

BACTERIAL PATHOGENESIS

The great escape

Although the initial phases of intestinal infection with *Vibrio cholerae* have been intensively scrutinized, the later stages of infection, which are characterized by mass detachment of bacteria from epithelial surfaces and movement into the intestinal lumen, have been largely overlooked. Alex Nielsen and Nadia Dolganov, working in the Schoolnik laboratory, now redress this balance by providing insights into this luminal migration, which takes place about 12 hours after infection and which they call the 'mucosal escape response'. They reveal that this phenomenon is accompanied by the induction of a specific genetic programme in *V. cholerae* that activates a range of motility and chemotaxis genes.

The authors used a combination of confocal microscopy, expression profiling and mutant analysis to study the mucosal escape response in a rabbit model of *V. cholerae* infection,

the ligated ileal-loop model. They first showed that 12 hours after inoculation in this model, at the time of bacterial detachment, *V. cholerae* replication slows to a rate similar to that of organisms entering stationary phase in culture. The stationary phase alternative sigma factor RpoS is required for the mucosal escape response, evidenced by the fact that an *rpoS* mutant strain did not detach from ileal-loop epithelial cells. HapR, an RpoS-controlled factor that downregulates virulence determinant expression and biofilm formation, is also required for this stage of cholera infection.

Expression profiling of stationary phase cultures and bacteria collected from the luminal fluid at the time of the bacterial detachment revealed the upregulation of genes involved in chemotaxis and bacterial motility, including those involved in flagella biosynthesis. Importantly,



81% of these upregulated genes were dependent on RpoS for their induction during stationary phase. The assembly of flagella and the activation of chemotactic function during the mucosal escape response would facilitate efficient movement of bacteria into the intestinal lumen in preparation for exit from the host.

BACTERIAL PHYSIOLOGY

Right on target



Cyclic-di-GMP has come to prominence recently as an important bacterial second messenger. The GGDEF/EAL proteins responsible for the synthesis and degradation of this small molecule are believed to function as global regulators in biofilm formation and motility. However, a recent paper from Regine Hengge and colleagues now reveals that some GGDEF/EAL proteins act on a single target.

Cyclic-di-GMP is synthesized by diguanylate cyclases (DGCs) containing a GGDEF amino-acid motif and degraded by phosphodiesterases (PDEs) containing an EAL motif. Bacterial genomes encode many GGDEF/EAL proteins, with the *Escherichia coli* genome encoding a total of 29. Earlier work in Hengge's laboratory had indicated that GGDEF/EAL-protein-encoding genes are over-represented in the general stress-response network controlled by the sigma factor σ^s .

Hengge and colleagues followed up this work by constructing single-copy *lacZ* fusions in putative

σ^s -controlled GGDEF/EAL genes. Positive control by σ^s was confirmed for five of the genes analysed and, interestingly, it seemed that although these five genes are under the overall control of σ^s , they each have distinct regulatory patterns that can respond to different environmental signals. Deletion mutants were then constructed for each of the five genes. The σ^s signalling network controls certain biofilm functions, and Weber *et al.* were interested in the signalling pathway that results in the formation of curli fimbriae. Of the deleted genes, only two, *ydaM* and *yciR*, were found to be involved in curli formation, and they were shown to have antagonistic effects — *ydaM* positively regulates curli formation whereas *yciR* is a negative regulator. Further biochemical analysis confirmed that YdaM has DGC activity and YciR has PDE activity.

Weber *et al.* then took a closer look at these two proteins to try and tease out their precise role in curli formation. The production of these adhesive protrusions is controlled

 HOST RESPONSE

Viral persistence: IL-10 is the key

Schoolnik and colleagues point out that these data conflict with previous studies showing that chemotaxis genes are downregulated in cholera patient rice-water stools, and they discuss the possible reasons for these contrasting results.

The effect of RpoS on virulence gene expression was also addressed in a series of experiments. The research team reasoned that the continued production of virulence factors after bacterial detachment would be ineffectual and so RpoS might downregulate virulence gene expression. In accordance, virulence assays confirmed that RpoS markedly downregulated cholera toxin production *in vitro*.

The authors conclude by proposing a model for the role of RpoS in *V. cholerae* infection, detailing its part in the genetic programme that integrates the mucosal escape response and the control of virulence and chemotaxis.

Shannon Amoils

ORIGINAL RESEARCH PAPER Nielsen, A. T., Dolganov, N. A. *et al.* RpoS controls the *Vibrio cholerae* mucosal escape response. *PLoS Pathog.* 2, e109 (2006)

by a complex feedforward regulatory cascade comprising σ and the regulators MlrA and CsgD. The authors found that the antagonistic control of curli formation by YdaM and YciR occurs at the level of *csgD* transcription.

In addition, rather than being global regulators, the GGDEF/EAL proteins YdaM and YciR function only on *csgD* transcription, as indicated by genome-wide transcription analysis. The authors speculate that this high selectivity for a single target might be the result of the 'functional sequestration' of these two proteins, and that this might contribute to the specificity of cyclic-di-GMP signalling in the presence of multiple GGDEF/EAL proteins.

Sheilagh Molloy

ORIGINAL RESEARCH PAPER Weber, H. *et al.* Cyclic-di-GMP-mediated signalling within the σ network of *Escherichia coli*. *Mol. Microbiol.* 62, 1014–1034 (2006)

FURTHER READING Romling, U., Gomelsky, M. & Galperin, M. Y. C-di-GMP: the dawning of a novel bacterial signalling system. *Mol. Microbiol.* 57, 629–639 (2005)

Persistent viral infections are responsible for some of the most devastating human diseases, including AIDS and hepatitis. Now, two research groups have concluded that elevated interleukin-10 (IL-10) production causes the immunosuppression that allows viruses to persist unchecked by the immune system. Moreover, by administering a therapeutic antibody to block IL-10 action, the ability of the immune system to clear persistent viruses can be restored.

Intense research has been directed towards understanding the mechanisms that underlie virus persistence. Cell-mediated immunity is essential for virus clearance, but persistent viruses can avoid this antiviral response by inducing immunosuppression. Strategies for inducing immunosuppression include physical deletion of virus-specific T-cell subsets, a useful tactic that is used by HIV-1, hepatitis B and C viruses and lymphocytic choriomeningitis virus (LCMV). Concomitant with the deletion of T-cell subsets is the loss of T-cell cytolytic functions, which results in an inability to kill virus-infected cells. The latest research from groups led by Michael Oldstone and Matthias von Herrath has established a clear link between cytokine production and T-cell function in persistent LCMV infection.

One extremely useful model system for probing the subtle interplay between virus replication and the host response during persistence is the infection of rodents with the arenavirus LCMV. Infection with LCMV can produce either an acute or a persistent infection depending on the virus strain, route of infection and the infectious dose. Both of the studies just published investigated the basis of persistence using this model.

Initial studies by Brooks *et al.* and Ejrnaes *et al.* revealed elevated concentrations of IL-10 in mice infected with a persistent LCMV strain compared with mice infected with an acute LCMV strain. When knockout mice that fail to produce IL-10 were infected with a persistent LCMV strain they had increased cytotoxic T-cell activity compared with wild-type mice. In addition, the IL-10 knockout mouse model that was infected with a persistent LCMV strain produced memory T-cells that enabled resistance to reinfection with LCMV.

By using an antibody to block signalling by IL-10 through the IL-10 receptor, a persistent LCMV infection was completely resolved.

Both groups showed that treatment

of persistently infected mice with the anti-IL-10 antibody led to the resolution of a persistent LCMV infection, even when the antibody was administered during established infection. This showed the pivotal importance of the cytokine IL-10 in the maintenance of persistent viral infections. Researchers will now be hoping to build on these findings to specifically define just how IL-10 modulates T-cell functions.

This research might herald the use of new therapies based on blocking IL-10 functions for the treatment of HIV-1 and hepatitis B and C virus infections. But the authors caution that the immunosuppressive role of IL-10 should be confirmed for each persistent virus before launching new therapeutic strategies, and that care must be taken in the design of therapeutic regimes, because modulating cytokine function could lead to immunopathology.

Susan Jones

ORIGINAL RESEARCH PAPERS Ejrnaes, M. *et al.* Resolution of a chronic viral infection after interleukin-10 receptor blockade. *J. Exp. Med.* 09 October 2006 (doi:10.1085/jem.20061462-12) | Brooks, D. G. *et al.* Interleukin-10 determines viral clearance or persistence *in vivo*. *Nature Med.* 15 October 2006 (doi:10.1038/nm1492)

