β_1 -Adrenoceptor mRNA level reveals distinctions between infantile hemangioma and vascular malformations

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BACKGROUND: Infantile hemangioma (IH) is the most frequent vascular tumor of early childhood. Recently, propranolol, a nonselective β_1 - and β_2 -adrenoceptor inhibitor, was introduced into the therapy of severe proliferating IH with excellent results. However, the underlying mechanism of action of propranolol is still unclear.

METHODS: We performed immunohistochemistry for cluster of differentiation 31 (CD31), D2-40, glucose transporter-1 (GLUT-1), and Ki67 in order to characterize 21 vascular anomalies (nine IH, seven venous malformations (VMs), and five lymphatic malformations (LMs)). Furthermore, we analyzed the expression of β_1 -, β_2 -, and β_3 -adrenoceptor mRNA in these specimens as well as in hemangioma-derived stem cells by quantitative real-time PCR (qPCR).

RESULTS: We show that the expression of β_1 -adrenoceptor mRNA is 10.7-fold higher in IH independent of the proliferative or regressive phase as well as 2.5-fold higher in hemangioma-derived stem cells as compared with β_2 -adrenoceptor mRNA. In LM, the expression of β_2 -adrenoceptor mRNA was ninefold higher than that of β_1 -adrenoceptor mRNA. VM showed low expression levels of all β -adrenoceptor mRNAs, and β_3 -adrenoceptor mRNA was hardly detectable in any specimens examined.

CONCLUSION: These results provide the first evidence of distinctions between IH and vascular malformations with regard to β -adrenoceptor subtype mRNA levels.

Recently, propranolol, a nonselective β -adrenoceptor inhibitor, was introduced into the therapy of severe proliferating infantile hemangioma (IH) (1). Propranolol treatment in IH results in fast growth inhibition and rapid tumor regression within a few days of therapy. This novel medical treatment has not been developed in a classic way; it was accidentally observed during corticosteroid therapy, the so far standard therapy for proliferating IH (2) when cardiac hypertrophy as a side effect is treated by propranolol. The molecular mechanism of propranolol on IH proliferation is unknown at present. A possible method of action could be vasoconstriction or induction of endothelial cell apoptosis (3). Inhibition of β -adrenoceptors has been suggested to have a negative impact on tumor development as well as a direct effect on endothelial cells by downregulation of angiogenesis factors such as vascular endothelial growth factor, matrix metalloproteinase-2, and matrix metalloproteinase-9 (4,5). A possible involvement of the β_2 -adrenoceptor/cyclic adenosine monophosphate pathway has been described in tumor vascularization of glioblastoma (4).

IH is the most frequent benign tumor of early childhood that shows specific biological features: after an initial phase of proliferation during the first year of life, a long-lasting period of spontaneous regression over several years is observed (6). The underlying molecular mechanism could not be completely determined because of a lack of models for IH. Until recently, neither IH cell lines nor animal models were available. Endothelial cells from IH show specific expression of glucose transporter-1 (GLUT-1) (7) and can be discriminated by this marker from endothelial cells of the vascular tumors and vascular malformations composed of veins, arteries, or lymphatic vessels. Furthermore, the clinical behavior, with lack of independent growth and regression in vascular malformations, clearly demonstrates a different biological background.

In this study, we studied the expression of β -adrenoceptor subtype mRNA in IH and vascular malformations, including venous malformations (VMs) and lymphatic malformations (LMs), to describe the molecular background of the new therapeutic approach with propranolol, a nonselective β -adrenoceptor inhibitor.

RESULTS

IHs of six children included in this study showed proliferation when surgery was performed. These children were younger than 1 y (**Table 1**). IH from three children older than 1 y had passed a period of regression. All endothelial cells from IH showed GLUT-1 expression by immunohistochemistry, which confirms the diagnosis (**Figure 1**, **Table 2**). The six proliferating IHs additionally showed 10–15% Ki67-positive endothelial cells, confirming the growth potential. In contrast, Ki67 expression was not detectable in the IHs from three older children.

The age of patients with surgically removed vascular malformations varied from 4 wk to 18 y. Localization was in the head and neck region as well as at the extremities, which is in accordance with the literature (8). All vascular malformations

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Patient	Vascular	Growth		Previous	Age at					
ID	anomaly	behavior	Localization	treatment	surgery					
1	IH	Proliferative	Retroauricular right	None	11 mo					
2	IH	Proliferative	Neck	None	5 mo					
3	IH	Proliferative	Occipital	None	5 mo					
4	IH	Proliferative	Face (glabella)	None	11 mo					
5	IH	Proliferative	Parietal	None	9 mo					
10	IH	Proliferative	Back	Laser therapy	8 mo					
6	IH	Regression	Right mandible	Cryotherapy	5.5 y					
7	IH	Regression	Right thoracic wall	Laser therapy	3.5 y					
13	IH	Regression	Parietal	None	2.5 y					
9	VM	Unchanged	Left thoracic wall	None	12 y					
11	VM	Unchanged	Left knee	None	5 y					
12	VM	Unchanged	Right cheek	None	11 y					
14	VM	Unchanged	Right thigh	None	10 mo					
15	VM	Unchanged	Left knee	None	12 y					
16	VM	Unchanged	Intra-abdominal	None	3 wk					
17	VM	Unchanged	Lower lip	None	17 y					
18	LM	Unchanged	Right axilla	None	10 mo					
19	LM	Unchanged	Right shoulder	None	18 y					
20	LM	Unchanged	Inguinal right	None	10 y					
21	LM	Unchanged	Left axilla	None	16 y					
22	LM	Unchanged	Cervical right	None	4 mo					

Table 1. Clinical characteristics of patients

IH: n = 9, VM: n = 7, and LM: n = 5.

IH, infantile hemangioma; LM, lymphatic malformation; VM, venous malformations.

showed cluster of differentiation 31 (CD31)–positive immunostaining of endothelial cells and all were GLUT-1 negative (**Figure 2, Table 2**). To discriminate between vascular malformations of blood and/or lymphatic vessels, D2-40 expression was determined and the result confirmed the clinical and radiological diagnosis identifying five LMs, which were D2-40 positive, and seven VMs.

Expression of β-adrenoceptor subtype mRNA was analyzed with quantitative real-time PCR (qPCR) and normalized to expression of ribosomal protein RPS29 mRNA (Figure 3). Overall, β-adrenoceptor mRNA density was one order of magnitude higher in IH and LM samples as compared with VM samples. IH showed 10.7-fold higher expression of β_1 as compared with β_2 -adrenoceptor mRNA (Figure 3a). β_2 -Adrenoceptor mRNA could be detected in all nine IH specimens, although at levels that were three orders of magnitude lower than β_1 -levels. β_1 -Adrenoceptor mRNA concentrations did not differ significantly between proliferative and nonproliferative IHs (proliferative 45 \pm 10 vs. nonproliferative 90 \pm 41 ADRB1 copies/1,000 RPS29 copies, P = 0.18). In VM specimens, mRNA levels for β_1 - and β_2 -adrenoceptors did not differ significantly (Figure 3b). In LM specimens, β_2 -adrenoceptor mRNA expression was ninefold higher than β_1 -adrenoceptor



Figure 1. Characterization of IH. (**a**) H&E staining (**a**) and immunohistochemistry for (**b**) GLUT-1 and (**c**) Ki67 in a proliferative IH as well as (**d**) H&E staining and immunohistochemistry for (**e**) GLUT-1 and (**f**) Ki67 in IH after spontaneous regression (original magnification ×10). Proliferative IH of patient ID 10 showing (**a**) multiple capillaries with (**b**) strong expression of GLUT-1 and (**c**) 10–15% Ki67-positive endothelial cells. The IH of patient ID 6 after spontaneous regression shows vessels with (**d**) bigger lumen, (**e**) GLUT-1 positive immunostaining, and (**f**) 1% Ki67-positive endothelial cells. GLUT-1, glucose transporter-1; H&E, hematoxylin and eosin; IH, infantile hemangioma.

mRNA expression (Figure 3c). In all specimens examined, expression of β_3 -adrenoceptor mRNA was very low as compared with β_1 - and β_2 -subtypes.

Finally, we had the opportunity to study β -adrenoceptor mRNA expression in hemangioma-derived stem cells (9). Again, β_1 -adrenoceptor mRNA was the predominant subtype expressed in the hemangioma-derived stem cells (**Figure 4**).

DISCUSSION

In this study, we could demonstrate for the first time that β_1 adrenoceptors represent the most abundant β -adrenoceptor subtype expressed in IH and in hemangioma-derived stem cells. By contrast, in LM, β_2 -adrenoceptors were the predominant subtype. Unfortunately, we could not study β -adrenoceptor expression on the cellular level in these specimens due to the lack of sufficiently specific antibodies and the condition of the material, which did not allow performance of *in situ* hybridization. Therefore, it is not clear whether the endothelium or other cellular components such as pericytes, lymphocytes, or fibroblasts in IH, VM, and LM, express the β -adrenoceptors. A recent study using a tissue microarray showed high expression of β_2 -adrenoceptors on the protein level in proliferating and involuting IH (10). Low receptor expression was noted in VMs. Unfortunately, β_1 -adrenoceptor expression was not investigated in that study, and therefore expression levels of receptor mRNA and protein cannot be directly compared with those of the current investigation. Furthermore, β_1 -adrenoceptors are abundant in the walls of arteries, and it is highly probable that in IH arteries could be present due to the abundant vascularization of the tumor. On the other hand, arteries are less abundant in VMs and LMs. In addition, it is not clear whether the β_1 -adrenoceptors found in IH are functional. These are all limitations of our study. However, the difference of mRNA levels is statistically significant and could therefore be relevant for the clinical behavior of the vascular anomalies.

Table 2. Results of immunohistochemistry for CD31, GLUT-1, D2-40,and Ki67 in 22 specimens of vascular anomalies

Patient	Vascular				
ID	anomaly	CD31	GLUT-1	D2-40	Ki67 (%)
1	Proliferative IH	Pos.	Pos.	Neg.	5–10
2	Proliferative IH	Pos.	Pos.	Neg.	10–15
3	Proliferative IH	Pos.	Pos.	Neg.	2–5
4	Proliferative IH	Pos.	Pos.	Neg.	5–7
5	Proliferative IH	Pos.	Pos.	Neg.	5–7
10	Proliferative IH	Pos.	Pos.	Neg.	10–15
6	Regressive IH	Pos.	Pos.	Neg.	1
7	Regressive IH	Pos.	Pos.	Neg.	1
13	Regressive IH	Pos.	Pos.	Neg.	2
8	VM	Pos.	Neg.	Neg.	Neg.
9	VM	Pos.	Neg.	Neg.	Neg.
11	VM	Pos.	Neg.	Neg.	1
12	VM	Pos.	Neg.	Neg.	1
14	VM	Pos.	Neg.	Neg.	Neg.
15	VM	Pos.	Neg.	Neg.	Neg.
16	VM	Pos.	Neg.	Neg.	2
17	VM	Pos.	Neg.	Pos.	Neg.
18	LM	Pos.	Neg.	Pos.	Neg.
19	LM	Pos.	Neg.	Pos.	1
20	LM	Pos.	Neg.	Pos.	1
21	LM	Pos.	Neg.	ND	Neg.
22	LM	Pos.	Nea.	Pos.	Nea.

CD31, cluster of differentiation 31; GLUT-1, glucose transporter-1; IH, infantile hemangioma; LM, lymphatic malformation; ND, not determined; Neg., negative; Pos., positive; VM, venous malformation.

Of note, cells derived from hemangiomas (9) also expressed β_1 -adrenoceptorsat2.5-foldhigherlevels than β_2 -adrenoceptors, suggesting that the β -adrenoceptor subtype profile identified in IH tumor samples is also representative of the subtype profile in the hemangioma cells.

 β -Adrenoceptors are members of the G protein–coupled receptor superfamily. Downstream activation of β -adrenoceptors



Figure 2. Characterization of vascular malformations. (**a**) H&E staining and immunohistochemistry for (**b**) CD31 and (**c**) GLUT-1 in a venous malformation and (**d**) H&E staining and immunohistochemistry for (**e**) CD31 and (**f**) D2-40 in a lymphatic malformation (original magnification ×10). VM of patient ID 16 presenting (**a**) large and dilated caverna with (**b**) CD31-positive, but (**c**) GLUT-1 negative endothelium. Erythrocytes in the vessel lumen are GLUT-1 positive. (**d**) The LM shows collapsed vessels that are (**e**) CD31 and (**f**) D2-40 positive. CD31, cluster of differentiation 31; GLUT-1, glucose transporter-1; H&E, hematoxylin and eosin; LM, lymphatic malformation; VM, venous malformations.



Figure 3. Expression of β_1 , β_2 , and β_3 -adrenoceptors in (**a**) human IH, (**b**) venous malformations, and (**c**) lymphatic malformation. (**a**) In IH, β_1 -adrenoceptor (*ADRB1*) mRNA was more abundantly expressed than β_2 - or β_3 -adrenoceptor mRNA. (**b**) In VM, β_1 - and β_2 -adrenoceptor mRNA were detected at similar levels. (**c**) In LM specimens, β_2 -adrenoceptor expression (*ADRB2*) was higher than expression of β_1 - and β_3 -adrenoceptors. **P* < 0.05, ***P* < 0.01, †*P* < 0.001. *ADRB1*, *ADRB2*, *ADRB3*, β_1 , β_2 , β_3 -adrenoceptor mRNA, respectively. IH, infantile hemangioma; LM, lymphatic malformation; VM, venous malformation.

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Figure 4. Expression of β_1 -, β_2 -, and β_3 -adrenoceptors in hemangiomaderived stem cells. β_1 -Adrenoceptor (*ADRB1*) mRNA was more abundantly expressed than β_2 - or β_3 -adrenoceptor mRNA. **P* < 0.05, ***P* < 0.01. *ADRB1*, *ADRB2*, *ADRB3*, β_1 -, β_2 -, β_3 -adrenoceptor mRNA, respectively.

Table 3. Primer sequences for quantitative real-time PCR analysis of human β -adrenoceptor subtype expression

Gene symbol		Primer $5' \rightarrow 3'$	Product length (bp)
Adrb1	Sense	CCG GGA ACA GGA ACA CAC	73
	Antisense	GAA AGC AAA AGG AAA TAT GTC TTG A	
Adrb2	Sense	CAG GAA GCC ATC AAC TGC TA	61
	Antisense	GGC TTG GTT CGT GAA GAA GT	
Adrb3	Sense	TTT TCT AAA CCC CAG CCT TG	59
	Antisense	CAC GGC ACC TGG ACA CTA C	
Rps29	Sense	GGT TCT CGC TCT TGT CGT GTC	100
	Antisense	ATA TCC TTC GCG TAC TGA CGG	

leads to activation of guanine nucleotide–binding proteins causing dissociation of G α -guanosine-5'-triphosphate and G $\beta\gamma$ subunits, thereby activating intracellular signaling pathways via adenylyl cyclase activation (11,12). Inhibition of this pathway by β -blockers has not been studied in detail with respect to inhibition of cell proliferation. Furthermore, data on inhibition of downstream signaling of β -adrenoceptors in endothelial cells from IH are not available and future studies using hemangiomaderived stem cells are warranted (9). However, models of the antiangiogenic effect of β -blockers by downregulation of angiogenesis factor expression or induction of endothelial cell apoptosis in IH have been postulated (3).

Our results demonstrate that IH, VM, and LM can be discriminated based on their difference in β -adrenoceptor subtype mRNA expression. Of note, until today only proliferating IHs have been reported to show growth inhibition by propranolol, a nonselective β -adrenoceptor antagonist. Interaction on the β_1 -adrenoceptor level could be a possible mechanism of action.

METHODS

Hemangioma Cells

Hemangioma-derived stem cells were obtained from Joyce Bischoff (Harvard Medical School, Boston, MA). The derivation, sources, and culture conditions for the hemangioma-derived stem cells have been detailed previously (9).

Tissue

Tissue specimens from IHs (n = 9), VMs (n = 7), and LMs (n = 5) of patients who underwent surgery from 2004 to 2008 at the University Medical Hospital Freiburg were available in our tumor tissue bank. Informed consent of the families was provided, and the study was approved by the ethics committee of the University Medical Center of Freiburg. **Table 1** summarizes the localization of the vascular anomaly, age at surgery, previous therapy, and growth stage.

Immunohistochemistry for Ki67, CD31, GLUT-1, and D2-40

Sections of 6 µm from formalin-fixed tissue material were provided to perform hematoxylin and eosin staining and immunohistochemistry. Antibodies against Ki67 (NeoMarkers, Fremont, CA, dilution 1:300), CD31 (NeoMarkers, dilution 1:200), GLUT-1 (Diagnostic BioSystems, Pleasanton, CA, dilution 1:50), and D2-40 (Covance, Emeryville, CA, dilution 1:100) were used following a standard protocol of the manufacturer of the automated immunohistochemistry system (A. Menarini Diagnostics, Berlin, Germany). Ki67 staining was defined as the percentage of positive nuclei from a total of 2,000 cells counted using an eyepiece grid. The positive nuclei were counted by one investigator (E.J.).

RNA Extraction, cDNA Synthesis, and $\beta\text{-}Adrenoceptor$ Expression Analysis by qPCR

Gene expression analysis from the available tissues was performed by qPCR as described previously (13). In brief, RNA was isolated from snap-frozen tissue using the RNeasy Mini Kit (Qiagen, Hilden, Germany). Total RNA (1 µg per probe) was reverse-transcribed according to the manufacturer's instructions (QuantiTect Rev. Transcription Kit; Qiagen). For qPCR, 30 µl of the amplification mixture (Quantitect SYBR Green kit; Qiagen) was used containing 20 ng of reverse-transcribed RNA and 300 nmol/l primers (MWG, Ebersberg, Germany) specific for the human β -adrenoceptor subtypes or the ribosomal protein RPS29. The cycling conditions were as follows: 15 s polymerase activation at 95 °C and 40 cycles at 95 °C for 15 s, at 58 °C for 30 s, and at 72 °C for 30 s. Results were normalized to RPS29 values. Primer sequences used for qPCR experiments are listed in **Table 3**.

Statistics

Data are presented as means \pm SEM of individual data points. Data were analyzed using one-way ANOVA followed by Bonferroni *post hoc* tests. A *P* value of <0.05 was considered statistically significant.

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