



“ ciliary and nuclear import pathways use similar mechanisms ”

The construction and maintenance of cilia requires intraflagellar transport along axonemal microtubules by the heterotrimeric kinesin-2 family complex KIF3A–KIF3B–KAP and the homodimeric *KIF17* motor. But how kinesin motors and their cargoes enter cilia is unknown. Dishinger *et al.* now uncover intriguing parallels between nuclear and ciliary import mechanisms by showing that ciliary entry of *KIF17* is regulated by the nuclear import protein importin β 2 and a ciliary–cytoplasmic gradient of the small GTPase *RAN*.

KIF17 was shown to accumulate in cilia in all mammalian cell lines tested, but deletion of the carboxy-terminal tail domain abolished ciliary localization of *KIF17*, suggesting that a ciliary localization signal (CLS) is present in this region. Surprisingly, constructs containing the tail region localized predominantly to the nucleus, which prompted the authors to look for a nuclear localization signal (NLS) in the *KIF17* tail. They identified two putative NLSs, one of which could also act as a CLS and is necessary and sufficient for ciliary targeting of *KIF17*.

Given that nuclear import requires recognition of NLSs by importin proteins, translocation through the nuclear pore complex and dissociation of the NLS–importin complex in the nucleus by GTP-bound *RAN*, Dishinger *et al.* verified the presence of *RAN*–GTP in cilia. To test whether ciliary *RAN*–GTP regulates *KIF17* import, they used a system for fast induction of *RAN* expression in the cytoplasm. Cytoplasmic expression of wild-type *RAN* or a *RAN* mutant

that was unable to bind GTP did not affect the ciliary localization of *KIF17*, whereas expression of a constitutively active *RAN*–GTP mutant reduced the number of cells containing ciliary *KIF17*. The authors confirmed these findings by fluorescence recovery after photobleaching (FRAP) analysis of ciliary *KIF17* in the presence or absence of the constitutively active *RAN*–GTP mutant. The FRAP data showed a drastic reduction in ciliary *KIF17* recovery when levels of this *RAN* mutant were increased. Together, these findings suggest that cytoplasmic *RAN*–GTP abolishes ciliary entry of *KIF17*.

So, do importin proteins feature in the ciliary entry of *KIF17*? Indeed, importin β 2 is present in the proximal region of the cilium and co-immunoprecipitates with *KIF17* but not with a *KIF17* mutant containing a defective CLS. This suggests that the interaction of *KIF17* with importin β 2 is necessary for ciliary import. Interestingly, no interaction between *KIF17* and importin β 1 was observed by immunoprecipitation, suggesting that importin β 2 alone is responsible for ciliary entry of *KIF17*.

This study provides the first direct evidence that ciliary and nuclear import pathways use similar mechanisms, and expands the role of *RAN* in intracellular transport.

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ORIGINAL RESEARCH PAPER Dishinger, J. F. *et al.* Ciliary entry of the kinesin-2 motor *KIF17* is regulated by importin- β 2 and *RanGTP*. *Nature Cell Biol.* 6 Jun 2010 (doi:10.1038/ncb2073)

FURTHER READING Verhey, K. J. & Hammond, J. W. Traffic control: regulation of kinesin motors. *Nature Rev. Mol. Cell Biol.* 10, 765–777 (2009)