CHARACTERIZATION OF SERUM ANTI-GLIADIN IGA IN CHILD-HOOD COELIAC DISEASE VERSUS OTHER GASTROINTESTINAL

35 DISEASES AND CONTROLS

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IMMUNOLOGICAL PROPERTIES OF GLIADIN PRESENTED VIA THE GUT

36 R Troncone, A Ferguson Gastro-Intestinal Unit, University of Edinburgh and Western General Hospital, Edinburgh, Scotland, UK When an antigen is first presented via the gut, either priming or suppression of the systemic immune response (oral tolerance) can Aim of this study was to establish if wheat gliadin result. behaves as an oral immunogen or tolerogen. BALB/c mice were reared on normal diet (with 0.3% gliadin) or gluten free diet (second or later generation). Antigen was fed either as normal diet or as pure gliadin weighed and presented in The immune status (tolerant or sensitized) was an agar pellet. then defined by the antibody and CMI response to parenteral immunization (ELISA and footpad swelling test). Mice on a normal diet or fed this diet for a week before immunization showed a significantly (p<0.05) reduced humoral response to gliadin compared to mice reared on gluten free diet. CMI was reduced to a lesser extent (p<0.1). Mice fed pure gliadin (5, 25 or 125 mg) one week before immunization had suppression of both antibody (p<0.02) and CMI (p<0.05) at the higher dose. 25 mg feed suppressed only the humoral response. These results indicate that wheat gliadin can act as an

effective oral tolerogen; however the type of immune response is influenced both by the amount and the form in which the antigen is presented to the gut.

> INTESTINAL MUCOSA OF COELIACS IN REMISSION IS UNABLE TO REMOVE TOXI-CITY OF GLIADIN PEPTIDES ON IN <u>VITRO</u> DEVELOPING FETAL RAT INTESTINE

37 CITV OF GLIADIN PEPTIDES ON IN VITRO DEVELOPING FETAL RAT INTESTINE AND CULTURED ATROPHIC COELIAC MUCOSA. <u>H.J.Cornell°,S.Auricchio*</u>, <u>G.de Ritis*,M. De Vincenzi°°,L. Majuri*,V. Raia*,V. Silano</u>°°. [®]Dept of Applied Chemistry, Royal Melbourne Institute of Technology,

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Subfraction 2R of fraction 9 of a wheat gliadin's peptic-tryptic-pancreatic digest is known to be toxic <u>in vivo</u> for coeliacs (H.J.Cornell et al., Clin.Chim. Acta 31,123,1982). We have found that fractions 9 and 2R agglutinate K 562(S) cells and inhibit the <u>in vitro</u> development of fetal rat intestine and the increase of enterocyte height ∞curring in organ culture of atrophic coeliac mucosa (0.1-0.5 mg/ml medium). Other peptide fractions of the gliadin digest are devoid of such <u>in vitro</u> effects. Fraction 2R, after incubation with morphologically normal small intestinal mucosa of coeliacs in remission and ultrafiltration, was still able to agglutinate K cells and was very active in both culture systems, at low concentration (0.1 mg/ml); on the contrary, fraction 2R was inactivated after incubation with normal mucosa. In intestinal mucosa of coeliacs in remission either a primary (or secondary) enzyme deficiency or some other mechanisms may explain these results which are compatible with the hypothesis that there is a mucosal defect in handling gliadin peptides in coeliac disease.

A-GLIADIN RELATED SYNTHETIC PEPTIDES AGGLUTINATE UNDIFFERENTIATED K

38 562 S CELLS AND AFFECT IN VITRO DEVELOPING FETAL RAT INTESTINE AND CULTURED ATROPHIC COELIAC MUCOSA. S. Auricchio*, A. Arcoo*, G. D'Auria*, G. de Ritis*, M. De Vincenzi**, G. Magazzů**, L. Maiuri*, V. Pavone*, V. Raia*, V. Silano**.

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Peptides from wheat gliadins, A-gliadin and prolamins from cereals toxic for coeliac patients agglutinate K 562(5) cells; they also damage <u>in vitro</u> cultured fetal rat intestine and atrophic coeliac mucosa. The largest common sequences among the <u>in vitro</u> active A-gliadin peptides were -Pro-Ser-Gln-Gln and -(Gln)₃-Pro-. The following peptides all containing the aminoacid sequence -(Gln)₃-Pro have been synthesized: the pentapeptide Tyr-(Gln)₃-Pro, its dimer and the rater and the eptapeptide Gln-Pro-Tyr-(Gln)₃-Pro in their free and N-acetylated forms and the Pyroglutamic derivate of the heptapeptide (Pyr 7). Pyr 7 agglutinated cells and inhibited the <u>in vitro</u> development of fetal rat intestine (medium's concentration 0.5-2 mg/ml); it was non toxic on the <u>in vitro</u> cultured coeliac atrophic mucosa. The N-acetylated form of the pentapeptide's tetramer (1 mg/ml) also damaged the atrophic coeliac mucosa in 3 cultured biopsies. These results suggest that the sequence -(Gln)₃-Pro when part of a larger peptide may be toxic <u>in vitro</u> for the atrophic coeliac mucosa.

AN IN VITRO MODEL SYSTEM FOR STUDYING THE BIOSYNTHESIS OF BRUSH BORDER MEMBRANE (BBM) HYDROLASES IN COELIAC DISEASE (CD). H.Y. Naim, P. Ambühl, E.E. Sterchi, H. Gaze, M.J. Lentze. Dept. of Pediatric Gastroenterology, University of Berne, Inselspital, Switzerland.

In an effort to mimic the in vitro situation in CD, the effect of gliadin peptides III and IV on the biosynthesis of a number cf BBM hydrolases was studied. Thus 5 biopsies from patients with CD in remission were labelled with ³⁵S-methionine for 24 hours in the presence or absence of gliadin and α -casein. Sucrase (SI), lactase (LPH), maltase (MGA), aminopeptidase N (APN), di-peptidylpeptidase IV (DPPIV) and angiotensin-converting enzyme (ACE) were immunoprecipitated with monoclonal antibodies and analyzed by SDS-PAGE, fluorography and densitometric scannings. Gliadin inhibitory effect on the synthesis of the hydrolases is expressed as % decrease in labelling intensity versus control. Thus SI synthesis was inhibited by 72%, that of MGA and APN by 85-88%, whereas DPPIV, ACE and LPH were completely absent in gliadin-treated biopsies. a-Casein affected the synthesis of SI, LPH and ACE by about 35%, APN was not affected whereas DPPIV was to about 55 %. These values are significantly lower than their counterparts in gliadin-treated biopsies. Conclusion: This in vitro system provides therefore a promising approach for studying the mechanisms involved in the induction of mucosal damage at the molecular level.

SELECTIVE BINDING TO MANNAN OF GLIADIN PEPTIDES WHICH ACCLUTINATE

40 UNDIFFERENTIATED K 562 S CELLS AND INHIBIT <u>IN VITRO</u> DEVELOPMENT OF FETAL RAT INTESTINE. <u>L. Maiuri°,S. Auricchio°,M. Cardelli*,G. de</u> <u>Ritis°,M. De Vincenzi*,F. Latte°,E. Mancini*,V. Raia°,V. Silano*</u>. °Clinical Pediatrics,II Medical School, University of Naples,Naples,

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Mannan and N,N'-diacetyl-chitobiose and N,N',N"-triacetyl-chitotriose prevent the agglutinating activity on K 562(S) cells and the damaging <u>in vitro</u> effect on fetal rat intestine and atrophic coeliac mucosa of mixtures of gliadin peptides and pure A-gliadin peptides (S.Auricchio et al.,J. Ped. Gastroenterol. and Nutr. 4,923, 1985). We separated by cromatography on mannan-Sepharose 4-B of peptic-tryptic(PT) digest of gliadins: 1) A small fraction (C) which was very active in agglutinating cells and inhibiting the <u>in vitro</u> development of fetal rat intestine (20 y/ml medium). Both effects were prevented by the sugars. 2) A much larger fraction (B) which is devoid of both activities. These results show that only a few peptides of a gliadin digest, which are specifically bound by mannan, are active in the two <u>in vitro</u> systems.

	Peptide fraction	Minimal concentration agglu- tinating all cells (mg/1)	Percentage of the total digest
B)	PT digest Fraction not bound	73	100
	Fraction bound by mannan mannan	No agglutination up to 4666	92-97
		0.8	0.15-0.2