not reveal any abnormalities in all four patients (data not shown). Semithin transverse sections from the distal sural nerve biopsy (FC703, 16 years) showed rather characteristic myelin abnormalities, with the loss of large myelinated fibers, and there was frequent evidence of onion bulb.

Discussion

We identified two combinations of likely pathogenic compound heterozygous *SH3TC2* mutations in four CMT4C patients. The CMT4C frequency was calculated as 2.02% in the demyelinating or intermediate patients without *PMP22* duplication (n = 198), which is similar or slightly lower than patients in Japan (1.76%) [6] and Germany (5.2%) [5]. The frequencies were 0.39% in the total Korean CMT Table 2 Clinical characterization of the four patients with compound heterozygous SH3TC2 mutations

Item/Patient ID	FC523	FC657	FC703	FC1080	
Mutation	G310E, E944G	G310E, E944G	G310E, G1091V	G310E, G1091V	
Sex	Female	Male	Male	Female	
Age at onset (years)	30	38	10	8	
Disease duration (years)	23	14	6	17	
Age at exam (years)	53	52	16	25	
Muscle weakness ^a					
Upper limb	+	+	+	+	
Lower limb ++		+	+	++	
Muscle atrophy ^b	U < L	L	L	U < L	
Sensory loss	V > P	V > P	V = P	V > P	
DTR	Absent	Absent	Absent	Absent	
Scoliosis	Yes	No	Yes	Yes	
Cranial nerve involvements	No	Deafness, diplopia, facial palsy	Deafness	Deafness, diplopia	
Foot deformities	Yes	Yes	Yes	Yes	
Wheel-chair bound No		No	No	No	
FDS/CMTNS	2/15	2/11	2/11	2/12	
Brain MRI	Normal	Normal	Normal	Normal	
Lower limb MRI	ND	ND	Fatty replacement	Fatty replacement	
Sural nerve biopsy	ND	Mixed type ^c	Demyelinating type	ND	
Nerve conduction stud	ly (Right/Left) ^d				
Median nerve					
CMAP (mV)	7.2/7.4	10.3/11.3	12.1/15.5	11.3/18.4	
MNCV (m/s)	30.0/30.1	38.0/36.2	27.0/27.8	36.9/34.9	
Peroneal nerve					
CMAP (mV) A/A		A/A	0.2/2.1	2.6/1.8	
MNCV (m/s)	A/A	A/A	13.8/17.5	28.1/30.6	
Sural nerve					
SNAP (µV)	2.9/1.6	7.0/A	2.1/A	7.1/7.1	
SNCV (m/s)	35.6/31.4	32.6/A	28.0/A	33.3/35.0	

A absent potential, CMAP compound muscle action potential, CMTNS CMT neuropathy score, DTR deep tendon reflexes, FDS functional disability scale, MNCV motor nerve conduction velocity, MRI magnetic resonance imaging, ND not done, P pain sense, SNAP sensory nerve action potential, SNCV sensory nerve conduction velocity, V vibration sense

^aUpper limb: + indicates intrinsic hand weakness 4/5 on medical research council (MRC) scale. Lower limb: ++ indicates ankle dorsiflexion <4/5 on MRC scale

^bU < L indicates lower limb predominant muscle atrophy, L indicates lower limb muscle atrophy alone ^cMixed type: both axonal and demyelinating neuropathies

^dNormal nerve conduction velocities: median nerve \geq 50.5 m/s, peroneal nerve \geq 41.2 m/s, sural nerve \geq 32.1 m/s; normal amplitude values: median nerve $\geq 6 \text{ mV}$, peroneal nerve $\geq 1.6 \text{ mV}$, sural nerve $\geq 6.0 \mu \text{V}$

cohort (n = 1035), and 0.79% in the demyelinating and intermediate patients (n = 504). The CMT4C frequency in the Korean cohort was relatively lower than other groups: 0.8% in USA [15], 1.7% in Germany [5], 0.47% in Japan [6], and 0.56% in United Kingdom [16]. Moreover, no homozygous mutation was found in the Korean patients, which may be partly due to a strict legal prohibition of consanguineous marriage.

Onset ages in the two patients with p.G310E and p. G1091V were 8–10 years; however, the other two patients with p.G310E and p.E944G had quite a late onset (30 and 38 years, respectively). Spine deformities appeared comparably early, similarly to previous reports [1, 3, 4, 6]. Cranial nerve involvement was evident in three patients; however, it occurred independently from the disease duration and neuropathy severity. Although some studies have suggested an early proximal involvement [1, 3], we could not find any proximal weakness. Based on the motor and sensory amplitudes, the lower limb nerves were definitively more compromised than the upper limb nerves, and the sensory nerves appeared to be more affected than the motor nerves. We found no correlation between nerve conduction slowing and disease duration which was also reported by other studies [1, 7, 16]. Interestingly, the FC703 patient showed electrophysiological and pathological evidence of intermediate type with variable degrees of axonal loss.

This study is the first report of Korean CMT4C families, and it will be helpful for molecular diagnoses of demyelinating and intermediate AR-CMT due to *SH3TC2* mutations.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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References

- Senderek J, Bergmann C, Stendel C, Kirfel J, Verpoorten N, De Jonghe P, et al. Mutations in a gene encoding a novel SH3/TPR domain protein cause autosomal recessive Charcot–Marie–Tooth type 4C neuropathy. Am J Hum Genet. 2003;73:1106–19.
- Gooding R, Colomer J, King R, Angelicheva D, Marns L, Parman Y, et al. A novel Gypsy founder mutation, pArg1109X in the CMT4C gene, causes variable peripheral neuropathy phenotypes. J Med Genet. 2005;42:e69.
- Azzedine H, Ravise N, Verny C, Gabreels-Festen A, Lammens M, Grid D, et al. Spine deformities in Charcot–Marie–Tooth 4C caused by SH3TC2 gene mutations. Neurology . 2006;67:602–6.
- Piscosquito G, Saveri P, Magri S, Ciano C, Gandioli C, Morbin M, et al. Screening for SH3TC2 gene mutations in a series of

demyelinating recessive Charcot–Marie–Tooth disease (CMT4). J Peripher Nerv Syst. 2016;21:142–9.

- Rudnik-Schöneborn S, Tölle D, Senderek J, Eggermann K, Elbracht M, Kornak U, et al. Diagnostic algorithms in Charcot–Marie–Tooth neuropathies: experiences from a German genetic laboratory on the basis of 1206 index patients. Clin Genet. 2016;89:34–43.
- Yuan JH, Hashiguchi A, Okamoto Y, Yoshimura A, Ando M, Shiomi K, et al. Clinical and mutational spectrum of Japanese patients with recessive variants in *SH3TC2*. J Hum Genet. 2018;63:281–7.
- Yger M, Stojkovic T, Tardieu S, Maisonobe T, Brice A, Echaniz-Laguna A, et al. Characteristics of clinical and electrophysiological pattern of Charcot–Marie–Tooth 4C. J Peripher Nerv Syst. 2012;17:112–22.
- Arnaud E, Zenker J, de Preux Charles A-S, Stendel C, Roos A, Medard J-J, et al. SH3TC2/KIAA1985 protein is required for proper myelination and the integrity of the node of Ranvier in the peripheral nervous system. Proc Natl Acad Sci USA. 2009;106:17528–33.
- Roberts RC, Peden AA, Buss F, Bright NA, Latouche M, Reilly MM, et al. Mistargeting of *SH3TC2* away from the recycling endosome causes Charcot–Marie–Tooth disease type 4C. Hum Mol Genet. 2010;19:1009–18.
- Laššuthová P, Mazanec R, Vondráček P, Sišková D, Haberlová J, Sabová J, et al. High frequency of *SH3TC2* mutations in Czech HMSN I patients. Clin Genet. 2011;80:334–45.
- Kim SJ, Nam SH, Kanwal S, Nam DE, Yoo DH, Chae JH, et al. BAG3 mutation in a patient with atypical phenotypes of myofibrillar myopathy and Charcot–Marie–Tooth disease. Genes. Genomics . 2018;40:1269–77.
- Nam SH, Hong YB, Hyun YS, Nam DE, Kwak G, Hwang SH, et al. Identification of genetic causes of inherited peripheral neuropathies by targeted gene panel sequencing. Mol Cells. 2016;39:382–8.
- Nam DE, Yoo DH, Choi SS, Choi BO, Chung KW. Wide phenotypic spectrum in axonal Charcot–Marie–Tooth neuropathy type 2 patients with KIF5A mutations. Genes Genom. 2018;40:77–84.
- Cipriani S, Phan V, Médard JJ, Horvath R, Lochmüller H, Chrast R, et al. Neuromuscular junction changes in a mouse model of Charcot–Marie–Tooth disease type 4C. Int J Mol Sci. 2018;19:pii: E4072.
- DiVincenzo C, Elzinga CD, Medeiros AC, Karbassi I, Jones JR, Evans MC, et al. The allelic spectrum of Charcot–Marie–Tooth disease in over 17,000 individuals with neuropathy. Mol Genet Genom Med. 2014;2:522–9.
- Murphy SM, Herrmann DN, McDermott MP, Scherer SS, Shy ME, Reilly MM, et al. Reliability of the CMT neuropathy score (second version) in Charcot–Marie–Tooth disease. J Peripher Nerv Syst. 2011;16:191–8.