

Pancreatic Phospholipase A₂ Contributes to Lung Injury in Experimental Meconium Aspiration

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ABSTRACT: To investigate the role of pancreatic (group I) secretory PLA₂ (sPLA₂-I) in the pathogenesis of meconium aspiration syndrome, human particulate meconium or its supernatant either before or after extraction of PLA₂-I was insufflated into rat lungs. In addition, the pulmonary effects of intra-tracheal human and bovine PLA₂-I were studied. Lungs with saline instillation served as controls. Intrapulmonary particulate meconium (both before and after PLA₂-I extraction), unlike meconium supernatant, resulted in markedly elevated lung tissue PLA₂ catalytic activity and human PLA₂-I concentrations when compared with controls. On the other hand, tissue concentrations of the group II PLA₂ remained unchanged in all meconium lungs. Pulmonary PLA₂-I concentrations further correlated positively with lung injury scores. Instillation of meconium-derived human PLA₂-I, at a concentration of one-third of that in particulate meconium, did not raise PLA₂ activity or concentrations of PLA₂-I or PLA₂-II in the lung tissue from the control level, but still resulted in significantly elevated lung wet/dry ratio and injury score. In contrast, insufflation of bovine pancreatic PLA₂ increased the lung tissue enzyme activity and wet/dry ratio from the control level, but had no effect on the type II PLA₂ concentration or lung injury score. Our data thus indicate that human pancreatic PLA₂, introduced in high amounts within aspirated meconium especially in particulate form, is a potent inducer of lung tissue inflammatory injury. (*Pediatr Res* 59: 641–645, 2006)

Perinatal aspiration of meconium may result in severe pulmonary failure with ventilation-perfusion mismatch in the lungs, hypoxemia and increase in pulmonary vascular resistance, associated with high morbidity and mortality in full- or post-term newborn infants (1). The pathophysiology of the neonatal meconium aspiration syndrome (MAS) is complex, but inflammation with pulmonary surge of cytokines, possibly through activation of alveolar macrophages, and accumulation of polymorphonuclear leukocytes in the pulmonary tissue is believed to be a central event in the development of acute tissue damage (1,2). This inflammatory reaction is connected with increased lung epithelial cell apoptosis, which may together with inactivation of the pulmonary surfactant and direct toxic effects of meconium significantly contribute to the lung injury process (1,3,4). Still, the clinical effects of

anti-inflammatory therapeutic approaches are unsatisfactory and do not consistently improve the outcome of severe complications of this disease (1). New advances into the pathophysiology and therapy of MAS are therefore needed.

Phospholipase A₂ (PLA₂) represents a family of ubiquitous enzymes that generate, by hydrolysing membrane phospholipids, biologically active FFA and lysophospholipids (5). In the lungs, activation of PLA₂ by direct or indirect insults may lead through arachidonic acid to formation of pro-inflammatory lipid mediators, including eicosanoids and platelet-activating factor (6,7). Since PLA₂ may also directly damage alveolar cells and inactivate surfactant (8,9), its activity is proposed to be important in the pathogenesis of acute inflammatory lung injury (6,8,10). The PLA₂ enzymes occur as cytosolic (cPLA₂) and secretory (sPLA₂) types. Among the mammalian secretory (extracellular) PLA₂'s two well-characterized enzymes have been identified: group I secreted by the pancreas (PLA₂-I) and group II (synovial) detected in inflammatory fluids (PLA₂-II) (5,10). High PLA₂ activity in inflammatory lung injury is generally connected with increased release of PLA₂-II originating from macrophages and platelets, but other types of PLA₂ enzymes may also play a modulating role (6). Previous studies in our laboratory in fact demonstrate that human meconium contains high amount of PLA₂-I and that the PLA₂ activity and concentration of PLA₂-I in the meconium-contaminated lung tissue is high (3). Although these results connected intra-tracheal insufflation of particulate meconium with pulmonary inflammation, there are also data suggesting an important role for the soluble fraction of human meconium in the initiation of acute inflammatory response in the insulted lungs (3,4). We hypothesized that PLA₂-I is present in both particulate and soluble fraction of meconium and that it contributes to the inflammatory and apoptotic lung injury induced by meconium aspiration, and therefore decided to investigate the pulmonary effects of meconium before and after extraction of PLA₂-I, and also the direct effects of intra-tracheal PLA₂-I. We additionally evaluated the contri-

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Abbreviations: MAS, meconium aspiration syndrome; PLA₂, phospholipase A₂; PLA₂-I, group I phospholipase A₂ (pancreatic); PLA₂-II, group II phospholipase A₂ (synovial); TUNEL, terminal deoxynucleotidyl transferase-mediated nick end-labeling

bution of the pulmonary PLA₂-II production on the meconium-induced lung injury.

MATERIALS AND METHODS

Animal preparation. Sixty-six male Sprague-Dawley rats (mean weight 320 g, SD 150 g) were studied. The animals were anesthetized with pentobarbital (60 mg/kg IP (intraperitoneal), Mebunat®, Orion-Farmos, Turku, Finland) and intubated through an incision in the neck. Lungs were then ventilated using a pressure-controlled ventilator (Baby-Bird, Bird Corp, Palm Springs, CA) for three hours (rate 30/min, peak inspiratory pressure 15 cmH₂O, positive end-expiratory pressure 1–2 cmH₂O, FiO₂ 0.70). The experiments were approved by the Committee of Animal Care in Research of the University of Turku.

Meconium preparation. Human meconium was collected from the first stools of healthy full-term neonates. The samples were frozen, later pooled, lyophilised and irradiated for sterility. Before instillation, meconium was diluted with sterile saline to a concentration of 65 mg/mL (particulate meconium), clinically corresponding to thick meconium in amniotic fluid (3). To separate supernatant, this particulate solution of meconium was spun down at 5,000 rpm for 20 min. in room air. Endotoxin in meconium was quantified using the *Limulus* Amebocyte Lysate Assay (LAL Kinetic-QCL, 50-650U; BioWhittaker/Cambrex Bio Science, Walkersville, M.D., USA). The meconium batch was sterile in culture and its endotoxin content was 24 ng/mg of meconium.

Study protocols: Protocol I: The effects of PLA₂-I in meconium. Lung injury was produced in 18 rats by intra-tracheal instillation (3 mL/kg) of human particulate meconium (65 mg/mL) (Group MP, *n* = 8) or supernatant of meconium (Group MS, *n* = 10). To study the pulmonary effects of PLA₂-I in meconium, two further groups of rats were instilled either with the same amount of particulate meconium after extraction of PLA₂-I by immunoabsorption (Group MP/E, *n* = 10) or supernatant of meconium after extraction of PLA₂-I (Group MS/E, *n* = 10). The mean concentration of PLA₂-I in particulate human meconium before extraction was 2280 µg/L (341, SD), and after extraction 1286 µg/L (462), corresponding to a mean decrease of 44% (19%). Similarly, the mean concentration of PLA₂-I in meconium supernatant before extraction was 2348 µg/L (1088), and 672 µg/L (591) after extraction, corresponding to a mean decrease of 71% (9%). Control animals received a bolus of sterile saline intra-tracheally (*n* = 7) (Control). To estimate the baseline PLA₂ activity and PLA₂-II enzyme concentration in rat lungs, five additional animals were anesthetized and the lungs studied without any insult or ventilation. All the instilled lungs were ventilated with 70% oxygen for 3 h. At the end of the experiments, thoracotomy was made and heparin was injected into the left ventricle to prevent blood from clotting. Subsequently, the lungs were perfused with 10 mL of sterile saline through a pulmonary artery catheter. The lungs were finally isolated and the lobes were separated for biochemical and histologic analysis and measurement of wet/dry ratio. The upper right lobe was deep frozen in liquid nitrogen and then put to the -70°C freezer.

Study protocols: Protocol II: The effects of meconium-derived or bovine group I PLA₂. To further study the pulmonary effects of PLA₂-I, the enzyme was purified from samples of human meconium. Due to technical difficulties, the concentration of PLA₂-I after purification from meconium remained on an average level of 840 µg/L (89). Three mL/kg of this solution was then instilled into the lungs of six rats (Group hPLA₂). In comparison, two other groups received a similar bolus of bovine PLA₂-I (Sigma Chemical Co. Chemicals). One group was given bovine PLA₂-I intra-tracheally at a concentration of 4 mg/mL, corresponding to the catalytic activity found in the particulate (65 mg/mL) meconium (*n* = 5) (Group bPLA₂/4). Catalytic activity of bovine PLA₂-I [1 mg/mL] was 1804 U/L (868). The other group received an intrapulmonary bolus of double concentration, 8 mg/mL, of bovine PLA₂-I (*n* = 5) (Group bPLA₂/8). The rats were similarly ventilated with 70% oxygen for 3 h and the lungs were studied as above at the end of the study.

PLA₂-I immunoabsorption. Immunoabsorption was used to reduce the PLA₂-I concentration in meconium. Particulate meconium was diluted in sterile saline in three steps to get the final concentration of 65 mg/mL. After every step diluted meconium was centrifuged [19 500 rpm (47 000 G)] for 30 min, and the supernatant was collected. Supernatant was then used for immunoabsorption in polystyrene tubes coated with rabbit anti-human PLA₂-I antibody (11) (25 µg/mL). This antibody is highly specific to human PLA₂-I (12), which is characterized and proved, e.g., in the immunoassay use (13). Incubation (45 min/tube) of the supernatant in the antibody-coated plastic tubes (1 mL/tube) was repeated 16 times and the solid (dry) phase of the meconium was finally diluted in the supernatant for measurement of the PLA₂-I concentration.

PLA₂-I purification from meconium. An immunoaffinity column was prepared for immunoabsorption of human PLA₂-I from the meconium. Briefly, a 5 mL sized HiTrap NHS-activated Sepharose column was prepared by immobilizing 13 mg of HiTrap protein A purified from human PLA₂-I-immunized rabbit antiserum as instructed by the manufacturer of both columns (Amersham Pharmacia Biotech AB, Uppsala Sweden).

Clarified supernatant of meconium was applied into the column that was equilibrated with 0.05 M sodium phosphate buffer pH 7.5. The column was washed with the same buffer and the attached PLA₂-I was released with 0.05 M glycine-hydrochloride buffer pH 2.7 containing 0.5 M NaCl. The purification was monitored with an UV-detector, 280 nm. The collected PLA₂-I fraction was instantly neutralized by dialyzing against sodium phosphate buffer pH 7.5 overnight at + 4°C. For the animal experiments the purified PLA₂-I was concentrated and the buffer changed to sterile saline solution with centrifugal filter device (Ultrafree-15, Millipore Bedford, MA).

PLA₂ concentrations and catalytic activity. The catalytic activity of PLA₂ and concentrations of human group I PLA₂ and rat group II PLA₂ were measured using the same protocols as described earlier (3,11,14). Briefly, frozen lung tissues were homogenized, centrifuged for 20 min with 1000 G, and the supernatant was separated and used for PLA₂ catalytic activity and concentration assays. In the activity measurement the substrate was prepared by mixing unlabelled 1,2-dipalmitoylphosphatidylcholine (Sigma Chemical Co., ST. Louis, MO) with 1-palmitoyl-2-[¹⁴C]-arachidonoylphosphatidylethanolamide (DuPont, Boston, MA) in a ratio of 6 mM/1.325 µM (250 nCi), dissolved in a mixture of chloroform and methanol (2:1), dried under a flow of nitrogen, and redissolved in 10 mL 0.1 M glycine buffer (pH 8.1). 10 µL samples of lung tissue supernatant were incubated with 100 µL of substrate buffer for three hours at 40°C. The reaction was stopped by adding 100 µL of Dole's reagent. Released [¹⁴C]-arachidonic acid was separated by SiO₂/water/heptane phase extraction, and detected by a liquid scintillation spectrometer (Wallac, Turku, Finland).

The concentrations of PLA₂-I and PLA₂-II were measured by time-resolved fluoroimmunoassays. PLA₂-I concentrations were measured in the used solutions of each animal group before experiments. After experiments, the human PLA₂-I and rat PLA₂-II concentrations were measured in the supernatant of the homogenized rat lung tissue. PLA₂-I immunoassay uses MAB-coated microtitre plates and polyclonal (rabbit) antibody as tracer. Both antibodies were raised against purified human pancreatic group I PLA₂. PLA₂-II immunoassay utilizes a polyclonal antibody, raised in rabbits against rat recombinant PLA₂-IIA, immobilized on the microtitre plate and also used as a detecting tracer antibody. Fluorescence was measured with an Arcus fluorometer (Wallac, Turku, Finland). Purified human pancreatic PLA₂ and rat recombinant PLA₂ were used as standards in the measurements. We have earlier shown that there is no cross-reaction between the human and rat group I PLA₂ (3).

Histologic examinations. Paraffin sections of the left lower lung lobe were stained with hematoxylin and eosin for light microscope analysis and assessed by a pathologist blinded to the grouping of rats. A score from 0 to 4 was assigned for three different characteristics: 1) the extent of leucocyte infiltration; 2) the amount of intra-alveolar leukocytes; and 3) the amount of exudative debris. The calculated total injury score means the sum of these scores (3).

In situ detection of TUNEL-positive cells. Terminal deoxynucleotidyl transferase-mediated nick end-labeling (TUNEL) for the detection of apoptotic cells was performed in paraffin wax sections, as described earlier (3,4,14). Lymphocytes undergoing apoptosis in the lymph nodes served as a positive control. TUNEL-positive cells were counted in lung sections stained with the antidigoxygenin antibody. A distinct color reaction within the cells was regarded to represent apoptotic DNA fragmentation. The results are expressed as the number of positive cells per mm² of tissue section area in at least ten fields of view of a ×10 objective lens. The *in situ* detection of free DNA 3'-ends is a well established method in the detection of apoptotic cellular changes and has been validated by simultaneous electrophoretic DNA analysis in pancreatic tissue (14).

Statistical analysis. One-way analysis of variance (ANOVA) was used to compare the data in different groups. If the overall ANOVA was significant, comparisons between the groups were made using the Tukey's *post hoc* test. For comparison of nonparametric data, Kruskal-Wallis one-way ANOVA was used, followed by the Dunn's *post hoc* test. Differences between two groups were evaluated by unpaired 2-tailed *t* test. A level of *p* < 0.05 was considered statistically significant. The results are expressed as mean (SD).

RESULTS

The effects of PLA₂-I in meconium. The PLA₂ catalytic activity in the unventilated rat lungs was lower than in the

saline-instilled control lungs [77 (16, SD) *versus*. 166 (53) mU/g lung tissue, $p < 0.01$], whereas pulmonary tissue PLA₂-II concentrations were at the same level [160 (47) *versus* 126 (65) ng/g lung tissue, respectively]. Administration of particulate meconium (both before and after PLA₂-I extraction) resulted in markedly elevated lung tissue PLA₂ catalytic activity and PLA₂-I concentrations, whereas insufflation of meconium supernatant (with or without PLA₂-I extraction) had no significant effect on these parameters, when compared with saline controls (Fig. 1). On the other hand, the concentrations of PLA₂-II in the lung tissue were not affected in any of the study groups (Fig. 1).

The lung tissue wet/dry-ratio was not elevated from the control level after instillation of meconium, except after insufflation of meconium supernatant with PLA₂ extraction (Table 1). In contrast, the pulmonary histologic injury scores in all meconium-insulted groups were increased, with the highest score, mainly due to elevated intrapulmonary leukocyte accumulation, after insufflation of particulate, non-manipulated meconium (Table 1). The lung injury score had further a direct correlation with pulmonary PLA₂-I content ($r = 0.426$, $p < 0.01$), but not with PLA₂ activity or PLA₂-II concentrations in the lung tissue. On the other hand, pulmonary PLA₂-I, unlike PLA₂-II, concentration correlated significantly with the lung PLA₂ activity ($r = 0.826$, $p < 0.0001$), whereas no correlations between these enzyme concentrations and lung tissue wet/dry ratio were found. The amount of TUNEL-positive cells in the lungs did not differ between the groups (Table 1).

The effects of meconium-derived or bovine PLA₂-I. Instillation of meconium-derived human pancreatic PLA₂, at a concentration of a third of that in particulate meconium, resulted in lung tissue-catalytic PLA₂ activity, and concentrations of PLA₂-I and PLA₂-II that were on the control levels (Table 2). Still, the lung tissue wet/dry ratio and injury score, but not the count of TUNEL-positive cells, were significantly higher than in the controls (Table 2). Actually, the wet/dry ratio was similar than after intra-tracheal meconium instillation, but the injury score was significantly ($p < 0.01$) lower than observed in the meconium-contaminated lung tissue.

Insufflation of bovine PLA₂-I with the activity corresponding to that in particulate meconium increased the lung tissue enzyme activity from the level observed in controls, but had no effect on the pulmonary PLA₂-II concentration, lung tissue

wet/dry ratio and injury score or number of TUNEL-positive cells (Table 2). Pulmonary instillation of a double amount of bovine PLA₂-I resulted in even higher lung enzyme activity and wet/dry ratio, but again had no effect on the PLA₂-II concentration, tissue injury score or number of TUNEL-positive cells (Table 2). In fact, the higher amount of intra-tracheal bovine PLA₂-I resulted in a higher ($p < 0.02$) lung tissue wet/dry ratio than what was observed after insufflation of particulate meconium or meconium supernatant. Human PLA₂-I concentrations were not measured in these lungs.

DISCUSSION

Our present data suggest that pancreatic PLA₂ within aspirated meconium is related to the intense lung inflammatory, but not apoptotic, reaction in the development of the meconium aspiration-induced tissue injury (1–4). It is further evident that lung PLA₂-II production is not stimulated after meconium contamination. Based on the present data, we cannot however totally exclude the influence of some other components of meconium on the present pulmonary findings. Especially the fact that PLA₂-I seems to be attached very firmly to the solid particles of meconium complicated its extraction procedure and hence also the interpretation of the results. Moreover, despite of reduction of pancreatic PLA₂ from particulate meconium (44%) and its supernatant (71%), enzyme activities and concentrations of PLA₂-I were similar before and after manipulation in the meconium-contaminated lungs. This may be explained, at least in part, by absorption of the introduced enzyme into the pulmonary circulation. We have in fact previously observed elevated human PLA₂-I concentrations in plasma during the first hours after intra-tracheal meconium administration in newborn piglets (P. Kääpä, unpublished observations). On the other hand, the pulmonary PLA₂-I content correlated directly with the lung injury score. Although intra-tracheal endotoxin may also contribute to the pulmonary reactions, endotoxin content of meconium was in low nanomolar range, which remains well below the amount required for prompt induction of significant pulmonary neutrophil influx. Alike, even high amounts of intra-tracheal endotoxin do not affect pulmonary PLA₂-I expression (15). Despite the experimental lung injuries in the present work were induced in fully developed lungs and the results therefore cannot be directly applied to neonatal pulmonary

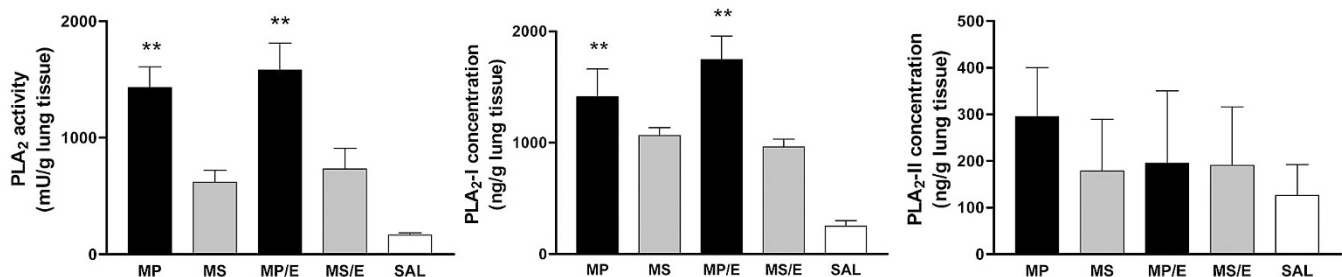


Figure 1. Tissue phospholipase A₂ (PLA₂) activity (mU/g lung tissue), and group I PLA₂ (PLA₂-I) and group II PLA₂ (PLA₂-II) concentrations (ng/g lung tissue) in rat lungs at 3 h after intra-pulmonary instillation of particulate meconium (MP, $n = 8$), particulate meconium after PLA₂-I extraction (MP/E, $n = 10$), supernatant of meconium (MS, $n = 10$), meconium supernatant after PLA₂-I extraction (MS/E, $n = 10$) and saline (control, $n = 7$). Mean(SD). ** $p < 0.01$ vs. saline.

Table 1. Lung tissue results

Group	Wet/dry ratio	Leukocyte infiltration	Alveolar leukocytes	Alveolar exudation	Injury score	TUNEL-positive cells
MP	5.8 (0.9)	1.4 (0.8)**	1.1 (0.7)*	1.9 (0.7)**	4.4 (1.7)**	0.5 (0.3)
MP/E	5.7 (1.0)	1.2 (0.6)*	0.4 (0.5)	1.0 (1.0)**	2.6 (1.9)**	0.8 (0.7)
MS	6.0 (0.7)	1.2 (0.8)*	0.9 (0.7)	1.2 (0.4)**	3.3 (1.3)**	0.6 (0.8)
MS/E	6.1 (0.4)*	1.2 (0.4)*	0.7 (0.5)	1.3 (0.5)**	3.0 (1.2)**	0.7 (0.9)
Control	4.8 (0.4)	0.0 (0.0)	0.0 (0.0)	0.0 (0.0)	0.0 (0.0)	0.2 (0.1)

* $p < 0.05$ vs. control; ** $p < 0.01$ vs. control.

Lung tissue wet/dry ratios, histological injury score and number of TUNEL-positive cells (/mm²) in rats instilled with particulate meconium (MP, $n = 8$), particulate meconium after group I phospholipase A₂ (PLA₂-I) extraction (MP/E, $n = 10$), supernatant of meconium (MS, $n = 10$), meconium supernatant after PLA₂-I extraction (MS/E, $n = 10$) and saline (control, $n = 7$). A score from 0 to 4 was assigned for the extent of leukocyte infiltration, amount of intra-alveolar leukocytes and amount of exudative debris. The calculated total injury score means the sum of these scores. Mean (SD).

Table 2. Lung tissue results

Group	PLA ₂ activity	PLA ₂ -I	PLA ₂ -II	Wet/dry- ratio	Injury score	TUNEL-positive cells
hPLA ₂	204 (55)	282 (65)	188 (54)	6.3 (0.5)**	1.3 (1.0)†	1.3 (1.0)
bPLA ₂ /4	1412 (404)*	ND	119 (15)	5.6 (0.6)	0.0 (0.0)	0.7 (1.0)
bPLA ₂ /8	1993 (426)**;¶	ND	139 (2)	7.1 (0.4)**;§	0.0 (0.0)	0.7 (0.5)
Control	166 (53)	250 (134)	126 (65)	4.8 (0.4)	0.0 (0.0)	0.2 (0.1)

* $p < 0.05$ vs. control; ** $p < 0.01$ vs. control; ¶ $p < 0.05$ vs. hPLA₂, § $p < 0.01$ vs. bPLA₂/4† $p < 0.01$ vs. all the other groups.

ND, not determined.

Lung tissue phospholipase A₂ (PLA₂) catalytic activities (mU/g lung tissue), group I PLA₂ (PLA₂-I) and group II PLA₂ (PLA₂-II) concentrations (ng/g lung tissue), wet/dry ratios, histological injury score and number of TUNEL-positive cells (/mm²) in rats after intra-tracheal instillation of meconium-derived human PLA₂-I (hPLA₂, $n = 6$), bovine PLA₂-I (bPLA₂/4, $n = 5$) at a concentration of 4mg/ml, corresponding to the catalytic activity found in the particulate (65mg/ml) meconium, bovine PLA₂-I at double concentration (bPLA₂/8, $n = 5$) and sterile saline (control, $n = 7$). Mean (SD).

changes, the airway alterations in newborns tend to be more severe than those in adult lungs in response to a pulmonary insult (16). Thus, based on our present data, it may be suggested that meconium aspiration-induced lung damage is exacerbated by the high amount of pancreatic PLA₂ in meconium.

Corroborating our present results, several lines of clinical and experimental evidence implicate that excess pulmonary PLA₂ activity, through generation of pro-inflammatory lysophospholipids and eicosanoids, may contribute to the progression of various inflammatory lung disorders (5–8). Accordingly, lung PLA₂ catalytic activity is significantly increased in adults with acute respiratory distress syndrome and after experimental pulmonary insults correlating with the severity of the pulmonary failure (15,17–19). High PLA₂ activity in inflammatory lung injuries is commonly connected with increased release of secretory PLA₂-II originating from macrophages and platelets (5,9,15). The present data in contrast indicate that meconium aspiration may provoke inflammatory lung damage through pulmonary introduction of high amount of PLA₂-I with no influence on pulmonary PLA₂-II production. Our earlier data accordingly indicate that high PLA₂ catalytic activity found in human meconium is mainly (in > 90%) due to high concentration of human PLA₂-I (3). Although some experimental studies propose that PLA₂-I may induce, receptor-mediated, local PLA₂-II expression (20), this was not obvious at least in the early phase of meconium-induced lung injury. The reason for the apparently lacking local production of PLA₂-II and interaction between group I and II enzymes in our lung injury model remains unclear, but may be related to the short study period or varying catalytic or receptor-mediated interaction between these enzymes in the

insulted lungs (17,20,21). On the other hand, we cannot exclude the involvement of other phospholipases within aspirated meconium (22) or products of pulmonary cytosolic PLA₂ activation in the development of lung injury in our model (23).

PLA₂-I, earlier considered as a digestive enzyme secreted from the pancreas, is today known to be expressed and to modulate cellular function in a variety of tissues, including rat and human lungs (24,25). This enzyme is further able to stimulate cytokine chemokine and eicosanoid production from pulmonary cells involved in inflammatory responses and it may also directly stimulate neutrophil secretory function (26–28). These effects are shown to be exerted by mechanisms that are independent of the catalytic enzyme activity and are mediated by the interaction of secretory PLA₂s with specific membrane receptors (27,28). Mammalian PLA₂-I unlike group II PLA₂ enzyme, is further able to inactivate pulmonary surfactant concentration-dependently through hydrolysis of phosphatidylcholine (29). Experimental investigations have additionally shown that exogenously administered PLA₂-I may induce most likely through enhanced pulmonary thromboxane A₂ synthesis, receptor-mediated contractile responses in the airways (30). Similarly increased pulmonary thromboxane production may contribute to the development of vascular hypertension and edema formation in the meconium-insulted lungs (1,31). The observed challenge of the meconium-aspirated lungs with high exogenous activity of PLA₂-I may thus participate in propagation of the pulmonary ventilation disturbances hypertensive response and inflammation with ensuing respiratory failure in newborns (1–3,22,26). The variation of PLA₂-I content and enzyme activity in particulate meconium and its supernatant may further explain the ob-

served differences in the lung inflammatory reactions after aspiration.

Due to technical difficulties in the extraction of PLA₂-I from human meconium, the amount of this enzyme used for intra-tracheal instillation studies remained low, but was still able to induce significant edema formation and some histologic injury in the exposed lungs. In contrast, bovine PLA₂-I, even though given at a concentration corresponding to a similar or even higher catalytic PLA₂ activity than found in particulate meconium did not cause any histologic injury, but induced, in a concentration-dependent manner, lung edema formation. These data, again, strengthen the view that human PLA₂-I, either in soluble form or within meconium, is able to promote not only edema formation, but also neutrophil influx in the insulted lungs. The reason for the different responses to human and bovine PLA₂-I remains unclear, but may be related to varying histamine or eicosanoid release, connected to pancreatic PLA₂-induced permeability increase, or divergent chemokine and cytokine production in the lungs (27,30,32,33). In fact, earlier investigations have shown that mammalian pancreatic PLA₂s, despite having highly homologous structures, have different activity toward various lipid substrates (34).

In conclusion, our data indicate that intrapulmonary aspirated meconium challenges the lungs with high human pancreatic PLA₂ concentration and activity and may thereby contribute to the pulmonary inflammatory, but not apoptotic damage. Especially aspiration of thick particulate meconium through its high concentration of PLA₂-I may have the most deleterious pulmonary effects. These findings may be amenable to development of new modes of more specific therapeutic approaches.

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