and IRVING SCHULMAN. Dept. of Ped., The Abraham Lincoln Sch. of Med., Univ. of Illinois, Chicago, IL.

nois, Chicago, IL. Platelet aggregation was studied in platelet-rich plasma (PRP) prepared from samples of maternal and cord blood taken immediately after delivery. When mothers had received no drugs for analgesia, adenosine diphosphate- and collagen-induced aggregation curves of platelets from the infants and from the mothers were virtually identical and were within the normal range. When mothers (18) had received drugs (acetylsalicylic acid, promethazine, alphaprodine and meperidine) prior to delivery, 17 of 18 of the cord blood samples showed a marked decrease in collagen-induced platelet aggregation. By contrast, in paired PRP samples from these mothers abnormalities of collagen-induced platelet aggregation were found in only 25%, and these changes were minimal. These findings suggested that the infants' platelets were more susceptible to the druginduced suppression of collagen aggregation than were platelets of the mothers. Dose response curves to promethazine added in vitro demonstrated a markedly increased susceptibility of newborn platelets when compared with those of the mothers. These results indicate that a variety of drugs given to mothers before delivery may alter platelet aggregation of the newborn infants, resulting in significant impairment of the plateletmediated phase of hemostasis.

34 Plasma 17-OH Progesterone in Maternal and Umbilical Cord Plasma in Children, and in Congenital Adrenal Hyperplasia (CAH): Application to Neonatal Diagnosis of CAH. MORRIS R. JENNER, MELVIN M. GRUMBACH and SELNA L. KAPLAN. Univ. of California, San Francisco Med. Center, San Francisco, CA.

The concentration of plasma 17a-OH progesterone (17-OHP) was determined utilizing a ligand binding radioassay at delivery in maternal (8) and umbilical venous (8) blood and in the blood of newborn infants (8), children (12), adult males (13), and pre- and post-treatment in 6 patients with CAH. Adult male values (mean =  $0.106 \ \mu g/100 \ ml$ ) are in agreement with the data of STROTT *et al.* [J.clin. Invest. 48: 930, 1969]. Mean concentration in maternal plasma at delivery was  $0.365 \,\mu\text{g}/100 \,\text{ml}$  and in cord plasma  $1.64 \,\mu\text{g}/100 \,\text{ml}$ . Twelve newborns (age 1–7 days) demonstrated a rapid fall in concentration of plasma 17-OHP within the first day of life (range <0.100–0.125  $\mu$ g/100 ml). Twelve normal children (age 5–11 years) had a mean level of 0.036  $\mu$ g/100 ml (range 0.016–0.059). In contrast, 5 children (age 4 days–7 years) and an adult (age 26 years) with untreated CAH had a mean value of 12.8 µg/100 ml (range 2.4–33.0 µg/100 ml). A 4-dayold female pseudohermaphrodite, the youngest subject with CAH, had a plasma 17-OHP of 5.7  $\mu$ g/100 ml. In 1 infant with CAH the concentration of plasma 17-OHP fell from 11.4  $\mu$ g% to 0.12 $\mu$ g% on cortisone therapy, and in a female, age 2–7/12 years, from 33.0  $\mu$ g% to 0.148  $\mu$ g/100 ml. In 4 patients with CAH, changes in plasma testosterone paralleled those in 17-OHP. The results suggest that estimation of plasma 17-OHP in a 0.2 ml sample by this method permits rapid identification of the 21 hydroxylase deficiency form of CAH by the 2nd day of life and is useful in assessing the adequacy of glucocorticoid treatment. The high concentration of plasma 17-OHP in cord blood is attributable to the capacity of the placenta to convert maternal and fetal steroid precursors to 17-OHP.

## INDEX OF ABSTRACTS

(Numbers following entries refer to abstract number)

AAGENAES, O. 23 adenoids 1 ADRIANZEN, T. B. 16 amniocentesis 25 amniotic fluid 4 antibody, rubella 7 ARBIT, J. 28 BAKER, L. 22 BEAUDRY, P. H. 27 BECK, J. 26 BERLIN, C. M. 25 bile 23 BLANC, W. 15 BLONSKY, E. R. 28 blood 33 BRADLEY, K. H. 32 brain 31 CALDWELL, B. M. 13

carbohydrate metabolism 11, 22 care, pediatric 2 CasseLI, S. 28 cell hybridization 18 central nervous system 28, 31 CHABOT, A. 10 CHACKO, C. M. 18 CHAPMAN, E. M. 14 child care 10, 13 children 26 cholestasis 23 chromosomes 25 COCHRAN, W. 9 COHEN, M. M. 17 community health program 10 CONDON, A. 3 congenital adrenal hyperplasia 34 congenital anomalies 25 congenital rubella 7 COOPER, L. Z. 7 CORBY, D. G. 33 CRAWFORD, J. D. 14 CUDERMAN, B. 23 cystine 32 cystinosis 32

deafness 7 deoxyadenosyl-B<sub>12</sub> 12 development 31 developmental pharmacology 33 diagnosis, prenatal 4 DIMAURO, S. 3 DORFMAN, A. 4

Embryo, mouse 17 endocardial fibroelastosis 24 enzyme 22 erythroblastosis 9 erythrocytes 6 exfoliation 29

Fertilization, in vitro 17 fetal transfusions 9 fetus 3 fibrinolysis 5 fibrin split products 5 FLORMAN, A. L. 7 fructose-1,6-diphosphatase 22

Galactose 1-phosphate uridyl transferase 18 genetic disease 12, 21 GIBSON, M.S. 33 GINSBERG-FELLNER, F. 11 GLASGOW, L.A. 29 glomerulonephritis 19 gluconeogenesis 22 glucose metabolism 31 glycolysis 3 GOODMAN, J.R. 26 GORDIS, L. 2 GRAHAM, G.G. 16 growth 16 growth 16 growth 16 growth failure 21 GRUMBACH, M. M. 34

HALSTEAD, S.B. 8 HAYEK, A. 14 hemoglobin SS disease 6 *H. influenza*, type b 20 HORSTMANN, D. M. 8 Hunter's syndrome 4 Hurler's syndrome 4 21-hydroxylase deficiency 34 hypoglycemia 22 hypoxemia 30

Immunity 1, 7, 8, 20 immunization 20 inanition 31 infant 22, 25 infant mortality rate 10 infants 9, 27, 28, 33, 34 infants, low birth weight 15 infection, intrauterine 15 intragenic complementation 18 intrauterine blood transfusion 9,28 intravascular coagulation 5 JACOBSON, C.B. 4, 25 Jankus, E. F. 24 Jenner, M.R. 34 KAPLAN, S.L. 34 Kazemi, H. 30 kidney 5, 19, 26 KNITTLE, J. L. 11 KRIVIT, W. 23 KRUGMAN, S. 7 KUPLIĆ, L.S. 5 LANGE, K. 19

LEONARD, A.S. 23 LIEBHABER, H. 8 lipocytes 11  $\beta$ -lipoproteinemia, hypo 21 liver 22, 23 lung 27 lung function 30 lymphedema 23 lymph flow, hepatic 23

## Index of Abstracts

lysergic acid diethylamide 25 Mahoney, M.J. 12 malnutrition 16 MARKOWITZ, M. 2 MATALON, R. 4 Mellish, M. E. 29 Mellman, W. J. 3 metabolic acidosis 22 methylmalonicaciduria 12 methylmalonyl-CoA mutase 12 monosodium glutamate 11 mucopolysaccharides 4 MUKHERJEE, A. 17 myocarditis 24 Nadler, H.L. 18 Naeye, R. 15 nephritis 26 nephritogenic antigen 19 newborn 33 newborns 20 Nogrady, B. 27 Noren, G.R. 24 Obesity 11 Одга, Р. L. 1 Он, W. 28 Olson, W.H. 32 Oski, F.A. 3 Outerbridge, E.W. 27 oxidative metabolism 31 PARKE, J. C., Jr. 20 PARTIN, J. C. 21 PEARSON, H. A. 6 PIEL, C. F. 26 placenta 33, 34 platelets 33 poliovirus 1 poverty 15 pregnancy 25 prematurity 15 17-OH progesterone 34 Race 15 RACHMELER, M. 18 radioiodine 14 renal disease 26 respiratory distress syndrome 27 Rh erythroblastosis 28 rheumatic fever 2 RICHMOND, J.B. 13 RISEBOROUGH, E. P. 30 ROBBINS, J. B. 20 RODRIGUES, L.P. 20 ROSENBERG, D.M. 8 ROSENBERG, L.E. 12 round heart disease 24 rubella 8

rubella vaccine 8 rubella virus vaccine 7 SAGEL, I. 19 SATTERTHWAITE, H.S. 31 Scheinman, J.I. 5 Schneerson, R. 20 Schubert, W.K. 21 Schulhoff, C. 9 SCHULMAN, I. 33 SCHULMAN, J. D. 32 Schwartz, A.D. 6 scoliosis 30 SEEGMILLER, J.E. 32 SHANNON, D.C. 30 SHARP, H.L. 23 sickle cell anemia 6 SIGSTAD, H. 23 skeletal development 11 Smith, A.L. 31 Sokoloff, L. 31 Spencer, R.P. 6 spleen 6 STALEY, N.A. 24 STAMBAUGH, M.C. 12 staphylococcal scalded skin syndrome 29 staphylococcus 29 STARK, A. 9 STERN, L. 27 STEVENSON, J.E. 24 STIEHM, E.R. 5 streptococcus 19 systemic lupus erythematosus 26 Thyroid cancer 14

thyrotoxicosis 14 tonsils 1 toxin 29 TRYGSTAD, C.W. 5 TURNER, M.D. 29 Ty, A. 19

UEHLING, D.T. 5 umbilical cord 34

Vaccine, H. influenza type b 20 VALENCA, L. M. 30 virus disease 26 vitamin B<sub>12</sub> 12

White villus disease 21 WINEGRAD, A. I. 22 WOLFSON, S. L. 6 WONG, V. G. 32

Zelson, J. H. 6 Zirbel, C. L. 33 Ziring, P. R. 7