The Apolipoprotein ε4 Allele Confers Additional Risk in Children with Familial Hypercholesterolemia

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ABSTRACT

Children with familial hypercholesterolemia (FH) exhibit substantial variance of LDL cholesterol. In previous studies, family members of children with FH were included, which may have influenced results. To avoid such bias, we studied phenotype in 450 unrelated children with FH and in 154 affected sib-pairs. In known families with classical FH, diagnosis was based on plasma LDL cholesterol above the age- and genderspecific 95th percentile. Girls had 0.47 \pm 0.15 mmol/L higher LDL cholesterol, compared with boys (p = 0.002). Also in girls, HDL cholesterol increased by 0.07 \pm 0.03 mmol/L per 5 y ($p_{\rm for}$ trend = 0.005; this age effect was not observed in boys. The distribution of apolipoprotein (apo) E genotypes was not significantly different between probands, their paired affected siblings, or a Dutch control population. Carriers with or without one $\epsilon 4$ allele had similar LDL and HDL cholesterol levels. Within the affected sib-pairs, the $\epsilon 4$ allele explained 72.4% of the variance of HDL cholesterol levels (-0.15 mmol/L, 95% confidence interval -0.24 to -0.05, p = 0.003). The effect of apoE4 on HDL cholesterol differed with an analysis based on probands or on affected sib-pairs. The affected sib-pair model used adjustment for shared environment, type of LDL receptor gene mutation, and a proportion of additional genetic factors and may, therefore, be more accurate in estimating effects of risk factors on complex traits. We conclude that the $\epsilon 4$ allele was associated with lower HDL cholesterol levels in an affected sib-pair analysis, which strongly suggests that apoE4 influences HDL cholesterol levels in FH children. Moreover, the strong association suggests that apoE4 carries an additional disadvantage for FH children. (*Pediatr Res* 53: 1008–1012, 2003)

Abbreviations

apo, apolipoproteinBMI, body mass indexFH, familial hypercholesterolemia

FH is an autosomal dominant disorder, strongly predisposing for premature coronary disease (1). In the Netherlands, heterozygous FH has a prevalence of approximately one in 400 individuals, making it one of the most common inherited disorders of metabolism (2). Inherited defects in the gene coding for the LDL receptor constitute the molecular basis for FH (3). As a result, LDL cholesterol is insufficiently taken up by its receptor and, subsequently, plasma LDL cholesterol levels are elevated (3).

In most FH patients, there is excessive deposition of cholesterol esters in the intima of the vasculature, leading to accelerated atherosclerosis and premature coronary artery disease. Although the clinical sequelae of atherosclerosis usually ensue in adult life, atherogenesis begins in early childhood. A significant positive correlation exists in childhood between serum LDL cholesterol and the extent of atherosclerosis in the coronary arteries and the aorta (4). Moreover, in adolescence, a strong relationship is observed between elevated cholesterol levels and the risk of subsequent coronary disease later in life (5).

However, among children with FH, a substantial variation in LDL cholesterol levels exists and was reported to be associated with age, gender, BMI, diet, and apoE phenotype (6). This suggests that the clinical phenotype in FH children may be influenced by a host of environmental and genetic factors.

Polymorphisms at the apoE locus are among the major factors affecting the variability of serum lipid levels in normal populations (7). Three common alleles, $\epsilon 2$, $\epsilon 3$, and $\epsilon 4$, code for three protein isoforms, apoE2, E3, and E4, resulting in six major phenotypes (8–10). The $\epsilon 4$ allele is associated with high, and the $\epsilon 2$ allele with lower total and LDL cholesterol levels (11–19). Furthermore, the $\epsilon 2$ allele has been associated with decreased cholesterol absorption (20), which then leads to

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increased cholesterol synthesis and up-regulation of the LDL receptor proteins (21, 22). Therefore, variation at the apoE gene locus influences LDL cholesterol metabolism and has been suggested to account for as much as 14% of the genetically determined variation in total serum cholesterol (7, 23).

However, in most previous studies in FH children, siblings were included and this selection on familial factors may have biased the analyses of the influences of additional factors. Therefore, we studied the clinical and biochemical phenotype in strictly unrelated FH children and in affected sib-pairs to estimate the contribution of age, gender, BMI, and apoE genotype to LDL and HDL cholesterol levels in these subjects. Here we report the results of our studies.

METHODS

Patients. A total of 450 children, 206 boys and 244 girls, were recruited from unrelated consecutive families. The FH parents had presented with the classical symptoms of FH (fasting plasma LDL cholesterol >4.9 mmol/L, and a personal or family history of tendon xanthomas and/or premature coronary artery disease) (24). In addition to vertical transmission from the FH parent, the diagnostic criteria in the children were based on fasting plasma LDL cholesterol above the age- and gender-specific 95th percentiles (25, 26). At present, the diagnosis of FH could be confirmed by molecular methods in 84% of the 450 unrelated probands. Sixty-nine different types of mutation were found in these 378 children [61 (16%) carried the frequent Dutch mutation N543H+2393del9, and 58 (15%) carried another frequent Dutch mutation, $1359-1(G \rightarrow A)$]. Children homozygous for FH were excluded. None of the children had diabetes mellitus, kidney, liver, or thyroid disease. At the time of sampling, the children did not use any medication known to influence lipid metabolism, except 13 girls who used oral contraceptives.

The study protocol was approved by the Institutional Review Board of the Academic Medical Center. DNA analysis was performed after informed consent was obtained.

Affected sib-pairs. Data were available for affected siblings (a brother or sister) of 154 index children. Out of the affected siblings of one particular family, we selected the second child that visited our outpatient Pediatric Lipid Clinic or by a random method (random numbers table) when more siblings visited the clinic together. The analyses were performed on the index child and one affected sibling only (154 concordant pairs). In 146 (95%) sib-pairs an LDL receptor gene defect could be demonstrated. In affected sib-pair analyses, the relatives were matched for the specific mutation in the LDL receptor gene, generation, and familial or environmental factors. In a matched multiple linear regression model, the effect of the ϵ 4 allele on the mean concentration of LDL and HDL cholesterol was estimated after adjustment for the difference in age, gender, and BMI within each affected sib-pair.

Controls. A control population of 2018 randomly selected 35-y-old Dutch males was used for comparison of the apoE allele frequencies. This group has been described in detail elsewhere (27).

Biochemistry. Blood samples were collected after at least a 12 h overnight fast. Plasma levels of cholesterol and triglycerides were determined by standardized enzymatic procedures (Roche Molecular Biochemicals, Mannheim, Germany) and HDL cholesterol in serum was measured by an automated method using polyethylene-glycol-modified enzymes and sulfated α -cyclodextrin (P-800 clinical chemistry analyzer, Roche Diagnostics, Basel, Switzerland) (28). LDL cholesterol levels were calculated using the Friedewald equation (29).

ApoA1 and apoB concentrations were assayed by an immuno-rate-nephelometric procedure using a polyclonal goatanti-human antiserum and were calibrated on World Health Organization-proposed international reference samples (30, 31). ApoE genotypes were identified by characteristic visible bands after amplification by PCR, restriction endonuclease digestion, and electrophoresis on 5% agarose gel, as described before (32).

Statistical analyses. The statistical analyses were performed by using SPSSWIN 10.0 (SPSS Inc., Chicago, IL, U.S.A.). Chi-square statistics were applied to test for Hardy-Weinberg equilibrium and to compare the apoE allele frequencies of the FH children with the Dutch control population. Lipoprotein levels are presented as mean \pm SEM in millimole per liter. The effects of age, gender, BMI, and apoE allele on the lipid profile were estimated simultaneously in a multiple linear regression analyses.

These effects were also estimated in affected sib-pairs by using a matched multiple linear regression model. The effects of differences in age, gender, BMI, and apoE allele on the differences in lipoprotein levels within the pairs were estimated simultaneously. These differences in age, gender, BMI, and apoE allele were not mutually related and were included concomitantly into the regression analyses. The determinant gender was scored: -1 when the index was a girl and the sibling a boy (38 pairs), 0 (identical gender, 84 pairs), or 1 (index a boy and sibling a girl, 32 pairs). In this way, adjustment for the exact differences in gender within the pairs was made.

RESULTS

General characteristics. A total of 450 consecutive, unrelated children with FH were recruited from one outpatient pediatric lipid clinic. The 206 boys and 244 girls had a mean age of 10.8 y [for boys, median age, 10.8 y (range, 2.0–18.7 y); for girls, median age, 10.6 y (range, 3.0–18.2 y); p = 1.0]. Twenty-seven children (6%) had already started smoking before their first visit. The girls had a mean BMI (± SEM) of 18.7 ± 0.3 kg/m² (median BMI, 17.9 kg/m²; range, 12.6–41.1 kg/m²) and the boys of 18.3 ± 0.2 kg/m² (median, 17.5 kg/m²; range, 13.1–30.2 kg/m²) (p = 0.2). Twenty-four children (5.5%) had palpable xanthomas. In 81 out of 244 girls (33%) menarche had occurred and 13 of them were using oral contraceptives.

Lipoproteins in 450 unrelated probands. Girls had a significantly higher plasma LDL cholesterol (5.99 \pm 0.10 mmol/L) compared with boys (5.54 \pm 0.10 mmol/L, p = 0.002). This difference did not change after adjustment for age and carrier

Table 1. ApoE genotype and ε allele frequencies
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0	FH children	Dutch controls	p
Genotype	(n = 450)	(n = 2018)	Value
E4E4	12	59	
E4E3	135	512	
E4E2	10	45	NS
E3E3	243	1128	
E3E2	50	261	
E2E2	0	13	
Allele	Frequency	Frequency	p Value
ε2	0.067	0.082	
ε3	0.746	0.750	NS
ε4	0.188	0.167	

status of at least one $\epsilon 4$ allele (0.47 \pm 0.15 mmol/L, p = 0.002). Even considering the wide range of ages among the unrelated children, we did not observe any influence of age on LDL cholesterol levels with or without adjustment for gender and $\epsilon 4$ carrier status (-0.05 \pm 0.10 mmol/L per 5 y, $p_{\text{for trend}} =$ 0.6). BMI and age were correlated ($r_{\text{partial}} =$ 0.57, p < 0.001) and had in similar models comparable effects on LDL cholesterol levels (data not shown).

The levels of HDL cholesterol were similar in boys and girls (both were 1.25 \pm 0.02 mmol/L; p = 1.0). In the entire group of children, the mean HDL cholesterol level showed no tendency over age (0.03 \pm 0.02 mmol/L per 5 y, $p_{\text{for trend}} = 0.1$). We repeated this analysis in boys and girls separately. In girls, HDL cholesterol increased with 0.07 \pm 0.03 mmol/L per 5 y ($p_{\text{for trend}} = 0.005$), whereas such an effect of age was not observed in boys. In a multiple regression model, menarche, oral contraceptives, and smoking did not explain the variation of HDL cholesterol in girls. An association was observed between triglyceride and HDL cholesterol levels, as expected. Additional adjustment for the triglyceride concentration did not change the results of the multivariate analyses.

No differences were observed in apoE genotype distribution ($\chi^2 = 7.198$, df = 5; p = 0.2) or allele frequencies ($\chi^2 = 4.091$,

df = 2; p = 0.1) between the children and a sample of 2018 Dutch males (Table 1). The results were similar after exclusion of the 13 apoE2 homozygotes from the Dutch control group. ApoE genotypes of the children and control subjects were in Hardy-Weinberg equilibrium ($\chi^2 < 4.551$, df = 3; p > 0.2). Carriers of the different apoE genotypes had similar fasting serum LDL cholesterol levels as shown in Table 2. Moreover, the carriers of an $\epsilon 4$ allele had similar LDL and HDL cholesterol levels to those of the carriers of other alleles (Table 2*A*). Adjustment for age, gender, and BMI did not change these results (data not shown).

ApoAI and apoB100 levels were available in 391 of the 450 unrelated children with FH. In Table 2*B*, the concentrations are shown according to apoE genotype and the presence of an $\epsilon 4$ allele. The apoB levels were lower in $\epsilon 2$ allele carriers. After exclusion of the $\epsilon 4 \epsilon 2$ carriers, the children with $\epsilon 4$ alleles had similar apoB levels compared with the children without $\epsilon 4$ alleles.

Lipoproteins in 154 affected sib-pairs. The contribution of the ϵ 4 allele to the variation of LDL and HDL cholesterol levels within the affected sib-pairs was estimated using a matched multiple linear regression model with adjustment for the differences in age or BMI, and gender. Because the affected siblings shared environment and an identical mutation in the LDL receptor gene with their probands, additional genetic factors are likely to explain the difference in LDL cholesterol levels. The presence of the $\epsilon 4$ allele did not contribute to the observed differences in LDL cholesterol levels (mean difference 0.16 mmol/L; 95% CI, -0.33 to 0.64 mmol/L; p = 0.2) within the affected pairs. The finding that apoB levels were lower in probands with an ϵ^2 allele could not be confirmed in the sib-pairs (-0.17 ± 0.12 g/L, p = 0.2), which supports our findings on LDL cholesterol levels. In the sib-pair analysis, we also did not find an effect of the presence of an ϵ 4 allele on apoB levels (0.003 \pm 0.01 g/L, p > 0.9).

Table 2A. Mean serum LDL- and HDL-cholesterol concentrations of 450 unrelated FH children according to apoE genotype or the
presence of the ε4 allele

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	E4E4 $(n = 12)$	E4E3 $(n = 135)$	E4E2 $(n = 10)$	E3E3 $(n = 243)$	E3E2 $(n = 50)$	p Value
LDL (mmol/L)	5.17 ± 0.36	5.88 ± 0.12	5.40 ± 0.28	5.84 ± 0.10	5.47 ± 0.23	0.2
HDL (mmol/L)	1.31 ± 0.08	1.25 ± 0.03	1.21 ± 0.08	1.25 ± 0.02	1.27 ± 0.03	0.9
		$\varepsilon 4 \ (n = 147)$		non $\varepsilon 4$ ($n = 293$)		p Value
LDL (mmol/L)		5.87 ± 0.12		5.78 ± 0.09		0.8
HDL (mmol/L)		1.25 ± 0.02		1.25 ± 0.02		1.0

Values are unadjusted mean serum concentrations \pm SEM; patients with the apo E4E2 genotype (n = 10) were excluded from the analyses.

Table 2B. Mean serum apolipoprotein AI and B100 concentrations of 391 of the 450 FH children according to apoE genotype or the presence of the $\varepsilon 4$ allele

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E4E4 $(n = 10)$	E4E3 $(n = 114)$	E4E2 $(n = 9)$	E3E3 $(n = 213)$	E3E2 $(n = 45)$	p Value
1.24 ± 0.05	1.27 ± 0.02	1.30 ± 0.06	1.27 ± 0.02	1.32 ± 0.03	0.7
1.54 ± 0.10	1.70 ± 0.03	1.39 ± 0.07	1.64 ± 0.03	1.50 ± 0.06	0.02
	$\varepsilon 4 \ (n = 124)$		non $\varepsilon 4$ ($n = 258$)		p Value
	1.27 ± 0.02		1.28 ± 0.01		0.6
	1.68 ± 0.03		1.62 ± 0.03		0.1
	1.24 ± 0.05	E4E4 (n = 10) E4E3 (n = 114) 1.24 ± 0.05 1.27 ± 0.02 1.54 ± 0.10 1.70 ± 0.03 $\varepsilon 4$ (n = 124) 1.27 ± 0.02	E4E4 (n = 10) E4E3 (n = 114) E4E2 (n = 9) 1.24 ± 0.05 1.27 ± 0.02 1.30 ± 0.06 1.54 ± 0.10 1.70 ± 0.03 1.39 ± 0.07 $\varepsilon 4$ (n = 124) 1.27 ± 0.02	E4E4 (n = 10) E4E3 (n = 114) E4E2 (n = 9) E3E3 (n = 213) 1.24 ± 0.05 1.27 ± 0.02 1.30 ± 0.06 1.27 ± 0.02 1.54 ± 0.10 1.70 ± 0.03 1.39 ± 0.07 1.64 ± 0.03 $\epsilon 4$ (n = 124) non $\epsilon 4$ (n = 258) 1.27 ± 0.02 1.27 ± 0.02 1.28 ± 0.01	E4E4 (n = 10) E4E3 (n = 114) E4E2 (n = 9) E3E3 (n = 213) E3E2 (n = 45) 1.24 ± 0.05 1.27 ± 0.02 1.30 ± 0.06 1.27 ± 0.02 1.32 ± 0.03 1.54 ± 0.10 1.70 ± 0.03 1.39 ± 0.07 1.64 ± 0.03 1.50 ± 0.06 $\epsilon 4$ (n = 124) non $\epsilon 4$ (n = 258) 1.27 ± 0.02 1.28 ± 0.01

Values are unadjusted mean serum concentrations \pm SEM; patients with the apo E4E2 genotype (n = 9) were excluded from the analyses.

 Table 3. Variance of mean paired differences in HDL-cholesterol levels as explained by specific determinants

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Determinant	HDL difference	%	p Value
ε4 allele	-0.15 (-0.24 to -0.05)	72.4	0.003
Age	-0.01 (-0.03 to -0.002)	7.2	0.02
Gender	-0.03 (-0.10 to 0.04)	14.7	NS
Unexplained	0.01 (-0.04 to 0.06)	5.7	NS

Values for HDL cholesterol are given as means in millimole per liter with 95% confidence interval in parentheses (The negative value of the difference in HDL cholesterol as a result of differences in ε 4 carrier status within the pairs is the result of a larger number of relatives with the ε 4 allele compared with the probands.)

Strikingly, the presence of the ϵ 4 allele explained 72.4% of the variance in HDL cholesterol levels (-0.15 mmol/L; 95% CI, -0.24 to -0.05; p = 0.003) (Table 3). In a similar model with identical results, differences in BMI significantly explained 14.5% of the variation of the mean paired differences in HDL cholesterol levels. Part of this may be attributed to an effect of differences in age, because the differences in age and BMI were correlated ($r_{\text{partial}} = 0.59$; p < 0.001). In agreement with our findings on HDL cholesterol levels, the ϵ 4 allele was associated with lower apoAI concentrations among the sibpairs (-0.12 ± 0.04 g/L, p = 0.002).

The distribution of the apoE genotypes was not significantly different between the probands, their paired affected siblings, and the Dutch control population. Nonetheless, the affected siblings were not in Hardy-Weinberg equilibrium, illustrating the dependency of this second sample coming from the same families ($\chi^2 = 8.365$, df = 3; p = 0.04).

DISCUSSION

We studied a large cohort of unrelated FH children with a wide age range and no concomitant disorders. The apoE genotype distribution and allele frequencies of our cohort did not differ from the general Dutch population and were in Hardy-Weinberg equilibrium (27). With these conditions met, it became possible to assess the impact of the apoE genotype on lipids and lipoproteins in these children.

In a wide range of populations, both adults and children, it has been convincingly demonstrated that the $\epsilon 4$ allele is associated with both increased LDL cholesterol and apoB, whereas the opposite is true for the $\epsilon 2$ allele (27, 33–38).

We could not, however, demonstrate a statistically significant effect of the $\epsilon 4$ allele on LDL cholesterol, HDL cholesterol, or triglycerides in our carefully recruited and unrelated pediatric FH cohort. Only the apoB levels were lower in $\epsilon 2$ allele carriers.

Our observations in these FH children suggest that the apoE genotypes have little influence on their lipid profiles. This is in contrast with normocholesterolemic children, whose $\epsilon 4$ alleles are associated with increased levels of LDL cholesterol (39–41). The variation of LDL cholesterol levels in healthy individuals is to a certain extent determined by the affinity of the different apoE isoforms for the LDL receptor. It is likely that in heterozygous FH, with 50% of LDL receptor activity, and LDL cholesterol levels twice the normal value, the subtle effects of apoE isoforms are nullified. A similar effect is seen with

variation in other genes affecting LDL cholesterol levels, such as cholesteryl ester transfer protein and microsomal triglyceride transfer protein (42). At young age, the presence of a LDL receptor gene mutation likely overrules other factors and influences the variation of LDL cholesterol levels in terms of a major gene. Our findings are similar to those in adults with FH, in whom the apoE phenotype has no influence on the lipid profile (43, 44). In adults, the type of mutation in the LDL receptor gene contributes to the variation of LDL and HDL cholesterol levels; moreover, it determines to a large extent the occurrence of tendon xanthomas (45). In our series of unrelated FH children, such mutation-related effects may have diminished the possibilities to quantify the contribution of the apoE genotype.

Besides these LDL locus effects, environmental factors may also contribute to the variation of the lipid profiles in FH children similar to normocholesterolemic children (39). Therefore, we also performed analyses in pairs of affected siblings. All affected siblings shared their environment and carried an identical mutation in the LDL receptor gene. As a result of matching these relatives, the effect of additional genetic factors to the variation of LDL and HDL cholesterol levels can be estimated independent of environment and type of LDL receptor gene mutation. In this model, the ϵ 4 allele did not explain variation of the LDL cholesterol levels, nor could the finding of lower apoB levels in $\epsilon 2$ allele carriers be confirmed in the sib-pairs. However, carriers of an ϵ 4 allele had lower HDL cholesterol levels and this allele explained >50% of the difference in HDL cholesterol levels within the affected pairs. The effect of the ϵ 4 allele on HDL cholesterol has therefore been underestimated in our series of unrelated children. This is probably a consequence of the contribution of the type of LDL receptor gene mutation to the variation of the HDL trait (45). In adults with FH, low HDL cholesterol levels associate with coronary artery disease (46). An ϵ 4 allele may therefore confer a serious disadvantage for FH children.

Moreover, a recent intriguing study showed that an ϵ 4 allele may have an untoward effect in survivors of myocardial infarction independent of lipoprotein levels (47).

The strength of the present study is that the influence of the ϵ 4 allele on the lipid profile was analyzed in both a series of probands and affected sib-pairs. As explained above, earlier studies contained both probands and relatives and, therefore, the findings may have been influenced by familial factors. These familial factors could consist of the mutation type in the LDL receptor gene, additional genetic factors, and environmental factors. Our analysis of a series of probands was relatively independent of such additional familial factors compared with the earlier studies. However, similar to these earlier studies, it may be biased by the type of mutation in the LDL receptor gene. The analysis of the affected sib-pairs avoids such bias by matching for the haplotype of the LDL receptor locus. In addition to matching based on the molecular defect in the LDL receptor locus, this analysis also adjusts for shared environment and shared genome. In the matched regression model, we further adjusted for differences in age, gender, and BMI to reduce the variance within the matched pairs caused by other sources than the apoE locus. The combination of analyses, in the present study, shows that the analysis of index cases is relatively insensitive to the influences of the apoE locus, whereas the affected sib-pair analysis allowed us to detect a remarkable lower concentration of HDL cholesterol in carriers of the $\epsilon 4$ allele compared with other alleles. Additional adjustment for differences in triglyceride levels did not change the results. In agreement with our findings on HDL levels, the $\epsilon 4$ allele was associated with lower apoAI levels among the sib-pairs.

In conclusion, the ϵ 4 allele did not cause variance of fasting LDL cholesterol level in children with FH. The carriers of this allele had significantly lower HDL cholesterol levels compared with the carriers of the other alleles. These effects were studied free from influences of the mutation type in the LDL receptor gene and other familial factors.

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