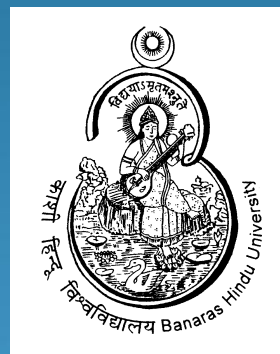


Purification and physicochemical studies of metalloprotease, cotinifolin from *Euphorbia cotinifolia*

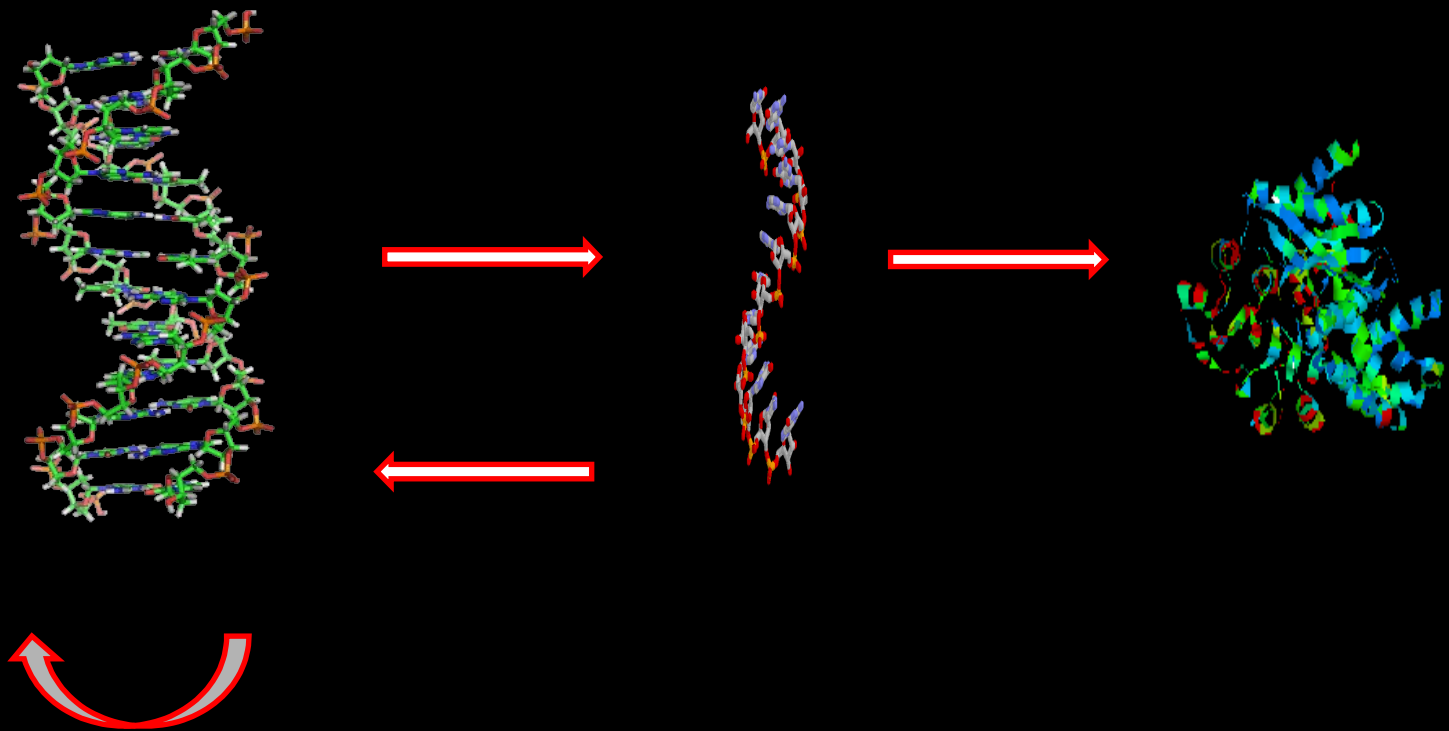


Reetesh Kumar



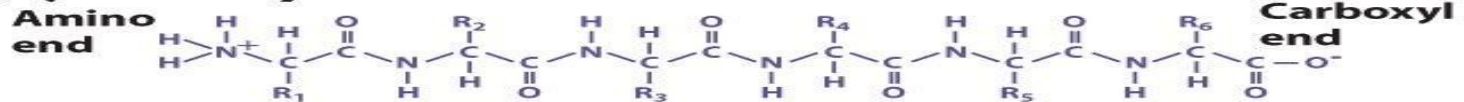
**Molecular Biology Unit
Institute of Medical Sciences
Banaras Hindu University**

Introduction of proteins and protein folding

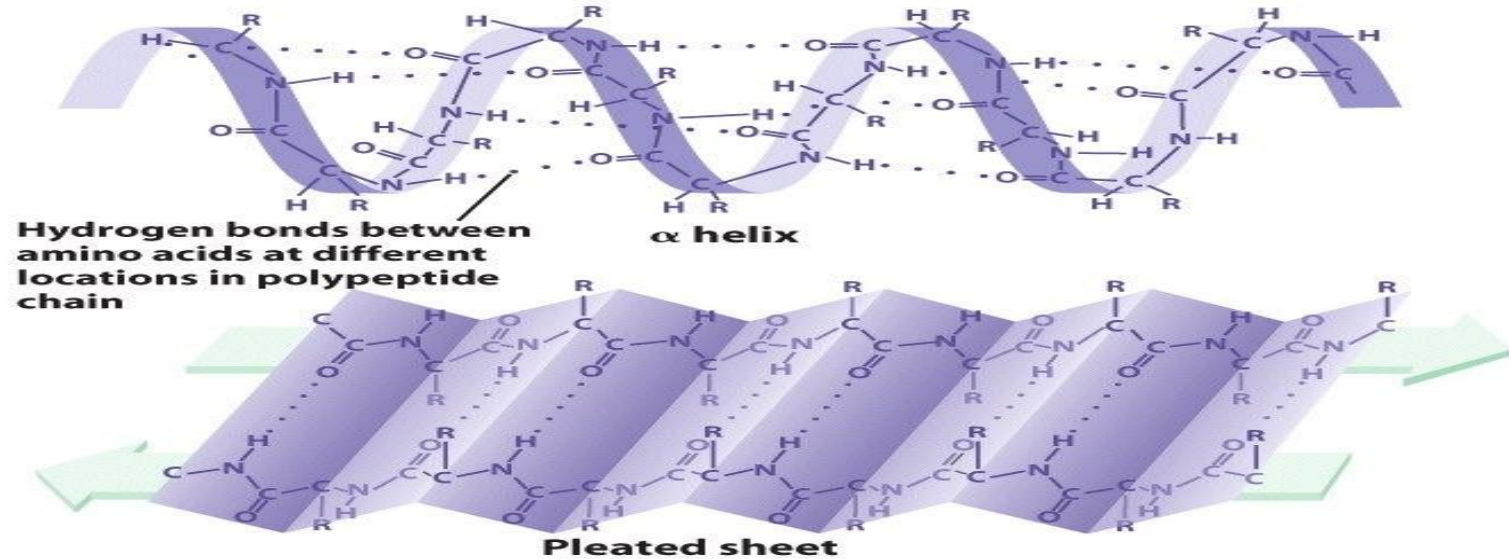


Protein, An introduction!!

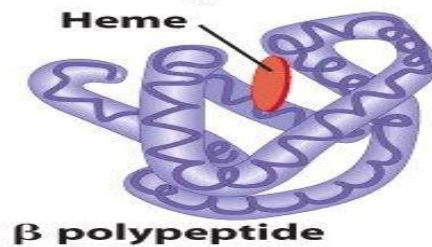
(a) Primary structure



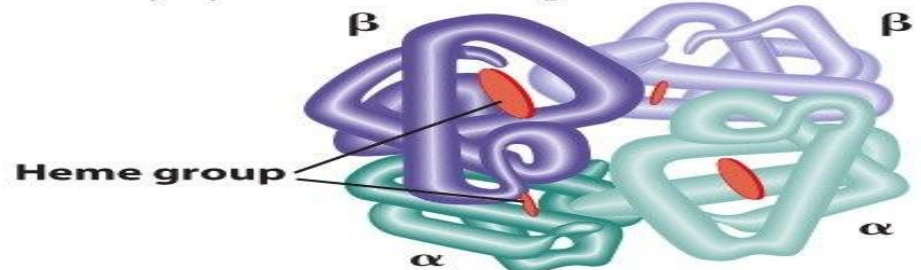
(b) Secondary structure



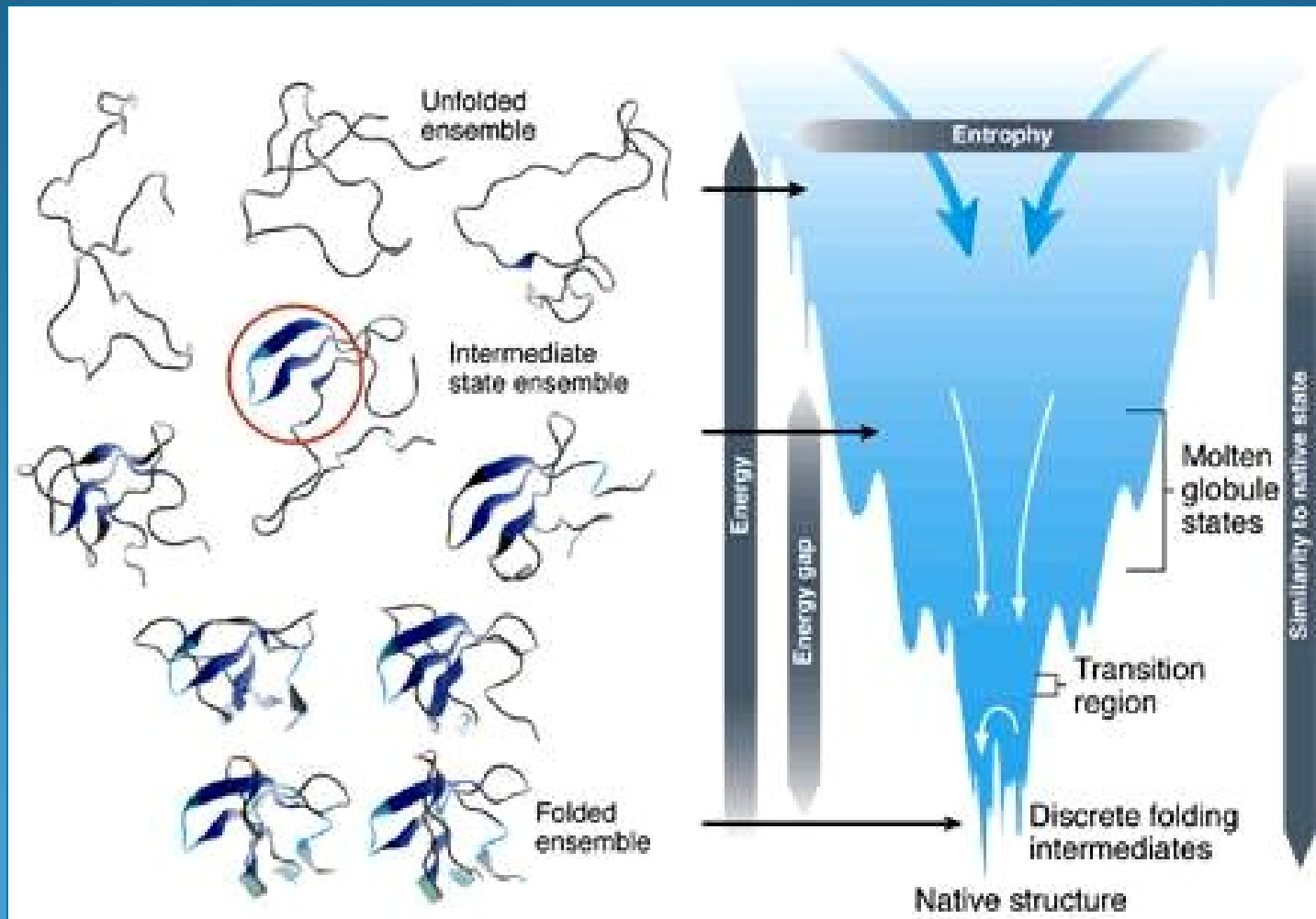
(c) Tertiary structure



(d) Quaternary structure



Protein folding funnel



Biochemical and spectroscopic characterization of a novel metalloprotease, cotinifolin from an antiviral plant shrub: *Euphorbia cotinifolia*

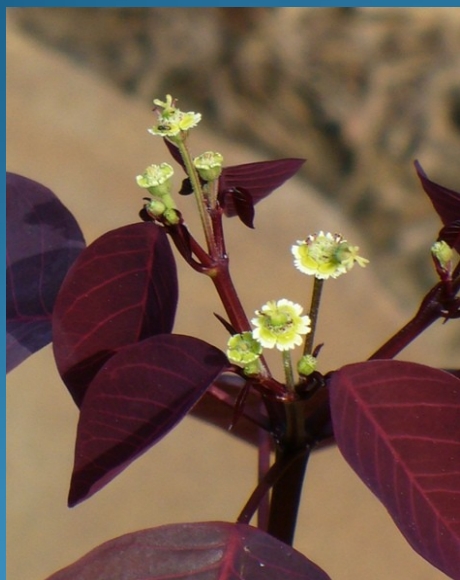
Euphorbia cotinifolia plant with medicinal implications

↓
Latex extracted from stem

↓
Using anion exchange chromatography and Hydrophobic interaction chromatography

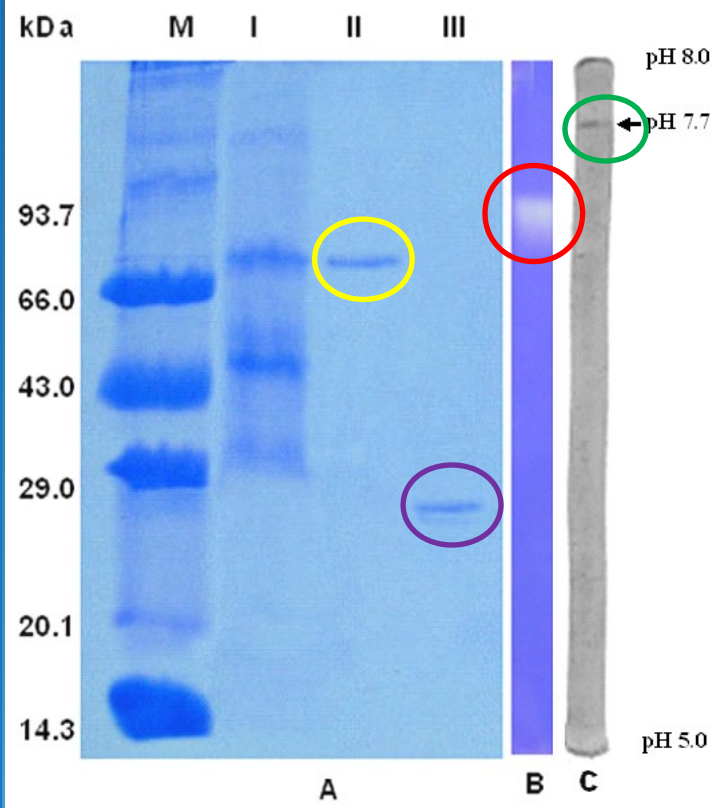
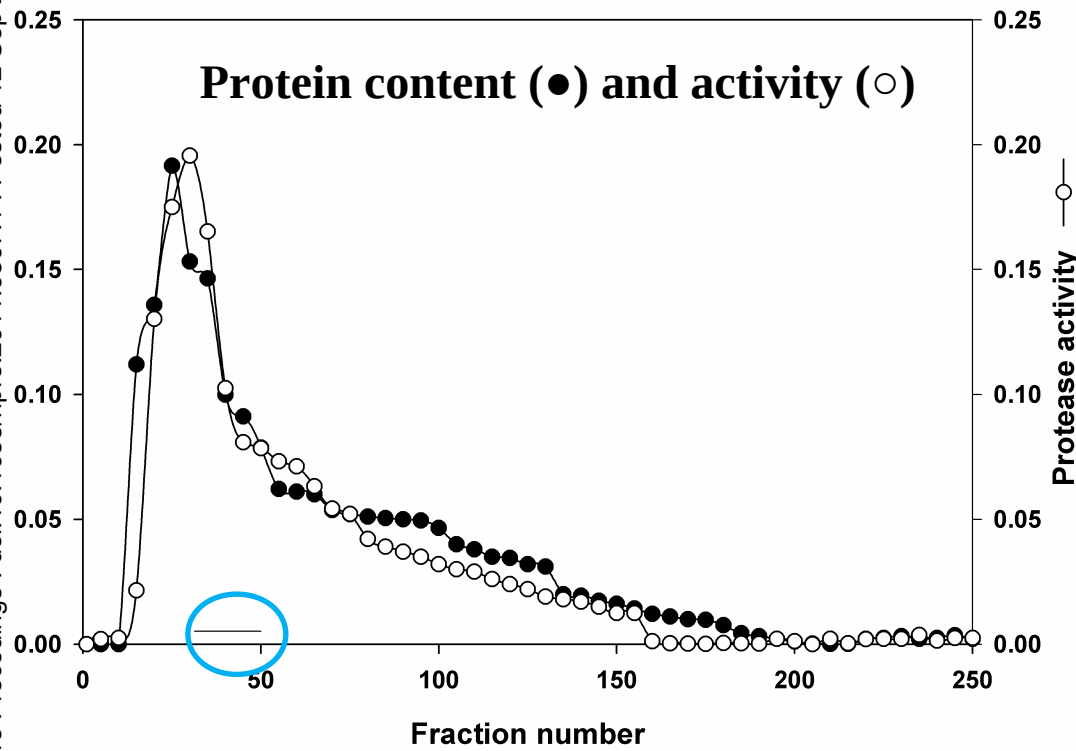
↓
Purification of Metalloprotease (Cotinifolin)

- pH, Temperature optima and Stability
- Effect of inhibitors
- Effect of metal ions
- Effect of substrate concentration on Reaction Velocity
- Effect of Chaotrops, Organic Solvents, detergents
- Autodigestion study
- Spectroscopic Studies: Absorbance, Fluorescence
Circular Dichroism



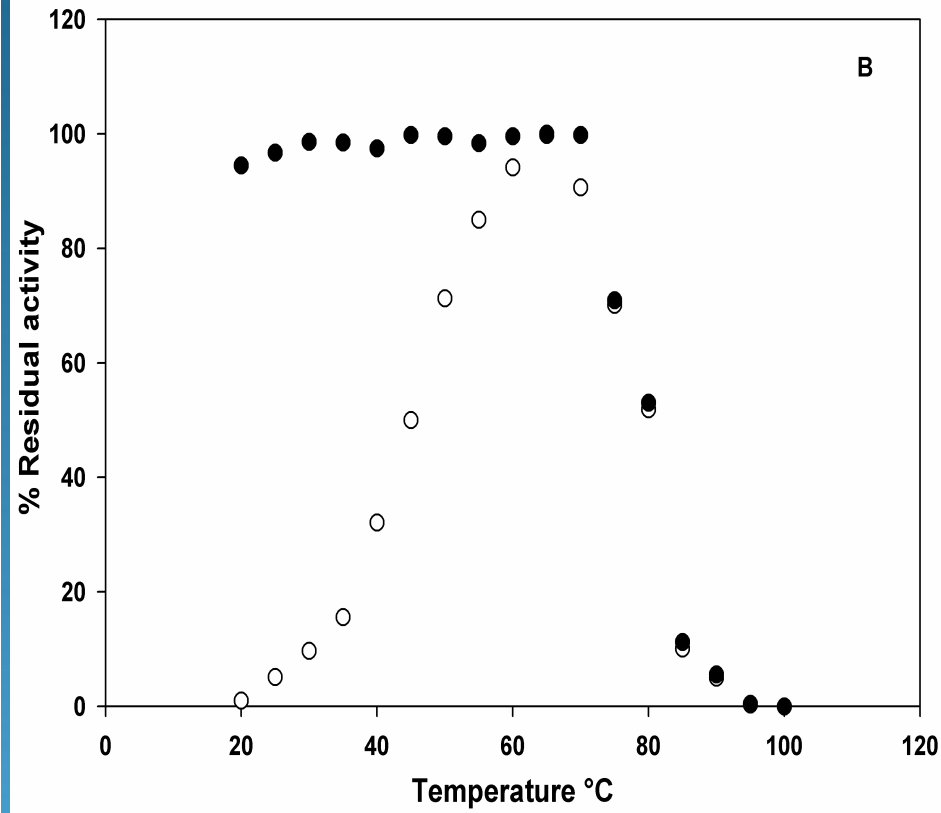
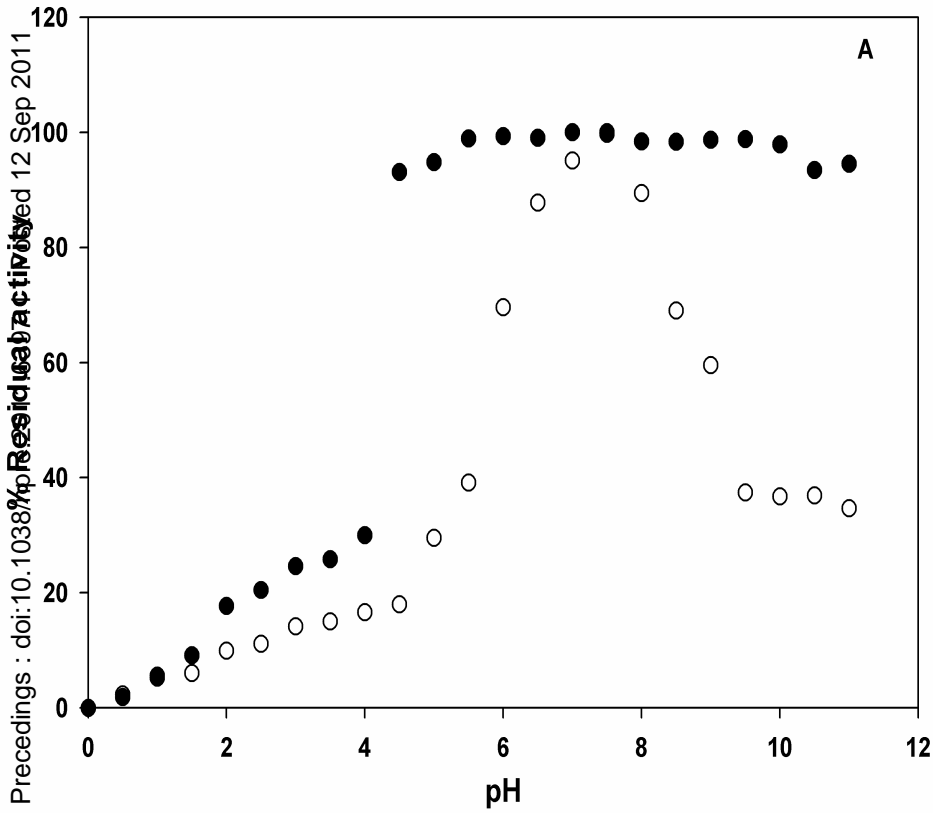
Purification of metalloprotease, Cotinifolin

Nature Precedings : doi:10.1038/npre20112809711.1 : Posted 12 Sep 2011



Steps	Total protein (mg)	Total activity (units)	Specific activity (units.mg ⁻¹)	Yield (%)
Crude extract	315	6160	19.55	100
DEAE Sepharose	115	2324	20.20	37.7
HIC column	18	418	23.22	6.78

pH, Temperature optima and Stability



Effects of pH Activity (○) Stability (●)

pH Optima 7.5

PH Stability 4 - 11

Effect of temperature Activity (○) Stability (●)

Temperature Optima 65 °C

Temperature Stability 20 – 75 °C

Nature Precedings : doi:10.1038/npre.2011.6397.1 : Posted 12 Sep 2011

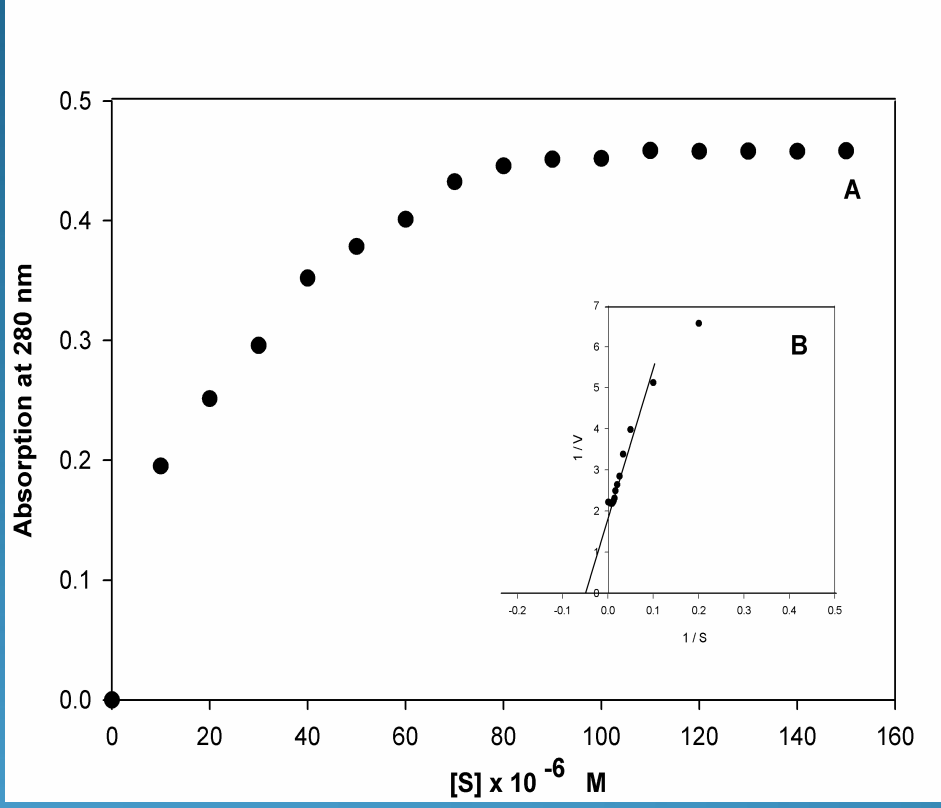
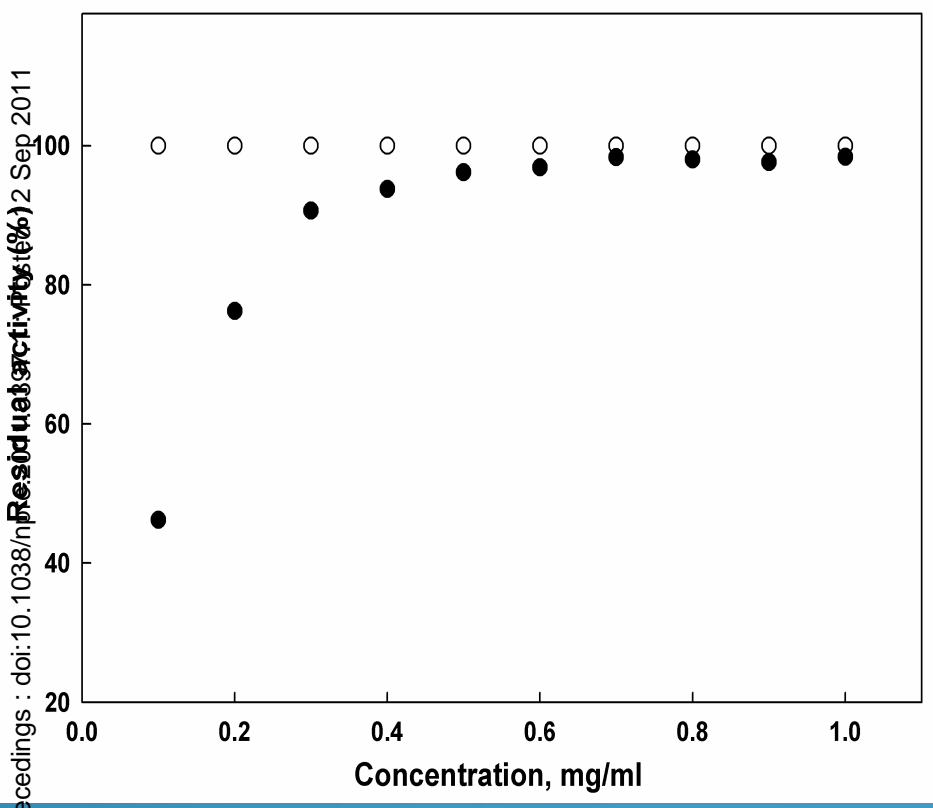
Chaotroph & Organic solvent	Concentration (M)	Residual activity (%)
Ethanol	50%	30
Methanol	50%	85
Iso propanol	50%	95
Butanol	50%	50
N-Hexane	50%	100
Acetonitrile	50%	100
Dioxane	50%	50
Triton X-100	3.5%	85
SDS	0.1%	50
Exilin	1%	50
GuHCl	1.5M	90
Urea	8M	90

Inhibitors	Concentration (M)	Residual activity (%)
IAA	10⁻³	80
PMSF	10⁻³	85
SBTI	10⁻³	90
o-phenanthroline	10⁻³	0
	10⁻⁴	5
	10⁻⁵	10
EGTA	10⁻³	60
	10⁻⁴	80
	10⁻⁵	100
EDTA	10⁻³	40
	10⁻⁴	55
	10⁻⁵	70

Metal ions	Concentration (M)	Protease activity (%)
Hg²⁺	10⁻³	100
Mg²⁺	10⁻³	75
Ca²⁺	10⁻³	80
Ba²⁺	10⁻³	95
Cs²⁺	10⁻³	75
Ni²⁺	10⁻³	125
Mn²⁺	10⁻³	95
Co²⁺	10⁻³	15
Zn²⁺	10⁻³	50
Cu²⁺	10⁻³	60

Enhanced by : Ni²⁺

Inhibited by: Co²⁺



Autodigestion of Cotinifolin

- Residual activity (●)
- Activity after 5 min (○) of incubation, taken as 100% activity

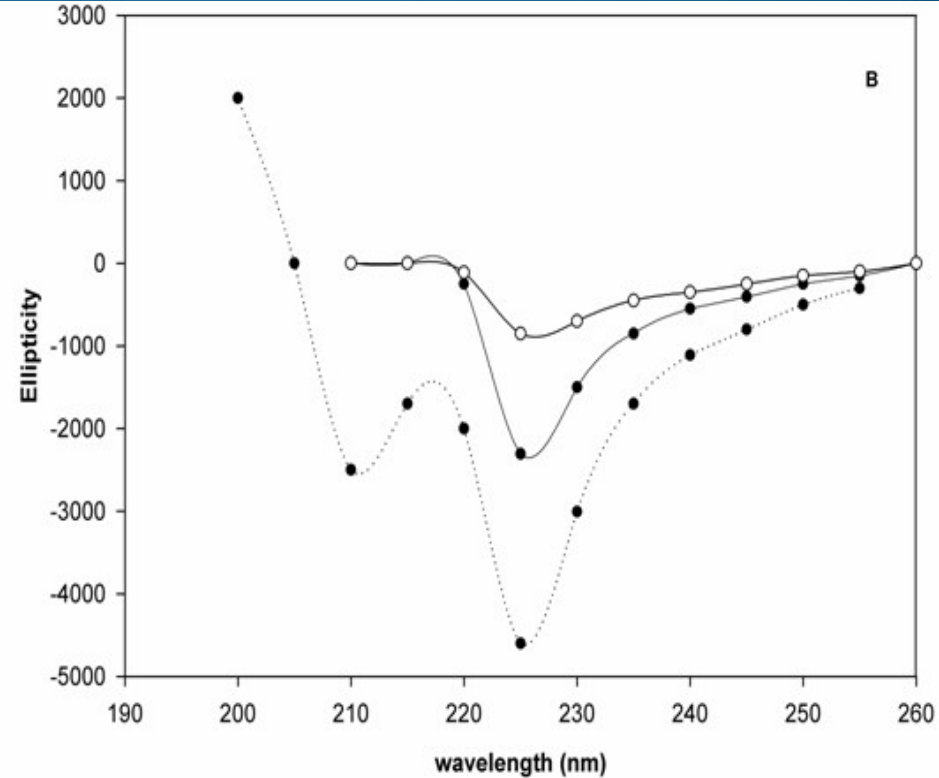
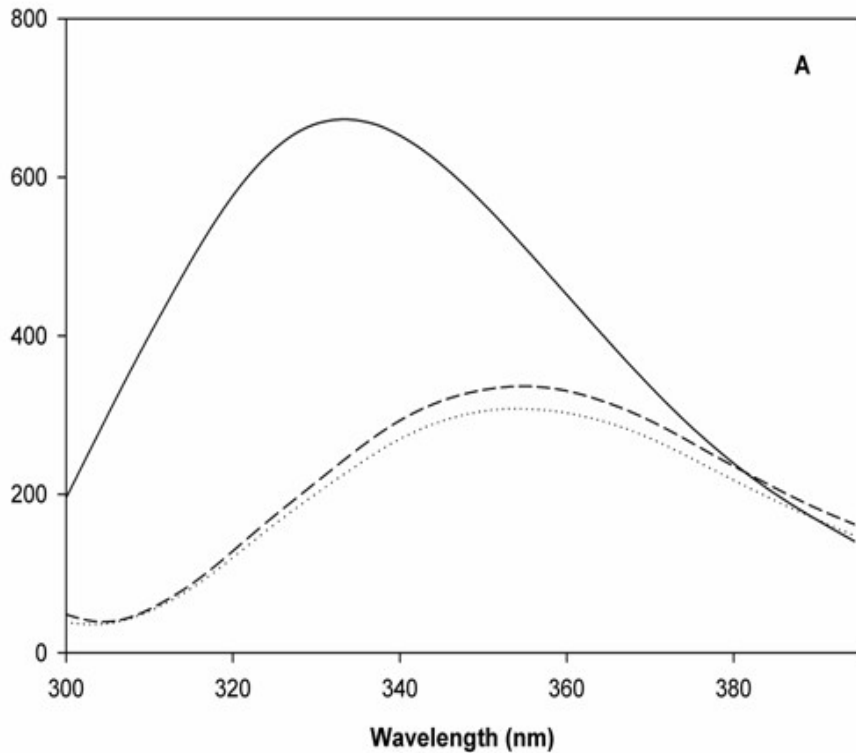
Effect of substrate concentration on reaction velocity

- A. Michaelis-Menten equation
- B. Lineweaver-Burk plot

$K_m = 20 \mu M \pm 0.5.$

Spectroscopic studies of metalloprotease, cotinifolin

Nature Precedings : doi:10.1038/npre2011060971r1e1 registered 12 Sep 2011



A. Intrinsic fluorescence spectra

Native (—)

Denatured conditions

3 M GuHCl (---)

6 M GuHCl(.....)

B. Circular dichroism spectrum

Native (---●---)

Denatured conditions

3 M GuHCl (-●-)

6 M GuHCl (-○-)

Summary

- Adequate amount of latex
- Easy economic purification
- Broad substrate specificity
- Stability against different temperature, pH
- Stability against different organic solvents
- Excellent model system to study structure-function relationship of other plant metalloprotease
- Crucial for food and biotechnological industries as well as protein folding studies.

Thank You