Cellular sources and immune functions of interleukin-9

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Abstract | Interleukin-9 (IL-9) has attracted renewed interest owing to the identification of its expression by multiple T helper (T_{μ}) cell subsets, including T_{μ}^2 cells, T_{μ}^2 cells, T_{μ}^2 cells, T_{μ}^2 cells, T_{μ}^2 cells and regulatory T (T_{Reg}^2) cells. Here, we provide a broad overview of the conditions that are required for cells to produce IL-9 and describe the cellular targets and nature of the immune responses that are induced by IL-9.

Interleukin-9 (IL-9) was originally identified in mice as a T cell growth factor and is a member of the common y-chain-receptor cytokine family, with other members including IL-2, IL-4, IL-7, IL-15 and IL-21. The IL9 gene loci of both humans and mice have a similar organization, consisting of five exons, and share a 55% amino acid homology at the protein level. The IL-9 receptor consists of the cytokine-specific IL-9 receptor α-chain (IL-9R α) and the γ -chain¹. IL-9-induced receptor activation promotes the crossphosphorylation of Janus kinase 1 (JAK1) and JAK3. This cross-phosphorylation leads to the downstream activation of signal transducer and activator of transcription (STAT) complexes, specifically STAT1 homodimers, STAT5 homodimers and STAT1-STAT3 heterodimers²⁻⁴ (FIG. 1). Although the contribution of IL-9 to the activation of these pathways in primary cells has not been fully elucidated, it may be of relevance in vivo.

Here, we summarize the cellular sources of IL-9, focusing principally on T cells. We also describe the main targets of IL-9 *in vivo* and discuss the downstream effects of IL-9-induced signalling on leukocyte function and overall immunity.

Cellular sources of IL-9

 $T_{\rm H}2$ cells. IL-9 was first linked to T helper 2 (T_H2) cells, which express IL-4, IL-5 and IL-13, following the report that levels of IL-9 expression were high in the T_H2-prone BALB/c mouse strain but low in the T_H1-prone C57BL/6 mouse strain during infection with *Leishmania major*. The same study showed that T cells were the main producers of IL-9 *in vivo* and that the levels of IL-9 expression correlated with the expansion of $T_{\rm H}^2$ cell populations. Futhermore, blockade of IL-4, which is a crucial mediator of $T_{\rm H}^2$ cell differentiation, could suppress IL-9 production⁵. Owing to the recent characterization of $T_{\rm H}^2$ cells (which express IL-9 but not the key signature cytokines of other $T_{\rm H}$ cell subsets, such as IL-4 ($T_{\rm H}^2$ cells), interferon- γ (IFN γ) ($T_{\rm H}^1$ cells) and IL-17 ($T_{\rm H}^{-1}$ 7 cells)) it is now unclear whether the correlation between IL-9 levels and $T_{\rm H}^2$ cell numbers is due to the direct production of this cytokine by



Figure 1 | **The IL-9 receptor signalling complex.** Interleukin-9 (IL-9) activates a heterodimeric receptor that consists of the IL-9 receptor α -chain (IL-9R α) and the γ -chain and promotes the crossphosphorylation of Janus kinase 1 (JAK1) and JAK3. This leads to the activation of signal transducer and activator of transcription 1 (STAT1), STAT3 and STAT5 and the upregulation of IL-9-inducible gene transcription. $\rm T_{H}2$ cells or whether $\rm T_{H}2$ -cell derived IL-4 supports the development of $\rm T_{H}9$ cells from either naive CD4⁺ T cells or $\rm T_{H}2$ cells. Currently, only one group has reported low but detectable co-expression of IL-4 and IL-9 during *in vitro* differentiation of T_{H}2 cells⁶. Surprisingly, to date there have been no reports addressing whether these cytokines are co-expressed by effector CD4⁺ T cell populations that differentiate *in vivo* during a T_{H}2-type immune response.

 $T_{\rm H}$ 9 cells. $T_{\rm H}$ 9 cells are a recently described subset of $\rm T_{\rm H}$ cells that express IL-9 but not other $\mathrm{T}_{_{\mathrm{H}}}$ cell lineage-specific cytokines or transcription factors7,8 (FIG. 2). The most definitive work that supports the existence and functional relevance of T₁₁9 cells in vivo has come from the study of mice with a T cell-specific deletion of the transcription factor PU.1. These mice do not develop IL-9-dependent allergic inflammation in the lungs and have only low levels of IL-9 in bronchoalveolar lavage fluid, but still develop normal numbers of T_{μ}^{2} cells. This finding indicates that, at least in this setting, $T_{\mu}9$ cells are the main T cell subset expressing IL-9 and that PU.1 is crucial for their development⁶. Another transcription factor that is associated with $\mathrm{T}_{\scriptscriptstyle H}9$ cell development both in vitro and in vivo is interferon-regulatory factor 4 (IRF4)⁹, which also regulates $T_{\rm H}^{2}$ and $T_{\rm H}^{17}$ cell differentiation¹⁰⁻¹². Characterization of $T_{_{\rm H}}$ 9 cells *in vitro* has shown that their generation is dependent on transforming growth factor- β (TGF β) and IL-4 (REF. 13). The addition of IL-25 along with these two cytokines can further enhance IL-9 production by T cells in vitro. T_H9 cells from IL-25-deficient mice have decreased IL-9 expression and reduced airway inflammation in a model of asthma14. There have also been conflicting reports that various cytokines, such as IL-1β, IL-6, IL-10, IL-12, IL-21, IFN α and IFN β , may have an additive effect in promoting T_{μ} 9 cell generation in the presence of TGF β and IL-4 in vitro^{15,16}. In addition, culturing human memory T cells in the presence of TGF β (alone or in combination with IL-1β, IL-4, IL-6, IL-12, IL-21 or IL-23) was shown to promote IL-9 production with or without co-expression of IFNy and IL-17 (REFS 16,17).

 T_{μ} 17 cells. There have also been reports that T₁₁17 cells can produce IL-9. In vivo differentiated $T_{\rm H}$ 17 cells, which were identified by their expression of either IL-17F or the T₁₁17-associated transcription factor retinoic acid receptor-related orphan receptor-yt (RORyt), could produce IL-9 after re-stimulation with phorbol 12-myristate 13-acetate and ionomycin¹⁸. In addition, some groups also reported co-expression of IL-17 and IL-9 during T_{H} 17 cell generation *in vitro* ^{18,19}. However, IL-23, which is important for the expansion of $T_{_{\rm H}}$ 17 cells after their initial differentiation, has been shown to inhibit the expression of IL-9 by T cells in mice19; therefore, this finding suggests that the expression of IL-9 by T₁₁17 cells could be transient and may decrease over time.

Human $T_{\rm H}17$ cells have also been shown to produce IL-9 *in vitro*. After multiple rounds of differentiation *in vitro*, some $T_{\rm H}17$ cells acquire expression of IL-9. In memory T cells, $T_{\rm H}17$ -promoting cytokines, such as IL-1 β and IL-21, can also enhance the co-production of IL-9 and IL-17. However, these cultures also contain IL-9-producing T cells that do not co-express IL-17 and are presumably bona fide $T_{\rm H}9$ cells¹⁶.

 T_{Reg} cells. There have been contradictory reports regarding the expression of IL-9 by T_{Reg} cells. Data from mice have shown that co-expression of forkhead box P3 (FOXP3) and IL-9 can occur in T_{Reg} cells that are found in tolerant allografts *in vivo*²⁰, as well as in purified T_{Reg} cell populations *in vitro*^{18,21}. However, other groups did not report co-expression of FOXP3 and IL-9 in T cell cultures that were generated *in vitro*^{8,19}. Similarly, there have been conflicting data from studies of T_{Reg} cells from healthy human donors^{15–17}. As one group showed that retinoic acid can downregulate IL-9 expression by T_{Reg} cells²², a possible explanation for the discrepancy between the data is that there may be differences in the levels of retinoic acid present in cultures *in vitro* and in the local microenvironments *in vivo*.

For all of the above CD4⁺ T cell subsets, TGF β seems to be the unifying factor for promoting IL-9 production; an inability to respond to TGF β *in vivo* prevents IL-9 expression by T cells⁸. The main exception to this observation is that TGF β does not seem to be necessary for IL-9 production by T_H2 cells; however, one possible explanation for this finding could be the presence of endogenous TGF β in the culture media that is used for T_H2 cell differentiation.

Mast cells as a source of IL-9. Although mainly studied as a target of IL-9, mast cells have also been reported to produce this cytokine. Production occurs in an autocrine manner in response to IL-9-induced signals and as a consequence of the crosslinking of IgE molecules on the surface of mast cells. Crosslinking of surface IgE molecules results in mast cell degranulation and leads to the immediate





release of numerous pre-formed mediators. Two of these products, histamine and IL-1 β , can promote further IL-9 production^{23,24}. As IL-9 is a growth factor for mast cells, it is thought that these pathways promote the survival and expansion of mast cells during an active immune response.

Natural killer T cells. Studies using natural killer T (NKT) cells from naive mice have shown that following stimulation with IL-2, but not IL-15, these cells can produce IL-9. IL-2 stimulation also led to some co-expression of IL-4, IL-5 and IL-13, but not IFN γ , by IL-9-producing NKT cells²⁵. In a mouse model of allergic airway inflammation, deficiency of CD1d-restricted NKT cells correlated with decreased IL-9 expression and reduced mast cell recruitment to the lungs. This finding suggested that NKT cells can promote IL-9 responses *in vivo*²⁶.

In addition, NKT cells that have undergone transformation to nasal NKT cell lymphoma cell lines also produce IL-9; in this setting, IL-9 functions as an autocrine growth factor. Histological sections from patients with nasal NKT cell lymphomas contained large numbers of infiltrating IL-9-producing NKT cells, which suggested that IL-9 may promote disease progression²⁷. It is currently unclear whether IL-9 can also have effects on other cell types in the nasal mucosa of these patients. For example, the effects of IL-9 on mast cells might contribute to cancer progression by promoting the production of angiogenic factors that increase the vasculature of the tumour.

Immune cell targets of IL-9

The downstream effects of IL-9 have been primarily associated with mast cells; however, this does not preclude IL-9 from exerting effects on other cell types. Here, we summarize the observed effects of IL-9 on mast cells and other cell types (FIG. 3). However, as there is currently no way to definitively confirm the expression of IL-9Ra on specific, rather than heterogenous, cell populations, it can be difficult to determine the relative importance of each individual cell type.

Mast cells. One of the main targets of IL-9 is the mast cell and, as mentioned earlier, initial studies described a role for IL-9 in promoting the expansion of mast cell populations²⁸. Subsequent work in mice that were deficient for both IL-9 and IL-9R α showed that IL-9 is not required for the generation of mast cell precursors, as the basal numbers of mast cells in these mice were normal. However, mice that were deficient for IL-9 or IL-9R α



Figure 3 | **Targets of IL-9 function.** Interleukin-9 (IL-9) has been shown to have various effects on different cell types. These effects include activating mast cells to secrete several products, including IL-13, which exerts its effects on the epithelial cells of the lung and gut. In addition, IL-9 seems to have a direct effect on regulatory T (T_{Reg}) cells, T helper 17 (T_{H} 17) cells and antigen-presenting cells (APCs). TGF β , transforming growth factor- β .

showed defective expansion and recruitment of mast cell populations in response to intestinal nematode infection or following the induction of experimental autoimmune encephalomyelitis (EAE)^{18,29}. IL-9 can induce mast cell production of TGF β , which can have pro-inflammatory downstream effects on neurons in a murine stroke model and on epithelial cells during intestinal inflammation^{30,31}. Following antigen-specific crosslinking of surface IgE molecules on mast cells, IL-9 can enhance mast cell expression of several cytokines, including IL-1β, IL-5, IL-6, IL-9, IL-10 and IL-13 (REF. 24). Of these cytokines, IL-5 and IL-13 are of particular interest, as the previously described direct effects of IL-9 in promoting eosinophilia and mucus production in the lung and the gut instead seem to be indirect effects mediated through the induction of these cytokines by IL-9 (REFS 32-36).

T cell subsets. There are some indications that IL-9 can target certain T cell subsets, specifically, $\rm T_{_H}17$ cells and $\rm T_{_{Reg}}$ cells. In the case of T_{μ} 17 cells, IL-9 seems to function as an autocrine growth factor that facilitates the expansion of $T_{H}17$ cell populations in vitro¹⁹. This is also supported by the decreased accumulation of $T_{\rm H}$ 17 cells that is seen in IL-9Ra-deficient mice during EAE18; however, because this is a global defect in vivo, the possibility that this is an indirect effect of IL-9 deficiency cannot be entirely dismissed. By contrast, IL-9 does not seem to facilitate the survival or expansion of T_{Reg} cells. Instead, standard suppression assays show that $T_{_{Reg}} \, cells$ from IL-9Ra-deficient mice are less able to inhibit the proliferation of effector CD4+ T cells in vitro19. This suggests that IL-9 can enhance the regulatory function of T_{Reg} cells through an unknown mechanism.

Antigen-presenting cells. Although a careful analysis of the specific antigenpresenting cell (APC) subsets that express the IL-9 receptor has not yet been carried out, there are indications that professional APCs are also targets of IL-9. During lipopolysaccharide-induced activation of a heterogenous population of macrophages and monocytes, IL-9 can promote the expression of TGFβ; this results in a decrease in the oxidative burst of these cells, as well as in decreased expression of tumour necrosis factor (TNF)37,38. In addition, the culture of peripheral blood mononuclear cells (PBMCs), which contain both T cells and monocytes, with IL-9 leads to

decreased production of IL-12 and IFN γ in response to treatment with extracts from *Mycobacterium tuberculosis*. Under these conditions, it was also shown that the addition of IL-9-specific blocking antibodies to the cultures upregulates PBMC production of IL-12 (REF. 39). Overall, these data suggest that IL-9 might limit the capacity of APCs to induce T_H1-type immune responses.

IL-9 and immunity

IL-9 can function as both a positive and negative regulator of immune responses (TABLE 1). In general, it seems that IL-9 has detrimental roles during allergy and autoimmunity. However, during parasitic infections, IL-9 can help to clear the pathogen, and during skin transplantation, IL-9 can promote the maintenance of a tolerant environment.

Allergy. During some allergic responses in the lung, which are traditionally thought of as being T_{H^2} cell-mediated, IL-9 contributes to disease by promoting mast cell expansion and production of IL-13, which in turn promotes the release of mucus that contributes to airway hyperresponsiveness^{33,35,40}. The initial influx of mast cells in this model is likely to be driven by IL-9 that is derived from NKT cells²⁶. In addition, T_H 9 cells also seem to contribute to disease in this model, as mice with PU.1-deficient T cells and a global IL-25 deficiency are protected from allergic airway inflammation^{6,14}. Similarly,

Table 1 Sources and targets of IL-9 during immune responses				
Immune model	Sources of IL-9	Targets of IL-9	Effects of IL-9	Refs
Allergy				
Allergic airway inflammation	NKT cells and T _H 9 cells	Mast cells	Promotes allergic inflammation	6,9,26, 33,36,40
Oral antigen-induced anaphylaxis	T _H 2 cells and T _H 9 cells	Mast cells	Promotes allergic inflammation	41
Autoimmunity				
EAE	$T_{_{\rm H}}$ 17 cells and $T_{_{ m H}}^{}$ 9 cells	Mast cells and T _H 17 cells	Promotes EAE	18, 42–44
	$T_{_{\rm H}}$ 17 cells and $T_{_{ m H}}^{}$ 9 cells	$T_{_{Reg}}cells$	Inhibits EAE	19
Type 1 diabetes	T _H 17 cells	Unknown	Unknown	16
Parasitic infection				
Lung infection with Schistosoma mansoni	T _H 2 cells and T _H 9 cells	Mast cells	No effect on granuloma formation	29
Intestinal infection with Trichuris muris	T _H 2 cells and T _H 9 cells	Mast cells	Promotes parasite expulsion	45–47
Transplantation				
Skin allograft transplantation	T _{Reg} cells	Mast cells and T _{Reg} cells	Promotes allograft tolerance	20,48
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EAE, experimental autoimmune encephalomyelitis; NKT, natural killer T; T_{H} , T helper; T_{Reo} , regulatory T.

during food allergy, IL-9 production (presumably by $T_H 2$ cells and/or $T_H 9$ cells) also contributes to mastocytosis and, indirectly, to increased intestinal permeability⁴¹.

Autoimmunity. In addition to having a pathogenic role in allergic responses in the lung and gut, IL-9 may contribute to the development of autoimmune disease. During EAE, both T_u17 cells and T_u9 cells have the capacity to produce IL-9. In addition, antibody-mediated blockade of IL-9, or IL-9Ra deficiency, attenuates disease progression18,42. This seems to be partly due to the direct effects of IL-9 on mast cells, as there is a decreased accumulation of these cells in the periphery¹⁸. However, in contrast to these findings, another study has reported that IL-9Ra-deficient mice develop more severe EAE¹⁹. Although these discrepancies cannot be readily explained, there is mounting evidence to suggest that IL-9 that is produced during EAE and other autoimmune diseases has pro-inflammatory effects; for example, the adoptive transfer of antigen-specific $T_H 9$ cells promotes EAE disease⁴³, and the decrease in EAE severity that is seen following the induction of oral tolerance is associated with reduced levels of IL-9 (REF. 44). In addition, patients with diabetes were shown to have increased levels of IL-9⁺ $T_{\rm H}$ 17 cells in the blood (REF. 16).

Parasitic infections. In the context of certain nematode infections, such as Trichuris muris infection, IL-9 can promote the isolation and/or expulsion of parasites from the gastrointestinal tract. In these circumstances, it is thought that T_{H}^{2} cells and/or $T_{\rm H}9$ cells are the main source of IL-9 and that the main targets of IL-9 are mucosal mast cells. IL-9 can stimulate the release of mast cell products that contribute to eosinophilia, mucus production, increased intestinal permeability and muscle contraction⁴⁵⁻⁴⁷. The net effect of these immune effector mechanisms is the physical expulsion of the parasite. By contrast, during lung parasitic infections, such as those caused by Schistosoma mansoni, IL-9-deficient mice do not show any functional defects in the formation of granulomas to isolate parasitic eggs, although decreased mucus production and mast cell accumulation are observed²⁹.

Transplant tolerance. During the transplantation of skin allografts, T_{Reg} cells, rather than T_{H} 9 cells, seem to be one of the major sources of IL-9, which is possibly involved

in the recruitment of mast cells that mediate tolerance without degranulation^{20,48}. IL-9 may also promote the ability of T_{Reg} cells that are present at the graft site to suppress ongoing immune effector responses against the skin allograft. Currently, it is unknown whether T_{Reg} cell-derived IL-9 has a central role in promoting tolerance at other transplantation sites.

Concluding remarks

Interest in IL-9 has been renewed recently, partly fuelled by reports that IL-9 can be expressed by several distinct T cell subsets, particularly following exposure to TGFβ. However, as much of this work has been based on in vitro studies, a re-examination of the main cell sources of IL-9 during different types of immune response in vivo is needed. This is particularly true of T_u2-type immune responses, as data from models of allergic lung inflammation suggest that $T_{H}9$ cells, rather than T_{H}^{2} cells, are the main source of IL-9 in this setting $^{6,14}\!\!.$ In addition, as $\rm T_{_H}17$ and T_{Reg} cells have been shown to be capable of producing IL-9 during autoimmunity and transplantation, the contribution of this cytokine to these processes also warrants further investigation¹⁸⁻²⁰.

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- Renauld, J. C. et al. Expression cloning of the murine and human interleukin 9 receptor cDNAs. Proc. Natl Acad. Sci. USA 89, 5690–5694 (1992).
- Bauer, J. H., Liu, K. D., You, Y., Lai, S. Y. & Goldsmith, M. A. Heteromerization of the γ chain with the interleukin-9 receptor α subunit leads to STAT activation and prevention of apoptosis. J. Biol. Chem. 273, 9255–9260 (1998).
- Demoulin, J. B. *et al.* A single tyrosine of the interleukin-9 (IL-9) receptor is required for STAT activation, antiapoptotic activity, and growth regulation by IL-9. *Mol. Cell Biol.* 16, 4710–4716 (1996).
- Demoulin, J. B., Van Roost, E., Stevens, M., Groner, B. & Renauld, J. C. Distinct roles for STAT1, STAT3, and STAT5 in differentiation gene induction and apoptosis inhibition by interleukin-9. *J. Biol. Chem.* 274, 25855–25861 (1999).
- Gessner, A., Blum, H. & Rollinghoff, M. Differential regulation of IL-9-expression after infection with *Leishmania major* in susceptible and resistant mice. *Immunobiology* 189, 419–435 (1993).
- Chang, H. C. *et al.* The transcription factor PU.1 is required for the development of IL-9-producing T cells and allergic inflammation. *Nature Immunol.* 11, 527–534 (2010).
- 7. Dardalhon, V. *et al.* IL-4 inhibits TGF- β -induced Foxp3 + T cells and, together with TGF- β , generates

IL-9 $^{\circ}$ IL-10 $^{\circ}$ Foxp3 $^{-}$ effector T cells. Nature Immunol. 9, 1347–1355 (2008).

- Veldhoen, M. et al. Transforming growth factor-β reprograms' the differentiation of T helper 2 cells and promotes an interleukin 9-producing subset. Nature Immunol. 9, 1541–1346 (2008).
- Staudt, V. *et al.* Interferon-regulatory factor 4 is essential for the developmental program of T helper 9 cells. *Immunity* 29 Jul 2010 (doi:10.1016/ j.immuni.2010.07.014).
- Brustle, A. *et al.* The development of inflammatory T_H-17 cells requires interferon-regulatory factor 4. *Nature Immunol.* 8, 958–966 (2007).
- Lohoff, M. *et al.* Dysregulated T helper cell differentiation in the absence of interferon regulatory factor 4. *Proc. Natl Acad. Sci. USA* 99, 11808–11812 (2002).
- Rengarajan, J. *et al.* Interferon regulatory factor 4 (IRF4) interacts with NFATc2 to modulate interleukin 4 gene expression. *J. Exp. Med.* **195**, 1003–1012 (2002).
- Schmitt, E. *et al.* IL-9 production of naive CD4⁺ T cells depends on IL-2, is synergistically enhanced by a combination of TGF-β and IL-4, and is inhibited by IFN-γ. J. Immunol. **153**, 3989–3996 (1994).
 Angkasekwinai, P., Chane, S. H., Thaoa, M., Watarai, H.
- Angkasekwinai, P., Chang, S. H., Thapa, M., Watarai, H. & Dong, C. Regulation of IL-9 expression by IL-25 signaling. *Nature Immunol.* 11, 250–256 (2010).
- Wong, M. T. *et al.* Regulation of human Th9 differentiation by type I interferons and IL-21. *Immunol. Cell Biol.* 88, 624–631 (2010).
- Beriou, G. *et al.* TGF-β Induces IL-9 production from human Th17 cells. *J. Immunol.* **185**, 46–54 (2010).
- Numan In17 Cells. J. Immunol. 185, 46–34 (2010)
 Putheti, P., Awasthi, A., Popoola, J., Gao, W. & Strom, T. B. Human CD4 memory T cells can become CD4+IL-9+ T cells. *PLoS ONE* 5, e8706 (2010)
- Nowak, E. C. *et al.* IL-9 as a mediator of Th17-driven inflammatory disease. *J. Exp. Med.* **206**, 1653–1660 (2009).
- Elyaman, W. et al. IL-9 induces differentiation of T_µ17 cells and enhances function of FoxP3+ natural regulatory T cells. Proc. Natl Acad. Sci. USA 106, 12885–12890 (2009).
- Lu, L. F. *et al.* Mast cells are essential intermediaries in regulatory T-cell tolerance. *Nature* 442, 997–1002 (2006).
- Liu, Y., Teige, I., Birnir, B. & Issazadeh-Navikas, S. Neuron-mediated generation of regulatory T cells from encephalitogenic T cells suppresses EAE. *Nature Med.* 12, 518–525 (2006).
- Hill, J. A. *et al.* Retinoic acid enhances Foxp3 induction indirectly by relieving inhibition from CD4+CD44^{hi} cells. *Immunity* 29, 758–770 (2008).
- Stassen, M. *et al.* Murine bone marrow-derived mast cells as potent producers of IL-9: costimulatory function of IL-10 and kit ligand in the presence of IL-1. *J. Immunol.* **164**, 5549–5555 (2000).
- Wiener, Z., Falus, A. & Toth, S. IL-9 increases the expression of several cytokines in activated mast cells, while the IL-9-induced IL-9 production is inhibited in mast cells of histamine-free transgenic mice. *Cytokine* 26, 122–130 (2004).
- Lauwerys, B. R., Garot, N., Renauld, J. C. & Houssiau, F. A. Cytokine production and killer activity of NK/T-NK cells derived with IL-2, IL-15, or the combination of IL-12 and IL-18. *J. Immunol.* 165, 1847–1853 (2000).
- Jones, T. G. *et al.* Antigen-induced increases in pulmonary mast cell progenitor numbers depend on IL-9 and CD1d-restricted NKT cells. *J. Immunol.* 183, 5251–5260 (2009).
- Nagato, T. *et al.* Expression of interleukin-9 in nasal natural killer/T-cell lymphoma cell lines and patients. *Clin. Cancer Res.* 11, 8250–8257 (2005).
- Renauld, J. C., Goethals, A., Houssiau, F., Van Roost, E. & Van Snick, J. Cloning and expression of a cDNA for the human homolog of mouse T cell and mast cell growth factor P40. *Cytokine* 2, 9–12 (1990).
- Townsend, J. M. et al. IL-9-deficient mice establish fundamental roles for IL-9 in pulmonary mastocytosis and goblet cell hyperplasia but not T cell development. *Immunity* 13, 573–583 (2000).
- Mesples, B., Fontaine, R. H., Lelievre, V., Launay, J. M. & Gressens, P. Neuronal TGF-β1 mediates IL-9/mast cell interaction and exacerbates excitotoxicity in newborn mice. *Neurobiol. Dis.* 18, 193–205 (2005).
- Miller, H. R., Wright, S. H., Knight, P. A. & Thornton, E. M. A novel function for transforming growth factor-β1: upregulation of the expression and

the IgE-independent extracellular release of a mucosal mast cell granule-specific β-chymase, mouse mast cell protease-1. *Blood* **93**, 3473–3486 (1999). Steenwinckel, V. *et al.* IL-9 promotes IL-13-dependent

- Steenwinckel, V. et al. IL-9 promotes IL-13-dependent paneth cell hyperplasia and up-regulation of innate immunity mediators in intestinal mucosa. J. Immunol. 182, 4737–4743 (2009).
- Steenwinckel, V. et al. IL-13 mediates in vivo IL-9 activities on lung epithelial cells but not on hematopoietic cells. J. Immunol. 178, 3244–3251 (2007).
- Temann, U. A., Ray, P. & Flavell, R. A. Pulmonary overexpression of IL-9 induces Th2 cytokine expression, leading to immune pathology. J. Clin. Invest. 109, 29–39 (2002).
- Whittaker, L. *et al.* Interleukin-13 mediates a fundamental pathway for airway epithelial mucus induced by CD4 T cells and interleukin-9.
 Am. J. Perspir CH. Mol. Biol. 27, 593–603 (2002)
- Am. J. Respir. Cell. Mol. Biol. 27, 593–602 (2002).
 Sitkauskiene, B. et al. Airway allergen exposure stimulates bone marrow eosinophilia partly via IL-9. Respir. Res. 6, 33 (2005).
- Pilette, C. *et al.* IL-9 inhibits oxidative burst and TNF-α release in lipopolysaccharide-stimulated human monocytes through TGF-β. *J. Immunol.* 168, 4103–4111 (2002).

- Pilette, C. *et al.* Oxidative burst in lipopolysaccharideactivated human alveolar macrophages is inhibited by interleukin-9. *Eur. Respir. J.* 20, 1198–1205 (2002).
- Wu, B. *et al.* IL-9 is associated with an impaired Th1 immune response in patients with tuberculosis. *Clin. Immunol.* **126**, 202–210 (2008).
- Cheng, G. et al. Anti-interleukin-9 antibody treatment inhibits airway inflammation and hyperreactivity in mouse asthma model. Am. J. Respir. Crit. Care Med. 166, 409–416 (2002).
- Osterfeld, H. et al. Differential roles for the IL-9/IL-9 receptor a-chain pathway in systemic and oral antigeninduced anaphylaxis. J. Allergy Clin. Immunol. 125, 469–476, e462 (2010).
- Li, H. *et al.* Neutralization of IL-9 amerilorates experimental autoimmune encephalomyelitis by decreasing the effector T cell population. *J. Immunol.* 30 Aug 2010 (doi:10.4049/jimmunol.1000986).
- Aug 2010 (doi:10.4049/jimmunol.1000986).
 Jager, A., Dardalhon, V., Sobel, R. A., Bettelli, E. & Kuchroo, V. K. Th1, Th17, and Th9 effector cells induce experimental autoimmune encephalomyelitis with different pathological phenotypes. *J. Immunol.* 183, 7169–7177 (2009).
- 44. Peron, J. P. *et al.* Oral tolerance reduces Th17 cells as well as the overall inflammation in the central nervous

system of EAE mice. J. Neuroimmunol. 25 Jun 2010 (doi:10.1016/j.neuroim.2010.06.002).

- Khan, W. I. *et al.* Modulation of intestinal muscle contraction by interleukin-9 (IL-9) or IL-9 neutralization: correlation with worm expulsion in murine nematode infections. *Infect. Immun.* 71, 2430–2438 (2003).
- Forbes, E. E. *et al.* IL-9- and mast cell-mediated intestinal permeability predisposes to oral antigen hypersensitivity. *J. Exp. Med.* **205**, 897–913 (2008).
 Richard, M., Grencis, R. K., Humphreys, N. E.,
- Richard, M., Grencis, R. K., Humphreys, N. E., Renauld, J. C. & Van Snick, J. Anti-IL-9 vaccination prevents worm expulsion and blood eosinophilia in *Trichuris muris*-infected mice. *Proc. Natl Acad. Sci.* USA 97, 767–772 (2000).
- de Vries, V. C. *et al.* Mast cell degranulation breaks peripheral tolerance. *Am. J. Transplant* 9, 2270–2280 (2009).

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Competing interests statement

The authors declare no competing financial interests.