

Steric Control in Ring-Opening Polymerization of 1,6-Anhydro Galactose Derivatives by Neighboring Group Participation

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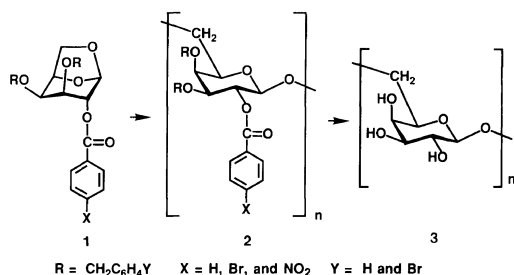
ABSTRACT: Four 1,6-anhydro- β -D-galactopyranoses having benzoyl derivatives in position 2 and benzyl derivatives in positions 3 and 4 were synthesized and polymerized. (1 \rightarrow 6)- β -D-Galactopyranan oligosaccharide derivatives ($DP_n < 7.6$) were obtained with PF₅ as initiator in dichloromethane at 0 to -40°C from 1,6-anhydro-2-*O*-benzoyl-3,4-di-*O*-benzyl- β -D-galactopyranose (**1a**), 1,6-anhydro-2-*O*-benzoyl-3,4-di-*O*-*p*-bromobenzyl- β -D-galactopyranose (**1c**), and 1,6-anhydro-2-*O*-*p*-nitrobenzoyl-3,4-di-*O*-*p*-bromobenzyl- β -D-galactopyranose (**1d**). 1,6-Anhydro-2-*O*-*p*-nitrobenzoyl-3,4-di-*O*-benzyl- β -D-galactopyranose (**1b**) gave a stereoirregular product of lower molecular weight in lower yield. The apparent rate of polymerization increased in the order of **1b** < **1a** < **1c** < **1d**. The formation of β -(1 \rightarrow 6)-linked configuration is explained by the mechanism in which the benzoyl group in position 2 took part in the steric retention of the anomeric center during the ring-opening polymerization. Substituent effects on the reactivities of the monomers and growing species are discussed on the basis of the mechanism.

KEY WORDS Synthetic Polysaccharide / Ring-opening Polymerization / Anhydro Sugar Derivatives / Neighboring Group Participation / (1 \rightarrow 6)- β -D-Galactopyranan/

Well-defined (1 \rightarrow 6)- β -linked galactopyranan is a useful model polysaccharide to study immunology and enzymology.^{1,2} We previously reported the synthesis of (1 \rightarrow 6)- β -D-galacto oligosaccharides *via* polymerization of 1,6-anhydro-2-*O*-benzoyl-3,4-di-*O*-benzyl- β -

D-galactopyranose (**1a**) and subsequent deprotection according to Scheme 1.^{3,4} The formation of β -(1 \rightarrow 6)-linkage was explained by a neighboring-group effect of the benzoyl group in position 2 on the steric control of the anomeric center in the propagation step.

This paper deals with ring-opening polymerization of the four different anhydrogalactoses having benzoyl derivatives in position 2 (Figure 1): 1,6-anhydro-2-*O*-benzoyl-3,4-di-*O*-benzyl- β -D-galactopyranose (**1a**); 1,6-anhydro-2-*O*-*p*-nitrobenzoyl-3,4-di-*O*-benzyl- β -D-galactopyranose (**1b**); 1,6-anhydro-2-*O*-benzoyl-3,4-di-*O*-*p*-bromobenzyl- β -D-galactopyranose (**1c**); 1,6-anhydro-2-*O*-*p*-bromobenzoyl-3,4-di-*O*-*p*-bromobenzyl- β -D-galactopy-



Scheme 1. Synthesis of (1 \rightarrow 6)- β -D-galactopyranan (3).

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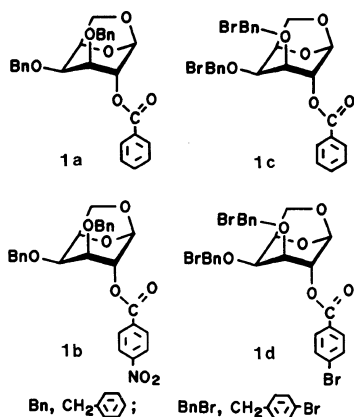


Figure 1. The structures of the monomers.

ranose (**1d**). Their polymerizations are investigated under several conditions and polymerization reactivities are discussed on the basis of the proposed mechanism.

EXPERIMENTAL

General Methods

200-MHz ^1H and 50-MHz ^{13}C NMR spectra were obtained on a Japan Electro-Optic Laboratory JNM-FX-200 Fourier transform NMR spectrometer for solutions in deuteriochloroform and $\text{Me}_2\text{SO}-d_6$ with tetramethylsilane as the internal reference. For deprotected polysaccharides, measurements were made on solutions in D_2O with acetone as a reference (2.07 ppm in ^1H NMR and 39.4 ppm in ^{13}C NMR). Optical rotations were determined with a JASCO DIP 181 digital polarimeter using a jacketed 1-dm cell. Gel-permeation chromatography was carried out on a Toso HLC-8020 high-speed chromatograph using TSK-gel GMH_{XL} \times 2 and G2000H_{XL}, G3000H_{XL}, G4000H_{XL}, and G5000H_{XL} columns (solvent, chloroform; polystyrene standards). Microanalysis was made on a Perkin-Elmer 240C elemental analyzer.

1,6-Anhydro-2-O-benzoyl-3,4-di-O-benzyl- β -D-galactopyranose (**1a**)

It was synthesized and purified by previous-

ly reported methods.⁵ mp, 93–94°C. $[\alpha]_{\text{D}}^{25} = +1.6^\circ$ (*c* 1.0, in chloroform). *Anal.* Calcd for $\text{C}_{27}\text{H}_{26}\text{O}_6$: C, 72.63%; H, 5.87%. Found: C, 72.59%; H, 5.83%.

^1H NMR (CDCl_3 , TMS): 8.07–7.28 (m, 15H, benzoyl- and benzyl- C_6H_5), 5.53 (s, 1H, H-1), 5.22 (s, 1H, H-2), 4.93 and 4.70 (2d, $J = 12.0$, 2H, benzyl- CH_2), 4.60–4.38 (m, 4H, H-5, H-6_{endo}, and benzyl- CH_2), 3.92 (m, 2H, H-3 and H-4), and 3.71 (d \times d, $J \approx 5.9$ and 6.1 Hz, 1H, H-6_{exo}).

^{13}C NMR (CDCl_3): 165.7 (C=O), 138.5 and 138.2 (*ipso*- C_6H_5 of benzyl), 133.8 (*para*- C_6H_5 of benzoyl), 129.9 (*ipso*- C_6H_5 of benzoyl), 130.2, 128.8, 128.7, 128.2, 127.9, (other C_6H_5 of benzoyl and benzyl), 100 (C-1), 74.9 (C-2), 73.6 (C-5), 73.4 (C-3), 73.1 (C-4), 71.2 and 71.1 (benzyl- CH_2), and 62.2 (C-6).

1,6-Anhydro-2-O-*p*-nitrobenzoyl-3,4-di-O-benzyl- β -D-galactopyranose (**1b**)

It was prepared from 1,6-anhydro-3,4-di-O-benzyl- β -D-galactopyranose.⁵ mp, 107.5–110.5°C. $[\alpha]_{\text{D}}^{25} = +23.8^\circ$ (*c* 1.0, in chloroform). *Anal.* Calcd for $\text{C}_{27}\text{H}_{25}\text{O}_8\text{N}$: C, 65.98%; H, 5.13%; N, 2.85%. Found: C, 66.01%; H, 5.16%; N, 2.72%.

^1H NMR (CDCl_3 , TMS): 8.24 (m, 4H, C_6H_4), 7.35–7.25 (m, 5H, C_6H_5), 5.53 (s, 1H, H-1), 5.21 (s, 1H, H-2), 4.90 and 4.70 (2d, $J = 12.1$, 2H, benzyl- CH_2), 4.64–4.42 (m, 4H, benzyl- CH_2 , H-5 and H-6_{endo}), 3.91 (m, 2H, H-3 and H-4), 3.72 (d \times d, $J \approx 6.2$ and 5.6 Hz, 1H, H-6_{exo}).

^{13}C NMR (CDCl_3): 163.2 (C=O), 150.6 (*para*- C_6H_4), 137.7 and 137.5 (*ipso*- C_6H_5), 134.6 (*ipso*- C_6H_4), 130.8 (*ortho*- C_6H_4), 128.2–127.4 (C_6H_5), 123.4 (*meta*- C_6H_4), 99.1 (C-1), 74.2 (C-2), 73.0 (C-5), 72.7 (C-3 and C-4), 71.5 and 70.8 (benzyl- CH_2), and 64.7 (C-6).

1,6-Anhydro-2-O-benzoyl-3,4-di-O-*p*-bromobenzyl- β -D-galactopyranose (**1c**)

It was prepared by treating 1,6-anhydro-3,4-di-O-*p*-bromobenzyl- β -D-galactopyranose with benzoyl chloride. mp, 79–81°C. $[\alpha]_{\text{D}}^{25} =$

+12.1° (*c* 1.0, in chloroform). *Anal.* Calcd for $C_{27}H_{24}O_6Br_2$: C, 53.66%; H, 4.00%. Found: C, 53.67%; H, 3.95%.

1H NMR ($CDCl_3$): 8.07–7.05 (m, 13H, benzoyl- C_6H_5 and *p*-bromobenzyl- C_6H_4), 5.53 (s, 1H, H-1), 5.17 (s, 1H, H-2), 4.87 and 4.60 (2d, $J=12.2$, 4H, benzyl- CH_2), 4.55 (d, $J=7.6$, 2H, H-6_{endo} and H-5), 4.42 (d, $J=3.2$, 1H, benzyl- CH_2), 3.89 (broad s, 2H, H-3 and H-4), and 3.72 (d × d, $J \approx 5.6$ and 5.4 Hz, 1H, H-6_{exo}).

^{13}C NMR ($CDCl_3$): 165.3 (C=O), 137.0 and 136.6 (*ipso*- C_6H_4 of *p*-bromobenzyl), 133.6 (*para*- C_6H_5 of benzoyl), 131.5, 131.3, 129.9, 129.3, 129.0, and 128.5 (*ortho*- and *meta*- C_6H_4 , and *ipso*-, *meta*-, and *ortho*- C_6H_5), 121.5 (*para*- C_6H_4), 99.5 (C-1), 74.3 (C-2), 72.9 (C-5), 72.8 (C-3), 71.7 (C-4), 70.3 and 69.9 (*p*-bromobenzyl- CH_2), and 64.6 (C-6).

1,6-Anhydro-2-*O*-*p*-bromobenzoyl-3,4-*O*-isopropylidene- β -D-galactopyranose

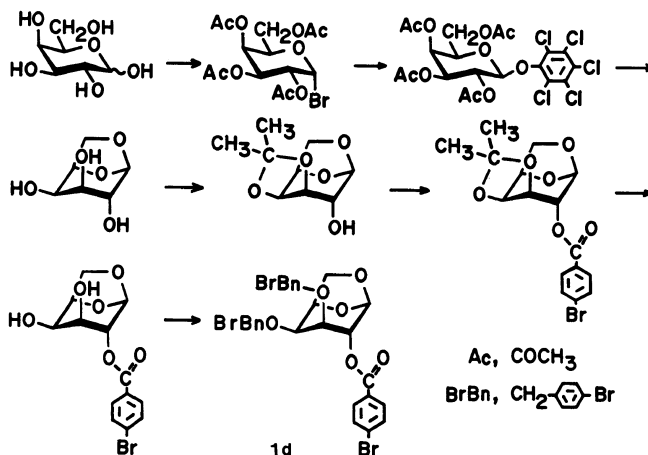
1,6-Anhydro- β -D-galactopyranose was prepared by a modified method⁷ *via* pentachlorophenyl 2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranoside. 1,6-Anhydro-3,4-*O*-isopropylidene- β -D-galactopyranose was prepared by a modification of the reported method.⁶ According to Scheme 2, pentachlorophenyl β -D-

galactopyranose was employed as an intermediate⁷ instead of phenyl β -D-galactopyranose.

A solution of 1,6-anhydro-3,4-*O*-isopropylidene- β -D-galactopyranose⁶ (16.6 g, 0.082 mol) in chloroform (400 ml) was treated with triethylamine (25 ml) and *p*-bromobenzoyl chloride (50 g, 0.228 mol) at room temperature for 6 h. Water (10 ml) was added and the mixture was stirred for 2 h and concentrated. The residue was dissolved in chloroform and washed with water and a saturated aqueous sodium hydrogen carbonate. The chloroform layer was concentrated to dryness, and the residue was crystallized from ethanol. Yield was 27 g (86%). mp, 147.5–148.5°C. $[\alpha]_D^{25} = +22.2^\circ$ (*c*, 1.0, in chloroform).

1H NMR ($CDCl_3$): 7.94 (d, $J=8.4$, 2H, *ortho*- C_6H_4), 7.59 (d, $J=8.4$, 2H, *meta*- C_6H_4), 5.49 (s, 1H, H-1), 5.12 (s, 1H, H-2), 4.59 (t, 1H, H-5), 4.51 (t, 1H, H-4), 4.24 (d, $J=6.9$, 1H, H-3), 4.19 (d, $J=7.5$, 1H, H-6_{endo}), 3.65 (d × d, $J=7.5$ and 5.1, 1H, H-6_{exo}), and 1.57 and 1.37 (2s, 6H, CH_3).

^{13}C NMR ($CDCl_3$): 164.4 (C=O), 131.8 and 131.3 (*ortho* and *meta* C_6H_4), 128.6 and 128.2 (*ipso* and *para* C_6H_4), 109.0 (>C<OO), 98.9 (C-1), 73.9 (C-2), 72.0 (C-3 and C-5), 69.1 (C-4), 63.3 (C-6), 25.7 and 24.2 (CH_3).



Scheme 2. Synthetic procedure of the monomer 1d.

1,6-Anhydro-2-O-p-bromobenzoyl-β-D-galactopyranose

1,6-Anhydro-2-O-*p*-bromobenzoyl-3,4-O-isopropylidene-β-D-galactopyranose) (27.3 g, 70.7 mmol) was treated with a mixture (250 ml) of trifluoroacetic acid and water (7:3 in volume) at room temperature for 30 min. Methanol (10 ml) was added and the mixture was concentrated to dryness and the residue was crystallized from ethanol. Yield was 20.6 g (86%). $[\alpha]_D^{25} = +56.6^\circ$ (*c* 1.0, in dimethyl sulfoxide).

$^1\text{H NMR}$ ($\text{Me}_2\text{SO}-d_6$): 7.90 (d, $J = 8.7$, 2H, *ortho*-C₆H₄), 7.76 (d, $J = 8.7$, 2H, *meta*-C₆H₄), 5.41 (s, 1H, H-1), 5.22 and 5.16 (d, $J = 3, 4$, 1H and d, $J = 6.8$, 1H, OH-3 and OH-4). These signals disappeared with a drop of H₂O), 4.84 (s, 1H, H-2), 4.36 (d, H-5 and H-6_{endo}), 3.85 (broad s, 2H, H-3 and H-4), and 3.50 (broad d × d, 1H, H-6_{exo}).

$^{13}\text{C NMR}$ ($\text{Me}_2\text{SO}-d_6$): 163.8 (C=O), 131.8 and 131.2 (*ortho*- and *meta*-C₆H₄), 128.3 (*ipso*-C₆H₄), 127.5 (*para*-C₆H₄), 98.1 (C-1), 74.0 (C-2), 73.7 (C-5), 67.9 (C-3), 64.6 (C-4), and 63.1 (C-6).

p-Bromobenzyl Trichloroacetimidate

The method^{8,9} reported for benzyl trichloroacetimidate was modified. Sodium hydride (110 mg, 2.7 mmol) was washed with hexane and dispersed in tetrahydrofuran (3 ml). *p*-Bromobenzyl alcohol (5.0 g, 27 mmol) in tetrahydrofuran (5 ml) was added to it in an atmosphere of nitrogen. The mixture was cooled at 0°C in an ice bath. Trichloroacetonitrile (2.7 ml, 26.7 mmol) was added through a dropping funnel for 15 min. The mixture was gradually warmed to room temperature over a period of 60 min and concentrated. Crude crystals deposited were dissolved into *n*-hexane containing a small amount of methanol. The supernatant was decanted and concentrated. Yield was 8.6 g (97%). The colorless crystal was stored in a refrigerator.

$^1\text{H NMR}$ (7.1% in CDCl₃, TMS, 200 MHz): δ 8.41 (s, 1H, NH), 7.50 (d, $J = 8.6$, 2H,

meta-C₆H₄), 7.30 (d, $J = 8.6$, 2H, *ortho*-C₆H₄), and 5.28 (s, 2H, CH₂).

$^{13}\text{C NMR}$ (7.1% in CDCl₃, TMS, 50 MHz): 162.1 (C=NH), 134.2 (*ipso*-C₆H₄), 131.4 (*meta*-C₆H₄), 129.1 (*ortho*-C₆H₄), 122.1 (*para*-C₆H₄), and 69.7 (CH₂).

1,6-Anhydro-2-O-p-bromobenzoyl-3,4-di-O-p-bromobenzyl-β-D-galactopyranose (1d)

1,6-Anhydro-2-O-*p*-bromobenzoyl-β-D-galactopyranose (1.1 g, 3.2 mmol) in dichloromethane (200 ml) was treated with *p*-bromobenzyl trichloroacetimidate (4.24 g, 12.8 mmol) in cyclohexane (50 ml) and trifluoromethanesulfonic acid (0.3 ml) at room temperature. After 26.5 h, the reaction was terminated with pyridine (6 ml). The mixture was washed with water (100 ml), a saturated aqueous sodium hydrogen carbonate solution (120 ml), and water (100 ml). The organic layer was concentrated, and chromatographed on silica gel with *n*-hexane–ethyl acetate (3:1 in volume) as eluent. Yield was 2.9 g (64%). The product was crystallized from ethanol. mp, 98–99.5°C. $[\alpha]_D^{25} = +33.9^\circ$ (*c* 1.0, in chloroform).

$^1\text{H NMR}$ data (CDCl₃, TMS): 7.90 and 7.60 (2d, $J = 8.5$, 4H, benzoyl-C₆H₄), 7.4–7.11 (m, 8H, benzyl-C₆H₄), 5.52 (s, 1H, H-1), 5.14 (s, 1H, H-2), 4.85 and 4.60 (2d, $J = 12.2$, 2H, benzyl-CH₂), 4.54 (d, $J = 7.3$, 2H, H-6_{endo} and H-5), 4.4 (d, $J = 2.2$, 2H, benzyl-CH₂), 3.87 (s, 2H, H-3 and H-4), and 3.72 (d × d, $J \approx 6.6$ and 5.6 Hz, 1H, H-6_{exo}).

$^{13}\text{C NMR}$ (CDCl₃, TMS): 164.4 (C=O), 136.7 and 136.4 (*para*-C₆H₄ of benzyl), 131.7, 131.3, 131.2, 131.1, 129.1, 128.8, and 127.9, (C₆H₄ of benzoyl and *ortho*- and *meta*-C₆H₄ of benzyl), 121.6, and 121.3 (*ipso*-C₆H₄ of benzyl), 99.2 (C-1), 74.2 (C-2), 72.8 (C-5), 72.7 (C-3), 71.7 (C-4), 70.5 and 69.9 (benzyl-CH₂), and 64.5 (C-6).

Polymerization^{3,4,10}

Each monomer was dried on calcium hydride in anhydrous dichloromethane in a polymerization vessel attached to a high

vacuum system. Polymerization was carried out with phosphorus pentafluoride at 0°C to -40°C and terminated with a cold mixture of methanol and pyridine. The product was chromatographed on silica gel with *n*-hexane-ethyl acetate (2:1 in volume) as eluent to remove monomer. The eluate was concentrated and the residue was freeze-dried from benzene.

Deprotection^{3,4,10}

A solution of polymer **2** (0.20 g) in a mixture of toluene (15 ml) and 1,2-dimethoxyethane (5 ml) was added to 30 ml of liquid ammonia at -33°C. Small pieces of freshly cut sodium metal were added to the mixture until the dark blue color of the solution persisted. A small amount of ammonium chloride and water (15 ml) were successively added dropwise. The mixture was stirred overnight at room temperature to evaporate ammonia. The water layer was passed through a column of Amberlite IR-120 (H⁺) resin. The eluate was concentrated by a rotary evaporator and the product was reprecipitated into methanol. The methanol-insoluble fraction was freeze-dried from water to give a white hygroscopic powder.

RESULTS AND DISCUSSION

Synthesis of 1,6-Anhydro-β-D-galactopyranose Derivatives

1,6-Anhydro-β-D-galactopyranose⁶ was prepared from galactose and then the hydroxyl groups were protected regiospecifically with benzyl and benzoyl derivatives. Monomers **1a**, **1b**, and **1c** were prepared according to the previously reported reaction route⁵ which includes early benzylation with benzyl halides and subsequent benzoylation.

Monomer **1d** was synthesized *via* the modified reaction route as shown in Scheme 2. The hydroxyl group in position 2 of 1,6-anhydro-3,4-*O*-isopropylidene-β-D-galactopyranose was benzoylated at first, followed by removal

of the isopropylidene group and subsequent benzylation of the resulting hydroxyl groups in positions 3 and 4. This route involves no extra steps of protection and deprotection of the hydroxyl group in position 2. Instead, *p*-bromobenzyl trichloroacetimidate was employed as the benzylation reagent. The *p*-bromobenzylation was operated under mild acidic conditions to avoid the removal of the benzoyl group.

Conformational Aspects of 1,6-Anhydro-β-D-galactopyranose Derivatives

Compounds **1a**—**1d** were obtained as crystals. The ¹H NMR spectral data were described in Experimental section. The assignments of some samples were based on the homo-decoupling technique. Although complete analysis has not been made, the chemical shifts and coupling constants were similar among the four compounds as follows. (1) Broad singlet signals of H-1 and H-2 appeared at 5.53 and ~5.2 ppm, respectively. (2) The signals at 3.72 ppm of H-6_{exo} were seemingly triplet, whose coupling constants were about 5 to 7 Hz due to $J_{\text{H-6}_{\text{exo}},\text{H-5}}$ and $J_{\text{H-6}_{\text{exo}},\text{H-6}_{\text{endo}}}$. (3) One of the benzyl-CH₂ signals were composed of an AB coupling system at 4.9 and 4.7 ppm ($J=12.1$ Hz). The former two characteristics are similar to those of skeletal 1,6-anhydro galactopyranose.¹¹ There are no detectable differences in ring distortion which may be correlated with the polymerizability.

Polymerization of 1,6-Anhydro-β-D-galactopyranose Derivatives

Table I summarizes the results of polymerization of **1a**—**1d**.

Polymerizations of **1c** using phosphorus pentafluoride as catalyst in dichloromethane proceeded at 0 and -20°C to give methanol insoluble products at moderate yields (expt. IT-6 and IT-8). There appeared a single anomeric C-1 carbon signal at 101.3 ppm, whose ¹³C-¹H coupling constant ($J_{\text{C-1},\text{H-1}}$) was 163.0 Hz assignable to the coupling be-

Table I. Polymerization of 1,6-anhydrogalactose derivatives (**1a**–**1d**)^a

Exp. no.	Monomer	[M] ₀	Initiator ^a	Solv.	Temp	Time	Yield	<i>M</i> _n ^b × 10 ⁻³	[α] _D ²⁵ ^c	β-Form ^d
		mol l ⁻¹			°C	h	%		deg	%
I-76	1a	1.5	PF ₅	CH ₂ Cl ₂	0	6	24	3.6	+32	~100
I-77	1a	1.5	PF ₅	CH ₂ Cl ₂	0	48	66	2.6	+27	~100
I-81	1a	1.5	PF ₅	CH ₂ Cl ₂	-20	6	1	—	—	—
I-89	1a	1.5	BF ₃ OEt ₂ ^g	CH ₂ Cl ₂	0	48	30	1.8	+38	~100
IT-22	1b	1.2	PF ₅	CH ₂ Cl ₂	0	48	16	1.6	—	—
IT-18	1b	^e	PF ₅	CH ₂ Cl ₂	-20	18	4	1.7	+46	—
IT-6	1c	1.2	PF ₅	CH ₂ Cl ₂	0	6	50	3.8	+45	~100
IT-8	1c	1.2	PF ₅	CH ₂ Cl ₂	-20	48	35	2.5	+27	~100
IT-9	1c	1.2	PF ₅	CH ₂ Cl ₂	-40	48	trace	2.0	—	—
IT-29	1c	^f	SbCl ₅ ^h	CH ₂ Cl ₂	0	18	36	1.8	+33	~100
IT-19	1c	1.2	PF ₅	CH ₃ C ₆ H ₅	0	10	9	4.4	—	~100
IT-20	1c	1.2	PF ₅	C ₃ H ₇ NO ₂	0	6	56	4.6	+38	~100
							28	2.3	+35	—
IT-25	1c	1.2	PF ₅	C ₃ H ₇ NO ₂	-20	12	36	3.6	—	~100
							17	1.3	—	—
IT-23	1d	1.2	PF ₅	CH ₂ Cl ₂	0	6	56	3.1	+36	<100
IT-24	1d	1.2	PF ₅	CH ₂ Cl ₂	-20	48	60	3.4	+22	~100
IT-26	1d	1.2	PF ₅	CH ₂ Cl ₂	-40	48	24	2.3	+25	~100

^a Initiator, PF₅, 10 mol%.^b By GPC, polystyrene standard.^c In chloroform at 25°C.^d By ¹³C NMR spectroscopy.^e Monomer, 1.0 mmol; CH₂Cl₂, 0.55 ml.^f Monomer, 1.0 mmol; CH₂Cl₂, 0.50 ml.^g Initiator, BF₃OEt₂, 30 mol%.^h Initiator, SbCl₅, 15 mol%.

tween the β- anomeric carbon and axial hydrogen atom.¹² The polymerization proceeded stereospecifically to give stereoregular polymers having β-(1→6)-linkage. Number-average molecular weights of the polymers estimated by GPC were 2500–3800, which correspond to \overline{DP}_n s of 4.1–6.3. Deprotection of the benzyl and benzoyl groups of the polymers was performed using sodium in liquid ammonia to give the polysaccharide **3** quantitatively. The β-(1→6)-linked structure of the products was confirmed by comparing the ¹³C NMR spectra with those of β-(1→6)-galactopyranan oligosaccharides reported in the literature.^{5,13}

When boron trifluoride diethyl etherate and antimony pentachloride were used as initiators (expt. I-89 and IT-29), the products were of

lower molecular weight.

Polymerizations of **1c** with PF₅ initiator in toluene and 1-nitropropane were compared with the polymerizations in dichloromethane. Polymerization was slow in toluene (expt. IT-19). Polymerization in 1-nitropropane gave products in rather high yields (expt. IT-20 and IT-25). However, there appeared two peaks in their GPC chromatograms, and two products with different structures were separated by silica gel chromatography. The higher molecular-weight product was β-(1→6)-galactopyranan derivative which was similar to those obtained in dichloromethane and toluene. The lower molecular-weight product had a little different NMR resonances, and its chemical structure has not been identified.

Polymerization of monomer **1d** proceeded

at 0°C, -20°C, and even -40°C. Only the β -anomeric C-1 carbon signal was observed at 100.8 ppm for the product IT-24 and IT-26, although the α -anomeric carbon signal was observed for the product (IT-23) obtained at 0°C. The number-average molecular weights were in the range of 2300–3400 (\overline{DP}_n s, 3.4–5.0).

Polymerization of **1b** was slow, and the yield and molecular weight of the product were low, compared to those of **1a**, **1c**, and **1d**. In the anomeric region of the ^{13}C NMR spectrum of the products (expt. IT-22 and IT-18) were observed two broad signals at 99.1 and 98.5 ppm. The polymer was of irregular structures.

Polymerizability

The polymerizability of **1** was lower than that of 1,6-anhydro-2,3,4-tri-*O*-benzyl- β -D-galactopyranose,^{6,14,15} the latter of which gave (1 \rightarrow 6)- α -D-galactopyranan derivative with high α -stereoregularity and high molecular weight. The polymerizabilities of the four monomers were in the order: **1b** < **1a** < **1c** < **1d**, as judged from the following comparison.

(1) Polymer yields at 0°C (expt. IT-22, I-77, IT-6, and IT-23).

(2) Apparent polymerizability as a function of temperature. Polymer could be obtained even at -40°C from **1d** (expt. IT-26).

(3) High β -stereoregularity of the polymers

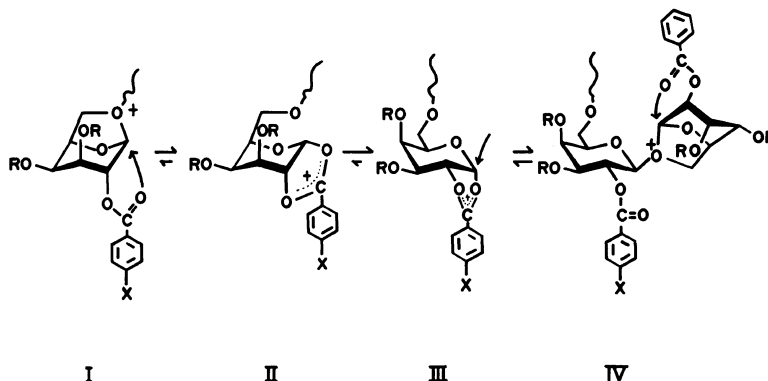
obtained from **1a**, **1c**, and **1d**, in contrast to the lack of stereoregularity of **1b**.

(4) For the products prepared from **1a** were observed unassignable ^{13}C NMR signals at 20–45 ppm, which disappeared after deprotection. Some side reactions might occur at the sites of the protecting groups of **1a**. Such signals were not detected for the polymers **1c**, **1d**, and also for the deprotected polymers.

Introduction of bromine atom to the para positions of the benzyl and benzoyl groups increased the rate of polymerization, but did not increase the degree of polymerization.

Reaction Mechanism

Scheme 3 illustrates the proposed propagation mechanism, in which the neighboring benzoyloxy group in position 2 takes part in the steric control of the anomeric reaction center to produce the β -(1 \rightarrow 6)-galactopyranan derivatives. The cyclic trialkyloxonium ion (I)¹⁴ is transiently formed at the growing terminal unit. It is immediately attacked by the carbonyl oxygen of the neighboring benzoyl substituent. As a result, the C-1 configuration is inverted and a dioxacarbenium (dioxolenium or benzoyloxonium) ion (II) is formed. The conformation of the pyranose ring may be converted from $^1\text{C}_4$ to $^4\text{C}_1$ form (III). Then, the incoming monomer can attack the C-1 carbon of III exclusively from the



Scheme 3. Neighboring-group participation in formation of (1 \rightarrow 6)- β -linkage.

opposite side of the dioxacarbenium ion, that is, the β -side of the pyranose ring. The C-1 configuration is inverted again, and the trialkyloxonium ion (IV) is regenerated.

The ${}^1\text{C}_4$ conformation of monomers **1a**–**1d** may be reasonably assumed on the basis of the above mentioned NMR data.¹¹ The chair form conformations are also adopted for the intermediate species I–IV, although the possibility of other transitional conformations is not ruled out. The species I is directly converted to III in the previously proposed scheme.^{3,4} In Scheme 3, however, the species II has been placed between I and III to take into account some conformational change like ${}^1\text{C}_4 \rightarrow {}^4\text{C}_1$. According to the examination of molecular models, the back-side attack of monomer to the reaction center of II seems to be inhibited owing to the steric hindrance.

The reactivities of the growing species toward monomers may be influenced by electronic nature of the substituents at the para position of the benzoyl and benzyl groups. It can be summarized from the following discussion that, on the whole, substituents with moderate electron-withdrawing characters must be introduced in order to enhance the overall reaction rate through I to IV.

In the step of I \rightarrow II, the ester carbonyl group acts as a nucleophile, and hence an electron-donating substituent on the 2-*O*-benzoyl group will accelerate the formation of the species II.¹⁶ In the latter III \rightarrow IV step, the same ester carbonyl group must act as a leaving group and, in this turn, an electron-withdrawing substituent is required. On the other hand, the electron-withdrawing substituent will lower the nucleophilicity of the acetal oxygen of anhydro ring and slow down the attack of the monomer toward the species III. Furthermore, the neighboring group participation (I \rightarrow II) also competes with the nucleophilic attack of the monomer toward the species I, the latter of which would result in the formation of α -anomeric sequences. The charge-bearing central carbon atoms of the benzoyl-

oxonium ion (II and III) can also become a reaction centers of nucleophilic attack.¹⁷ The attack would yield quite different by-products instead of the normal ring-opening products.

Much slower polymerization of **1** than that of 1,6-anhydro-2,3,4-tri-*O*-benzyl- β -D-galactopyranose is rationalized by the following assumption. The dioxolenium ion III is thermodynamically more stable than the trialkyloxonium ion I and the rate-determining step is III \rightarrow IV. As a result, the nucleophilic attack of the monomer to III becomes slow. The relative stability of species III is also responsible for the low degree of polymerization of the polymers **2**. In these respects, the neighboring group participation contributes to steric control but not to rate enhancement, that is, no anchimeric assistance.

It is also worth to note that a larger amount of Lewis acid PF_5 as initiator was needed to polymerize the monomers **1** than 1,6-anhydro-2,3,4-tri-*O*-benzyl- β -D-galactopyranose. Coordination of PF_5 to basic oxygen atoms was increased in the presence of the ester functions and reduced by electron-withdrawing substituents on benzoyl and benzyl groups.¹⁸

As judged from the lower polymerization rate and the lower stereoregularity of **1b**, *p*-nitrobenzoyl substituent was too strong an electron-withdrawing group. First, the nucleophilicity of the acetal oxygen of the monomer **1b** was decreased. Second, the activation energy of the neighboring group participation (I \rightarrow II) might be elevated to allow the nucleophilic attack of the monomer toward the species I.

Bromine substituent of benzoyl group of **1d** was more suitable to retain the β -anomeric configuration and increase the overall polymerization rate. The effects of bromo substituents of the benzyl groups in position 3 and 4 also must be considered. According to the comparison of **1a** and **1c**, the bromo substituents increased the polymer yield but not the degree of polymerization. We assume that the initiation step¹⁹ is accelerated.

1,6-Anhydro- β -D-galactopyranose derivative is known to possess lower ring-opening reactivity than other anhydro sugar skeletons.¹⁴ More highly reactive skeletal structures may be possible to give higher molecular weight polymers in higher yields.

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