Water-soluble complex formation of fullerenes with a biocompatible polymer

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Fullerenes (C_{60} or C_{70}) and water-soluble poly(2-(methacryloyloxy)ethyl phosphorylcholine) (PMPC_n) were mixed by physical means to prepare water-soluble fullerene/PMPC_n complexes. The ultraviolet-visible absorption spectra confirmed the presence of aqueous solutions with high fullerene concentrations. The fullerene/PMPC_n complexes were characterized using light scattering measurements, small-angle X-ray scattering measurements and transmission electron microscopic observations. The complexes generated singlet oxygen upon visible light irradiation.

Polymer Journal (2016) 48, 999-1005; doi:10.1038/pj.2016.60; published online 8 June 2016

INTRODUCTION

Fullerenes have been investigated for applications such as novel pharmaceuticals and cosmetics.¹ C₆₀ fullerenes and their derivatives have been reported to possess activity as enzyme inhibitors,² antivirals,³ DNA scission agents,⁴ radical quenchers⁵ and photodynamic therapy (PDT) agents.⁶ In addition, C₆₀ and C₇₀ fullerenes have been proposed as photosensitizers for PDT to eliminate cancerous tissue because fullerenes can efficiently generate active oxygen upon visible light irradiation.⁷ However, biological applications of fullerenes have been limited because the solubility of fullerenes in water is extremely low.8 Various C₆₀ fullerene derivatives have been synthesized to improve water solubility,9 and methods to improve the solubilization of fullerenes in water have been studied.^{10,11} In addition, solubilization of C₆₀ in water has been investigated using inclusion complexes composed of C₆₀ and host molecules, such as cyclodextrin and calixarene,^{12–15} micelles,¹⁶ liposomes¹⁷ and poly(N-vinylpyrrolidone) (PVP).^{18,19} In particular, PVP can form water-soluble complexes with fullerenes simply by grinding them together in a mortar, which results in charge transfer interactions. Complexation with PVP can better solubilize fullerenes at high concentrations compared with other methods.

Successful PDT requires the accumulation of the photosensitizer only around cancerous tissue. Vascular endothelium in cancerous tissue contains pores on the order of tens to hundreds of nanometers (nm), which are not observed in healthy tissue. If the size of a drug is less than several nanometers, then the drug is removed quickly through kidney clearance. If the size of a drug is >400 nm, then it is removed quickly by macrophages. However, a drug with a size on the order of tens to a hundred nanometers can accumulate in cancerous tissue through an enhanced permeation and retention effect.²⁰ Drugs within this size range can remain in cancerous tissue for a long period because the lymphatic vessels around cancerous tissue are undeveloped with an incomplete collection mechanism. Thus PDT photosensitizers accumulated around cancerous tissue can damage and kill only cancer cells by the active oxygen generated upon light irradiation.

The vinyl monomer, 2-methacryloyloxyethyl phosphorylcholine (MPC), has a pendant phosphorylcholine group, which is ubiquitous on the surface of cell membranes. The phosphorylcholine group has a betaine structure and a hydrophilic nature.^{21,22} The MPC homopolymer (Figure 1a) PMPC_n is well studied as a biomaterial because it shows inhibition of both blood coagulation and immunoresponses.²³

In this study, PMPC_n with a well-controlled structure, was prepared via reversible addition–fragmentation chain transfer (RAFT) radical polymerization. Fullerene and PMPC_n powders were mixed using an agate mortar to form water-soluble fullerene/PMPC_n complexes. The complexation of fullerenes with PMPC_n can solubilize fullerenes in water (Figure 1b). The generation of singlet oxygen from the complexes upon visible light irradiation was confirmed by photooxidation of 9,10-anthracendipropionic acid (ADPA). Use of the complex in PDT was attempted because the water-soluble complexes are biocompatible owing to the surface PMPC_n chains (Figure 1c).

EXPERIMENTAL PROCEDURE Materials

MPC was purchased from NOF Corp. (Tokyo, Japan), which was produced using a previously reported method.²¹ 4,4'-Azobis(4-cyanovaleric acid) (V-501, 98%) and poly(acrylic acid) (PAA, molecular weight $(M_w) = 2.50 \times 10^4$) from Wako Pure Chemical (Osaka, Japan) were used as received. 4-Cyanopentanoic

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Received 3 February 2016; revised 25 April 2016; accepted 26 April 2016; published online 8 June 2016

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Figure 1 (a) Chemical structure of PMPC_n. (b) Conceptual illustration of the preparation of the fullerene/PMPC_n complex. (c) Application of the fullerene/PMPC_n complex for PDT. PMPC, poly(2-(methacryloyloxy)ethyl phosphorylcholine); PDT, photodynamic therapy. A full color version of this figure is available at the *Polymer Journal* online.

acid dithiobenzoate (CPD) was synthesized according to previously reported methods.²⁴ Methanol was dried over 4-Å molecular sieves and purified by distillation. The fullerenes (C₆₀: 98% and C₇₀: \geq 99%) from Sigma-Aldrich (St Louis, MO, USA) were used as received without further purification. Poly (methacrylic acid) (PMA) was prepared by neutralization of poly(sodium methacrylate) (M_w = 9.50 × 10³) from Sigma-Aldrich (St Louis, MO, USA). ADPA was prepared according to a method previously reported.²⁵ Water was purified using a Millipore (Billerica, MA, USA) Milli-Q system.

Preparation of polyMPC (PMPC_n)

The MPC homopolymer was synthesized according to a modified version of previously reported methods.²³ MPC (10.8 g, 36.7 mmol), CPD (94.7 mg, 0.339 mmol) and V-501 (47.6 mg, 0.170 mmol) were dissolved in a mixed solvent (36.7 ml) of water and methanol (4/1 v/v). The solution was degassed by purging with Ar gas for 30 min. Polymerization was performed at 70 °C for 6 h. After the reaction, ¹H nuclear magnetic resonance (NMR) data indicated that the conversion was 100%. The polymerization mixture was dialyzed against pure water for 2 days.²⁶ PMPC_{113} was recovered by a freeze-drying technique (9.30 g, 85.0%). The number-average degree of polymerization (DP) and the number-average molecular weight estimated from NMR (Mn(NMR)) for $PMPC_{113}$ were 113 and 3.36×10^4 , respectively. The number-average molecular weight $(M_n(GPC))$ and molecular weight distribution (M_w/M_n) estimated from gel-permeation chromatography (GPC) were 1.93×10^4 and 1.17, respectively. Low-molecular-weight PMPC₆₄ (DP = 64, $M_n(NMR) = 1.89 \times 10^4$, $M_w/M_n = 1.23$) and high-molecular-weight PMPC₃₆₂ (DP = 362, $M_n(NMR) = 1.07 \times 10^5$, $M_{\rm w}/M_{\rm n} = 1.97$) were prepared using methods similar to those described above.

Preparation of fullerene/PMPC_n complexes

A typical preparation of a fullerene/PMPC₁₁₃ complex is as follows: PMPC₁₁₃ (10 mg, 2.98×10^{-4} mmol, $M_n(\text{NMR}) = 3.36 \times 10^4$, $M_w/M_n = 1.17$) and C_{60} (11 mg, 15.3×10^{-3} mmol) powders were mixed in an agate mortar and then vigorously mulled for 30 min. Water (10 ml) was added to the powder mixture. The final polymer concentration (C_p) was adjusted to $1.0 \text{ g} \text{ l}^{-1}$ by adding water. Insoluble material was removed by centrifugation at 6000 r.p.m. for 1 h. All sample solutions were filtered with a 0.45-µm membrane filter. Other concentrations of aqueous C_{60} and C_{70} solutions were prepared using methods similar to those described above.

Measurements

¹H NMR spectra were obtained with a Bruker BioSpin (Billerica, MA, USA) DRX-500 NMR spectrometer operating at 500 MHz. The sample solutions for ¹H NMR measurements were prepared using D₂O. The GPC measurements were performed using a refractive index detector equipped with a Shodex (Tokyo, Japan) 7.0- μ m bead size GF-7 M HQ column (exclusion limit ~ 10⁷) operated at 40 °C under a flow rate of 0.6 ml min⁻¹. A phosphate buffer (pH 9) containing 10 vol% acetonitrile was used as the eluent. The M_n (GPC) and Mw/Mn values were calibrated using standard sodium poly(styrene sulfonate) samples. Ultraviolet-visible (UV-vis) absorption spectra were recorded on a Jasco (Tokyo, Japan) V-630 UV-vis BIO spectrophotometer with a 1.0-cm path length quartz cell. Dynamic light scattering (DLS) measurements were performed using a Malvern (Worcestershire, UK) Zetasizer nano ZS at 25 °C. A He-Ne laser (4.0 mW at 632.8 nm) was used as the light source. Sample solutions were filtered using a 0.45-µm pore size membrane filter. The data were analyzed by the Malvern Zetasizer 6.20 software. Static light scattering (SLS) measurements were performed using an Otsuka Electronics Photal (Osaka, Japan) DLS-7000HL light scattering spectrometer. A He-Ne laser (10.0 mW at 632.8 nm) was used as the light source. Sample solutions for light scattering measurements were filtered with a 0.45-µm pore size membrane filter. The weight-average molecular weight (M_w) , the z-average radius of gyration (R_g) , and the second virial coefficient (A_2) values were estimated from the following relationship:

$$\frac{KC_{\rm p}}{R_{\rm \theta}} = \frac{1}{M_{\rm w}} \left(1 + \frac{1}{3} R_{\rm g}^{2} q^{2} \right) + 2A_{2}C_{\rm p} \tag{1}$$

where R_{θ} is the difference between the Rayleigh ratio of the solution and that of the solvent, $K = 4\pi^2 n^2 (dn/dC_p) 2/N_A \lambda^4$, where dn/dC_p is the refractive index increment against C_p , N_A is Avogadro's number and q is the magnitude of the scattering vector. The q value was calculated from $q = (4\pi n/\lambda)\sin(\theta/2)$, where n is the refractive index of the solvent, λ is the wavelength of light source (=632.8 nm) and θ is the scattering angle. By measuring R_{θ} for a set of C_p and θ , the values of M_w , R_g and A_2 were estimated from Zimm plots. The known Rayleigh ratio of toluene was used to calibrate the instrument. The values of dn/dC_p at 633 nm were determined using an Otsuka Electronics Photal DRM-3000 differential refractometer. The dn/dC_p values for C_{60} /PMPC₁₁₃ and C_{70} /PMPC₁₁₃ were 0.722 and 0.833 ml g⁻¹, respectively. Small-angle X-ray scattering (SAXS) measurements were performed on a BL40B2 beamline at SPring-8, Hyogo, Japan. A $30 \times 30 \text{ cm}^2$ imaging plate (Rigaku R-AXIS VII, Tokyo, Japan) detector was used and was placed 4.0 and 2.0 m away from the

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sample. The wavelength of the incident beam (λ) was 0.10 nm. This setup provided a *q* range of 0.04 – 3.5 nm⁻¹, where *q* is the magnitude of the scattering vector defined by $q = (4\pi n/\lambda)\sin\theta$ with a scattering angle of 2 θ . For the SAXS measurements, we used a vacuum sample chamber to remove the effects of the background from the window material and air. The X-ray transmittance of the samples was measured with an ion chamber located in front of the sample and a Si photodiode (Hamamatsu Photonics S8193, Shizuoka, Japan) placed after the sample. Detailed experimental procedures are reported elsewhere.^{27,28} Transmission electron microscopy (TEM) was performed using a JEOL (Tokyo, Japan) JEM-2100 at an accelerating voltage of 200 kV. Samples for TEM observations were prepared by placing one drop of an aqueous solution of the fullerene/PMPC₁₁₃ complex on a copper grid coated with support films. The samples were stained with phosphotungstic acid.

Generation of singlet oxygen

Photoirradiation was performed using an Asahi Spectra (Tokyo, Japan) Max-301 with light from a 300-W Xe lamp passing through a 420-nm cutoff filter. The light intensity was 6.5 mW cm⁻² at 420 nm. A quartz cuvette with a 1-cm pass length was used. ADPA bleaching experiments were performed to confirm that singlet oxygen was generated by fullerene/PMPC₁₁₃ complexes in water, where ADPA was generated as a singlet oxygen acceptor. Aqueous solutions of ADPA and fullerene/PMPC₁₁₃ complexes were mixed, and the combined solution was irradiated at \ge 420 nm. The reaction was monitored by recording a decrease in the intensity of the absorption peak at 400 nm owing to ADPA as a function of the irradiation time.

RESULTS AND DISCUSSION

Characterization of PMPC_n

MPC was polymerized via a RAFT-controlled radical polymerization method using CPD; the MPC conversion was 100%. The DP and M_n (NMR) values estimated from NMR for PMPC₁₁₃ were 113 and 3.36×10^4 , respectively (Table 1). The M_n (GPC) and M_w/M_n values estimated from GPC were 1.93×10^4 and 1.17, respectively. Low-molecular-weight (M_n (NMR) = 1.89×10^4) and high-molecular-weight PMPC_n (M_n (NMR) = 1.07×10^5) were also prepared. After polymerization, the percentage of conversion (*x*) was monitored using ¹H NMR before purification, which can be estimated by comparing the integral intensity ratios between vinyl protons and pendant methylene protons. The value of M_n (theoretical) was calculated using the following equation:

$$M_{\rm n}(\text{theory}) = \frac{[\text{MPC}]_0}{[\text{CPD}]_0} \times \frac{x}{100} \times \text{MW}_{\rm MPC} + \text{MW}_{\rm CPD}$$
(2)

where $[MPC]_0$ is the initial molar concentration of MPC, $[CPD]_0$ is the initial molar concentration of CPD, MW_{MPC} is the molecular weight of MPC and MW_{CPD} is the molecular weight of CPD.

Proton NMR spectra for $PMPC_{113}$ in D_2O were obtained (Supplementary Figure S1). Signals in the 7.45–7.95 p.p.m. region were attributed to the dithiobenzoate group. The DP of $PMPC_{113}$ was calculated as 113 from the area intensity ratio of peaks attributed to pendant methylene protons at 3.65 p.p.m. and terminal phenyl

Table 1 Molecular cha	aracteristics of	PMPC _n
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Polymer	M_n (theory)×10 ⁻⁴	DP	$M_n(NMR) \times 10^{-4}$	$M_n(GPC) \times 10^{-4}$	M _w /M _r
PMPC ₆₄	1.89	64	1.89	0.806	1.23
PMPC ₁₁₃	3.23	113	3.36	1.93	1.17
PMPC ₃₆₂	12.0	362	10.7	4.53	1.97

Abbreviations: DP, number-average degree of polymerization; M_n (GPC), number-average molecular weight estimated from gel-permeation chromatography; M_n (NMR), number-average molecular weight estimated from ¹H NMR; M_n (theoretical), number-average molecular weight estimated from Equation (2); M_w/M_n , molecular weight distribution; PMPC, poly(2-(methacryloyloxy)ethyl phosphorylcholine).

protons. The value of $M_n(NMR)$ for PMPC_n can be calculated from the corresponding DP, which is close to M_n (theoretical). The GPC peak for PMPC₁₁₃ was unimodal (Supplementary Figure S2). The M_w/M_n value for PMPC₁₁₃ was 1.17, suggesting that PMPC₁₁₃ had a well-controlled structure. The M_n (GPC) value was an estimated value because poly(sodium *p*-styrenesulfonate) was used as a standard for the calibration.

Preparation and characterization of fullerene/PMPC_n complexes

The sample preparation method is important for the solubilization of fullerenes in water using PMPC_n. Fullerene and PMPC_n powders were ground with a mortar and pestle for 30 min to achieve homogeneity.²⁹ The relationship between the amount of fullerene solubilized in water using PMPC_n and the duration of grinding was determined. The amount of fullerene solubilized was not increased by grinding for > 30 min. Therefore, the grinding time was kept constant at 30 min to prepare the fullerene/PMPC_n complexes. Insoluble matter was removed by centrifugation. All sample solutions were filtered with a 0.45-µm membrane filter.

To measure UV-vis absorption spectra for C_{60} /PMPC₁₁₃ and C_{70} /PMPC₁₁₃ complexes in water with varying fullerene concentrations, the sample solutions were diluted with water because the absorbance from the original aqueous complex solutions was too high to measure using a UV-vis absorption spectrometer (Figure 2). The original absorbance was calculated from the absorption spectra and the dilution ratio. The C_{60} /PMPC₁₁₃ complex indicated C_{60} absorption peaks at 215, 265 and 340 nm, which increased until $[C_{60}] \leq 1.1 \text{ g} \text{ l}^{-1}$ while increasing the C_{60} feed concentration. The C_{70} absorption peaks at 215, 240 and 384 nm for the C_{70} /PMPC₁₁₃ complex increased until $[C_{70}] \leq 0.5 \text{ g} \text{ l}^{-1}$ while increasing the C_{70} feed concentration.



Figure 2 UV-vis adsorption spectra for (a) C_{60} /PMPC₁₁₃ and (b) C_{70} /PMPC₁₁₃ complexes at various fullerene concentrations in water. The original solutions were diluted to 1/100 because the absorbance was too high. PMPC, poly(2-(methacryloyloxy)ethyl phosphorylcholine); UV-vis, ultraviolet-visible. A full color version of this figure is available at the *Polymer Journal* online.

These absorption peaks were similar to those for C_{60} and C_{70} in organic solvents.¹⁶ These observations indicate that $PMPC_n$ can solubilize C_{60} and C_{70} in water.

The relationship between the feed amount of fullerene and the amount of fullerene dissolved in water using PMPC₁₁₃ is shown in Figure 3. The solubilized amounts were calculated from the absorbance and extinction coefficients for C_{60} (68.0 Lg⁻¹ cm⁻¹ at 340 nm) and C_{70} (40.4 Lg⁻¹ cm⁻¹ at 384 nm).¹⁸ The maximum concentrations of C_{60} and C_{70} in water using PMPC₁₁₃ at $C_p = 1 \text{ g } \text{I}^{-1}$ were 0.67 and 0.49 g l⁻¹, respectively. The solubilized amounts of fullerenes using PMPC₁₁₃ were higher than those in previous reports. The reported concentration of C_{60} in water using surfactants, such as lecithin, was 0.012 g l⁻¹.¹⁶ Yamakoshi *et al.*¹⁸ reported that solubilized C_{60} and C_{70} concentrations using PVP in water were 0.4 and 0.2 g l⁻¹, respectively.

Although the pendant phosphorylcholine groups in PMPC_n are hydrophilic, the main chain of methacrylate is slightly more hydrophobic than the pendant group. The methacrylate main chain in PMPC_n may interact with hydrophobic fullerene. To confirm hydrophobic interactions between the main chain and fullerene, solubilization tests for C_{60} in water were performed using commercially available PMA ($M_w = 9.50 \times 10^3$) and PAA ($M_w = 2.50 \times 10^4$) powders. PMA at $C_p = 1 \text{ g l}^{-1}$ dissolved C_{60} in water with $[C_{60}] = 0.37 \text{ g}^{1-1}$. In contrast, PAA could not dissolve C_{60} in water. These results suggest that a methacrylate-type main chain structure may be extremely important for the formation of a fullerene/PMPC_n complex. The amount of fullerene solubilized by PMA was less than that by PMPC_n. Thus the solubilization of fullerenes may require hydrophobic interactions between the methacrylate polymer



Figure 3 Plots of solubilized amounts of C_{60} ($^{\circ}$) and C_{70} ($^{\wedge}$) as a function of feed concentration of C_{60} or C_{70} . A full color version of this figure is available at *Polymer Journal* online.



Figure 4 R_h of C₆₀/PMPC₁₁₃ (\circ) and C₇₀/PMPC₁₁₃ complexes (\triangle) as a function of C_p . C_p , polymer concentration; PMPC, poly(2-(methacryloyloxy) ethyl phosphorylcholine); R_h , hydrodynamic radius. A full color version of this figure is available at the *Polymer Journal* online.

backbone and fullerene as well as interactions between the pendant phosphorylcholine groups in PMPC_n and fullerene. To confirm the influence of the $M_{\rm w}$ of PMPC_n, solubilization of C_{60} in water was performed using PMPC_n with a low $(M_{\rm n}({\rm NMR}) = 1.89 \times 10^4)$ and a high $(M_{\rm n}({\rm NMR}) = 1.07 \times 10^5)$ $M_{\rm w}$. The maximum concentrations of C_{60} in water using PMPC₆₄ and PMPC₃₆₂ at $C_{\rm p} = 1$ g l⁻¹ were 0.76 and 0.44 g l⁻¹, respectively. The maximum C_{60} concentrations in water increased as the $M_{\rm w}$ of PMPC_n decreased. For every polymer chain, the number of solubilized C_{60} molecules can be calculated from the molar concentrations of polymer and solubilized C_{60} . The number of C_{60} molecules that could be solubilized by a single polymer chain of PMPC₆₄, PMPC₁₁₃, PMPC₃₆₂ and PMA was 20, 31, 65 and 7, respectively.

The stability of the complex during dilution with water was studied. The hydrodynamic radius (R_h) values for the complexes were plotted as a function of C_p (Figure 4). The R_h values for the complexes were kept constant and were independent of C_p in the region of 1–0.05 gl⁻¹. The complexes were stable even at 0.05 gl⁻¹upon dilution of C_p . When C_p was diluted to a concentration <0.05 gl⁻¹, DLS data could not be obtained owing to low scattering intensity.

To characterize the complexes in more detail, SLS measurements were performed for C₆₀/PMPC₁₁₃ and C₇₀/PMPC₁₁₃ complexes in water (Supplementary Figure S3). The results from light scattering measurements are summarized in Table 2. The values of $M_{\rm w}$ and $R_{\rm g}$ for the C₆₀/PMPC₁₁₃ complex were larger than were those for C70/PMPC113. The A2 values for C60/PMPC113 and C70/PMPC113 were 1.19×10^{-3} and 2.77×10^{-3} cm³·mol g⁻², respectively. The complex using C₆₀ had a small A₂, which indicates that the solubility of C₆₀/PMPC₁₁₃ in water was lower than that of C₇₀/PMPC₁₁₃.^{30,31} The R_g/R_h value is related to the shape and polydispersity of aggregates in solution. The theoretical R_g/R_h values for rigid hard spheres and spherical aggregates are 0.78 and 1.0, respectively.^{32–35} The R_g/R_h values for random coil and ellipsoidal aggregates are 1.3-1.5. Thread-like and low-density aggregates with a high polydispersity index indicate larger Rg/Rh values. The shapes of the fullerene/ PMPC₁₁₃ complexes may be spherical because the R_g/R_h values for the complexes were close to 1. The densities of C₆₀/PMPC₁₁₃ and C_{70} /PMPC₁₁₃ complexes calculated from the apparent M_w and R_g values estimated from SLS were 0.103 and 0.064 g cm⁻³, respectively. The maximum number of fullerenes for every PMPC₁₁₃ chain was calculated from the molar ratio of PMPC₁₁₃ and fullerene in water using UV-vis absorption data. The number of C_{60} and C_{70} in one PMPC₁₁₃ polymer chain was 31 and 20, respectively. The aggregation number (N_{agg}) for the complex was calculated from the apparent M_w for the complex and the $M_{\rm w}$ of a single polymer chain with the accompanying fullerenes. The N_{agg} was defined as the total number of PMPC₁₁₃ chains in one complex. The N_{agg} values for C_{60} /PMPC₁₁₃

Table 2 Dynamic and static light scattering data for fullerene/ PMPC113 complexes in water at 25 °C

				$A_2 imes 10^3$ (mol		
Samples	R _h (nm)	$M_w(SLS) \times 10^{-6}$	R _g (nm)	ml g ⁻²)	$R_g R_h$	N _{agg}
C ₆₀ /PMPC ₁₁₃	25.1	5.37	27.6	1.19	1.10	96
C70/PMPC113	23.2	1.99	23.2	2.77	1.00	40

Abbreviations: A_2 , the second virial coefficient; M_w (SLS), weight-average molecular weight estimated with static light scattering; N_{agg} , aggregation number of the fullerene/PMPC₁₁₃ complex; PMPC, poly(2-(methacryloploxy)ethyl phosphorylcholine); R_g , z-average radius of gyration; R_h , hydrodynamic radius.

 M_{agg} values are calculated from the M_{w} of the complex determined by SLS and the M_{w} of the corresponding single PMPC_n chain.



Figure 5 (a) SAXS profiles and (b) Guinier plots for C₆₀/PMPC₁₁₃ and C₇₀/PMPC₁₁₃ complexes in water. *a*: shift factor, *q*: the magnitude of the scattering vector and *l*(*q*): scattering intensity.



Figure 6 TEM images for (a) C₆₀/PMPC₁₁₃ and (b) C₇₀/PMPC₁₁₃ complexes. PMPC, poly(2-(methacryloyloxy)ethyl phosphorylcholine); TEM, transmission electron microscopy.



Scheme 1 Photooxidation of ADPA by singlet oxygen. ADPA, 9, 10-anthracendipropionic acid.

and C_{70} /PMPC₁₁₃ complexes were 96 and 40, respectively. The number of C_{60} and C_{70} fullerenes included in one complex was 2995 and 784, respectively.

To confirm the structure of the complex, we performed SAXS measurements (Figure 5). The R_g values for PMPC/C₆₀ and PMPC/C₇₀ were 18 and 20 nm, respectively, as obtained from Guinier plots. These values are smaller than those obtained from SLS measurements. As X-rays scatter owing to nanoscale electron density inhomogeneities within samples, they effectively highlight the clustered fullerene parts in the complex.³⁶ The R_g of C₇₀/PMPC was larger than that of C₆₀/PMPC, as estimated from SAXS measurements. This observation suggests that the aggregate formed from C₇₀ in the complex was swollen owing to incorporation of PMPC chains and water into the aggregate.

TEM measurements were performed to confirm the size and shape of the fullerene/PMPC₁₁₃ complex (Figure 6). The C_{60} /PMPC₁₁₃ and

 C_{70} /PMPC₁₁₃ complexes had a distorted spherical shape. The average diameters estimated from TEM images for C_{60} /PMPC₁₁₃ and C_{70} /PMPC₁₁₃ complexes were 29 and 24 nm, respectively. The sizes estimated from TEM were slightly smaller than were those estimated from light scattering measurements because the complexes may shrink during the drying process while preparing TEM samples.

Generation of singlet oxygen

Fullerenes can be excited by light, and the energy is subsequently transferred to ground-state oxygen to efficiently generate singlet oxygen. The photooxidation of ADPA by singlet oxygen can be applied for the detection of generated singlet oxygen atoms using a photosensitizer (Scheme 1).25 To confirm the generation of singlet oxygen atoms from fullerene/PMPC_n complexes upon visible light irradiation, ADPA was used to detect the singlet oxygen atoms in water. Photooxidation of ADPA to generate ADPA endoperoxide by singlet oxygen is indicated by observing a decrease in the UV-vis absorption peaks of ADPA. Changes in the UV-vis absorption spectra of ADPA were measured in the presence of fullerene/PMPC_n complexes in water upon visible light irradiation at \ge 420 nm, as shown in Figure 7. The polymer concentration was diluted to 0.021 gl-1 in water because the absorbance of fullerenes was too high for the original solutions at $C_p = 1 \text{ g } l^{-1}$. The concentrations of C_{60} and C_{70} were 0.0141 and 0.0103 gl⁻¹, respectively. The concentrations of ADPA used in the C₆₀ and C₇₀ experiments were 0.0377 and $0.0439 \text{ g} \text{ l}^{-1}$, respectively. The UV-vis spectra were calibrated by

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Figure 7 Changes in UV-vis absorption spectra of ADPA in the presence of (a) C_{60} /PMPC₁₁₃ and (b) C_{70} /PMPC₁₁₃ complexes in water upon visible light irradiation (\geq 420 nm). Fullerene absorptions were subtracted from the original spectra for calibration. Irradiation times are indicated in the figure. ADPA, 9,10-anthracendipropionic acid, PMPC, poly(2-(methacryloyloxy)ethyl phosphorylcholine); UV-vis, ultraviolet-visible. A full color version of this figure is available at the *Polymer Journal* online.

subtracting the fullerene absorption. The absorbance of ADPA in the presence of the complex decreased with increasing irradiation time. After 180 min of irradiation, the absorbance values at 399 nm of ADPA in the presence of C_{60} /PMPC₁₁₃ and C_{70} /PMPC₁₁₃ complexes were 0.186 and 0.160, respectively. This observation indicates that C_{70} /PMPC₁₁₃ can oxidize ADPA more efficiently than C_{60} /PMPC₁₁₃. The amounts of singlet oxygen generated from C_{70} /PMPC₁₁₃ were higher than those from C_{60} /PMPC₁₁₃ by light irradiation above 420 nm, presumably because the absorption wavelength of C_{70} /PMPC₁₁₃ was longer than that of C_{60} /PMPC₁₁₃.

CONCLUSIONS

Hydrophilic and biocompatible PMPC_n with a well-controlled structure was prepared via RAFT-controlled radical polymerization. Fullerene/PMPC_n complexes were prepared by mixing fullerene and PMPC_n powders. Fullerenes could be solubilized in water using PMPC_n with a higher concentration than those reported previously. Hydrophobic interactions of the main chain with fullerene and interactions between the pendant phosphorylcholine and fullerene may be important for fullerene solubilization. The R_h values for C₆₀/PMPC₁₁₃ and C₇₀/PMPC₁₁₃ complexes were 25.1 and 23.2 nm, respectively. The number of C₆₀ and C₇₀ fullerenes in one complex was 2995 and 784, respectively. Singlet oxygen was generated from fullerene/PMPC_n complexes upon visible light irradiation > 420 nm. These fullerene/PMPC_n complexes are promising candidates for application in PDT.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

This work was financially supported by a Grant-in-Aid for Scientific Research (No. 25288101) from the Japan Society for the Promotion of Science (JSPS) and the Cooperative Research Program 'Network Joint Research Center for Materials and Devices' (No. 2015467). All SAXS measurements were performed at the SPring-8 beamline (2016A1242, 2016A1619).

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Supplementary Information accompanies the paper on Polymer Journal website (http://www.nature.com/pj)