Chromosomal microarray analysis in clinical evaluation of neurodevelopmental disorders-reporting a novel deletion of *SETDB1* and illustration of counseling challenge

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BACKGROUND: The pathogenicity of copy number variations (CNV) in neurodevelopmental disorders is supported by research literature. However, few studies have evaluated the utility and counseling challenges of CNV analysis in clinic.

METHODS: We analyzed the findings of CNV studies from a cohort referred for genetics evaluation of autism spectrum disorders (ASD), developmental disability (DD), and intellectual disability (ID).

RESULTS: Twenty-two CNV in 21 out of 115 probands are considered to be pathogenic (18.3%). Five CNV are likely pathogenic and 22 CNV are variants of unknown significance (VUS). We have found seven cases with more than two CNV and two with a complex rearrangement of the 22q13.3 Phelan-McDermid syndrome region. We identified a new and *de novo* 1q21.3 deletion that encompasses *SETDB1*, a gene encoding methylates histone H3 on lysine-9 (H3K9) methyltransferase, in a case with ASD.

CONCLUSION: We provide evidence to support the value of CNV analysis in etiological evaluation of neurodevelopmental disorders in autism genetics clinic. However, interpretation of the clinical significance and counseling families are still challenging because of the variable penetrance and pleiotropic expressivity of CNV. In addition, the identification of a 1q21.3 deletion encompassing *SETDB1* provides further support for the role of chromatin modifiers in the etiology of ASD.

The discovery of copy number variations (CNV) in patients with various neurodevelopmental disorders, including autism spectrum disorders (ASD), developmental disability (DD), and intellectual disability (ID), has supported the use of CNV analysis in clinical genetics practice (1,2). Endorsed by the American College of Medical Genetics and the American Academy of Pediatrics (3), CNV analysis using array-based comparative genomic hybridization (Array-CGH) or chromosomal microarray analysis (CMA) is now routinely performed in clinical genetics laboratories. A consensus statement,

published in 2010, recommended CMA as the first-tier clinical diagnostic test for individuals with DD and/or congenital anomalies (4). Evidence accumulated in the research literature has strongly supported the pathogenic role of rare and *de novo* CNV in ASD, DD, and ID (5,6). Numerous large-scale research studies have identified CNV in various genomic loci in patients with neurodevelopmental disorders, primarily ASD (7,8). However, for the majority of CNV reported in the research literature, the calling for pathogenicity of a given CNV is determined by a statistical comparison of its frequency in cases and controls. Few studies have assessed the clinical utility and counseling challenges of genome wide CNV analysis in the clinic. In this study, we report the yield and specific findings of CNV analysis in 115 patients referred to the Duke Autism Genetics Clinic for clinical genetics evaluation of ASD and DD/ID. Most importantly, we report a novel deletion of chromosome 1q21.3 encompassing the H3K9 methyltransferase *SETDB1* gene, and two cases (#37 and #38) of complex chromosomal rearrangements involving the Phelan-McDermid syndrome (PMS) region, located at chromosome 22q13.3, which is associated with ASD and ID.

RESULTS

Study Subjects

CMA was performed in 115 out of 165 cases (69.7%) referred for genetics evaluation of ASD/DD/ID (Figure 1). The reasons varied for why CMA was not performed for 50 of the cases. The lack of insurance coverage and parental consent were two major issues encountered. CMA was performed in 83 males and 32 females with male to female ratio of 2.6 to 1. The mean age at the time of evaluation was 5.7 year, ranging from 18 mo to 15.1 year with a 95% confidence interval of 5–6.3 year. There were 66 cases (66/115, 57.4 %) with a confirmed primary diagnosis of ASD and 49 cases (49/115, 42.6%) with the primary diagnosis of DD or ID at the time referred for clinical genetics evaluation. The clinical evidence to support these diagnoses varied. DSM-IV, DSM-5, ADOS, and ADI-R were used to

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Figure 1. The summary of genetic evaluation of 165 cases with autism spectrum disorders, intellectual disability, and developmental disability in genetics clinic.

support the diagnosis of ASD. IQ and DQ scores were used for diagnosis of ID and DD respectively. The referral providers include general pediatrician (33%), developmental pediatrician (41%), neurologists (24.8%), others including child psychiatrists and clinical psychologists (1.2%).

Basic Genetic Evaluation

Prior to referral to the Autism Genetics clinic, the primary care providers or other subspecialty providers had performed a basic genetic evaluation for some cases. These included chromosome analysis, and basic biochemical screening for disorders related to inborn errors of metabolism. In the Autism Genetics Clinic, molecular testing for fragile X syndrome was recommended for all cases and completed in >93% of cases. Molecular testing for Rett syndrome and tuberous sclerosis complex (TSC) was requested if the clinical presentations suggested these diagnoses. We have confirmed two cases of fragile X syndrome (2/165, 1.2%), three cases of TSC (3/165, 1.8%), and one case (1/165, 0.6%) of Rett syndrome (Figure 2) (Table 1).

CNV

Copy number variant analysis was performed in clinical laboratories using three different array platforms. The BlueGenome CytoChip v2 comparative genomic hybridization array was used in 18 cases (15.7%), the Affymetrix v6.0 single-nucleotide polymorphism array was used in 69 cases (60%), and the Affymetrix Cytoscan HD array was used in 28 cases (24.3%). These analyses reported a total of 49 pathogenic or potentially clinically relevant CNV in 39 out of 115 patients (Figure 1). CNV considered to be not clinically relevant, or benign, were not reported by clinical laboratories. Thirty-two cases had a single CNV (20 deletions and

12 duplications), and 7 patients had more than one CNV (4 cases with 2 CNV and 3 cases with 3 CNV). Parental tests, including both father and mother, were performed for 33 of 49 CNV. Six CNV (6/33, 18.2%) were *de novo*, and 27 CNV (27/33, 82.8%) were inherited. Seven CNV (7/49, 14.3%) were detected in the mother but the father was not tested, and parents were not available to be tested for nine CNV. The clinical relevance of the 49 CNV identified was assessed first by the clinical laboratories based on the size, inheritance, gene content, and previous clinical and research reports in published literature. We performed an additional assessment based on further literature review, genome and gene function annotation following the recommendations by others in literature (9,10). In some cases, the further research studies in extended family members were conducted. We categorized the CNV as “pathogenic” if the same CNV had been previously implicated in a known deletion or duplication syndrome, or a similar disease phenotype with good evidence in literature. For CNV that are rare, novel, and likely disease causing, we defined them as “likely pathogenic” based on assessment of size of CNV, inheritance pattern, and functional annotation of the genes within the deleted or duplicated interval of CNV. CNV lacking sufficient evidence to support their pathogenicity were classified as “Variants of Unknown Significance” (VUS). We concluded that 22 CNV (17 single CNV and 5 CNV found in 4 probands with more than one CNV) are “pathogenic” because the same CNV has been associated with ASD, ID, and/or DD in previous reports in literature. Five CNV (five single CNV) were “likely pathogenic” and 22 CNV were “VUS” (Table 1). Because there is evidence in literature supporting that the 15q11.2-q13.1 duplication, identified in #33, can cause mild ASD and ID (11,12), we classified it as pathogenic (see detail discussion

Figure 2. A patient (case #18) with a 260-kb deletion at 1q21.3 that disrupts the *SETDB1* gene. (a) A facial profile of patient. Noted for slightly deep set of eyes, mild hypotelorism, small chin, and short neck. (b) A local view of the 17p13.3 deletion from single-nucleotide polymorphism (SNP) array. (c) The diagram of deletions of *SETDB1* and other genes in 260kb deleted interval. (d) Fluorescence *in situ* hybridization confirmation for the deletion in proband but absence in both parents.

below). However, the pathogenicity of this duplication is not certain in this family. Other genetic defects may also contribute to the clinical presentations of both siblings.

Cases With Single CNV

Pathogenic CNV. The single CNV found in 17 probands were considered to be pathogenic (Table 1). These CNV included the maternal deletions of 15q11-q13, and deletions of 1p36.2, 16p11.2, 2q32.1, 3q29, and 22q11.3 that are frequently reported in the literature. For the CNV that are known to be pathogenic and have a known parental origin, 9 out of 13 (69.2%) were inherited from reported healthy and normal parents. This is higher than that of inherited CNV reported in other research studies in literature (13,14) although this may also be biased because parental testing was not always done in other cases.

Likely pathogenic CNV. Five cases have single CNV that are likely pathogenic including copy number loss in chromosomal regions 1q21.3 and 16p12.1 and copy number gain in 5q12.1-q12.3, 7q31.1, and 15q26.3. The 1q21.3 deletion is *de novo* but the others are inherited. These CNV have neither been reported to associate with ASD or ID consistently nor are present in a large number of controls. A *de novo* 260 Kb deletion of 1q21.3 was found in an 11-year-old Caucasian male with ASD, seizures, developmental delay, and mild facial dysmorphisms including small chin and flat nasal bridge, and flat feet (case #18, Figure 2). This CNV has not been previously reported in the literature. There are 10 genes (*ARNT*, *SETDB1*, *CERS2*, *ANXA9*, *FAM63A*, *PRUNE*, *CDC42SE1*, *BDNF*, *MLLT11*, *C1orf56*) mapped in the deleted interval. *CERS2* encodes a ceramide synthase (15) and *SETDB1*

encodes a histone methyltransferase which methylates histone H3 on lysine-9 (H3K9) (16). The function of the other genes is either unrelated to brain function or poorly characterized. For the other three CNV gains in this category, there are case reports describing individuals with variable clinical presentations including ID and DD (17–20). However, the correlations or causal roles of these CNV with particular clinical phenotypes have not been firmly established. We also identified a maternally inherited duplication of 17p12, a known cause for Charcot-Marie-Tooth disease type 1A (CMT1A), in the case (#30) of 9-year-old boy with dysmorphic face, hypospadias, short stature, DD, and ID but no sign of neuropathy at the time of evaluation. The ASD, DD, and ID have not been reported in individuals with CMT1A in the literature. Although it is possible that this duplication may contribute to some clinical features in the proband, a more plausible interpretation is that this is simply an incidental finding.

VUS. Ten of the CNV identified are novel and rare, and their clinical significances are currently unknown. None of these CNV have been reported in the literature or in reference database. The size of these CNV is generally <2 Mb and the inheritance is either *de novo* or inherited from healthy parents. The functions of genes within these CNV are either unknown or not revealing based on the review of literature. Therefore, it is not possible to assign a causal role to these CNV at this point.

Cases With Multiple CNV or a Complex Genomic Rearrangement

Seven cases had CNV in more than one chromosomal region. Four cases had two CNV and three cases had three CNV (Table 1). Two cases with three CNV (#37 and #38) had copy

Table 1. CNV identified from individuals referred for clinical genetics evaluation in autism clinic

Single CNV										
Case	Chromosome location	Deletion/ duplication	Position	Size, kb	Assembly	Parent of origin	Primary diagnosis	Novel CNV	Clinical significance	Karotype
Pathogenic										
1	1p36.31	Deletion	4,901,696-10,067,420	5,165	hg18	Mother normal ^a	DD	–		
2	1p36.32	Deletion	4,075,966-11,812,055	7,700	hg18	<i>De novo</i>	DD	–	–	NA
3	2q32.1	Deletion	186,401,261-186,791,896	391	hg18	NA	DD	–	–	NA
4	3q29	Deletion	193,800,001-199,501,827	5,701	hg18	Maternal	DD	–	NA	–
5	14q12	Deletion	25,488,077-28,883,347	3,400	hg18	NA	ID	–	–	NA
6	15q11.2-q13.1	Deletion	20,085,783-26,714,565	6,600	hg18	Maternal	Angelman Syndrome	–	–	NA
7	15q13.2-q13.3	Deletion	RP11-38E12->RP11-164K24	900	hg17	Maternal	DD	–	–	NA
8	15q13.3	Deletion	30,073,735-32,446,830	2,370	hg19	Maternal	ASD	–		
9	15q26.1	Duplication	90,486,082-90,967,701	482	hg18	Maternal	ASD	–	–	NA
10	16p11.2	Deletion	29,425,200-30,085,308	660	hg19	<i>De novo</i>	DD	–	–	NA
11	16p13.11	Deletion	RP11-489O1	200	hg18	Maternal	DD	–	NA	NA
12	16p13.11	Deletion	14,899,719-16,508,304	1,600	hg19	Maternal	ASD	–	–	–
13	16p13.12	Deletion	12,664,828-12,877,151	212	hg17	NA	CHARGE Syndrome	–	–	NA
14	18q22.1	Deletion	62,098,937-62,364,101	265	hg18	Mother normal ^a	ASD	–	–	–
15	22q11.21	Deletion	17,173,935-19,941,337	2,700	hg18	<i>De novo</i>	ASD	–	–	–
16	22q11.21	Duplication	17,173,935-20,164,236	3,000	hg18	Mother normal ^a	ASD	–	–	NA
17	Xp11.4	Deletion	38,310,495-38,402,885	92	hg18	Maternal	DD	–	–	NA
Likely pathogenic										
18	1q21.3	Deletion	NA	260	hg19	<i>De novo</i>	ASD	+	–	–
19	5q12.1-q12.3	Duplication	61,718,070-64,507,614	2,800	hg18	Paternal	ASD	+	–	–
20	7q31.1	Duplication	109,999,318-110,819,951	821	hg19	Maternal	DD	–	–	NA
21	15q26.3	Duplication	97,226,481-98,882,636	1,600	hg18	Maternal	ASD	–	–	NA
22	16p12.1	Deletion	21,883,228-22,338,859	456	hg18	Paternal	ASD	–	–	NA
Variant of unknown significance (VUS)										
23	4q22.2	Duplication	94,645,324-95,366,771	721	hg18	Mother normal ^a	ASD	–	NA	NA
24	6p21.1	Deletion	44,968,645-45,091,828	123	hg18	Maternal	DD	–	–	NA
25	6p21.2	Duplication	RP3-350J21->RP1-278E11	1,000	hg17	Maternal	DD	–	–	–

Table 1. Continued on next page

Table 1. Continued

Case	Chromosome location	Deletion/duplication	Position	Size, kb	Assembly	Parent of origin	Primary diagnosis	Novel CNV	Clinical significance	FMR1	MECP2	Karotype
26	8p22	Duplication	13,400,616-14,700,476	1,299	hg19	NA	ASD	-	-	-	-	-
27	11q22.3	Deletion	103,313,942-103,367,161	53	hg19	NA	ASD	-	-	-	NA	NA
28	12q24.11	Duplication	109,501,323-109,603,748	100	hg18	Maternal	DD	-	-	-	-	-
29	13q12.11	Deletion	19,695,316-20,002,412	307	hg18	Maternal	DD	-	-	NA	NA	NA
30	17p12 ^b	Duplication	14,035,412-15,423,228	1,380	hg18	Maternal	DD	-	-	-	NA	NA
31	18q12.2	Duplication	32,161,410-32,635,174	474	hg18	NA	DD	+	-	-	NA	NA
32	Xq21.31	Duplication	RP11-192B18	2,700	hg17	Maternal	ASD	-	-	-	NA	-
Multiple CNV												
33	5q35.3; 15q11.2-q13.1	Deletion; duplication	179,815,908-26,234, 399,180,459,915; 21,192,943	644; 5,041	hg17	De novo; paternal	ASD	-/-	Unclear; pathogenic ^c	-	-	-
34	1q21.1; 1q21.1	Duplication; deletion	144,083,908-144,503,409; 144,643,813-146,297,795	420; 1,600	hg18	Paternal; De novo	DD	+/+	Unclear; pathogenic	-	NA	NA
35	9p21.3-p21.2; Yp11.2	Deletion; deletion	24,863,950-26,314,013; 6,460,193-7,035,072	1,400; 575	hg18	Maternal; paternal	DD	-/-	Unclear; unclear	NA	NA	NA
36	16q24.3; 21q22.13	Duplication; deletion	88,195,201-88,504,477; 37,291,678-37,390,702	309; 99	hg18	Paternal; paternal	ASD	-/-	Unclear; unclear	-	NA	N/A
37	1p31.1; 16p13.2; 22q13.33	Deletion; duplication; deletion	75,731,050-76,402,593; 8,753,360-9,202,843; 49,320,608-49,581,309	672; 449; 260	hg18	Maternal; mother normal; mother normal	Phelan- McDermid syndrome	-/-; -/-	Unclear; pathogenic; pathogenic	-	-	-
38	22q13.31; 22q13.31; 22q13.32-q13.33	Duplication; deletion; deletion	44,744,569-45,629,066; 45,629,066-46,956,456; 49,059,493-51,197,838	884; 1,330; 2,100	hg19	NA; NA; NA	Phelan- McDermid syndrome	-/-; -/-	Unclear; unclear; Pathogenic	-	NA	NA
39	4q28.1-q28; 12p13.33 Xq28	Deletion; duplication; duplication	126,784,753-127,148,514; 733,738,880,267; 153,444,055-154,533,675	363; 156; 1,089	hg18	Paternal; maternal; maternal	ASD	-/-; -/-	Unclear; unclear; Unclear	-	NA	NA

^aNot present in mother but father was not available for study. ^bThe CNV gain is the known cause for the Charcot-Marie-Tooth type 1A. This is likely an incidental finding. ^cThe paternal duplication has been implicated in individuals with mild or moderate ASD and ID in literature. See discussion for the clinical relevance in this case in text. ASD, autism spectrum disorders; CNV, copy number variations; ID, intellectual disability; NA, not available; -/-, negative; +, positive.

number loss of the PMS region located at 22q13.3. In addition to this, CNV in the PMS region, case #37, has a copy number loss at 1p31.1 and a copy number gain at 16p13.2. In the 22q13.3 PMS region of this patient, a 260kb deletion began at linear position 49,320,608 and ended at 49,581,309 (www.genome.ucsc.edu; hg19). This interval overlaps 12 known

genes including *SHANK3*, a causative gene for ASD and a key player in PMS (21,22). The proband's clinical features include profound speech delay, motor development delay, severe intellectual disability, repetitive behaviors, significant self-injurious behavior, and aggression. Interestingly, her social behavior and social communication appear much less affected compared to other individuals with PMS. In the 1p31.1 region, a 672 kb maternally inherited loss was identified that includes 11 known or predicted genes, 4 of which, *ACADM*, *RABGGTB*, *MSH4*, and *ST6GALNAC3*, are listed in OMIM. In the 16p13.2 region, a 449 kb copy number gain was found that includes 10 known or predicted genes, including *ABAT*, *PMM2*, and *USP7* which are listed in OMIM. Chromosome analysis and fluorescence *in situ* hybridization analysis showed no evidence of an unbalanced chromosomal translocation. The deletion at 1p31.1 was confirmed to be present in the mother; however, the duplication of 16p13.2 and the deletion of 22q13.3 were not. Unfortunately, the father was not available to determine the inheritance of the CNV in 22q13.3 and 16p13.2. Case #38 was a 13-year-old boy with severe ASD, nonverbal, seizures, and global developmental delay. CNV analysis revealed a complex rearrangement involving the 22q13.3 PMS region as diagrammed in Figure 3. A proximal interstitial duplication of 884kb in the 22q13.31 region (44,744,569 to 45,629,066) is adjacent to an interstitial deletion of at least 1.3 Mb in the 22q13.31 region (45,629,259 to 46,956,456), followed by a

Figure 3. A diagram of complex chromosomal rearrangements in the 22q13 Phelan-McDermid region in case #38. The position for each CNV and genes disrupted in deleted or duplicated interval is diagrammed. *SHANK3*, the key gene for Phelan-McDermid syndrome, is highlighted.

Figure 4. A patient (case #33) with CNV of 5q35.3 deletion and the 15q11-q13 duplication. (a) A three generation pedigree. The proband has severe end of ASD and young brother has moderate end of ASD. The proband carries a *de novo* 640kb deletion at 5q35.3 region (red bar) and paternally inherited 5 Mb duplication (blue bar) of the 15q11-q13 region. Sibling brother, sister, mother, paternal grandmother and paternal grandfather do not carry either CNV. (b) Local view of 5q35.3 deletion by CMA. (c) Local view of 15q11-q13 duplication by CMA. (d) Interphase fluorescence *in situ* hybridization (FISH) of the proband shows the duplication of 15q11-q13 probe. Magnification is 100x. (e) Metaphase FISH of the proband shows the duplication of 15q11-q13. Magnification is 100x. (f) Metaphase FISH of father shows the same duplication of 15q11-q13. Magnification is 100x.

Figure 5. The proposed practice algorithm for interpreting copy number variations in clinic.

region of normal copy number and then a terminal deletion of at least 2.1 Mb of the 22q13 region (49,059,493 to 51,197,838) (hg19). The terminal deletion includes the *SHANK3* gene. These rearrangements were *de novo* and similar rearrangements have not been reported in the literature.

One of the cases with two CNV (#37) has a copy number gain in the 15q11-q13 Angelman syndrome (AS) and Prader-Willi syndrome region (PWS). The proband and his male sibling were affected by ASD (Figure 4). The older brother was more severely affected than the younger sibling. The older sibling carries a 5 Mb interstitial duplication of the 15q11.2-q13.1 AS and PWS region that was inherited from his healthy and normal father. The proband's younger affected brother and unaffected sister did not carry this duplication. This duplication was most likely a *de novo* event in the father because the paternal grandparents did not carry the same duplication in blood leukocyte DNA although the possibility of germline mosaicism could not be ruled out. The older sibling also carries a *de novo* 644 kb interstitial deletion of 5q35.3. Five OMIM genes including *CNOT6*, *SCGB3A1*, *FLT4*, *MGAT1*, and *BTNL3* were mapped in the deleted interval of 5q35.3 region. *MGAT1* is known to be involved in brain function by encoding a N-acetylglucosaminyltransferase, an enzyme for the glycosylation of proteins (1). However, screening for a glycosylation disorder by carbohydrate-deficient transferring analysis was unremarkable. Maternal duplication of 15q11-q13 is strongly associated with ASD while the pathogenic role of the same duplication of paternal origin in ASD and ID has not been

firmly established (11,12,23,24). The deletion of 5q35.3 has not been reported in literature to be associated with any disease phenotype.

DISCUSSION

Clinical benefit of CNV analysis

Based on the results of CNV analysis, performed in clinical laboratories, and additional research analysis for some cases, we detected "pathogenic" CNV in 21 out 115 individuals (18.3%) with ASD and/or ID and DD referred to our Autism Genetics Clinic (20 cases with single and 1 case with two CNV). Our detection rate of pathogenic CNV is slightly higher than that of 10–15% reported in other research studies (4,13). The difference is likely due to ascertainment bias of our patient cohort as it is possible that the patients referred to genetics clinic are probably more severely affected. A significant number of cases in which CNV are considered to be pathogenic and parental origins are known in our cohort were inherited (9/13, 46.1%). The frequency of inherited CNV appears higher than expected. Unfortunately, the parental origins of CNV are not fully investigated in many research studies of genome-wide CNV analysis for ASD in the literature (8,25). Most of the parents carrying the CNV identified in their affected children do not have significant medical or mental health problems based on review of available medical records although careful clinical and neuropsychiatric evaluation have not been conducted. Therefore, counseling for these CNV is challenging even if the evidence supporting the pathogenicity of these CNV is strong in the

research literature. In research studies, the “pathogenicity” of a given CNV is typically made based on the statistical value from population-based case and control studies. Translation of these research findings into the clinic and counseling families is clearly not straightforward if the CNV are inherited from healthy parents.

A Novel CNV Containing Histone Methyltransferase Associated With ASD and ID

The finding of a *de novo* CNV loss in the 1q21.3 region is novel and has not been reported in the literature. There are 10 genes (*ARNT*, *SETDB1*, *CERS2*, *ANXA9*, *FAM63A*, *PRUNE*, *CDC42SE1*, *BNIP1*, *MLLT11*, *C1orf56*) mapped in the deleted interval. However, the functions for these genes, except *CERS2* and *SETDB1*, are either not related to brain function or have not been characterized. *CERS2* encodes ceramide synthase, and ceramides form the lipid backbone of all complex sphingolipids. *Cers2* homozygous mutant mice have a short life span, liver dysfunction, and generalized and symmetrical myoclonic jerks (15). The defects in ceramide metabolism are implicated in a list of neurometabolic diseases. However, these diseases exclusively follow autosomal recessive inheritance in humans and the clinical relevance of the haploinsufficiency of *CERS2* is probably not significant unless there is a pathogenic mutation in other allele that is missed. On the other hand, deletion of *SETDB1*, a histone methyltransferase, is particularly interesting. *SETDB1* is a promising candidate gene for ASD/ID because of its role in epigenetic modification and brain function (26). In mice, *Setdb1* deficiency modifies the neuronal phenotypes caused by deficiency of the Rett syndrome protein MECP2 (27) and regulates the expression of NMDA receptor 2B (28), a gene implicated in cognitive function and epilepsy. Functional studies *in vitro* strongly support a role for *Setdb1* in synaptic development and function (27,28). In humans, an in-frame 3 bp deletion in *SETDB1* was reported in an individual with ASD (29). Furthermore, *de novo* loss of function (LOF) or likely gene-disrupting mutations (LGD) in a list of genes encoding chromatin modifier proteins have been identified in individuals with ASD from several large-scale whole-exome sequencing studies (30). Although pathogenic mutations in *SETDB1* have not been identified in these studies, the LOF and LGD mutations in genes regulated by *SETDB1* are frequently found in ASD individuals. These data provide evidence supporting the pathogenicity of haploinsufficiency of *SETDB1* in ASD and ID.

Clinical Relevance of Multiple CNV in Clinic

We found seven families (6.1%) with a complex pattern of multiple CNV, illustrating the challenges related to interpretation of the clinical relevance of CNV. For example, our case #33 that has an inherited paternal duplication of 15q11.2-q13.1 and *de novo* deletion of 5q35.3 in one but not the other affected sibling is particularly interesting. While maternal duplication of 15q11-q13 has been consistently implicated in severe ASD and ID (31,32), the clinical relevance of paternal duplication of 15q11-q13 is much less clear and has been

associated with a wide range of clinical presentations from normal to mild or moderate ASD and ID (31,33). In this family, the healthy father carried a *de novo* 15q11.2-q13.1 duplication that presumably derived from the paternal chromosome 15. It is difficult to establish that the paternal duplication of 15q11.2-q13.1 is the cause of ASD for the severely affected sibling based on the available data in the literature. However, the presence of the *de novo* 5q35.3 deletion may support a two hit model as suggested by others (34,35); the combination of the paternal duplication of 15q11.2-q13.1 and 5q35.3 deletion may be responsible for the symptoms in the more severely affected sibling. The 5q35.3 deletion contains *MGAT1*, a gene encoding an N-acetylglucosaminyltransferase I and involving protein glycosylation. Although *MGAT1* has not been associated with any genetic disease, deficiency of many other enzymes required for glycosylation cause a group of rare disorders, congenital disorders of glycosylation (CDG), in which developmental delay is an almost universal feature. Because all known CDG follow an autosomal recessive inheritance pattern, the significance of haploinsufficiency of *MGAT1* is not clear. However, in a two hit model, the plausible hypothesis is that the deletion of 5q35.3 may modify the paternal duplication of 15q11-q13 and contribute to the more severe clinical presentation of the older sibling. Because of the rarity of these CNV, it is challenging, if not impossible, to test this hypothesis directly in humans. Alternatively, both CNV of 15q11.2-q13.1 and 5q35.3 are not pathogenic, and other genetic defects, such as point mutations in another gene or genes, or epigenetic factors may be responsible for the clinical presentation in both siblings.

The two cases with three CNV, that both involve 22q13.3 PMS region, are also interesting. In the case #37, the 1p31.1 deletion is inherited from the unaffected mother. The inheritance patterns for 16p13.2 and 22q13.3 could not be determined because the father was unavailable. The deletion of 22q13.3 is a definitive cause for ASD and deletion of the *SHANK3* gene specifically is a major cause of the Phelan-McDermid syndrome. The *de novo* duplication of 16p13.2 has been associated with ASD in a recent report (36) and also found rarely in a control population (37). The patient's overall clinical presentation is consistent with PMS. Surprisingly, the patient's social interaction and communication are quite intact. Because ASD and impaired social interaction and communication are prominent features associated with PMS (38–40), this raised an interesting question whether her unique clinical phenotypes are modified by CNV of 1p13.3 and 16p13.2. The deletion of 1p13.1 was inherited from the patient's unaffected mother and has not been associated with any disease phenotype; therefore, it is less likely that this deletion will have a major role in her clinical presentation. An interesting question is whether the combination of 22q13.3 deletion and 16p13.2 duplication are responsible for her unique behavioral features, which are not apparently more severe than in cases carrying only one of these CNV. Alternatively, it can be hypothesized that the 1p31.1 deletion may modify the contribution of the deletion of 22q13.3 and the duplication of 16p13.2 to her presentation, or a more

provocative question is whether CNV of 16p13.2 and 1p31.1 may function as a protective allele to *SHANK3* deficiency in 22q13.3 CNV.

A Practical Algorithm and the Challenge of Interpreting and Counseling CNV Findings in Clinic

The clinical benefit and technical merit of using genome-wide CNV analysis to detect many well-defined deletion and duplication syndromes are clear in both research and clinical literature (2,10). However, there are few studies that systematically address the challenges of interpreting the CNV results and the accompanying counseling issues including the psychosocial and economic impact to families. For newly-identified CNV associated with ASD and ID, the evidence supporting their pathogenicity is mostly drawn from population-based studies. In the population-based research studies, while variable penetrance and pleotropic expressivity have been frequently observed and extensively discussed, clinical decision-making for individual cases is usually not addressed during the research phase of the studies. However, the situation is different for clinical CNV analysis because physicians will have to interpret the findings and counsel the family accordingly and in a timely manner. For this reason, we propose a practice algorithm for interpreting CNV findings in clinic (Figure 5). The counseling and classification of pathogenic CNV that are known to cause well defined deletion or duplication syndromes such as Angelman and Phelan-McDermid syndromes are straight forward. However, for the examples of 16p11.2 deletion, 15q11-q13 paternal duplication, and 15q13.3 deletion, the clinical decision making and counseling was challenging. Evidence from research literature clearly supports the classification of these CNV to be pathogenic. However, because these CNV can be inherited from an apparently healthy parent or *de novo*, and have variable penetrance or pleotropic effects, the interpretation of pathogenicity and counseling in the family with a severely affected proband and CNV inherited from a healthy parent is challenging. The counseling in prenatal setting is even more complex. This is best illustrated in case #10 with the deletion of 16p11.2 (ref. (41)). Because the deletion of 16p11.2 has been also found in healthy individuals in low frequency, the prenatal counseling for this family has been challenging. As illustrated in our findings from a small number of cases in this cohort, the clinical CNV analysis will discover a majority of CNV that are rare and novel but they fall into the second or third categories as described above. Families may have to deal with the uncertainty or psychological stress of being informed that their child has a CNV of unknown clinical relevance or the result is difficult to interpret. In other cases, there may also be an economic impact. Unfortunately, these questions have not been addressed systematically despite CNV analysis having been performed in clinics routinely for about a decade. A more comprehensive study is clearly warranted in this direction including assessing the long-term psychosocial impact of this testing to the families and patients.

Conclusion

Our data of CNV analyses from a cohort of 115 individuals with ASD/ID/DD referred to the Autism Genetics Clinic support the clinical utilization of CNV analysis. The overall positive rate for pathogenic CNV in this cohort is 18.3% but the clinical relevance of a significant number of CNV remains unclear. Our novel finding of deletion of *SETDB1* in a patient with classical autism provides the further support for the role of chromatin modifiers in the etiology of ASD. Our finding of more than one rare and novel CNV in individuals adds to the complexity for interpreting the clinical relevance of these CNV. Despite the strong evidence supporting the pathogenicity of many CNV from research studies, the clinical significance of these CNV, and counseling about these findings in clinics remain challenging because of variable penetrance and pleotropic effects. Further studies may be warranted to fully evaluate medical, social, psychological, and economical benefits related to CNV analysis in the clinical evaluation of neurodevelopmental disorders.

METHODS

Study Subjects and Case Review

A total of 165 patients carrying a diagnosis of ASD and/or ID and DD were recruited consecutively from the Autism Genetics Clinic at Duke Children's Hospital from 2010 to 2014. Only the medical records for the cases with positive CMA were then reviewed. The age of these patients ranged from 18 mo to 15.1 year. The diagnosis of ASD/DD/ID was made by each patient's referring clinician. Additional study was conducted for selected cases. The study was approved by the Duke University Health System Institutional Review Board. Written informed consent was obtained from the patients for publication of their individual details and accompanying images in this manuscript. The consent form is held by Y.H.J. and available at Duke University.

CMA or Array-Based Comparative Genomic Hybridization (Array-CGH)

CMA or Array-CGH was performed in one of the following clinical laboratories: the Duke University Cytogenetics Laboratory (Durham, NC), the Baylor College of Medicine Molecular Genetics Laboratory (Houston, TX), or LabCorp genetics laboratory (Research Triangle Park, NC). The following array platforms were used: the Duke University Cytogenetics Laboratory, BlueGnome CytoChip v2 (bacterial artificial chromosome array) (Illumina, San Diego, CA), the Duke University Cytogenetics Laboratory and LabCorp, Affymetrix human genome-wide SNP 6.0 array (Affymetrix, Santa Clara, CA), and the Baylor College of Medicine Molecular Genetics Laboratory Affymetrix Cytoscan HD array (42,43). The BlueGnome CytoChip v2 consists of ~4,400 bacterial artificial chromosome clones representing greater than 90 clinically defined syndrome loci and all 41 unique subtelomeric regions. The Affymetrix human genome-wide SNP 6.0 array consists of 1.8 million genetic markers at a median inter-marker spacing of less than 700 nucleotides. Approximately half of these markers provide SNP genotype calls in addition to copy number state information. The Affymetrix Cytoscan HD array consists of nearly 2.7 million genetic markers incorporating 743,304 SNP probes as well as 1,953,246 nonpolymorphic CNV probes. For the array hybridization, 500 ng of genomic DNA isolated from peripheral blood was enzymatically digested, amplified, and purified. Purified products were fragmented, labeled, and hybridized to the array. After washing (Affymetrix Fluidics Station 450), arrays were scanned (Affymetrix GeneChip Scanner 3000) and the analysis was performed using the Affymetrix Genotyping Console 3.0.2 software. CNV calling and interpretation were extracted from the reports from the individual clinical laboratories.

Fluorescence *In Situ* Hybridization

Fluorescence *in situ* hybridization analysis was performed using bacterial artificial chromosome clone RP11-11. as a probe for the 1q21.3 region, RP11-451H23 (BlueGenome) as a probe for the 5q35.3 region and the Abbott Molecular dual color probe for the Prader-Willi syndrome critical region (SNRPN) at 15q11-q13 and a distal locus for identification of chromosome 15.

AUTHOR CONTRIBUTIONS

Q.X. and Y.H.J. conceived and conducted the study; I.G., H.L., and C.R. contributed CNV analysis; W.P.P., A.M., and Y.H.J. contributed to review the medical records; Q.X., J.G., X.X., W.P.W., A.M., and Y.H.J. wrote the manuscript; all authors read and approved the final manuscript.

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