

synthesis of new proteins was markedly reduced in the axon terminals of *Xenopus laevis* neurons expressing full-length FUSALS-FTD

Some forms of amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD) are characterized by intraneuronal assemblies of mutant variants of RNA-binding proteins, such as FUS. However, whether and how these assemblies might contribute to neurotoxicity in ALS and FTD is unknown. Now, Murakami *et al.* show that pathological *FUS* mutations promote the transition of liquid droplets of FUS into a gel that can trap ribonucleoproteins (RNPs), thereby disrupting RNP granule function and protein synthesis.

The authors generated *Caenorhabditis elegans* that expressed either wild-type human FUS (FUS^{WT}) or one of five variants of mutant human FUS that have been associated with ALS or FTD (FUS^{ALS-FTD}) under a neuron-specific promoter. In contrast to FUS^{WT}-expressing worms, which showed no neurotoxicity, FUS^{ALS-FTD}-expressing worms exhibited age-dependent motor deficits and reduced lifespans, and the severity of this phenotype correlated with the abundance of intraneuronal FUS^{ALS-FTD} assemblies.

Previous work had indicated that the amino-terminal 'low-complexity' (LC) domain of FUS enables the protein to aggregate. Here, worms in which truncated FUSALS-FTD variants that lacked the LC domain were specifically expressed in neurons had fewer FUS assemblies and showed less-severe motor impairment and reductions in lifespan than did worms expressing full-length

FUS^{ALS-FTD} variants. Moreover, neuronal expression of the FUS^{ALS-FTD} LC domains led to more-severe neurotoxicity than did expression of the FUS^{WT} LC domain. Therefore, the LC domains of pathological FUS mediate neurotoxicity.

Next, the authors characterized the biophysical nature of the FUSALS-FTD assemblies. Live imaging of cultured mammalian neurons expressing vellow-fluorescent protein-tagged variants of FUS revealed that FUSWT resides as liquid droplets in the nucleus, whereas FUSALS-FTD droplets are present not only in the nucleus but also in the cytoplasm. The authors were able to form liquid droplets of FUS in vitro by cooling solutions of recombinant FUS variants to 4°C. On warming back to 23 °C, the droplets of FUSWT almost all dissolved, whereas more than half of the droplets of $FUS^{\mbox{\tiny ALS-FTD}}$ remained. When droplets of FUS LC domains were cooled, they formed a gel that could be 'melted' by warming. However, whereas FUSWT LC domains could undergo four or five of these liquid-gel cycles before irreversibly gelling, droplets of FUSALS-FTD LC domains formed an irreversible gel that was more viscous than the FUSWT LC domain gel after only two cycles. Thus, FUSALS-FTD variants have a higher propensity than FUSWT for phase transitioning from a monomeric, dissolved form to an irreversible fibrous gel form.

The authors investigated the functional effects of FUS^{ALS-FTD} gel formation on RNP granules.

They cooled mixtures of FUSALS-FTD or FUSWT and the RNPs survival motor neuron (SMN) and STAU1, and used single-molecule imaging to track the diffusion of SMN and STAU1 particles. On melting the gels, SMN and STAU1 in the FUSWT mixture could diffuse freely. By contrast, the FUSALS-FTD mixture contained irreversible gel assemblies that prevented SMN and STAU1 from moving, suggesting that the FUSALS-FTD gel may 'trap' RNPs. This effect seems to disrupt RNA translation, as the synthesis of new proteins was markedly reduced in the axon terminals of *Xenopus laevis* neurons expressing full-length FUSALS-FTD compared with neurons expressing FUSWT.

This study provides evidence that, whereas reversible liquid droplets and gels formed by FUS^{WT} support physiological RNP granule function, pathological variants of FUS can form irreversible gel assemblies that may trap RNPs, and thus could impair RNP granule function and disrupt protein synthesis. This represents a possible mechanism for neurodegeneration in the forms of ALS and FTD that are associated with mutant RNA-binding proteins.

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