

NMR Measurement of Identical Polymer Samples by Round Robin Method V. Determination of Degree of Polymerization for Isotactic Poly(methyl methacrylate) Having a *t*-Butyl End Group

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ABSTRACT: Research Group on NMR, The Society of Polymer Science, Japan, has continued assessments of the reliability of NMR measurements of polymers. In the present study, ¹H and ¹³C NMR data for isotactic poly(methyl methacrylate) (*it*-PMMA) having a *t*-butyl end group were collected on 27 NMR spectrometers by a round-robin method, to survey the accuracy of NMR spectroscopic determination of degree of polymerization (*DP*) of the polymer by the end group analysis as well as other basic NMR parameters such as chemical shift and spectral resolution. Five samples of *it*-PMMA whose *DP*'s range from 53 to 5200 were prepared with *t*-BuMgBr in toluene. ¹H and ¹³C NMR spectra were measured in nitrobenzene-*d*₅ at 110 °C. The standard deviations (σ) for chemical shift measurements of ¹H and ¹³C NMR signals were 0.001–0.004 ppm and 0.04–0.18 ppm, respectively. A singlet signal due to *t*-butyl group at initiating chain end of the *it*-PMMA is separately observed from the signals due to the monomeric units in ¹H NMR spectra measured in nitrobenzene-*d*₅. Thus *DP* of the polymer can be determined by the equation of $DP = 3 \times [\text{OCH}_3]/[t\text{-C}_4\text{H}_9]$. The value of σ for *DP* determination by ¹H NMR increased as *DP* increased (6.4–23.4%). However, the averaged values of *DP* agreed well with those determined by SEC. ¹³C NMR analysis of *DP* was possible for the *it*-PMMA with *DP* up to *ca.* 250. The values agreed with those determined by ¹H NMR spectroscopy. ¹H NMR determination of *DP* by 5 runs using single spectrometers (100 and 500 MHz) gave much better reproducibility; σ 3.9–9.2% and 0.7–3.4% for the measurements with 100 and 500 MHz instruments, respectively.

KEY WORDS ¹H NMR / ¹³C NMR / Poly(methyl methacrylate) / Round Robin Method / End Group Analysis / Reliability / Chemical Shift / Resolution / *DP* / Precision / Accuracy /

Research group on NMR, the Society of Polymer Science Japan (SPSJ), has made assessments on the reliability of NMR measurements of polymers. In a series of this cooperative research work the reliability of chemical shift, signal intensity, spin-lattice relaxation time, nuclear Overhauser enhancement factor, and quantitative analysis of copolymer composition have been investigated for ¹H and ¹³C NMR measurements of polymer samples and the results have been reported in this journal.^{1–4}

In the present work the reliability of degree of polymerization (*DP*) determined by end-group analy-

sis was studied by round robin method for poly(methyl methacrylate) (PMMA). The Committee on Molecular Weight and Molecular Weight Distribution of the Society of Polymer Science Japan reported previously the results of measurements of molecular weight and its distribution by round robin method.⁵ The present study is the NMR version of round robin test of *DP*. The PMMA of several *DP*'s were prepared through the living polymerization with *t*-C₄H₉MgBr in toluene at –60 °C and found to contain a *t*-C₄H₉ group at the initiating chain end of the macromolecule.⁶ The ¹H and ¹³C NMR data were collected from 27 (100–500 MHz) and

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Table I. *it*-PMMA samples prepared in toluene with *t*-C₄H₉MgBr at -60 °C

Sample	DP	$\overline{M}_w/\overline{M}_n^a$	Tacticity(%) ^b		
			<i>mm</i>	<i>mr</i>	<i>rr</i>
1	53.4	1.095	96.8	3.0	0.2
2	120.6	1.118	96.7	3.1	0.2
3	256	1.125	97.5	2.4	0.1
4	977	1.379	95.3	3.6	1.1
5	5200	1.41	94.0	3.0	3.0

^aDetermined by SEC. ^bDetermined by ¹H NMR.

23 (50–125 MHz) spectrometers, respectively. The *DP* values were determined from the ¹H NMR signal intensities of *t*-C₄H₉- and -OCH₃ groups in the polymer and the obtained values were compared with those measured by size exclusion chromatography (SEC) for the accuracy.^{††} The precisions of *DP* determination by round robin method for ¹H NMR were 6.4–23.4% depending on the magnitude of *DP*; the values increased with increasing *DP* values. The precision of determination by a single 500 MHz spectrometer was 0.7–3.4% which is much better than that by round robin method. ¹³C NMR analysis of the *DP* was also possible for the *it*-PMMA with *DP* up to about 250 although the fluctuation of the data is rather large.

EXPERIMENTAL

Five samples of isotactic (*it*-) PMMAs used in the measurements were prepared by the living polymerizations with *t*-C₄H₉MgBr in toluene at -60 °C,⁶ and tacticity, *DP* and $\overline{M}_w/\overline{M}_n$ values of the samples are listed in Table I.

SEC chromatography of the PMMAs was performed on a JASCO FLC-A10 chromatograph equipped with a Shodex A-80 M SEC column (50 cm × 2) using tetrahydrofuran as an eluent. RI and LALLS detectors were used. The molecular weight calibration was made using standard polystyrenes.

The PMMA samples were dissolved in nitrobenzene-*d*₅. The solutions were filtered under dry nitrogen to remove insoluble materials. The filtered solution of each sample was degassed, and sealed under nitrogen in 5 mm NMR sample tubes. The concentrations of the solutions were 10 wt vol⁻¹% for samples 1–4 and 5 wt vol⁻¹% for sample 5. Five sets of five sample tubes were circulated among collaborating test sites. The research work was carried out by several non-members as well as SPSJ members.

The number of spectrometers used was 27 for ¹H NMR measurements (100–500 MHz) and 23 for ¹³C

Table II. Conditions for the measurements of NMR spectra

	¹ H	¹³ C
Solvent	NB- <i>d</i> ₅ ^a	NB- <i>d</i> ₅ ^a
Conc. wt vol ⁻¹ % ^b	10, 5	10
Temp.(°C)	110	110
Obs.range(ppm)	15	250
Pulse width (°)	45	45
Pulse repetn. (s)	10	10
Number of scans	32–5700	3000–32000
Decoupling	–	NNE ^c

^aNitrobenzene-*d*₅. ^bThe concentration of solutions were 10 wt vol⁻¹% for the samples 1–4 and 5 wt vol⁻¹% for sample 5. ^cGated decoupling without NOE.

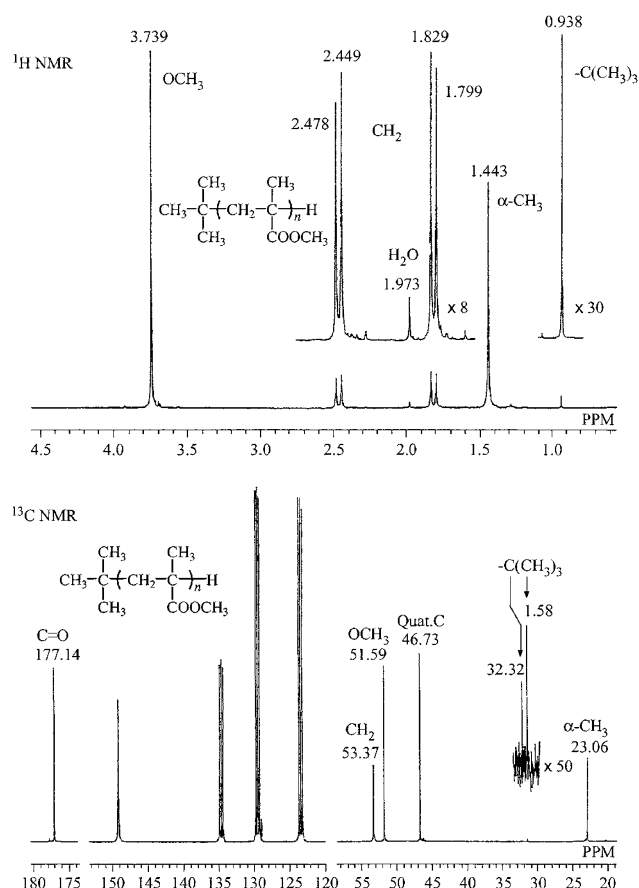


Figure 1. 500 MHz ¹H and 100 MHz ¹³C NMR spectra of *it*-PMMA (Sample 1 in Table I) in nitrobenzene-*d*₅ at 110 °C. The chemical shift values indicated in the figure are the averaged ones obtained from the determinations by round robin method.

NMR measurements (50–125 MHz). The conditions for ¹H and ¹³C NMR measurements are summarized in Table II. Typical ¹H and ¹³C NMR spectra of the sample are shown in Figure 1. The peak assignments are depicted in the figure. ¹H and ¹³C chemical shifts referred to the signals due to the residual meta protons (8.091 ppm) and ipso carbon (149.10 ppm) of nitrobenzene-*d*₅, respectively and were converted into

^{††}This strategy is an extension of the idea appearing in the paper of Demarquay *et al.*,⁷ in which *DP* of PVC has been determined by comparison of the NMR intensity for the polymer main chain with that for the *t*-C₄H₉ group from the initiator, *t*-C₄H₉MgCl.

Table III. Line width (ppm)^a of OCH₃ proton signal in the ¹H NMR spectra of *it*-PMMA

Freq. (MHz)	n ^b	Sample ^c				
		1	2	3	4	5
100	2	0.2071(1.3)	0.2132(3.4)	0.2065(4.2)	0.2543(2.4)	0.2522(5.7)
200	1	0.1258(-)	0.1456(-)	0.1429(-)	0.1301(-)	0.2568(-)
270	9	0.1172(14.6)	0.1215(15.6)	0.1169(15.6)	0.1302(21.1)	0.1831(17.8)
300	1	0.0977(-)	0.0998(-)	0.0951(-)	0.1035(-)	0.1763(-)
360	1	0.0907(-)	0.0933(-)	0.0920(-)	0.1040(-)	0.1600(-)
400	7	0.0962(16.7)	0.0959(7.9)	0.0958(6.4)	0.0911(17.7)	0.1616(17.9)
500	6	0.1004(9.6)	0.0981(6.9)	0.0885(12.7)	0.0960(17.2)	0.1512(15.3)

^aThe line width at the level of ¹³C-satellite. ^bNumber of spectrometers used. ^cFigures in parentheses represent the precision (%).

TMS scales.

The precision of measurements is represented by the percent ratio of the standard deviation to the number averaged value of the determinations.

RESULTS AND DISCUSSION

Reliability of Chemical Shift Measurement and Spectral Resolution

Averaged values of ¹H and ¹³C NMR chemical shifts of *it*-PMMA (Sample 1) are indicated in Figure 1. The precisions of chemical shift measurements for this *it*-PMMA sample were found to be ±0.001–0.004 ppm for ¹H NMR and ±0.04–0.18 ppm for ¹³C NMR. The both values are smaller than the corresponding values obtained for the atactic PMMA in the round robin experiments in 1987¹ (±0.0036–0.0071 ppm for ¹H NMR and ±0.054–0.307 ppm for ¹³C NMR). This may be due to the facts that *it*-PMMA shows sharp and well-separated peaks as compared with atactic one and that the performance and stability of NMR instruments have been improved recently.

In order to evaluate the spectral resolutions of ¹H NMR spectra, the line widths for OCH₃ proton resonances at the ¹³C-satellite level were measured. The results are shown in Table III. The line widths in ppm decreased generally with increasing frequency of measurements regardless of the samples. Since the isotacticities of the PMMA are very high and accordingly the signals consist of almost single component, the results obtained here indicate that the spectral resolutions are enhanced with increasing frequency of measurement. Sample 5 having a *DP* of 5200 showed a large spectral width as compared with other four samples, which may be due to the high viscosity of the solution. Relatively large values of precision may be partly due to the fact that the maximum value of spectral resolution is difficult to attain for the solution of polymer sample.

Spectral resolution should be enhanced at higher temperatures. 500 MHz ¹H NMR spectra of sam-

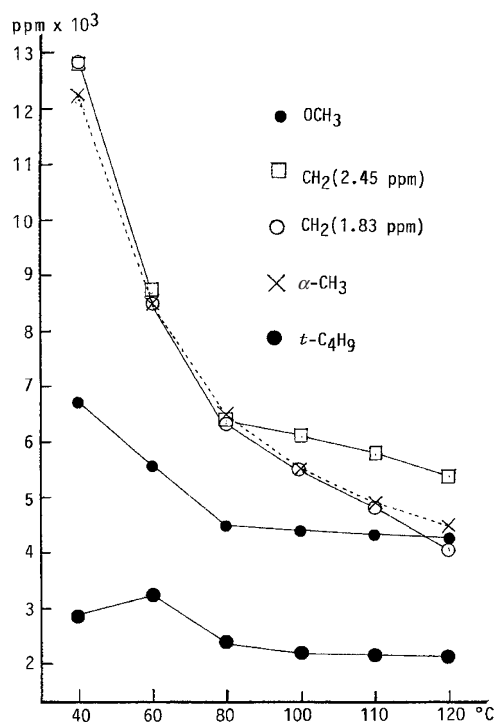


Figure 2. Line widths of 500 MHz ¹H NMR spectrum of sample 1 at various temperatures.

ple 1 were measured at various temperatures (Figure 2). With an increase in temperature the line widths of main-chain CH₂ and α -CH₃ proton signals decrease greatly up to 80 °C and then gradually above 80 °C while those of *t*-C₄H₉ proton signal hardly change with temperature. The plots for OCH₃ signal are in-between. The results indicate that the segmental mobility of the back bone chain is restricted at lower temperatures as compared with the substituent group apart from the main chain such as –OCH₃ group and with the substituent at the chain end. The appropriate temperature of measurement should be above 80 °C.

Reliability of Temperature Set-Up in the Spectrometer Used

It is important for understanding some of the NMR parameters such as temperature-dependent chemical

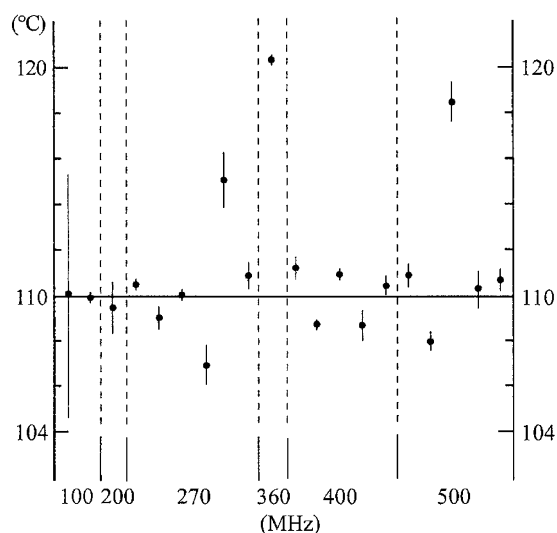


Figure 3. Accuracy and precision of the temperature of measurement of NMR spectrum. The vertical lines transfixing the circles represent the fluctuation of the temperature of sample solutions.

shift and relaxation times to note whether or not the real temperature of the sample solution meets the temperature set on the instrument used. In order to check this point for the data collected by round robin method, the chemical shifts of a small amount of water contained in the sample solutions were investigated. The relationship between the ^1H chemical shift of the water and the temperature of the sample solution was determined for the five individual test samples from the ^1H NMR measurement at one of the test sites (Hatada's laboratory) using a 100 MHz NMR spectrometer (JNM-FX100). Using the calibration curve thus obtained for each sample, the temperature of the sample solution at each ^1H NMR measurement was evaluated from the chemical shift of the water. The range and averaged values of the temperature for five sample solutions were determined for each spectrometer and are shown in Figure 3. The fluctuation of the temperatures for the measurements of five sample solutions installed in a given spectrometer was within $\pm 0.8^\circ\text{C}$ with a few exceptions; the range of the fluctuation is slightly but meaningfully smaller than that ($\pm 1.0^\circ\text{C}$) obtained in the previous work in 1995.⁴ The averaged values of the solution temperatures for respective instruments were in the range from 106.2 to 111.3 $^\circ\text{C}$ with three exceptions. The averaged value for all the instruments (110.4 $^\circ\text{C}$) was almost the same as the temperature set on the instrument; 110 $^\circ\text{C}$.

The temperature of the sample solution of the instrument operated at 360 MHz was higher than the set up temperature by over 10 $^\circ\text{C}$. This may be the reason why the spectrum showed sharp signals as compared with those obtained by other spectrometers (see Figure 3 and Table III).

Table IV. ^1H and ^{13}C spin-lattice relaxation time T_1 (s) of *it*-PMMA samples measured at 500 and 125 MHz, respectively at 110 $^\circ\text{C}$

Hydrogen	Chem. shift (ppm)	Sample				
		1	2	3	4	5
OCH ₃	3.74	1.38	1.35	1.36	1.36	1.32
CH ₂	2.48	0.376	0.363	0.361	0.364	0.360
CH ₂	2.45	0.371	0.364	0.360	0.364	0.361
CH ₂	1.83	0.335	0.321	0.323	0.327	0.321
CH ₂	1.80	0.330	0.321	0.322	0.326	0.322
α -CH ₃	1.44	0.424	0.407	0.399	0.408	0.408
<i>t</i> -C ₄ H ₉	0.94	1.30	1.24	1.30	1.24	1.23

Carbon	Chem. shift (ppm)	Sample				
		1	2	3	4	5
CO	177.1	2.19	2.42	2.30	2.23	2.07
CH ₂	53.4	0.255	0.252	0.244	0.246	0.246
OCH ₃	52.0	1.57	1.56	1.55	1.46	1.57
Quat.C	46.7	3.65	3.62	3.76	3.77	3.62
<i>t</i> -C ₄ H ₉	31.6	1.44	1.41	~1.6	–	–
α -CH ₃	23.1	0.465	0.473	0.462	0.456	0.461

Determination of DP by ^1H and ^{13}C NMR and Their Reliability

The DP values of the PMMA samples were determined by the intensity ratio of OCH₃ proton signal to *t*-C₄H₉ proton one and of α -CH₃ carbon signal to CH₃ carbon signal of *t*-C₄H₉ group. To obtain accurate signal intensity pulse repetition time must be sufficiently long for the magnetization to relax to an equilibrium state. With a 90 $^\circ$ pulse, pulse repetition time of $5T_1$ allows the magnetization to recover more than 99% of an equilibrium value. The ^1H and ^{13}C spin-lattice relaxation times (T_1) of all the PMMA samples were measured at 500 and 125 MHz, respectively at 110 $^\circ\text{C}$ and are shown in Table IV. Most of the T_1 values decreased slightly in the order from sample 1 to sample 5, probably due to the decreasing segmental mobility with increasing molecular weight. The ^1H and ^{13}C T_1 's for OCH₃, α -CH₃, and *t*-C₄H₉ groups are all less than 2 s (Table IV). The T_1 's except for that of C=O group decrease with a decrease in the frequency of measurement.² Accordingly, the pulse repetition time of 10 s and the pulse width of 45 $^\circ$ used in the measurements are very suitable for quantitative determination.

All the respective determinations of DP measured by ^1H NMR are listed in Table V. Although the fluctuation of data seems rather large, the precisions for the samples 1, 2, and 3 [DP determined by SEC, DP (SEC)=53.4–256] remain within 9%. The values of precision exceed 10% for samples 4 and 5 [DP (SEC)=977–5200]. The averaged values of DP for all the samples agree well with the DP values obtained by

Table V. ¹H NMR determination of *DP* of *it*-PMMA sample

Freq. (MHz)	Sample ^a										
	1	2	3	4	5	6	7	8	9	10	
1	100	50.8	(-9.3)	128.6	(-4.2)	229.0	(-15.5)	1007	(13.4)	5111	(-4.5)
2	100	51.9	(-7.3)	122.0	(-9.9)	283.0	(4.4)	811	(-8.8)	4286	(-19.9)
3	200	46.8	(-16.4)	119.0	(-11.3)	217.4	(-19.8)	750	(-15.5)	—	—
4	270	54.4	(-2.9)	131.0	(-2.4)	258.6	(-4.6)	929	(4.6)	4918	(-8.1)
5	270	61.2	(9.3)	142.9	(6.5)	272.7	(0.6)	1000	(12.6)	3000	(-43.9)
6	270	52.6	(-6.1)	124.5	(-7.2)	252.1	(-7.0)	888	(0.0)	4770	(-10.9)
7	270	57.6	(2.9)	151.5	(12.9)	329.0	(21.4)	1163	(31.0)	6667	(24.6)
8	270	58.5	(4.5)	140.0	(4.3)	278.0	(2.6)	646	(-27.3)	6211	(16.1)
9	270	56.6	(1.1)	136.4	(1.6)	260.9	(-3.7)	904	(1.8)	4918	(-8.1)
10	270	58.3	(4.1)	125.5	(-6.5)	256.4	(-5.4)	974	(9.7)	6110	(14.2)
11	270	57.6	(2.9)	137.0	(2.1)	275.2	(1.5)	940	(5.9)	5848	(9.3)
12	270	55.4	(-1.1)	132.2	(-1.5)	260.9	(3.7)	915	(3.0)	5445	(1.7)
13	300	51.6	(-7.9)	120.0	(-10.6)	246.0	(-9.2)	680	(-23.4)	4080	(-23.8)
14	360	53.8	(-3.9)	128.8	(-4.0)	300.0	(10.7)	600	(-32.4)	3333	(-37.7)
15	400	55.4	(-1.1)	136.0	(1.3)	270.1	(-0.3)	924	(4.1)	4107	(-23.3)
16	400	58.8	(5.0)	143.0	(6.6)	250.0	(-7.7)	750	(-15.5)	—	—
17	400	56.1	(0.2)	135.0	(0.6)	268.0	(-1.1)	955	(7.5)	5700	(6.5)
18	400	59.1	(5.5)	139.0	(3.6)	286.0	(5.5)	739	(-16.8)	6464	(20.8)
19	400	56.0	(0.0)	129.9	(-3.2)	263.2	(-2.9)	875	(-1.5)	5263	(-1.7)
20	400	56.6	(1.1)	135.1	(-0.7)	275.2	(1.5)	949	(6.9)	6148	(14.9)
21	400	54.9	(-2.0)	132.0	(-1.6)	305.0	(12.5)	943	(6.2)	7040	(31.5)
22	500	51.0	(-8.9)	126.7	(-5.6)	244.0	(-9.9)	843	(-5.1)	4815	(-10.0)
23	500	56.2	(0.4)	133.0	(-0.9)	275.0	(1.5)	1060	(19.4)	6880	(28.6)
24	500	56.6	(1.1)	132.0	(-1.6)	270.0	(-0.4)	683	(-23.1)	3243	(-39.4)
25	500	62.8	(12.1)	142.0	(5.8)	306.0	(12.9)	1070	(20.5)	—	—
26	500	60.0	(7.1)	160.0	(19.2)	301.0	(11.1)	974	(9.7)	8090	(51.2)
27	500	61.0	(8.9)	140.2	(4.5)	283.0	(4.4)	1000	(12.6)	6000	(12.1)
Average		56.0		134.2		271.0		888		5352	
Precision(%)		6.4		6.7		8.9		15.4		23.4	
<i>DP</i> (SEC) ^b		53.4		120.6		256.0		977		5200	

^aFigures in parentheses represent the values of $100 \times [(\text{an individual determination}) - (\text{average})]/(\text{average})$. ^b*DP* determined by SEC.

Table VI. ¹H NMR Determination of *DP* of *it*-PMMA samples—Data sorted out in terms of frequency of measurement

Freq. (MHz)	n ^b	Sample ^a				
		1	2	3	4	5
100	2	51.4(1.1)	125(2.6)	256(10.5)	909(10.8)	4699(8.8)
200	1	46.8(-)	119(-)	217(-)	750(-)	—
270	9	56.9(4.2)	136(5.9)	272(8.2)	928(14.0)	5320(19.3)
300	1	51.6(-)	120(-)	246(-)	680(-)	4080(-)
360	1	53.8(-)	129(-)	300(-)	600(-)	3330(-)
400	7	56.7(2.6)	136(3.0)	274(5.9)	876(9.9)	5790(16.2)
500	6	57.7(6.7)	139(7.7)	280(7.4)	938(14.5)	5806(28.8)
<i>DP</i> (SEC) ^c		53.4	120.6	256	977	5200
100 ^d		52.3(3.9)	122(5.6)	263(8.3)	1020(6.0)	5420(9.2)
500 ^d		50.5(1.3)	119(3.1)	246(0.9)	912(0.7)	4750(3.4)

^aFigures in parentheses represent the precision (%). ^bNumber of instruments used. ^cDetermined by SEC. ^dData were collected from the five runs of single spectrometer.

SEC if the experimental errors of both measurements are taken into account.

The fluctuation of the data must be small when the measurements are made at one test site using a single spectrometer. The results of the *DP* determinations

from the single ¹H NMR spectrometers (Osaka University) operated at 100 and 500 MHz, respectively, are shown in Table VI (the bottom two lines). The precisions remain within 10% for all the samples even in the data obtained from a 100 MHz spectrometer. Precisions

for 500 MHz NMR data are less than at most 3.4%. The averaged *DP* values for all the samples agreed very well with the *DP* determined by SEC within the experimental errors. The intensity ratio of the peaks due to the OCH₃ protons to *t*-C₄H₉- group for sample 5 is about 1780:1 on average. The precision and accuracy for the determination of the ratio of this magnitude have been found to be 8.6 and 2.9% for 100 MHz NMR and 4.3 and 2.9% for 500 MHz NMR.⁸ Thus, the *DP* of the polymer having an end group of definite chemical structure can be determined precisely and accurately by ¹H NMR up to the *DP* value at least 5000. In the present study by round robin method the numbers of scans used for sample 5 were from 32 to 5700. The maximum value of *DP* that could be measured by ¹H NMR would be increased more if the number of scans would be increased.

The data of *DP* in Table V were sorted out based on the frequency of measurements and the results are shown in Table VI. The precision and the agreement of the NMR data with SEC data do not seem to depend strongly on the frequency of measurement although there exists a trend that the values of precision increased with increasing frequency. The same trend for intensity measurement in ¹H NMR was observed in our previous work.⁹

The nuclear Overhauser effect (NOE) value of every carbon of PMMA differs from each other, particularly at higher frequency of measurement and removal of NOE should be needed for the accurate determination of peak intensities.² For the purpose, the ¹³C NMR spectra were measured by gate decoupling without NOE. From the gate-decoupled ¹³C NMR spectra, the *DP* values of the samples 1, 2, and 3 were obtained using the signal intensities of methyl carbons of α -CH₃ and *t*-C₄H₉ groups and are shown in Table VII by sorting the data in terms of the frequency. *DP* determinations for samples 4 and 5 are almost impossible in the measurements of 5000 scans (about 14 h) due to the low S/N ratios of CH₃ carbon signal of *t*-C₄H₉- group. Although the values of precisions for the data shown in Table VII are large as compared with those for the ¹H NMR data in Tables V and VI, the averaged values of *DP* agreed with those obtained by ¹H NMR and SEC.

In conclusion the *DP* of the PMMA having a *t*-C₄H₉- group at the chain end can be determined accurately

Table VII. ¹³C NMR Determination of *DP* of *it*-PMMA samples^a

Freq. (MHz)	Sample 1		Sample 2		Sample 3	
	<i>DP</i>	n ^b	<i>DP</i>	n ^b	<i>DP</i>	n ^b
50	60.6(—)	1	—	—	—	—
67.5	48.3(16.1)	9	123(20.6)	8	285(16.9)	6
75	62.1(—)	1	116(—)	1	—	—
90.6	60.5(—)	1	169(—)	1	—	—
100	54.6(11.1)	4	128(8.5)	4	240(9.5)	2
125	48.5(5.7)	4	128(16.1)	4	262(0.4)	2
Ave.	51.5(14.8)	20	126(18.3)	18	271(15.8)	10

^aFigures in parentheses represent the precision (%). ^bNumber of spectrometers.

and precisely by ¹H NMR spectroscopy up to the *DP* of about 5000 in the measurements with 32–5700 scans and repetition time of 10 s. In the case of ¹³C NMR the maximum value of *DP* that can be measured practically is around 300.

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