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# Pathobiont peacekeepers

Chronic inflammatory diseases of the gut, such as inflammatory bowel disease, are often potentiated by the same microbial species that colonize healthy individuals, such as the *Helicobacter* species. It has been unclear how the pro-inflammatory effector T cell response to these so-called pathobionts is restrained during normal gut homeostasis. Now, Littman and colleagues describe a population of pathobiont-specific peripherally derived regulatory T (pT<sub>reg</sub>) cells that selectively restrain pathobiont-driven inflammation in healthy mice.

*Helicobacter hepaticus* stably colonizes the large intestine of wild-type mice but causes T helper 17 (T<sub>H</sub>17) cell-mediated inflammation in mice with a deficiency of IL-10-mediated immune tolerance. Using this model, the authors cloned *H. hepaticus*-specific T cell receptor (TCR) sequences from the inflammatory T<sub>H</sub>17 cells and identified immunodominant peptide epitopes of *H. hepaticus*. Using mice transgenic for one of the cloned TCRs (for example, HH7-2tg mice) or MHC class II

“MAF is a crucial cell-intrinsic factor for the differentiation of *H. hepaticus*-specific pT<sub>reg</sub> cells”

tetramers loaded with *H. hepaticus* peptide, they could track *H. hepaticus*-specific T cells *in vivo*. When naive HH7-2tg T cells were transferred into wild-type mice colonized with *H. hepaticus*, they accumulated in the large intestinal lamina propria and differentiated mainly into forkhead box protein P3 (FOXP3)<sup>+</sup> pT<sub>reg</sub> cells. By contrast, when HH7-2tg T cells were transferred into colonized IL-10-deficient mice, they mainly differentiated into T<sub>H</sub>17 cells with pro-inflammatory T<sub>H</sub>1 cell-like features. Similar differentiation profiles were observed for the adoptive transfer of other *H. hepaticus*-specific T cell clones.

The transcription factor MAF was shown to be highly enriched in the HH7-2tg pT<sub>reg</sub> cells. In *H. hepaticus*-colonized mice with a conditional deletion of *Maf* in FOXP3<sup>+</sup> cells, the proportion of T<sub>reg</sub> cells in the large intestine was markedly decreased, with a concomitant increase in the number of T<sub>H</sub>17 cells. Furthermore, *H. hepaticus*-specific T cells were mainly T<sub>H</sub>17 cells rather than T<sub>reg</sub> cells in the mice with conditional *Maf*

deletion in T<sub>reg</sub> cells. Most of these T<sub>H</sub>17 cells expressed MAF, which indicates that they do not arise from the MAF-deficient T<sub>reg</sub> cells. Thus, the results suggest that MAF is a crucial cell-intrinsic factor for the differentiation of *H. hepaticus*-specific pT<sub>reg</sub> cells that control a separate population of *H. hepaticus*-specific T<sub>H</sub>17 cells. MAF-deficient pT<sub>reg</sub> cells were also functionally impaired in terms of defective expression of IL-10 and T<sub>reg</sub> cell signature genes.

In support of a role for MAF-expressing T<sub>reg</sub> cells in controlling spontaneous inflammatory responses to pathobionts, *H. hepaticus*-colonized mice with conditional deletion of MAF in T<sub>reg</sub> cells had signs of spontaneous colitis. Similar results were observed for mice with impaired signal transducer and activator of transcription 3 (STAT3) or transforming growth factor- $\beta$  signalling in T<sub>reg</sub> cells, both of which are required for MAF expression. Finally, the authors showed that *H. hepaticus*-specific T<sub>reg</sub> cells were better at suppressing colitis than polyclonal T<sub>reg</sub> cells in an adoptive transfer model, which highlights an important role for pathobiont-specific pT<sub>reg</sub> cells in addition to thymus-derived ‘natural’ T<sub>reg</sub> cells in maintaining gut homeostasis.

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