Relationship between Brain Blood Flow and Carotid Arterial Flow in the Sheep Fetus

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ABSTRACT. The present study investigates whether changes in total brain blood flow can be reliably estimated by changes in carotid arterial blood flow in fetal and perinatal lambs. We therefore compared carotid arterial blood flow, measured with implanted transit-time ultrasound transducers, with brain blood flow, measured by radioactive microspheres in fetal lambs during normal oxygenation and during pulmonary ventilation with oxygen, with PO₂ ranging from levels normal for the healthy fetus to levels normally seen postnatally. Cerebral perfusion pressure was modified over a wide range to alter brain blood flow: it was decreased by balloon occlusion of the brachiocephalic trunk and increased by a balloon occluder around the aortic isthmus. Carotid arterial blood flow and brain blood flow were closely related (r = 0.97, p < 0.0001). The relationship was not altered at different levels of oxygenation. However, measurements during higher cerebral perfusion pressures, obtained during aortic isthmus occlusion, had a negative influence on the agreement between carotid arterial blood flow and brain blood flow. When excluding values obtained by aortic isthmus occlusion, changes of 20% or more in brain blood flow could be predicted with carotid arterial blood flow within a confidence limit of 95%. Blood flow measurements in the carotid artery may be useful to estimate changes in brain perfusion. (Pediatr Res 35: 329-333, 1994)

Abbreviations

MCBP, mean carotid blood pressure Qbrain, blood flow to brain Qextracerebrai, blood flow to skin of the head and skull Qskin, blood flow to skin of the head Qskull, blood flow to skull Qcar, carotid arterial blood flow ID, inner diameter OD, outer diameter

The lamb fetus is frequently used in models to evaluate changes in antenatal and transitional Q_{brain} (1, 2). Cerebral perfusion usually has been measured by repeated injections of radioactive microspheres (3–5). This method has the advantages of reliability and the ability to determine regional as well as total Q_{brain} . The main disadvantage is the limited number of blood flow measurements that can be performed. An increasing number of studies

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investigating the pattern of fetal and neonatal Q_{brain} have used ultrasound-derived changes in the carotid artery as a measure of changes in Q_{brain} (6, 7). Although regional Q_{brain} cannot be determined with this method, frequent estimations of Q_{brain} can be made. Moreover, in long-term experimental studies, changes in total Q_{brain} can be monitored with ultrasonic flow transducers around the carotid arteries. Also, in clinical studies, ultrasoundderived changes in arteries supplying the brain, such as the carotid artery, are used to investigate the relationship between brain perfusion and various antenatal and perinatal complications, such as growth retardation and perinatal asphyxia (8–10). However, until now, no studies were performed to validate the relationship between changes in total Q_{brain} and Q_{car} .

We examined the relationship between changes in Q_{car} , measured with the transit-time ultrasound technique (11), and changes in radioactive microsphere-determined total Q_{brain} . We hypothesized that a close relationship and a good agreement existed between changes in flow measured by the two methods. This study was performed in fetal lambs before and during pulmonary ventilation and oxygenation to increase Po₂ to levels seen postnatally. Changes in Q_{brain} were induced by varying cerebral perfusion pressure over a wide range, including values well below and above the previously described limits of cerebral autoregulation (12, 13).

MATERIALS AND METHODS

Animals and surgical preparation. Before the initiation of these studies, all surgical and experimental procedures were approved by the Animal Care and Use Committee at the University of California, San Francisco. We studied 10 fetal sheep with gestational ages of 124–135 d and weights of 2550–4665 g. The ewes were fasted for 24 h before surgery. After local anesthesia with 2% lidocaine, low epidural anesthesia was achieved with 4 mL of 1% tetracaine hydrochloride (Pontocaine HCl; Breon Laboratories, New York, NY). Polyvinyl catheters (1.3 mm ID, 2.3 mm OD) were advanced from hind limb vessels into the ewe's descending aorta and inferior vena cava. NaCl (0.9% solution) was infused continuously into the ewe, and 50–100 mg of ketamine hydrochloride (Vetalar; Parke-Davis, Morris Plains, NJ) was administered to the ewe as necessary to maintain sedation.

The pregnant horn of the uterus was exposed through a midline incision in the ewe's abdomen, and a uterine incision was made overlying the fetal hind limbs. Local anesthetic (0.25% lidocaine hydrochloride) was administered before each fetal skin incision. Polyvinyl catheters (0.75 mm ID, 1.2 mm OD) were placed in the fetal descending aorta and inferior vena cava via the hindlimb vessels. The fetal skin and uterine incisions were sutured. Through a thoracotomy in the third intercostal space, catheters were placed in the brachiocephalic trunk from the internal thoracic artery (0.75 mm ID, 1.2 mm OD) and directly into the left atrium (20-gauge i.v. catheter, Becton Dickinson, Mountain View, CA) in those fetuses in which microsphere-determined

Q_{brain} measurements were planned during pulmonary ventilation and oxygenation (described later). Then silicone rubber inflatable balloons were placed around the aortic isthmus and the brachiocephalic trunk. The fetal chest was sutured in layers. The fetal head and neck were exposed through the same uterine incision. A 3.5-mm endotracheal tube was connected by an Y-tube to two 100-cm lengths of flexible polyvinyl tubing (3.2 mm ID, 6.4 mm OD). The tubing was filled with 0.9% saline, and the trachea was intubated. A second catheter was placed in the amniotic cavity and connected to the ventilation tubing to allow trachea fluid to flow to the amniotic cavity postoperatively. After exposing the carotid arteries in the neck, we placed appropriately sized transonic flow transducers (Transonic Systems, Inc., Ithaca, NY) around the carotid arteries to measure carotid blood flow (mL/ min). In four fetuses, a flow transducer was placed around both carotid arteries, and in the remaining six, only a single carotid artery was used for measuring carotid blood flow. The left forelimb was then exposed, and a polyvinyl catheter (0.75 mm ID, 1.2 mm OD) was passed from the brachial artery to the brachiocephalic trunk, distal to the balloon occluder, to measure arterial blood pressure and for blood sampling. All incisions were sutured, the vascular catheters were filled with heparin sodium solution (1000 IU/mL) and sealed, and all catheters and ventilation tubing were exteriorized to the maternal flank and protected by a cloth pouch. Antibiotics (400 mg gentamicin, 1 million IU penicillin) were given into the amniotic cavity and maternal vein on the day of surgery and each day thereafter. The animals were allowed 24-48 h to recover from surgery. Earlier studies in our laboratory on lamb fetuses with comparable gestational age and surgery showed that this period was adequate for hemodynamic stabilization.

Physiologic measurements. Fetal blood pressures were monitored continuously with Statham P23D6 strain-gauge transducers, and Beckman R 612 (Beckman Instruments, Inc., Palo Alto, CA) or Gould 2800S (Gould, Inc., Cleveland, OH) polygraphs. Fetal carotid blood pressure was calculated with amniotic cavity pressure as zero reference. Fetal heart rate, measured by a cardiotachometer, which was triggered by an arterial pulse wave, was also measured continuously. Blood samples for determination of pH, Po₂, PCO₂ (corrected for fetal temperature) and Hb (model 158 blood gas analyzer, Corning Medical, Medfield, MA; model OSM2 hemoximeter) were obtained from the carotid artery.

 Q_{car} was measured with the transit-time technique with ultrasonic flow transducers and ultrasonic transit-time blood flowmeter (Transonic Systems Inc.). The transonic flow transducer does not require close contact with the vessel. Reliable calibration and reproducibility have been shown in earlier studies (11, 14).

Blood flow to the head was measured with radionuclide-labeled microspheres as described previously (3). Microspheres, 15 μ m in diameter and labeled with ¹¹⁴In, ¹⁵³Gd, ⁵⁷Co, ⁵¹Cr, ¹¹³Sn, ⁸⁵Sr, ⁹⁵Nb, or ⁴⁶Sc were injected into the fetal inferior vena cava during baseline conditions and into the left atrium during ventilation with oxygen over a 30-s period while reference samples were obtained at a rate of 4 mL/min from the brachiocephalic trunk for 1.00-1.25 min. The microsphere injections were not associated with changes in heart rate or blood pressure. The volume of blood removed for analysis and for flow reference samples was replaced with an equal volume of a fetal donor or, in a minority of cases, with maternal blood. Blood to the head was determined as described previously in detail (3). Briefly, after completion of the study, the head was removed; separated into skin, skull, and brain tissue; weighed; and placed in formalin. The tissue was then carbonized in an oven at 350°C, ground into a coarse powder, placed into plastic vials to a uniform height of 3 cm, and counted in a 1000-channel multichannel pulse-height analyzer (Norland Corp., Fort Atkinson, WI). The activity of each isotope in the head was determined by the least-squares method as described previously (15). Q_{skin}, Q_{skull}, and Q_{brain} were calculated by comparing radioactivity of microspheres in the

tissue with flow rate of the appropriate reference sample from the brachiocephalic trunk and expressed as mL/100 g/min. Microsphere-determined cerebral blood flow was expressed in mL/min to compare Q_{brain} with Q_{car} .

The tubes connected to the endotracheal tube were opened and tracheal fluid was allowed to drain to provide ventilation to the fetus. These tubes were connected to a specially designed positive-pressure ventilator. On expiration, a solenoid in series with the expiratory tubing opened, and the expired air from the fetus was flushed from the tubing to the outside. This provided a small dead space for ventilation.

Experimental procedure. Ten fetuses were investigated during two conditions in a randomized order, with (n = 5) or without (n = 5) rhythmic pulmonary ventilation and oxygenation. The fetuses had ventilation with gases of various oxygen concentrations to achieve a range of arterial oxygen tensions. Po₂ in the ventilated fetuses ranged from values normal for the healthy lamb fetus (n = 2) to values normal for the (preterm) newborn lamb $(n = 3; Po_2 > 6 \text{ kPa})$. Surfactant (Exosurf, Burroughs-Wellcome Co., Research Park, NC) was instilled into the trachea before initiation of ventilation. Earlier studies in our laboratory have shown that changes in Q_{brain}, initiated by ventilation with oxygenation of the lamb fetus, are comparable to those occurring at birth (16).

After reaching a stable baseline level during each condition, carotid arterial pressure was decreased in steps of 5–10 mm Hg by inflation of the brachiocephalic trunk balloon. At each step, when a steady state (heart rate, carotid blood pressure) had been reached, blood samples were taken, hemodynamic parameters and Q_{car} registered, and blood flow to the head measured by microspheres. We aimed to decrease the cerebral perfusion pressure well below the lower limit of cerebral autoregulation for the fetal lamb of the gestational age studied (40–45 mm Hg) (12, 13). The brachiocephalic balloon was then released, and after reaching steady state again, cerebral perfusion pressure was increased by steps of 10 mm Hg by inflation of the aortic isthmus balloon until maximal inflation; all procedures were then repeated. Eight comparisons were made between Q_{car} and Q_{brain} in each fetus.

Statistical analysis. To investigate the relationship between Q_{brain} , determined by microspheres, and simultaneously determined Q_{car} , and the effect of oxygenation on this relationship, we used a multiple linear regression model with the following equation:

 $Y = b_0 + b_{Qbrain} Q_{brain} + b_{PO_2} PO_2 + b_{Qbrain} \times PO_2$

$$\cdot Q_{\text{brain}} \times PO_2 + \sum_{k=1}^{13} b_{1_k} L_k$$

where Y is the dependent variable Q_{car} and b_0 its overall mean value. Q_{brain} is the first independent variable, and its coefficient, b_{Qbrain} , defines the slope of the Q_{ear} - Q_{brain} relationship. The second (Po₂) and third ($Q_{brain} \times Po_2$) independent variables are action variables, whose coefficients, b_{Po_2} and $b_{Qbrain \times Po_2}$, indicate the independent effect of PO₂ on Q_{car} (affecting the intercept of the Q_{car} - Q_{brain} relationship) and the interactive effect of PO₂ and Q_{brain} on Q_{car} (indicating the effect of ventilation and oxygenation on the slope of the Q_{car} - Q_{brain} relationship), respectively. Finally, to correct for interanimal variability, 13 dummy variables (D_1 - D_{13}) were introduced for the 14 brain halves (see also Results) in 10 animals and included in this analysis (17). To determine the statistical significance of any variable, an F test was performed by dividing the mean square of that variable by the mean square residual.

To assess the agreement between Q_{brain} and Q_{car} as indicators for changes in cerebral perfusion, we used the method described by Bland and Altman (18). This method calculates the mean difference (d) between both measurements and the SD of the differences (s). From these data, the limits of agreement (d ± 2s) can be estimated. We therefore normalized the data per animal;

	Lowest MCBP	Baseline MCBP	Highest MCBP
Fetuses without ventilation			
MCBP (mm Hg)	19 ± 7	$52 \pm 13^{+}$	$81 \pm 8 \ddagger$
pH	7.36 ± 0.04	7.36 ± 0.02	7.38 ± 0.04
PCO ₂ (kPa)	7.6 ± 0.9	7.7 ± 0.7	7.3 ± 0.5
PO2 (kPa)§	3.1 (1.7–3.3)	3.1 (1.9–3.5)	3.2 (1.9-3.3)
Qextracerebral	$17 \pm 15 (12 \pm 10)$	$65 \pm 39 (40 \pm 21)$	$84 \pm 42 (47 \pm 27)$
Qbrain	$12 \pm 4 (14 \pm 9)$	$49 \pm 25 (102 \pm 59)^{\dagger}$	$58 \pm 21 (120 \pm 54)^{\dagger}$
% of Q _{brain} of total flow	41	43	41
Fetuses with ventilation			
MCBP (mm Hg)	22 ± 3	$53 \pm 10^{+}$	$78 \pm 12 \ddagger$
pН	7.40 ± 0.07	7.40 ± 0.07	7.43 ± 0.04
Pco ₂ (kPa)	6.0 ± 2.0	6.4 ± 0.6	6.4 ± 1.1
PO2 (kPa)§	11.7 (2.4–20)	7.7 (2.5–8.7)	5.9 (2.3-24.5)
Qextracerebral	$15 \pm 5 (10 \pm 5)$	$41 \pm 18 (26 \pm 9)$	$49 \pm 19 (32 \pm 12)$
Qbrain	$13 \pm 3 (23 \pm 5)$	$31 \pm 9 (56 \pm 16)^{\dagger}$	$44 \pm 15 (89 \pm 32)^{\dagger}$
% of Q _{brain} of total flow	45	43	47

Table 1. MCBP, pH and arterial blood gases, and radioactive microsphere-determined blood flow (in mL/min) to $Q_{extracerebral}$ and Q_{brain} at lowest, baseline, and highest MCBP values*

* Values are means ± 1 SD. Numbers in parentheses represent Q_{extracerebral} and Q_{brain} expressed as mL/100 g/min (means ± 1 SD).

 $\dagger p < 0.05 vs$ low MCBP.

p < 0.05 vs low and baseline MCBP.

Because oxygenation of fetuses with ventilation ranged from Po₂ values normal for the fetus to those normal for newborn lamb, the median and range for this variable has been given.

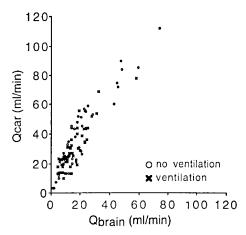


Fig. 1. Individual values of Q_{car} as a function of ipsilateral radioactive microsphere-determined Q_{brain} (n = 14).

values of Q_{car} and Q_{brain} during baseline level were considered to be 100%, whereas the other values were expressed as the percentage change from these baseline values.

Differences of blood gases, pH, MCBP, Qbrain, and Qextracerebral

between lamb fetuses without ventilation and with ventilation/ oxygenation were compared with the *t* test. Differences of these variables among baseline, lowest, and highest MCBP values were compared by one-factor analysis of variance for repeated measurements, followed by the Newman-Keuls test when a significant difference was found. Statistical significance was assumed as p < 0.05.

RESULTS

No differences were found for Pco_2 , pH, Hb, or heart rate between the fetuses without ventilation or with ventilation/ oxygenation at any MCBP level or among baseline, highest, and lowest MCBP values. Median Po_2 values in the fetuses with ventilation were higher than in those without ventilation. Table 1 summarizes these data. This table also shows microspheredetermined $Q_{\text{extracerebral}}$ (Q_{skin} and Q_{skull}) and Q_{brain} in the fetuses without ventilation and with ventilation/oxygenation at the different cerebral perfusion pressures. No significant difference was seen in the relationship between $Q_{\text{extracerebral}}$ and Q_{brain} among the different levels of perfusion pressure. The amount of blood perfusing the brain was always 41-47% of the total amount of blood flowing to the head regardless of cerebral perfusion pressures and whether the fetus had ventilation. Blood flow to the

Table 2. Results of multiple linear regression analysis of Q_{car} dependent variable on three independent variables*Independent

Independent variables	Coefficient	SEM	SC	t value	p value	
Intercept	8.07	2.93	8.07	2.76	0.01	
Q _{brain}	1.43	0.12	0.89	12.17	< 0.001	
PO ₂	-0.003	0.06	-0.01	-0.06	0.95	
$Po_2 \times Q_{brain}$	0.001	0.002	0.06	0.61	0.55	
ANOVA table	df	SS	MS	F value	p value	
Regression	16	37171.19	2323.20	53.92	<0.0001	
Residuals	80	3447.03	43.09			
Total	96	40618.23				

* The three independent variables were microsphere-determined Q_{brain} , Po_2 (whose coefficient b_{PO_2} indicate the independent effect of this variable on Q_{car}), and the interaction variable $Po_2 \times Q_{\text{brain}}$ (whose coefficient $b_{PO_2} \times Q_{\text{brain}}$ indicate the interactive effect of this variable on the slope of the $Q_{\text{car}}-Q_{\text{brain}}$ relationship); b_0 is the overall intercept. Animal variability was included in the model by introducing dummy variables, coded by effects, for the 14 brain-halves in 10 animals. The regression equation was statistically significant (r = 0.97, p < 0.0001). SC, standard coefficient; ANOVA, analysis of variance; SS, sum of squares; MS, mean square.

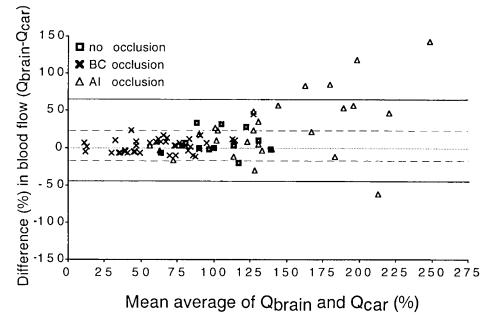


Fig. 2. Difference in changes (%) between Q_{car} and ipsilateral radioactive microsphere-determined Q_{brain} as a function of the mean average of Q_{car} and Q_{brain} . The two *solid horizontal lines* indicate the lower and upper limit of agreement (d ± 2s) when all measurements are included; the two *hatched horizontal lines* indicate the lower and upper limit of agreement without the measurements made after inflation of the aortic isthmus balloon. *BC*, Brachiocephalic; AI, aortic isthmus.

brain in relation to tissue weight was much higher than to extracerebral tissue. Ventilation and oxygenation tended to decrease cerebral and extracerebral blood flow, but these decreases were not significant. During baseline and highest arterial pressures, blood flow to extracerebral and cerebral tissue was significantly higher as compared with lowest carotid pressures.

In the four fetuses in which blood flow was measured in both carotid arteries, left and right Q_{car} was not significantly different (mean ± 1 SD; left carotid artery, 43 ± 17 mL/min; right carotid artery, 46 ± 18 mL/min). A close relationship was found in the fetuses between left and right carotid blood flow with a slope of the regression line close to 1 (r = 0.89, p < 0.001, y = 0.92x + 8.26).

Because blood flow was not measured in both carotid arteries in all fetuses, we divided the brain into left and right halves and compared Q_{car} with Q_{brain} (expressed as absolute flow, mL/min) in the ipsilateral halves of the brain. In this manner we compared 14 brain halves with the blood flow in 14 ipsilateral carotid arteries. Figure 1 shows the relationship between Qcar and the hemicerebral flow (r = 0.92, p < 0.001, y = 1.49x + 7.82). The multiple linear regression model, as described in Material and Methods, investigated the effects of oxygenation on the relationship and incorporated the animal variability. Table 2 shows the results: oxygenation had no effect on the intercept or the slope of the regression line of this relationship, whereas animal variability was not significant. Analysis of residuals reveiled a homogeneous distribution of the residuals versus either the fitted or the dependent variable (Table 2). Using the same multiple linear regression model, we found a similar close relationship between left or right Q_{car} and total Q_{brain} (r = 0.94, p < 0.0001; the coefficient b_{Qcar} was 0.85). Also, oxygenation here had no effect on intercept or slope of the regression line of the relationship. In this analysis nine dummy variables (D_1-D_9) were introduced for the 10 animals to correct for interanimal variability.

For several reasons, thoroughly discussed by Bland and Altman (18), correlation between two methods does not give us information about the error with which one can predict one measurement given the other. Therefore, we assessed the agreement between Q_{car} and Q_{brain} , which is shown in Figure 2. Taking all values into account, we found that agreement was poor; limits of agreement (d \pm 2s) ranged from -44% to +64%. However, when values obtained by aortic isthmus occlusion were excluded, the agreement improved considerably, with limits of agreement ranging from -19% to +25% (Fig. 2; *hatched horizontal lines*).

DISCUSSION

The present study in the fetal lamb shows a close relationship between changes in radioactive microsphere-determined total Q_{brain} and changes in Q_{car}. Oxygenation of the lamb fetus to levels normally seen in the newborn lamb did not alter this relationship. Of importance to potential clinical application is the fact that the proportion of total blood flow to the head that was directed to the brain tissue was similar over a wide range of cerebral perfusion pressures, including values well below and above the estimated limits of cerebral autoregulation. This situation was true for lamb fetuses without ventilation and those with ventilation (41-43% and 43-47%, respectively). In this respect it is important to state, however, that we investigated only healthy sheep. In case of brain injury (*e.g.* brain edema), the proportion between extracranial and intracranial blood flow may be changed. The agreement between both measurements, excluding values obtained during aortic isthmus occlusion, indicates that changes in Q_{brain} of 20% or more could be predicted from changes in Q_{car} within the 95% confidence interval in the lamb during fetal and transitional periods. This provides the advantages of monitoring brain perfusion continuously rather than intermittently, as with radioactive microspheres, as well as the ability to observe changes in brain perfusion as a function of time. If this relationship in the lamb were applied to the human fetus and infant, it would provide an opportunity to assess changes in Qbrain by measuring Q_{car} with noninvasive techniques. Although different in some respects, blood supply to the brain in both the lamb and the human fetus and newborn is predominantly provided by the carotid arteries (19-21). In the sheep, the carotid arteries predominantly supply the brain via the carotid rete, from which a short large vessel supplies all the blood to the circle of Willis, whereas the vertebral-basilar vascular system has no role or only a minor one (19, 20). Indeed, occlusion of both carotid arteries in the sheep has been shown to cause virtually complete ischemia with loss of electrical cortical activity in about 8 s (20). Eighty to 90% of the blood flow to the human brain is normally supplied

by the carotid arteries (22). In adults, Risberg et al. (21) showed a significant relationship between Doppler ultrasound-measured common Q_{car} velocity and ¹³³Xenon inhalation-measured Q_{brain}. In our study, actual Qcar was measured. Doppler ultrasonography, mostly used in the clinical studies, measures blood flow velocity and not absolute flow. To calculate absolute flow, it is necessary to measure the cross-sectional area of the vessel and the average blood flow velocity. Velocity is related to absolute flow if the internal diameter of the vessel does not change. Although major changes in diameter of carotid arteries do not usually occur, there are small changes during the cardiac cycle and during changes of cerebral perfusion pressure (23-25). Moreover, Sonesson and Herin (26) clearly showed that Doppler-measured blood flow velocity in large intracranial arteries in newborn lambs underestimated changes in actual Q_{brain} induced by changes in arterial carbon dioxide tension, probably because of a concomitant change in internal diameter of the arteries under investigation.

The reason for the relatively poor agreement between Q_{car} and Q_{brain} when measurements obtained during aortic isthmus occlusion are included is not apparent. A possible explanation is that at higher cerebral perfusion pressures, collaterals between vertebral and ventral spinal arteries may be recruited. These collaterals may become important shunts for the cerebral blood supply via the normal small basilar artery, which normally does not contribute to the blood supply of the circle of Willis (19, 20).

In conclusion, transit-time ultrasound-monitored Q_{car} (mL/ min) shows a close linear relationship with radioactive microsphere-determined blood flow to the brain in the fetal lamb. This relationship is not altered by pulmonary ventilation and oxygenation of the fetus. The assessment of agreement showed that changes in Q_{brain} larger than 20% can be monitored by this method, although aortic balloon occlusion-induced higher cerebral perfusion pressures have a negative influence on the agreement between Qcar and actual Qbrain.

REFERENCES

- 1. Rudolph AM, Heymann MA 1967 The circulation of the fetus in utero. Methods for studying distribution of blood flow, cardiac output and organ blood flow. Circ Res 21:163-184
- 2. Papile L, Rudolph AM, Heymann MA 1985 Autoregulation of cerebral blood flow in the preterm fetal lamb. Pediatr Res 19:159-161
- 3. Heymann MA, Payne BD, Hoffman JIE, Rudolph AM 1977 Blood flow measurements with radionuclide-labeled particles. Prog Cardiovasc Dis 20:55-79
- 4. Thiringer K, Hrbek A, Karlsson K, Rosen KG, Kjellmer I 1987 Postasphyxial

cerebral survival in newborn sheep after treatment with oxygen free radical scavengers and a calcium antagonist. Pediatr Res 22:62-66

- 5. Rosenberg AA, Murdaugh E, White CW 1989 The role of oxygen free radicals in postasphyxia cerebral hypoperfusion in newborn lambs. Pediatr Res 26:215-219
- 6. Wladimiroff J, Tonge H, Stewart P 1986 Doppler ultrasound assessment of cerebral blood flow in the human fetus. Br J Obstet Gynaecol 93:93-99
- 7. van Bel F, Klautz RJM, Schipper IB, Teitel DF, Baan J 1993 Auto-regulatory ability of cerebral circulation before and after indomethacin in the newborn lamb. Pediatr Res 34:178-181
- 8. Wladimiroff JW, van de Wijngaard JAGW, Degami S, Noordam MJ, van de Eyck J, Jonge MM 1987 Cerebral and umbilical arterial blood flow velocity wave form in normal and growth-retarded pregnancies. Obstet Gynecol 69:705-709
- 9. Lingman G, Marsal K 1989 Noninvasive assessment of cranial blood circulation in the fetus. Biol Neonate 56:129-135
- 10. van Bel F, Van de Bor M, Stijnen T, Baan J, Ruys JH 1987 Cerebral blood flow velocity pattern in healthy and asphyxiated newborns: a controlled study. Eur J Pediatr 146:461-467 11. Dobson AD, Sellers AF, McLeod FD 1986 Performance of a cuff-type blood
- flowmeter in vivo. J Appl Physiol 21:1642-1648
- 12. Szymonowicz W, Walker AM, Yu VY, Stewart ML, Cannata J, Cussen L 1990 Regional cerebral blood flow after hemorrhagic hypotension in the preterm, near-term, and newborn lamb. Pediatr Res 28:361-366
- Purves MJ, James IM 1969 Observations on the control of cerebral blood flow in the sheep fetus and newborn lamb. Circ Res 25:651–667
- 14. Hartman J, Koernier J, Lancaster L, Gorczynski R 1985 In vivo calibration of a transit-time ultrasound system for measuring ascending aorta volume flow. Pharmacologist 27:127-132
- Baer RW, Payne BA, Verrier ED, Vlahakes GJ, Molodowitch D, Uhlig PN, Hoffman JIE 1984 Increased number of myocardial blood flow measurements with radionuclide-labeled microspheres. Am J Physiol 246:H418-H434
- 16. Iwamoto HS, Teitel DF, Rudolph AM 1987 Effect of birth-related events on blood flow distribution. Pediatr Res 22:634-640
- 17. Slinker BK, Glantz SA 1990 Missing data in two-way analysis of variance. Am J Physiol 258:R291-R297
- 18. Bland JM, Altman DG 1986 Statistical methods for assessing agreement between two methods of clinical measurement. Lancet 1:307-310
- 19. Baldwin BA, Bell FR 1963 Blood flow in the carotid and vertebral arteries of the sheep and calf. J Physiol (Lond) 167:448-462 20. May NDS 1977 The Anatomy of the Sheep: A Dissection Manual, 3rd Ed.
- University of Queensland Press, Queensland, Australia, pp 237-240
- Risberg J, Smith P 1980 Prediction of hemispheric blood flow from carotid velocity measurements: a study with the Doppler and ¹³³Xenon inhalation techniques. Stroke 11:399-402
- Roberts B, Hardesty WH, Holling MC, Reivich M, Toole JF 1964 Studies on extracranial cerebral blood flow. Surgery 56:826-839
 Kontos HA, Wei EP, Navari RM 1978 Responses of cerebral arteries and
- arterioles to acute hypotension and hypertension. Am J Physiol 234:H371-H378
- 24. Ulmatsu S, Yang A, Preziosi TJ 1983 Measurement of carotid blood flow in man and its application. Stroke 14:256-262
- 25. Faraci FM, Heistad DD 1990 Regulation of large cerebral arteries and cerebral microvascular pressure. Circ Res 66:8-17
- 26. Sonesson SF, Herin P 1988 Intracranial arterial blood flow velocity and brain blood flow during hypocarbia and hypercarbia in newborn lambs: a validation of range-gated Doppler ultrasound flow velocimetry. Pediatr Res 24:423-426