

Probable circular permutation in the flavin-binding domain

Sir—Liepinsh *et al.*¹ have recently proposed an evolutionary link between electron transport and proteolysis based upon a structural relationship of the FMN binding protein (FMN-bp) from *Desulfovibrio vulgaris* (Miyazaki F) to chymotrypsin and related proteases. Having examined the sequence and structure of FMN-bp, I found no evidence that FMN-bp may be descended from a so far hypothetical single-domain precursor of the protease superfamily.

Firstly, unlike the serine proteases which have evolved into a vast and widespread sequence family, FMN-bp is an orphan and therefore is not likely to be of such an ancient origin. Secondly, the FMN-bp fold belongs to a different class of the β -sheet barrels than the fold of chymotrypsin domains. The β -sheet barrels are classified by two integer numbers: n , the number of strands, and S , the shear number^{2,3}. In the FMN-bp barrel, $n = 6$ and $S = 10$, whereas, in both chymotrypsin barrels, $n = 6$ and $S = 8$ (defined as in ref. 3, the shear number

is always even due to the periodicity of β -structure). The distinction in shear number is not always distinguished by present automated methods for structural similarity searches. It has only a minor effect on the number of spatially superimposable main chain atoms and their root-mean-square (r.m.s.) deviation; however it results in significant differences in the packing of side chains in the barrel interior and the overall barrel geometry³.

I argue here instead that FMN-bp probably has a recent origin and evolved from the ferredoxin reductase⁴ superfamily of flavoenzymes. Inspection of β -barrel folds in the SCOP database⁵, has readily identified this superfamily as a putative parent family for FMN-bp. Known structures of this superfamily also include phthalate dioxygenase reductase, nitrate reductase, cytochrome b_5 reductase and flavodoxin reductase⁶, cytochrome P450 reductase⁷ and flavohemoglobin⁸. The FAD-binding domain of reductases adopts a β -barrel fold of the same class as the FMN-bp barrel ($n = 6$, $S = 10$). The topologies of the FAD-binding domain and FMN-bp are different, but they can be related by a circular permutation (Fig. 1). (To convert the topology of FAD-binding domain into the FMN-bp topology, the end of strand 6 to the beginning of strand 1 would have to be connected with a peptide linker, while the connection between strands 2 and 3 would have to be cut to create the new termini). This permutation also will bring the flavin-binding sites of FMN-bp and FAD-binding domain into the same topological location. The binding modes for

FMN and the equivalent part of FAD are very similar (Fig. 1): the cofactor molecule bridges the gap between two strands (strands 4 and 5 in the FAD-binding domain and their counterparts strands 2a and 3 in FMN-bp). The isoalloxazine ring and ribityl chain hydrogen bond to free main chain groups in these strands and make non-polar contacts to side chains in the barrel interior. The phosphate group binds at the N terminus of α -helix that caps each barrel.

Structural evidence accumulated over the past years suggests that circularly permuted proteins can and do occur naturally⁹. It is further supported here by the structural and functional similarity between FMN-bp and the FAD-binding domain that are very likely to share a common ancestor. The FMN-bp fold probably has been created by a recent circular permutation event in one of the FAD-binding domains, therefore its superficial structural similarity to the serine protease fold is convergent rather than divergent.

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- Liepinsh, E., Kitamura, M., Murakami, T., Nakaya, T. & Otting, G. *Nature Struct. Biol.* **4**, 975–979 (1997).
- McLachlan, A.D. *J. Mol. Biol.* **128**, 49–79, 1979.
- Murzin, A.G., Lesk, A.M. & Chothia, C. *J. Mol. Biol.* **236**, 1369–1400, 1994.
- Karplus, P.A., Daniels, M.J. & Herriott, J.R. *Science* **251**, 60–66, 1991.
- Murzin, A.G., Brenner, S.E., Hubbard, T. & Chothia, C. *J. Mol. Biol.* **247**, 536–540, 1995.
- Ingelman, M., Bianchi, V. & Eklund, H. *J. Mol. Biol.* **268**, 147–157, 1997.
- Wang, M. *et al. Proc. Natl. Acad. Sci. U.S.A.* **94**, 8411–8416, 1997.
- Ermler, U., Siddiqui, R.A., Cramm, R. & Friedrich, B. *EMBO J.* **14**, 6067–6077, 1995.
- Lindqvist, Y. & Schneider, G. *Curr. Opin. Struct. Biol.* **7**, 422–427, 1997.

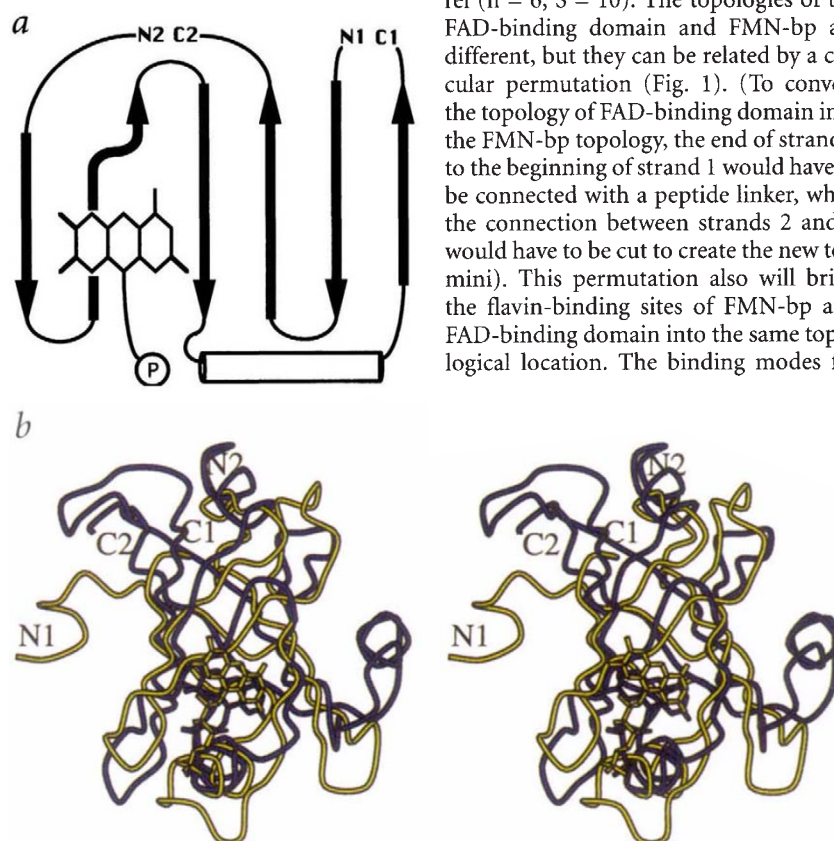


Fig. 1 a, Topology diagram of the flavin-binding β -barrel fold. Arrows represent β -strands and cylinder represents the capping α -helix. In the FAD-binding domain, the protein chain begins at N1, continues through the gap between C2 and N2 and ends at C1, whereas, in the FMN-binding protein, it begins at N2 and ends at C2. The position of the flavin-binding site is indicated by simplified picture of FMN. **b**, Stereo view showing a superposition of the backbones of the FAD-binding domain from phthalate dioxygenase reductase (yellow; PDB entry 2PIA, residues 1–100) and FMN-bp (blue; 1AXJ, coordinates courtesy of G. Otting). In this superposition, 65 pairs of $C\alpha$ atoms in all of the six barrel strands and the capping helix give an r.m.s. deviation of 2.3 Å. The FMN molecules bound to each structure shown in stick representation.