Efficacy of regulatory T-cell immunotherapy: are inflammatory cytokines key determinants?

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We read with interest the recent article in *Nature Reviews Immunology* by Maria-Grazia Roncarolo and Manuela Battaglia, which summarizes our current knowledge of regulatory T cells and the possibilities of using such cells in immunotherapeutic protocols for preserving or restoring tolerance to self antigens and alloantigens in humans¹. Because of the low numbers of antigen-specific regulatory T cells in the circulation and the difficulties in obtaining them, the most favoured approach for regulatory T-cell immunotherapy is the *in vitro* expansion of polyclonal regulatory T cells. As described in this Review, from the perspective of preventive therapy, regulatory T-cell immunotherapy, for example, represents a promising approach to avoid transplant rejection. However, considering the intrinsic defects in regulatory T-cell functions in several autoimmune diseases, the approach of regulatory T-cell immunotherapy proposed by the authors might not be efficacious for curing ongoing acute and chronic inflammatory and autoimmune diseases. We therefore suggest an alternative strategy for regulatory T-cell immunotherapy, which involves a primary step of induction of functionally fit regulatory T cells in patients, followed by *in vitro* expansion of these induced polyclonal regulatory T cells for therapeutic purposes.

Several recent studies have analysed the role and function of naturally occurring forkhead box P3 (FOXP3)⁺CD4⁺CD25⁺ regulatory T cells (referred to as natural T_{Reg} cells) in experimental models of arthritis and in patients with rheumatoid arthritis, both at acute and chronic stages of the disease². One of the major findings is that the frequency of natural T_{Reg} cells is higher in the inflamed joints than in peripheral blood of patients with rheumatoid arthritis. The reasons for the increased frequencies of these cells in the inflamed synovia are not known. In addition to preferential homing to synovia from peripheral blood, it is possible that the T_{Reg}-cell population expands within the synovia under the influence of cytokines². Nevertheless, the occurrence

of persistent inflammation in rheumatoid joints despite the presence of increased T_{Reg}-cell numbers indicates that these cells are ineffective in controlling the ongoing inflammatory response. Although activated human dendritic cells are susceptible to suppression by natural T_{Reg} cells *in vitro*^{3,4}, pro-inflammatory cytokines, in particular tumour-necrosis factor (TNF), secreted by activated monocytes and macrophages can suppress the functions of natural T_{Reg} cells and render them defective^{5–7}. This inhibition has been shown to involve signalling through TNF receptor type II, which is constitutively expressed by unstimulated natural T_{Reg} cells and is upregulated by TNF⁶. TNF-mediated inhibition of suppressive function has also been associated with a decrease in the expression of FOXP3 by natural T_{Reg} cells isolated from patients with active rheumatoid arthritis. These results may explain why most studies involving the adoptive transfer of natural T_{Reg} cells in experimental settings have shown therapeutic efficacy only at early stages of arthritis or when used as a preventive therapy, but have not been effective in curing established disease^{1,8–10}.

Consistent with the inhibitory effect of TNF on natural T_{Reg}-cell function, treatment of patients with rheumatoid arthritis with TNF-specific monoclonal antibodies has been shown to be associated with a significant increase in the number of peripheral-blood regulatory T cells in patients who responded to this treatment compared with those who did not respond^{5,6}. However, this increase was due to the generation of a newly differentiated population of regulatory T cells, which exert their suppressive function by secreting transforming growth factor- β and interleukin-10 (IL-10)¹¹. The absence of CD62L expression on these cells marked them as phenotypically distinct from the natural T_{Reg} cells present in patients with active rheumatoid arthritis. Whereas such 'induced' CD62L[−] regulatory T cells display potent suppressor capacity, the CD62L⁺ natural T_{Reg} cells remained defective in patients treated with TNF-specific monoclonal antibodies. These results suggest that natural

T_{Reg} cells are defective in rheumatoid arthritis, and perhaps also other autoimmune and inflammatory diseases such as multiple sclerosis and type 1 diabetes^{11–13}. Therefore, immunotherapy approaches involving the adoptive transfer of these defective natural T_{Reg} cells after expansion *ex vivo* might not be beneficial, as such cells are unlikely to be efficient at restoring self tolerance in patients with established disease.

Given the intrinsic defects in the natural T_{Reg}-cell compartment in several autoimmune diseases, therapeutic modulation of the regulatory T-cell compartment *in vivo* would appear a more plausible and promising approach. We therefore propose that an important strategy for restoring tolerance in patients with established autoimmune diseases would be to inhibit the activity or production of pro-inflammatory cytokines *in vivo* through pharmaceutical or biological agents¹⁴, such as TNF-specific antibody, and to induce functional regulatory T cells *de novo*⁸. It is however worth noting that patients with rheumatoid arthritis were found to relapse shortly after withdrawal of TNF-specific therapy¹⁵. This indicates that despite reducing the joint inflammation and inducing functional regulatory T cells, tolerance imparted by therapeutic monoclonal antibody therapy is of short duration. Nevertheless, functional regulatory T cells are already induced in these patients as a consequence of neutralization of inflammatory environment by monoclonal antibodies. We therefore suggest that such functionally competent, induced regulatory cells may then be considered for immunotherapy protocols to achieve long-term immune tolerance.

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