

CELL CYCLE

Destruct and arrest

The spindle-assembly checkpoint (SAC) blocks chromosome segregation until all sister chromatids are properly attached to the mitotic spindle. It does so by inactivating *CDC20* (also known as Slp1 and Fizzy) — an essential activator of the ubiquitin-ligase anaphase-promoting complex or cyclosome (*APC/C*), which promotes the degradation of proteins that arrest the cell cycle. Two studies now provide insights into how the SAC inhibits *CDC20* and reveal a new role for *CDC20* in *APC/C*-mediated protein degradation.

Using nocodazole (a microtubule-depolymerizing agent) to activate the SAC in live-imaging studies of human cells, Nilsson *et al.* observed that a fluorescent *CDC20* fusion protein

was degraded in response to SAC. Degradation of *CDC20* started at the time of nuclear envelope breakdown, which coincides with the time when *APC/C* recognizes its early mitotic substrates. This implies that *CDC20* might be a target of *APC/C*. Indeed, the use of small interfering RNAs (siRNAs) against *APC/C* or the SAC protein *MAD2* blocked *CDC20* degradation, which suggests that *CDC20* is degraded by *APC/C* in response to the SAC.

Size-exclusion chromatography and quantitative immunoblotting experiments in nocodazole-treated cells revealed that *CDC20* primarily interacts with the SAC component *BUBR1* and not with *MAD2*, as previously thought.

A mutant version of *CDC20* that could not be ubiquitylated could bind to the SAC protein complex and be released from it when the SAC was chemically inhibited. However, it was not degraded by *APC/C* and was unable to maintain the SAC-induced mitotic arrest. These findings contradict a previous model that suggests that ubiquitylation of *CDC20* is needed to release it from the SAC complex and to activate *APC/C*. By contrast, Nilsson *et al.* show that ubiquitylation of *CDC20* causes its degradation, which is required to maintain the SAC-associated mitotic arrest.

CDC20-family proteins recognize *APC/C* substrates through a carboxy-terminal WD40 repeat domain and recruit them to *APC/C* for degradation. Kimata *et al.* found that *Nek2A*, an *APC/C* substrate that interacts

directly with *APC/C* independently of *Cdc20*, was not degraded in *Xenopus laevis* egg extracts that were depleted of *Cdc20*. This suggests that *Cdc20* has an additional role besides substrate recruitment. By adding mutated versions of *Cdc20* back to the egg extracts, the authors found that the amino-terminal C-box domain of *Cdc20* facilitates *Nek2A* degradation. By contrast, the WD40-repeat domain was required for the degradation of 'canonical' substrates that are recruited to *APC/C* by *Cdc20*.

The ability of purified *APC/C* from *Cdc20*-depleted egg extracts to ubiquitylate *Nek2A in vitro* was restored by adding the C-box domain of *Cdc20* to the reaction. By contrast, the ubiquitylation of canonical substrates required the WD40 repeat domain. However, canonical substrates that were directly fused to the C-box domain of *Cdc20* were efficiently ubiquitylated by *APC/C*, thereby revealing a new role for *Cdc20* in promoting substrate ubiquitylation by *APC/C*.

These studies represent important steps in understanding how the activity of *APC/C* is regulated during the cell cycle.

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ORIGINAL RESEARCH PAPERS Nilsson, J. *et al.* The *APC/C* maintains the spindle assembly checkpoint by targeting *Cdc20* for destruction. *Nature Cell Biol.* **10**, 1411–1420 (2008) | Kimata, Y. *et al.* A role for the Fizzy/*Cdc20* family of proteins in activation of the *APC/C* distinct from substrate recruitment. *Mol. Cell* **32**, 576–583 (2008)
FURTHER READING Peters, J.-M. The anaphase promoting complex/cyclosome: a machine designed to destroy. *Nature Rev. Mol. Cell Biol.* **7**, 644–656 (2006)

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