

Helicobacter pylori Antigen in Stool Is Associated With SIDS and Sudden Infant Deaths due to Infectious Disease

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ABSTRACT: Infection with *Helicobacter pylori* has been proposed to be a common cause of Sudden Infant Death Syndrome (SIDS). We investigated the frequency of *H. pylori* infection in 160 infant deaths and 156 live controls by means of the *Helicobacter pylori* stool antigen (HpSA) immunoassay. Histology was performed in 26 randomly selected cases. *H. pylori* antigen was detected in 8% (12/156) of the live controls compared with 25% (30/122) of SIDS cases ($p < 0.001$), 53% (9/17) of deaths due to infection ($p < 0.001$), and 9% (1/11) of accidental/violent deaths ($p = 0.60$). In the classic age peak for SIDS, 1–5 mo, 31% (21/67) of SIDS cases were HpSA positive compared with 1.5% (1/68) of live controls ($p < 0.001$). Rod-like immunoperoxidase positive *H. pylori* organisms were identified in 7/12 HpSA positive gastric antrum sections compared with 2/14 HpSA negative ($p = 0.038$). Significantly elevated IL-6 levels in cerebrospinal fluid representing signs of central immune stimulation were demonstrated in HpSA positive SIDS victims compared with HpSA negative victims ($p = 0.045$). Detection of *H. pylori* antigen in stool is associated with SIDS and deaths due to infections. We hypothesize that *H. pylori* infection in infancy may be involved as the triggering pathogen for sudden death during the first 5 month after birth. (*Pediatr Res* 64: 405–410, 2008)

Infection with *Helicobacter pylori* is involved in the pathogenesis of acute gastritis and peptic ulcers and is associated with gastric adenocarcinoma and MALT lymphoma (1). In recent years, a variety of extradigestive disorders, including cardiovascular diseases, autoimmune disorders, and liver diseases have also been associated with *H. pylori* infection (2). In 1997, a possible link between *H. pylori* and Sudden Infant Death Syndrome (SIDS) was hypothesized (3). In a case-control study, Kerr *et al.* (4) reported a highly significant association between the detection of *H. pylori* DNA sequences in various organ tissues and SIDS. The study was, however, heavily debated and criticized for design and methodology (5). Later case-control studies have failed to confirm a role for *H. pylori* in SIDS (6–8).

The incidence of SIDS dropped dramatically following the intervention programs directed against prone sleeping and smoking in pregnancy, however the underlying mechanism involved in SIDS still remains unknown. Disturbed homeostasis of the immune system as a response to infection in the aerodigestive tract may play a role as trigger event for the

death mechanism in a large proportion of the SIDS cases (9,10). Increased concentrations of IL-6 in cerebrospinal fluid (CSF) have been found in SIDS victims (11), and elevated IL-6 levels is found to be associated with immunoreactivity in the larynx and with symptoms of cold before death (12). Mucosal immune activation in the duodenum has also been shown in SIDS (13). Epidemiologic studies have disclosed that SIDS is associated with symptoms of cold before death, winter season, and occurrence of respiratory epidemic infections (14,15).

We have recently presented a study of the prevalence of *H. pylori* stool antigen (HpSA) in healthy Norwegian infants (16). In this follow-up study, we have investigated the frequency and significance of *H. pylori* infection in SIDS victims and other sudden unexpected infant deaths.

METHODS AND MATERIALS

Subjects. The study population consisted of 160 cases of sudden unexpected deaths in infancy (SUDI) below 1 y of age (boys/girls = 96/64) from the southeastern region of Norway investigated at the Institute of Forensic Medicine in Oslo between 1993 and 2004. Only infants of ethnic Nordic citizens were included in the study. The postmortem investigation involved evaluation of circumstances of death, review of medical and family history, full-body radiographical examination, toxicology, and a thorough autopsy with extensive histologic and microbiological examinations, including neuropathological examination. Autopsies were performed by the same forensic pathologists (ÅV, TOR, MAR, ASP); median time interval from death to autopsy was 19 h (range 5–77). Based on the Nordic SIDS criteria (17,18), the cases were categorized as either pure SIDS (no cause of death revealed), borderline SIDS (significant, yet nonlethal findings revealed), or explained deaths (Table 1). The latter were categorized as violent deaths, infectious deaths, or deaths due to noninfectious disease (Table 1). The control group consisted of 156 healthy children (boys/girls = 75/81) from the South-eastern part of Norway, recruited by consecutive sampling in two maternity clinics (Marienlyst, city of Drammen and Frogn, City of Drobak) from Oct 2003 to Nov 2004 and has been presented in detail previously (16). Informed consent was obtained from the parents. The study was approved by the Committee for Medical Research Ethics in Norway.

Sample collection and preparation. Stool specimens from the controls were collected from the diapers upon admission to the maternity clinic, as described previously (16). From the patients, stool specimens were obtained during autopsy by a “milking procedure” of the dissected rectal colon. Specimens were stored at -80°C before analysis. Samples of cerebrospinal fluid were obtained by suboccipital puncture. Gastric antrum tissue specimens were also obtained during autopsy, fixed directly in cold ethanol, and embedded in paraffin.

HpSA ELISA. For detection of *H. pylori*, the Premier Platinum *H. pylori* stool antigen ELISA kit, HpSA (Meridian Bioscience Inc., Ohio, USA), was

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Abbreviations: CSF, cerebrospinal fluid; H & E, hematoxylin and eosin staining; HpSA, *Helicobacter pylori* stool antigen; *H. pylori*, *Helicobacter pylori*; IL-6, interleukin-6; SIDS, sudden infant death syndrome; SUDI, sudden unexpected death in infancy

Table 1. HpSA detection rate and classification of subjects investigated

Category	No	Sex M/F	Age in days median (range)	HpSA positive	p-value*
SIDS	83	48/35	111 (7–323)	17 (20%)	0.004
Borderline SIDS	39	25/14	76 (11–287)	13 (33%)	<0.001
16 slight infection				5	
14 nonlethal conditions				5	
9 circumstances of death				3	
Infectious deaths	17	15/2	91 (15–357)	9 (53%)	<0.001
9 pneumonia				5	
3 septicemia				1	
5 other (meningitis, myocarditis, peritonitis)				3	
Non-infectious disease	10	1/9	95 (7–345)	2 (20%)	0.20
4 heart failure (malformations)				1	
2 lung failure (malformation and immaturity)				0	
2 epilepsy (birth-related cerebral injury)				0	
1 inborn immunodeficiency (DiGeorge syndr.)				0	
1 Ileus/peritonitis (Meckels diverticulum)				1	
Accidental/inflicted death	11	7/4	200 (9–333)	1 (9%)	0.60
6 asphyxic deaths				1	
4 head injury				0	
1 starvation/dehydration				0	
Live infants	156	75/81	75 (8–360)	12 (8%)	
Total	316	171/145	90 (7–360)	54 (17%)	

* Chi-squared analysis compared to live control infants.

performed according to the instructions of the manufacturer. Briefly, diluted stool samples were incubated in microwells coated with rabbit antibodies recognizing *H. pylori* antigens and then with rabbit anti-*H. pylori* antibody conjugate in the presence of color developing solution. The OD values were determined at 450 nm wavelength. Tests were regarded positive when OD \geq 0.160. All samples were tested twice using separate test tubes and blinded for diagnosis.

Histology and immune staining. Gastric antrum tissue specimens from 30 randomly chosen cases (first 15 HpSA positive and first 15 age-matched negative cases in the dataset) were fixed directly in cold ethanol and embedded in paraffin. Parallel sections were cut at 4 μ m and either stained with hematoxylin and eosin (H & E), Giemsa or mounted on clean polysine slides and processed for immune staining: Dried, deparaffined and rehydrated slides were subjected to pretreatment with Proteinase K for 8 min and incubated with ChemMate Peroxidase Blocking Solution in room temperature for 10 min. The slides were subsequently incubated with the polyclonal rabbit anti-*H. pylori* antibodies (Dilution 1:40, Batch B-0471, DAKO, Glostrup, Denmark) for 30 min at room temperature. Immunostaining was performed with the ChemMate EnVision Detection Kit Peroxidase/DAB Rb/Mo (K5007; DAKO); incubation with polymer for 30 min, incubation with substrate for 5 min and toning in 0.5% copper sulfate for 5 min at room temperature. Counterstaining was finally briefly performed with hematoxylin.

The tissue sections were evaluated blindly by a specialist in microbiology and pathology (ÅV) who had no information about case history or stool test findings. Four sections (3 HpSA positive and 1 HpSA negative) were ineligible for evaluation due to technical errors in the paraffin embedding. H & E stains of the remaining 26 sections were reviewed and semiquantitated for degree of autolysis and chronic and active inflammation. Giemsa and immunoperoxidase stains were reviewed at high (630 \times) power light microscopy for the presence of *H. pylori* in the surface mucous layer and within gland crypts of the lamina propria. Tissue sections were reevaluated blindly by the same observer and the reproducibility was good having the same result in 24 of 26 reexamined cases.

IL-6 measurement. IL-6 concentrations in CSF were measured by an ELISA kit (R&D Systems Inc., Minneapolis, USA), utilizing 100 μ L CSF and performed with OD determination as previously described (12).

Microbial cultures. In all patients investigated in this study, microbial aerobic/anaerobic cultures of blood and CSF as well as organ tissues of lung, liver, spleen, and kidney were taken at autopsy. To disclose possible associations between *H. pylori* antigen detection and other bacteria, the culture reports from the infectious deaths ($n = 17$) and the borderline SIDS cases with possible infection ($n = 16$) were investigated.

Statistical analyses. The χ^2 analysis was used for comparison of categorical variables between groups. When the expected values were <5 , the Fisher's exact test was applied. The Mann-Whitney U-test was used for comparison of continuous variables. The significance level was set at values of $p < 0.05$.

RESULTS

H. pylori stool antigen in cases and controls. *H. pylori* antigen in stool was detected in 53% of infants who died due to an infectious disease, and 25% of the SIDS cases (pure and borderline SIDS conjoined), compared with 8% of the age-matched live control infants ($p < 0.001$ for both groups) (Table 1). Furthermore, positive HpSA tests were found in 20% ($p = 0.20$) of the deaths due to noninfectious disease and in 9% of the violent deaths ($p = 0.60$).

Histology. Examination of histology was problematic due to tissue autolysis (Table 2). The foveolar epithelium and covering mucus-layer were partially or totally lost. Presence of both rods and coccoid bacteria were detected in 50% (13/26) of specimen. Scattered curved and rod-like structures stained positive by specific immune staining were observed, localized on the luminal side of foveolar epithelial cells, and in glandular crypts (Fig. 1). The pathologist, who was unaware of the HpSA results, confirmed the presence of *H. pylori* bacteria in 7/12 gastric specimen of HpSA positive infants compared with 2/14 HpSA negative ($p = 0.038$) (Table 2). No signs of acute inflammation were observed in any of the sections investigated, but in five of the cases a few foci with lymphocytes in the basal layer of the mucosa were observed, however not related to positive immune staining (Table 2).

H. pylori and epidemiologic variables. Twenty-six percent (24/92) of SIDS victims below 6 mo of age were HpSA positive, compared with 7% (2/30) of SIDS victims in the second half year of infancy ($p = 0.24$). In the classic peak age group, 1–5 mo, 31% (21/67) of the SIDS cases were HpSA positive compared with 1.5% (1/68) of the live controls ($p < 0.001$) (Fig. 2). Looking at all infant deaths, a significantly higher proportion of the infants dying in the cold season were HpSA positive, compared with those who died in the summer season (33% (30/90) and 17% (12/70), respectively, $p = 0.021$). In the live controls, no difference was found between

Table 2. Histology examination of 26 cases of sudden death in infancy

Cause of death	Sex	Age (mo)	HE staining			Giemsa	Hp immune staining	HpSA stool test
			Autolysis*	Inflammation**	Bacteria			
SIDS	M	1	(+)	++	Rods and cocci	Rods and cocci	pos	pos
SIDS	F	1	+++	+	Rods and cocci	Rods and cocci	pos	pos
SIDS	F	2	+++	+	Rods and cocci	Rods and cocci	pos	pos
SIDS	M	2	+++	+	—	Rods	pos	pos
SIDS	M	3	++	+	—	Rods	pos	pos
Pneumonia	M	4	+	+	Rods and cocci	Rods and cocci	pos	pos
Pneumonia	F	3	++	+	Rods	Rods and cocci	pos	pos
SIDS	M	4	+	+	Rods	Rods	pos	neg
SIDS	F	2	+	+	—	Rods and cocci	pos	neg
SIDS	F	1	+	+	Rods and cocci	Rods and cocci	neg	pos
SIDS	F	2	+++	+	Rods	Rods	neg	pos
Lung failure	F	1	+++	+	Rods and cocci	Rods and cocci	neg	pos
Septicemia	M	11	+++	+	Rods	Rods	neg	pos
Pneumonia	M	3	+	++	—	Rods	neg	pos
SIDS	F	1	+++	++	Rods and cocci	Rods and cocci	neg	neg
SIDS	M	1	+++	+	Rods and cocci	Rods and cocci	neg	neg
SIDS	M	3	+	+	Rods and cocci	Rods and cocci	neg	neg
SIDS	M	9	+++	+	Rods and cocci	Rods and cocci	neg	neg
SIDS	M	4	++	+	—	Rods and cocci	neg	neg
SIDS	F	1	+++	+	Rods	Rods	neg	neg
SIDS	M	1	+++	+	Rods	Rods	neg	neg
SIDS	F	2	++	+	Rods	Rods	neg	neg
SIDS	M	5	+++	+	—	—	neg	neg
Accidental	M	5	++	++	Rods	Rods	neg	neg
Accidental	F	8	+++	+	Rods	Rods	neg	neg
Accidental	M	8	+++	++	Rods	Rods	neg	neg

* Degree of autolysis: [+] areas with some preserved surface and foveolar epithelium, gland crypts well preserved; [++] most foveolar epithelium lost, gland crypts present; [+++] all superficial and foveolar compartment lost. Only basal layers of gland crypts present.

** Degree of inflammation: [+] scattered inflammatory cells; [++] one or a few foci with inflammatory cells; [+++] severe inflammation.

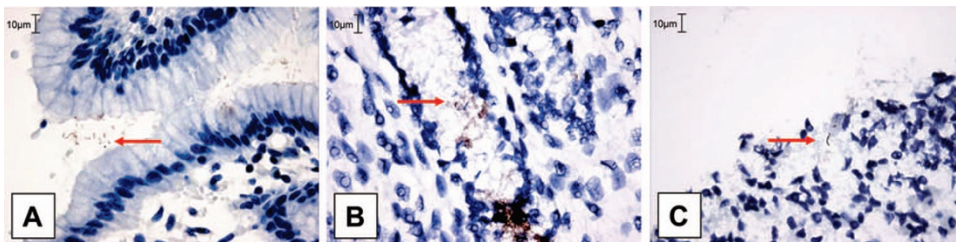


Figure 1. Photomicrographs (450X) of anti-*H. pylori* immunoperoxidase staining in gastric tissue. A: positive control. B–C: cases of SIDS. Red arrows point at curved and rod-like structures with positive immune staining (*H. pylori* bacteria).

stool samples collected in the winter season, 9% (8/92) and in the summer season, 6% (4/64), $p = 0.57$. Other epidemiologic factors for SIDS, such as male gender, prone sleeping position, or bed sharing did not show any association with the HpSA findings (Table 3).

H. pylori and microbial cultures. Pathogens of possible significance (i.e., pure growth of streptococci, *Klebsiella pneumoniae*, staphylococci, or enterococci) growing in CSF and/or heart blood specimens were detected in 12 out of 33 cases of infant deaths classified as due to infection or as borderline SIDS with signs of infection insufficient to explain the cause of death. Forty-two percent of the cases with findings of such pathogens were HpSA positive, compared with 38% of the cases without ($p = 0.8$ N.S.).

H. pylori and IL-6. IL-6 concentrations in CSF were measured in 147/160 infant deaths. In HpSA positive cases, the IL-6 levels were significantly higher (median 32 pg/mL; interquartile range (IQR) 153) compared with HpSA negative cases (median 9 pg/mL; IQR 32, $p = 0.006$) (Fig. 3). The

difference was also significant for SIDS tested separately: Median IL-6 levels in HpSA positive and negative SIDS cases were 28 pg/mL (IQR 153) and 10 pg/mL (IQR 33), respectively ($p = 0.047$).

DISCUSSION

In 1997, Pattison and Marshall (3) put forward the hypothesis of a possible relationship between *H. pylori* infection and SIDS. They hypothesized that aspiration of gastric juice containing large amounts of urease from *H. pylori*, could react with plasma urea to produce ammonia toxicity and cause respiratory arrest. Three years later, Kerr *et al.* (4) detected *H. pylori* DNA sequences by means of PCR in 28 of 32 SIDS cases compared with only one of 8 controls and concluded that a causative role for *H. pylori* in SIDS was likely. The study was heavily debated (5), but was followed by only a few small-number studies, which were unable to confirm Kerr’s results (6,8,19). Elitsur *et al.* (8) investigated 25 SIDS cases

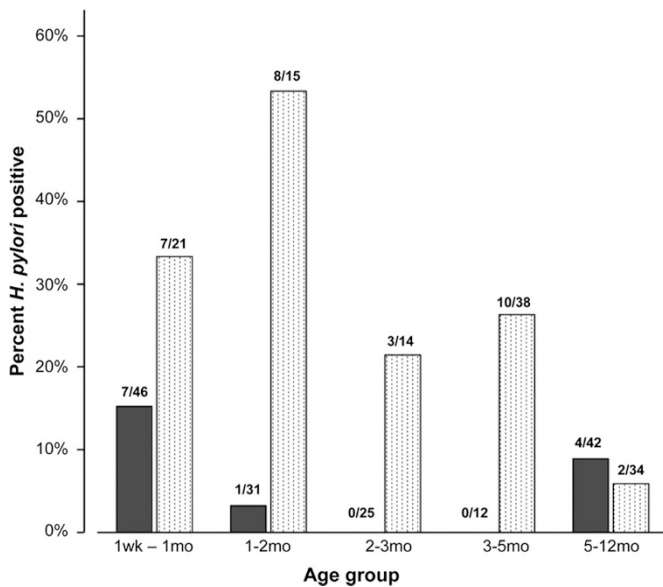


Figure 2. *H. pylori* antigen detection rate in SIDS* (white dotted columns) and live control infants (shaded columns) at different age groups. *Both pure SIDS and borderline SIDS are included.

and detected *H. pylori* DNA in gastric or tracheal specimen from six of these cases. The presence of *H. pylori* organisms was, however, not confirmed by histology or immunohistochemistry. Ho *et al.* (6) detected *H. pylori* DNA by PCR in gastric tissue samples in 54% (9/17) of SIDS cases and 57% (4/7) of controls, but were unable to confirm the results by urease test, bacterial culture, or immunohistochemical methods. Recently, Løddenkötter *et al.* (7) investigated gastric tissue specimen from 94 cases of SIDS with an accurate PCR technique, finding traces of the *H. pylori* specific ureC gene in only two of these cases. In conclusion, though *H. pylori* DNA has been detected by PCR in gastric or respiratory tissue to a variable extent, only Kerr's study is supportive of an association between *H. pylori* and SIDS. Moreover, *H. pylori* infection has not been documented histopathologically in SIDS in any of the studies.

The present study is the first to investigate stool specimens, demonstrating a significant association between the presence of *H. pylori* stool antigen and sudden infant deaths. The method of choice, the HpSA test, is a well-recognized procedure

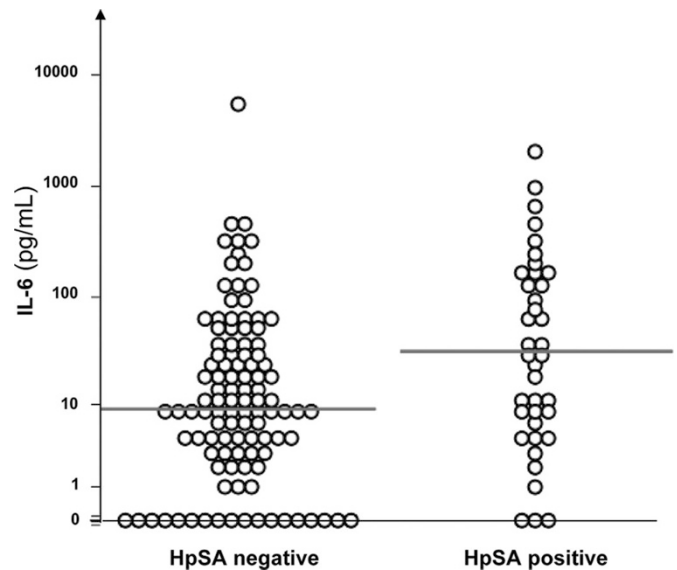


Figure 3. Cerebrospinal fluid IL-6 concentrations in HpSA negative and positive cases of sudden deaths in infancy (SUDI). Horizontal lines represent median IL-6 levels in the two groups, 9 and 32 pg/mL respectively ($p = 0.006$).

for detecting *H. pylori* infection in pediatric populations (20,21), utilizing polyclonal antibodies that target *H. pylori* specific proteins. Although a monoclonal kit has now been introduced by the manufacturer, the HpSA kit has been widely used and a high diagnostic accuracy has been described in both high and low prevalence groups (20,21). In the Nordic countries, the prevalence of *H. pylori* infection in children is most likely very low (16,22). The low frequency of HpSA positive tests in our controls (8%) thus favors the reliability of the test. Correspondingly, the fact that only one of 11 infants (9%) in the group of accidental deaths were HpSA positive yields an indication that the test behaves similarly in stool samples from live and dead infants.

Comparing SIDS with other subclasses of SUDI is complicated. Infants that die suddenly and unexpectedly due to an infection or other disease are likely, in some manner, to be predisposed to the fatal outcome. Accidental deaths rarely occur in infancy, but the age distribution is different from in SIDS (Table 1). Epidemiologic risk factors for SIDS like maternal smoking, winter seasonality and social deprivation

Table 3. Epidemiological variables and autopsy findings in HpSA positive and negative SIDS* cases

	HpSA positive		HpSA negative		p-value
	Mean	N = 30	Mean	N = 92	
Male gender	53%	(16/30)	62%	(57/92)	0.40
Cold season	67%	(20/30)	50%	(46/92)	0.11
Prone sleeping	40%	(10/25)	60%	(47/78)	0.08
Bed sharing	39%	(11/28)	32%	(29/91)	0.47
Cold at time of death	45%	(10/22)	30%	(23/77)	0.17
Fever at time of death	16%	(3/19)	7%	(5/71)	0.36
Maternal smoking	65%	(11/17)	65%	(32/49)	0.96
Intrathoracic petechia	67%	(20/30)	72%	(66/92)	0.60
Age at time of death**	63 days		112 days		0.003
Maternal age**	23.5 yr		26.0 yr		0.28

* SIDS and borderline SIDS conjoined.

** Age presented with median values.

are not restricted to SIDS, but are also associated with other natural causes of sudden infant death (23). On the other hand, except for the variability of *H. pylori* detection with age, no associations were found between *H. pylori* status and epidemiologic factors (Table 3). Unfortunately, the dataset did not include sufficient information with regard to socio-economic status to enable valid comparison analyses. Thus, it cannot entirely be ruled out whether the higher incidence of *H. pylori* detection rate in SIDS and deaths due to infections reflects lower hygienic standards. However, the socio-economic imbalance is less pronounced in Norway than in other western countries and in a previous study from the same region, Arnestad *et al.* (24) were unable to find a difference in socio-economic status between SIDS and age-matched live controls.

Acute infection with *H. pylori* does not necessarily result in chronic infection (25,26). Serological studies indicate that particularly the acquisition of *H. pylori* infection in early infancy is associated with a high rate of subsequent loss of bacteria (22,27). We recently reported a high proportion of HpSA positive stool samples in healthy Norwegian newborn infants delivered vaginally (16). The unanticipated result was confirmed by the detection of *H. pylori* DNA by PCR in several of the samples. We hypothesized that *H. pylori* transmission from mother to child is likely to occur during the baby's passage through the birth canal. On the other hand, the HpSA detection rate in infants older than 1 week of age is low. Thus, in most normal individuals the encounter with *H. pylori* does not seem to result in a permanent colonization of the gastric mucosa. Development of chronic infection and *H. pylori* related disease is determined by host-pathogen interactions (25). The significance of *H. pylori* virulence factors has been widely studied (28), but individual susceptibility and environmental factors probably play an equally important role (29). Polymorphisms in host genes that encode immune modulating factors have been shown to affect *H. pylori* related diseases (30). Wu *et al.* found that IL-10 polymorphisms and smoking were individual risk factors for the development of gastric cancer. Interestingly, the most important environmental risk factor for SIDS after prone sleeping is maternal smoking (24) and IL-10 polymorphisms (31) have also been implicated in SIDS and sudden infant deaths due to infectious disease.

There is a discrepancy between our findings and previous studies that have investigated gastric specimen from SIDS by immunohistochemical methods. Neither Elitsur *et al.* (8) or Ho *et al.* (6) identified *H. pylori*-like organisms in any of the samples investigated. On conventional H & E stained gastric tissue sections, Ho *et al.* (6) reported occasional thick rods and clusters of cocci, but no curved rod-like bacteria characteristic of *H. pylori*. Immunoperoxidase staining gave also negative results. Unavoidably, postmortem gastric tissue is subject to autolysis shortly after death. In our material, the presence of rod-like *H. pylori* positive stained structures was scarce and void of inflammatory changes in the surrounding mucosa. A majority of the tissue samples were collected during autopsy performed within 24 h after death. However, manifest autolysis was observed in almost all of the sections. Thus, inflammatory changes may have been present before death in several

of the cases, although we may have been unable to detect it due to tissue autolysis.

The drop in SIDS rate following the intervention programs directed against prone sleeping and smoking in pregnancy, predominantly involved young infants between 1 and 5 mo of age, whereas older infants were less affected (14). The mucosal immunity is developing rapidly during the first months of life (32–34) and an activation of mucosal immunity in the aerodigestive tract has been demonstrated in SIDS victims (13,35). In the present study detection of *H. pylori* stool antigen also varied significantly with age; cases below 5 mo of age being accountable for the difference between SIDS and live controls. The HpSA results from the youngest SIDS cases were comparable to the group of infectious deaths, whereas the results from the SIDS cases older than 5 mo of age were similar to the live controls (Fig. 2). The highest *H. pylori* detection rate was found among infants that died suddenly and unexpectedly due to infections. We postulate that *H. pylori* infection in infancy in a Norwegian population represent a biomarker of increased vulnerability to infections and increased risk for sudden death before 5 mo of age. We have previously reported that a significant proportion of SIDS cases have elevated CSF levels of the pro-inflammatory cytokine IL-6, produced in response to microbial products (36), analogous to infants that died due to infection (11). Disturbances in the immunologic homeostasis caused by a combination of genetic predisposition, vulnerable developmental stage, and a trigger event may induce the death mechanism in a large proportion of SIDS cases (9,10,37). Such a trigger event may be a slight infection with stimulation of the mucosal immune system (9). The association between *H. pylori* detection and central immune stimulation may indicate that *H. pylori* may be involved as the triggering pathogen.

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