

NOTES

Solubility of Cellulose Acetate Prepared by Different Methods and Its Correlations with Average Acetyl Group Distribution on Glucopyranose Units

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Very recently, experimental determination of the substituent group distribution towards three hydroxyl groups directly attached to C_2 , C_3 , and C_6 positions in a glucopyranose unit of cellulose derivatives $\langle f_k \rangle$ ($k=2, 3, 6$) has attracted keen attention, because this distribution has been shown to govern some important physical and physiological properties. Kamide, Okajima, and their co-workers¹⁻³ demonstrated that the anticoagulant activity of sodium cellulose sulfate (NaCS) against human blood and the absorbency of sodium carboxymethyl cellulose (NaCMC) against water and physiological saline water are significantly influenced by their $\langle f_k \rangle$, in particular $\langle f_2 \rangle + \langle f_3 \rangle$ for NaCS and $\langle f_6 \rangle$ for NaCMC. They¹⁻³ also showed that NaCS and NaCMC with different $\langle f_k \rangle$ can be prepared under adequate conditions, and cellulose acetate may not be an exception.

In 1981 Kamide and Okajima⁴ attempted to evaluate $\langle f_k \rangle$ of cellulose acetate (CA) by assigning three peaks in carbonyl carbon region to C_2 , C_3 , and C_6 positions. Later, Miyamoto *et al.*⁵ estimated $\langle f_k \rangle$ of CA based on different assignments of the peaks of three

carbonyl carbons. Kowsaka and his collaborators⁶ showed that the theoretical basis of the assignment of carbonyl carbon peaks in the past studies on CA, especially when the total degree of substitution $\langle F \rangle$ is below 3, is not always very reliable, due to unavoidable overlapping of carbonyl carbon peaks originated from mono-, di-, and tri-substituted glucopyranose units. They succeeded to give a reasonable assignment of carbonyl carbon peaks of tri-substituted glucopyranose unit by applying a low-power selective spin decoupling method to acetyl methyl proton located at the specific carbon position.⁶

Generally, CA having $\langle F \rangle = 1.7-2.5$ dissolves at room temperature readily in various organic solvents, including acetone, whereas it never dissolves in water at any temperature. Kamide *et al.*⁷ synthesized a series of CA with low $\langle F \rangle$ by adopting a method of hydrolysis reaction of CA with $\langle F \rangle = 2.46$ with hydrochloric acid, confirming that CA prepared thus is water-soluble at "room temperature" in the $\langle F \rangle$ range of *ca.* 0.4 to 0.9. They established, for well-fractionated CA with $\langle F \rangle = 0.49$, relationships between the limiting

viscosity number $[\eta]$, the radius of gyration $\langle S^2 \rangle_z^{1/2}$ and the weight-average molecular weight \bar{M}_w , estimating the Flory's viscosity parameter Φ , the unperturbed chain dimension A and the conformation parameter σ .

Four years later Miyamoto *et al.*⁸ investigated the water-solubility of CA with low $\langle F \rangle$, which are prepared by (1) the hydrolysis of CA sample with $\langle F \rangle = 2.94$ in aq. acetic acid (D series) or in dimethylsulfoxide (DMSO) (H series) and (2) the acetylation of cellulose dissolved in dimethylacetamide (DMAC)/lithium chloride (LiCl) mixture (A series). They concluded that only CA samples with $\langle F \rangle$ of 0.5–1.1 prepared by method (1) are completely water-soluble at 20°C. Based on analysis of $\langle f_k \rangle$ of the water-soluble and -insoluble parts, estimated from ¹³C and ¹H NMR spectra, they showed $\langle f_6 \rangle > \langle f_3 \rangle$ or $\langle f_2 \rangle$ for H and A series, but $\langle f_2 \rangle \simeq \langle f_3 \rangle \simeq \langle f_6 \rangle$ for D series.

In this article we clarify the effects of the preparing conditions on $\langle f_k \rangle$ and the relative ratio of various substituted glucopyranose units of CA and to obtain the relations between $\langle F \rangle$, $\langle f_k \rangle$ ($k = 2, 3, \text{ and } 6$) and the solubility of CA, discussing the dissolution mechanism of cellulose derivatives in solvents. For this purpose, we prepared nine CA samples by (1) a direct acetylation method of cellulose solution (one-step method) and (2) the acid-hydrolysis of cellulose triacetate with $\langle F \rangle = 2.92$ (two-step method), as described in the

previous paper,⁷ and measured ¹³C NMR spectra of the samples in deuterated dimethylsulfoxide (DMSO-*d*₆).

EXPERIMENTAL

Preparation of Cellulose Acetate Samples

One-Step Method. Softwood pulp (cellulose I, the viscosity-average molecular weight $\bar{M}_v = 1.7 \times 10^5$, estimated from by the Mark-Houwink-Sakurada (MHS) equation proposed by Brown and Wikström⁹ for cellulose in cadoxen solution at 25°C) was immersed in 5 *N*-sulfuric acid at 60°C for 80 min to give a hydrolyzed cellulose (cellulose I, $\bar{M}_v = 8 \times 10^4$). Five g of this cellulose sample were dipped in 200 g of *N,N*-dimethylacetamide (DMAC) (guaranteed grade), heated at 165°C for 30 min with reflux, added 16 g of LiCl at 100°C, cooled down to 20°C and maintained at that temperature with vital mechanical agitation to give optically a clear and colorless transparent solution. By adding various amounts of acetic anhydride and pyridine (as a catalyst) to the solution and stirring it at 20°C for 4–6 h, CA samples were prepared. Table I summarizes the detailed preparing conditions, such as the amounts of acetic anhydride and pyridine added to the system and the reaction time. The resultant CA was completely precipitated with methanol, washed with methanol, dried *in vacuo*, dissolved in DMSO at 20°C, reprecipitated, rewashed with methanol and dried *in*

Table I. Acetylation condition, $\langle F \rangle$, and $\langle f_k \rangle$ of CAs (one-step)

Sample code	Ac ₂ O ^a mol	Pyr. ^a mol	react. time h	\bar{M}_v 10 ⁵	$\langle F \rangle^b$	$\langle f_2 \rangle$ + $\langle f_3 \rangle^c$	$\langle f_6 \rangle^c$	$\langle f_6 \rangle$ $\langle F \rangle^c$	20°C water soluble part %	Solubility ^d			
										20°C water	20°C DMAC	80°C DMAC	80°C DMSO
I-1	6.0	2.0	4.0	2.6 ^e	0.38	0.13	0.27	0.71	—	× ^g	×	×	○ ⁱ
I-2	8.0	2.0	4.0	2.6 ^e	0.57	0.20	0.37	0.65	3.3	×	×	×	○
I-3	10.0	2.0	5.0	2.6 ^e	0.68	0.27	0.41	0.60	—	×	△	△	○
I-4	12.0	3.0	6.0	2.2 ^f	0.93	0.36	0.57	0.61	2.9	×	△	○	○
I-5	15.0	3.0	6.0	2.8 ^f	1.24	0.54	0.70	0.56	2.1	△ ^h	△	○	○

^a mol mol⁻¹ glucose unit. ^b From titration. ^c From NMR. ^d Judged visually. ^e From $[\eta]$ in DMSO. ^f From $[\eta]$ in DMAC. ^g Insoluble. ^h Highly swelling. ⁱ Soluble.

Solubility of Cellulose Acetates

Table II. \bar{M}_w , $\langle F \rangle$, and $\langle f_k \rangle$ of CAs (two-step)

Sample code	$\bar{M}_w(\bar{M}_v)$ 10 ⁵	$\langle F \rangle^a$	$\langle f_2 \rangle + \langle f_3 \rangle^b$	$\langle f_6 \rangle^b$	$\langle f_6 \rangle / \langle F \rangle^b$	20°C water soluble part %	Solubility ^c		
							20°C water	20°C DMAC	80°C DMSO
CTA-0	2.32	2.92	1.89	1.03	0.35	0	× ^e	○ ^f	○
II-1	1.20	2.46	1.59	0.86	0.35	0	×	○	○
II-2	0.46 ^d	1.75	1.14	0.61	0.35	0	×	○	○
II-3	0.41 ^d	0.80	0.53	0.27	0.34	100	○	○	○
II-4	0.80	0.68	0.45	0.23	0.34	100	○	○	○

^a From titration. ^b From NMR. ^c Judged visually. ^d \bar{M}_v from $[\eta]$ in DMAC. ^e Insoluble. ^f Soluble.

vacuo again. The five samples prepared in this way were subjected to further NMR measurements. In the fifth column of the table, the viscosity-average molecular weight \bar{M}_v of the samples, estimated from the hypothetical MHS equation of CA/DMAC or CA/DMSO system at 25°C, is compiled. The k and a values in the MHS equations for estimating \bar{M}_v for CA having given $\langle F \rangle$ were derived from relationships between $\langle F \rangle$ and k , a values in the MHS equations for various CA established by Kamide *et al.*^{7,10-12} $\langle F \rangle$, determined by the neutralization titration method, of these CA samples is also given in the sixth column of the table.

Two-Step Method. An unfractionated cellulose triacetate sample with $\langle F \rangle$ of 2.92 was deacetylated in acidic media in the same manner, as described in the previous papers^{7,10-12} to obtain four CA samples with different $\langle F \rangle$. Table II summarizes the data of the weight-average molecular weight \bar{M}_w by the light scattering method and $\langle F \rangle$ by the titration method of four CA samples prepared by the two-step method. In the table, the data on the original CTA sample are also included.

¹³C Nuclear Magnetic Resonance Spectra

Proton-noise-decoupled ¹³C NMR spectra were recorded for all ten CA samples in DMSO-*d*₆ at the polymer concentration of 7 wt% at 50.15 MHz on a JEOL FX-200 pulse-Fourier-transform NMR spectrometer at

100°C. Here, the detailed operating conditions are the same as used before.⁶ Slight decrease in solution viscosity was observed during NMR measurements of 14 h, implying that CA degraded to some extent in DMSO at 100°C.

Solubility Determination

To 0.25 g of CA sample 5 ml of distilled water was added at room temperature. The mixture was stirred using a home mixer, subjected to centrifuge and the insoluble part was dried *in vacuo* and weighed. The weight percentage of the water-soluble part of CA is indicated in the tenth column of Table I and the seventh column of Table II. The water-solubility was also judged in a qualitative manner visually. As was also the solubility of CA against DMAC and DMSO. The results are compiled in the last columns of Tables I and II.

RESULTS AND DISCUSSION

Figure 1 shows the ¹³C{¹H} NMR spectra in *O*-acetyl carbonyl carbon region of CTA ($\langle F \rangle = 2.92$) and CA (one-step method, sample code I-1—I-5). For CTA, three large and sharp peaks, attributable to the substituent groups at C₆, C₃, and C₂ positions in trisubstituted (*i.e.*, fully acetylated) glucopyranose unit, were observed, as noted before, at 169.8, 169.0, and 168.6 ppm, respectively. For CA (one-step method, $\langle F \rangle = 0.38$), three

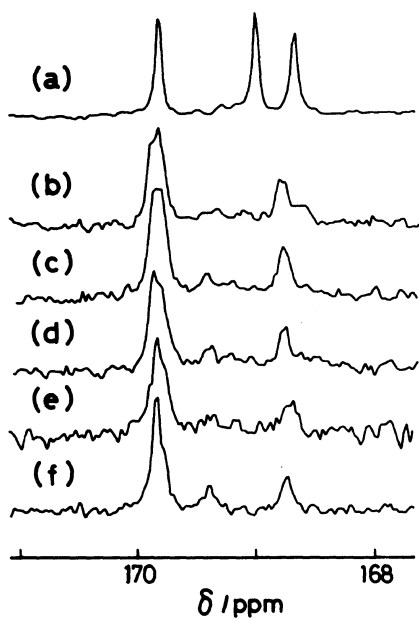


Figure 1. $^{13}\text{C}\{^1\text{H}\}$ NMR spectra in *O*-acetyl carbonyl carbon region of CTA and CA (one-step method) in $\text{DMSO}-d_6$. (a), CTA-0 ($\langle F \rangle = 2.92$); (b), I-5 (1.24); (c), I-4 (0.93); (d), I-3 (0.68); (e), I-2 (0.57); (f), I-1 (0.38).

peaks were observed at 169.9, 169.4, and 168.8 ppm and they can be assigned for the *O*-acetyl carbonyl carbon of C_6 -mono-, C_3 -mono-, and C_2 -mono-substituted glucopyranose units, respectively. It is obvious from Figure 1 that the difference in chemical shift of *O*-acetyl carbonyl carbon at C_6 or C_2 position is only of the order of 0.1–0.2 ppm between tri-substituted and mono-substituted glucopyranose units (that is, the *O*-acetyl carbonyl carbon of tri-substituted glucopyranose units is located at higher magnetic field than that of mono-substituted ones). But, the situation is somewhat different for the carbonyl carbon at C_3 position. There is a large (0.4 ppm) change between tri- and mono-substituted glucopyranose units.

CA samples with $\langle F \rangle = 0.57$ –1.24 can have di-substituted glucopyranose units. *O*-acetyl carbonyl carbon peaks on C_6 position for C_6 , C_2 -di- and C_6 , C_3 -di-substituted ones may appear around 169.8 ppm (C_6 in tri-

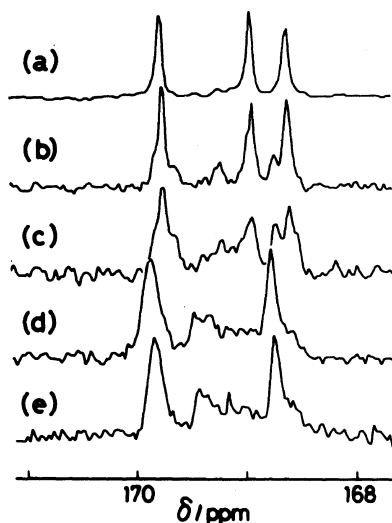


Figure 2. $^{13}\text{C}\{^1\text{H}\}$ NMR spectra in *O*-acetyl carbonyl carbon region of CTA and CA (two-step method) in $\text{DMSO}-d_6$. (a), CTA-0 ($\langle F \rangle = 2.92$); (b), II-1 (2.46); (c), II-4 (1.75); (d), II-3 (0.80); (e), II-4 (0.68).

substituted one) because additional substitution to C_2 or C_3 may not influence greatly the *O*-acetyl carbonyl carbon at C_6 magnetically. Complicated peaks between 169.0 and 169.7 ppm might originate from *O*-acetyl carbonyl carbons at C_3 and possibly at C_2 for C_3 , C_6 -di- and C_2 , C_3 -di-substituted ones.

Figure 2 shows the $^{13}\text{C}\{^1\text{H}\}$ NMR spectra of *O*-acetyl carbonyl carbon region of CTA ($\langle F \rangle = 2.92$) and CA (two-step method, sample code II-1–II-4). The spectra conspicuously changed with decreasing $\langle F \rangle$. When $\langle F \rangle$ decreased from 2.92 to 2.46, peaks appeared at 169.7, 169.2, and 168.8 ppm. These envelopes may originate from *O*-acetyl carbonyl carbons on C_6 , C_3 , and C_2 for di-substituted glucopyranose units, respectively. For CA with $\langle F \rangle = 0.68$, which may be mainly constituted of mono-substituted ones, *O*-acetyl carbonyl carbon peaks of C_6 -mono-, C_3 -mono-, C_2 -mono-substituted at 169.9, 169.4, and 168.8 ppm were again clearly observed. It should be noted again here that the spectra patterns between 169.0 and 169.7 ppm are very complicated especially for CA with $\langle F \rangle = 0.68$

and 0.80. Assuming that the peak positions of the carbonyl carbon peaks due to di-substituted glucopyranose units are not far from the corresponding peaks of mono- and tri-substituted glucopyranose units, we can divide roughly the whole spectra in a carbonyl carbon region into three regions: the region between 170—169.6 ppm for C_6 , 169.6—168.9 ppm for C_3 , and 168.9—168.3 ppm for C_2 . It should be remembered that this technique might give a convenient basis for estimating $\langle\langle f_k \rangle\rangle$ from ^{13}C NMR spectra, but it is not so rigorous. For example, two peaks groups assigned for C_3 and C_2 positions have an in-negligible possibility of mutual overlapping, which can be realized, for example, by noting that at least more than 6 peaks are observed experimentally in the above defined carbonyl carbon region at the so-called C_3 position, although the maximum number of corresponding peaks, theoretically expected, is only four (one for mono-, for tri-, and two for di-substituted glucopyranose units). Here, the theoretical number of peaks is taken by assuming that neighbouring glucopyranose unit does not affect *O*-acetyl carbonyl carbon on C_3 in question. In contrast to this, the carbonyl carbon peak group at C_6 position is absolutely separated from those at other positions. Then, $\langle\langle f_6 \rangle\rangle$ and $(\langle\langle f_2 \rangle\rangle + \langle\langle f_3 \rangle\rangle) (= \langle\langle F \rangle\rangle - \langle\langle f_6 \rangle\rangle)$ can be accurately estimated, regardless of its $\langle\langle F \rangle\rangle$, from ^{13}C NMR carbonyl carbon region spectra of CA with help of $\langle\langle F \rangle\rangle$ (by the titration method).

Comparison of Figures 1 and 2 indicates that the probability of substituting the hydroxyl group at C_6 position with acetyl group is higher in one-step method for CA with same $\langle\langle F \rangle\rangle$. Below $\langle\langle F \rangle\rangle = 1.00$, the probability of substitution at C_3 or fraction of C_3 -mono-substituted one for one-step CA is far lower than that of two-step CA.

In Tables I and II, $\langle\langle f_6 \rangle\rangle$ and $(\langle\langle f_2 \rangle\rangle + \langle\langle f_3 \rangle\rangle)$ data for all CA samples thus estimated are collected.

Figure 3 shows the plot of $\langle\langle f_6 \rangle\rangle$ against $\langle\langle F \rangle\rangle$

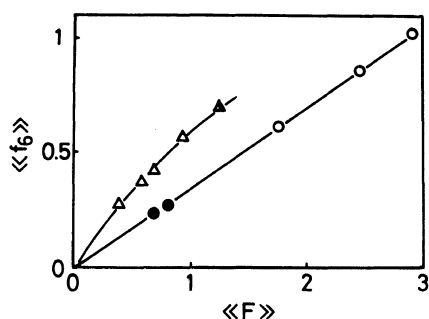


Figure 3. Plot of $\langle\langle f_6 \rangle\rangle$ of CA against $\langle\langle F \rangle\rangle$. (Δ , Δ), one-step method; (\circ , \bullet), two-step method; unfilled marks, water-insoluble; half-filled marks, highly swelling in water; filled marks, water-soluble.

for two series of CA samples (one-step and two-step methods). In the figure, the unfilled mark denotes water-insoluble and the half-filled mark, partially soluble and highly-swelling in water and the filled mark, completely water-soluble. From Tables I and II together with Figure 3, the following relations hold: $\langle\langle f_6 \rangle\rangle / \langle\langle F \rangle\rangle = 0.65 \pm 0.06$ for CA (one-step method) and $\langle\langle f_6 \rangle\rangle \approx (\langle\langle f_2 \rangle\rangle + \langle\langle f_3 \rangle\rangle) / 2$ for CA (two-step method).

In the one-step method, the hydroxyl group at C_6 position is 3—4 times reactive against acetic anhydride than that at C_2 or C_3 position. Whereas, in the two-step method, the acetyl groups at three different carbon positions are almost equally hydrolyzed when dissolved in acidic media. In other words, the reactivity of acetylation depends strongly on the carbon positions, to which the hydroxyl group is located, but that of deacetylation does not significantly depend on whether the acetyl group is derived from the primary hydroxyl or secondary group.

All CA samples in this paper are soluble in DMSO at 80°C and CA samples prepared in two-step method are also soluble in DMAC at 20°C. For many years, we have regarded DMAC as a common solvent for CA having various $\langle\langle F \rangle\rangle$ and in fact the measurements of the solution properties of CA have been carried out in the DMAC system. But, CA sam-

ples prepared in the one-step method were found not to dissolve in DMAC at 20°C. This is the first demonstration of the solubility study on CA. It is also evident from the tables that the solubility of CA samples against water and DMAC is different from sample to sample. Our previous study⁷ revealed that \bar{M}_w dependence of solubility of CA prepared in the two-step method was not found up to $\bar{M}_w = 1.45 \times 10^5$, showing that \bar{M}_w is not essential for the solubility of CA.

A comparison of sample codes I-1 and II-3, both having the same $\langle\langle f_6 \rangle\rangle (=0.27)$ and different $\langle\langle F \rangle\rangle$, shows that even if $\langle\langle f_6 \rangle\rangle$ is the same the solubility against water and DMAC is better for the polymer having larger $(\langle\langle f_2 \rangle\rangle + \langle\langle f_3 \rangle\rangle)$. A comparison of sample codes I-3 and II-4 leads us to the conclusion that, CA with larger $(\langle\langle f_2 \rangle\rangle + \langle\langle f_3 \rangle\rangle)$ exhibits a better solubility if compared at the same $\langle\langle F \rangle\rangle$. Therefore, it seems doubtlessly reasonable to consider that the solubility of CA is predominantly governed by $(\langle\langle f_2 \rangle\rangle + \langle\langle f_3 \rangle\rangle)$. However, CA samples (I-5, II-3) having almost same $(\langle\langle f_2 \rangle\rangle + \langle\langle f_3 \rangle\rangle) (\approx 0.54)$ have different solubility against water and DMAC. As mentioned before, CA sample codes II-3 which are completely water-soluble have higher probability of substitution at C₃ or higher fraction of C₃-mono-substituted glucopyranose units than CA sample codes I-5. Therefore, the minimum necessary conditions in which the CA sample dissolves in water and polar solvent such as DMAC seems to be that the degree of substitution at C₃ should be higher than a given threshold value. The threshold value cannot be exactly estimated but from Figures 1 and 2 it may be around 0.2 for water. That is, substitution at C₃ is one/five glucopyranose units. The introduction of an hydrophobic group at C₃ may break down the intramolecular hydrogen bond between O₃ ··· O₅. For two series of the CA sample, the solubility power of DMAC is somewhat larger than water. This may be due to the difference in solvation power to *O*-acetyl and

hydroxyl groups of the solvents. These experimental results agree well with those by Miyamoto *et al.*⁸ who demonstrated that CA having higher $\langle\langle f_6 \rangle\rangle$ than $\langle\langle f_2 \rangle\rangle$ and $\langle\langle f_3 \rangle\rangle$, synthesized by one-step method (*i.e.*, A series samples in their paper), does not dissolve in water at room temperature and they proposed that $\langle\langle f_2 \rangle\rangle$ and $\langle\langle f_3 \rangle\rangle$ play key roles in the water-solubility of CA. But, they did not give any explanation for their results. Note that $(\langle\langle f_2 \rangle\rangle + \langle\langle f_3 \rangle\rangle)$ or $\langle\langle f_3 \rangle\rangle$ or fraction of C₃-mono-substituted glucopyranose units is, of course, not a single parameter controlling water solubility.

Concerning the reactivity of cellulose towards acetic anhydride in DMAC/LiCl medium, Miyamoto *et al.*⁸ considered that the strength of hydrogen bonding by C₂ and C₃ hydroxyl groups plays an important role, as compared with that of C₆ hydroxyl group. They base their discussion on Gagnaire *et al.*'s claim¹³ that the intramolecular hydrogen bonding between the hydroxyl group at C₃ position and a heterocyclic oxygen atom in neighbouring glucopyranose unit of cellulose may exist even in the solution. Note that Gagnaire *et al.*'s assertion was formed only indirectly by observing the constancy of ¹H NMR spectra peak position of hydroxyl proton at C₃ position (4.6 ppm) of cellobiose (not cellulose!) dissolved in dimethylacetamide (DMAC)/LiCl system. It is well-recognized that ¹³C NMR information on polysaccharides depends significantly on the degree of polymerization (DP) in an extremely low DP region. Kamide and Saito¹⁴ showed experimentally, that the unperturbed chain dimension of cellulose derivatives, including CA, increases with the polarity of the solvent (dielectric constant ϵ) and both the substituent groups and remaining hydroxyl group of cellulose derivatives solvate with the solvent and the degree of solvation is closely correlated with ϵ . These facts explicitly demonstrate the impossibility of the existence of the intramolecular hydrogen bonding of CA in the

solvents. Accordingly, the selective acetylation at C₆ position for the cellulose in DMAC/LiCl system does not indicate difference in strength of intramolecular hydrogen bondings between the hydroxyl groups at C₂, C₃, and C₆ positions and a heterocyclic oxygen atom. The experimental data, summarized in Table II, suggest that a primary hydroxyl group at C₆ position is highly reactive to an acetylation reagent than secondary hydroxyl groups at C₂ and C₃ positions as in the case of low molecular weight organic compounds.¹⁵

Kamide *et al.*³ demonstrated that, as long as NaCMC does not dissolve in the liquids, the absorbency can be accurately determined by $\langle\langle f_6 \rangle\rangle$ alone. They explained this fact as follows: the introduction of a bulky substituent (carboxymethyl group) at the C₆ position destroys the intermolecular hydrogen bonds between the hydroxyl group at C₆ position and a heterocyclic oxygen atom in a neighbouring glucopyranose unit and also widens the distance between molecular chains, resulting in destruction of the intermolecular hydrogen bonds, creating a wide space to readily receive the absorbed liquids. It should be noted that the results by Kamide *et al.*³ for NaCMC do not mean that the simple introduction of the bulky substituent group into C₆ position is an important step to make cellulose soluble in water or aqueous solutions. Carboxymethyl group has the same molecular volume as an acetyl group and then, the introduction of these groups into C₆ positions is expected to bring about the destruction of intermolecular hydrogen bonding to the same degree. If the destruction of hydrogen bonding associated with C₆ position is a single universal factor controlling the water-solubility, CA with high $\langle\langle f_6 \rangle\rangle$ is should be water soluble. Unfortunately, this is not true. It is interesting to note that for NaCMC, the carboxymethyl group is more hydrophilic than the hydroxyl group and in CA the acetyl group is less hydrophilic than the hydroxyl group. Then, the affinity of the substituent group against

water seems another important factor.

Of course, an attempt to directly correlate the chemical structure, as characterized in terms of $\langle\langle f_2 \rangle\rangle$, $\langle\langle f_3 \rangle\rangle$, and $\langle\langle f_6 \rangle\rangle$, of cellulose derivatives with their solubility is too simple and rough. The solubility should be mainly determined by the supermolecular structure of the solid (*i.e.*, the crystalline and amorphous phases, intra- and intermolecular hydrogen bonds) and the latter is vitally controlled by the chemical structure, provided that the heterogeneity of the chemical structure in glucopyranose unit can be ignored and the solid is formed by a similar procedure. Recently, Kamide *et al.*¹⁶ pointed out that the solubility of cellulose in a 10 wt% aqueous NaOH at 4°C could generally be correlated to the relative amount of the high magnetic field envelope of C₄ carbon NMR peak (*i.e.*, that of the region where intramolecular hydrogen bonds are at least partly broken). At the present time, the supermolecular structures of CA samples are not fully understood and remain open for further study.

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