

1 **The “Minimum Information about an ENvironmental Sequence” (MIENS)**  
2 **specification**

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113 **Summary**

114 **We present the Genomic Standards Consortium’s (GSC) “Minimum Information**  
115 **about an ENvironmental Sequence” (MIENS) standard for describing marker**  
116 **genes. Adoption of MIENS will enhance our ability to analyze natural genetic**  
117 **diversity across the Tree of Life as it is currently being documented by massive**  
118 **DNA sequencing efforts from myriad ecosystems in our ever-changing biosphere.**

119

120 **Big Data need Standards**

121 The term Big Data is increasingly being used to describe the vast capacity of high-  
122 throughput experimental methodologies, especially next-generation sequencing, to  
123 generate data <sup>1,2</sup>. Sharing and re-use of such data, and translating such data into  
124 knowledge, requires widely-adopted standards that are best developed within the auspices  
125 of international working groups <sup>3</sup>. Here we describe a new standard, developed by a large  
126 and diverse community of researchers, to describe one of the most abundant and useful  
127 types of sequence data – that of marker gene data sets.

128

129 ***The wealth of marker gene data sets***

130 The adoption of phylogenetic marker genes as molecular proxies for tracking and  
131 cataloguing the diversity of microorganisms has revolutionized the way we view the  
132 biological world, and provided us with insights into how life has evolved and how  
133 different organisms are genetically related to each other. In the 1970s, studies of small  
134 subunit (SSU) ribosomal RNA (rRNA) genes from environmental samples led to the  
135 discovery of the domain *Archaea* <sup>4</sup> and to the proposal for a three domain classification of  
136 life <sup>5</sup>. Following Darwin's insight that all life is related, SSU rRNA gene surveys allow  
137 organisms from any communities, no matter how diverse, to be compared using the same  
138 universal phylogenetic tree. This rRNA gene-based molecular approach to characterizing  
139 natural communities of organisms provided, for the first time, culture-independent access  
140 to the diversity and distribution of microorganisms '*in situ*'. As a result, we are now  
141 acutely aware that the vast majority (90-99%) of microorganisms have evaded isolation  
142 using existing cultivation methods <sup>6-8</sup>.

143 Over the past three decades, the 16S rRNA, 18S rRNA and internal transcribed spacer  
144 gene sequences (ITS) from *Bacteria*, *Archaea*, and microbial *Eukaryotes* have provided  
145 deep insights into the topology of the tree of life <sup>9-12</sup> and the composition of communities  
146 of organisms that live in diverse environments, which range from deep sea hydrothermal  
147 vents to ice sheets in the Arctic <sup>13-24</sup>.

148 Numerous other phylogenetic marker genes have also proven useful <sup>25</sup>: Currently, around  
149 40 such phylogenetic marker genes are in wide use, representing well-conserved,  
150 housekeeping genes that include initiation factors, for example, RNA polymerase  
151 subunits (*rpoB*), DNA gyrases (*gyrB*), DNA recombination and repair proteins (*recA*) and  
152 heat shock proteins (*HSP70*) <sup>10,26</sup>. Most of these genes support or complement the  
153 currently accepted topology of the Tree of Life. Combinations of these genes can also be  
154 used in multi-locus sequence typing (MLST) approaches, increasing phylogenetic  
155 resolution and differentiating between closely related species of the same genus <sup>27,28</sup>.

156 Marker genes can also reveal key metabolic functions rather than phylogeny; examples  
157 include nitrogen cycling (*amoA*, *nifH*, *ntcA*) <sup>29,30</sup>, sulfate reduction (*dsrAB*) <sup>31</sup> or  
158 phosphorus metabolism (*phnA*, *phnI*, *phnJ*) <sup>32-34</sup>.

159 The molecular approach has been extended beyond microorganisms by its application to  
160 phylogeny and systematics of higher *Eukaryotes*. The Barcode of Life Initiative (BOLI)  
161 has adapted the molecular approach with the standardized use of a specific gene  
162 sequence: the 680 base-pair region of mitochondrial cytochrome c oxidase I (“COI”), as a  
163 means of rapid species identification and discrimination <sup>35</sup>.

164 In this paper we collectively define all of these different phylogenetic and functional  
165 genes (or gene fragments) as ‘marker genes’ as they are used to profile natural genetic



166 diversity across the Tree of Life, and argue that a small amount of additional effort  
167 invested in describing them with specific guidelines in our public databases will  
168 revolutionize the types of studies that can be performed with these large data resources.  
169 This effort is timely, given the need to determine how climate change and various other  
170 anthropogenic perturbations of our biosphere are affecting biodiversity, and how marked  
171 changes in our cultural traditions and lifestyles are affecting human microbial ecology.

172

173 *The collective value of marker gene sequences*

174 The quality and quantity of marker gene sequence data used to make phylogenetic  
175 assignments, to infer metabolic traits, and to assess biogeographic distributions continues  
176 to increase rapidly due to the availability of next generation sequencing (NGS)  
177 technologies powering the ability to study increasingly complex and/or divergent  
178 ecosystems.

179 Some recent large-scale molecular studies have endeavored to use environmental  
180 variables to explain the diversity and distribution of microbes. For example, a clear  
181 correlation between phylogenetic similarity and similar living conditions was observed  
182 using data in available SSU sequence repositories and culture collections <sup>36</sup>. In addition,  
183 two separate global environmental studies established a latitudinal diversity gradient for  
184 marine *Bacteria* <sup>37,38</sup>. Furthermore, it was shown that temporally-driven environmental  
185 factors, such as temperature and nutrients, correlate with local seasonal succession of  
186 marine microbial communities <sup>39</sup>. In a cross-habitat study, salinity and pH have been  
187 suggested to influence bacterial and archaeal community compositions, respectively <sup>40,41</sup>.  
188 In a different habitat, the human body, it has been suggested that the microbial

189 community composition varies systematically across body habitats, individuals and time  
190 <sup>42</sup>.

191 For multicellular organisms, modeling approaches to predict global distributions of  
192 marine species have been applied in projects such as AquaMaps <sup>43</sup>. Combination of such  
193 efforts with the potential of COI to unveil historical processes may successfully be  
194 applied in determining factors responsible for the contemporary geographic distributions  
195 of these organisms <sup>44</sup>.

196 Unfortunately, only a few of these large-scale environmental surveys of biodiversity and  
197 biogeography have relied on *existing* marker gene sequence data sets found in the public  
198 databases <sup>36,40,45</sup>. Mainly due to the lack of specific guidelines, most marker gene  
199 sequences in databases are sparsely annotated with the information that would be  
200 required to underpin data integration, comparative studies, and knowledge generation.  
201 Even with complex keyword searches, it is currently impossible to reliably retrieve  
202 marker gene sequences that have originated from certain environments or particular  
203 locations on Earth; for example, all sequences from ‘soil’ or ‘freshwater lakes’ in a  
204 certain region of the world.

205 With appropriate sequence and contextual data integration and analysis new potential  
206 explanations for observed distribution and abundance patterns of microorganisms can be  
207 unraveled. For example, in human health and the study of epidemiology, it would also be  
208 desirable to have additional contextual data to help monitor the origins and regional  
209 spreading of pandemics <sup>46</sup> and study the variation of the human microbiota <sup>47-49</sup>.

210 Combining clinical and environmental datasets could provide new insight into where the  
211 trillions of bacteria that inhabit our body come from, and could help predict new

212 outbreaks of disease or assist in understanding the normal ecology of occasional  
213 pathogens. Already known correlations of some microbial taxa in different environments,  
214 such as depth in the marine environment <sup>50,51</sup>, and pH in the soil environment <sup>52</sup>, can be  
215 extended further. Finally, micro- and macro-organismal taxonomic knowledge can be  
216 greatly enhanced with the preservation of ‘voucher specimens’, which serve as the basis  
217 of study and as a reference. These may be cultures, tissue lines, DNA, or even images,  
218 depending on the organisms and the traits involved. The literature is filled with  
219 discoveries that could not be validated due to lack of vouchers.

220

### 221 **The MIENS Specification**

222 Few of the publicly available marker gene datasets contain contextual information about  
223 the environment such as geographic location, sampling time, habitat, or about  
224 experimental procedures used to obtain the DNA sequences. Such information may or  
225 may not be available in associated publications but the ‘costs’ in terms of time and energy  
226 to collect this by hand or with semi-automated systems from the literature are prohibitive  
227 <sup>53</sup>. Public databases of the International Nucleotide Sequence Database Collaboration  
228 (INSDC; comprising of DDBJ, ENA, and GenBank; <http://www.insdc.org>) depend on  
229 information submitted by authors to enrich the value of these sequences. We argue that  
230 the only way to change the current practice is to establish a standard of reporting that  
231 requires contextual data to be deposited at the time of sequence submission <sup>3</sup>. The  
232 adoption of such a standard would elevate the quality, accessibility, and utility of  
233 information that can be collected from INSDC.

234 Here we present a reporting guideline for marker genes (MIENS: Minimum Information

235 about an ENvironmental Sequence), which is based on the “Minimum Information about  
236 a (Meta) Genome Sequence” (MIGS/MIMS) specification issued by the Genomic  
237 Standards Consortium (GSC) <sup>54</sup>. Since its proposal at the sixth GSC meeting in 2008 <sup>55</sup>,  
238 the consortium has been working to build a consensus on an ideal and minimum set of  
239 contextual data that should be reported for marker genes retrieved from the environment.  
240 The proposed MIENS standard (Table 1) extends the MIGS/MIMS specification for  
241 genomes and metagenomes by adding two new report types, a “MIENS-survey” and a  
242 “MIENS-culture”, the former being the checklist of choice for uncultured diversity  
243 marker gene surveys, the latter designed for marker gene sequences obtained from  
244 cultured organisms or any material identifiable via voucher specimens.

245 A specific focus of the extended requirements is the sets of measurements and  
246 observations describing particular habitats, termed ‘environmental packages’.

247 The MIENS checklist adopts and incorporates the standards being developed by the  
248 Consortium for the Barcode of Life (CBOL) ([http://www.barcoding.si.edu/PDF/  
249 DWG\\_data\\_standards-Final.pdf](http://www.barcoding.si.edu/PDF/DWG_data_standards-Final.pdf)). Therefore, the specification can be universally applied  
250 to any marker gene, from SSU rRNA to COI, to cultured and uncultured organisms, to all  
251 taxa and to studies ranging from single individuals to complex communities.

252 The MIENS checklist was developed by collating information from several sources and  
253 evaluating it in the framework of the existing MIGS/MIMS specification. These include  
254 four independent community-led surveys, examination of the parameters reported in  
255 published studies and examination of compliance with optional features in INSDC  
256 documents. The overall goal of these activities was to design the backbone of MIENS  
257 specification that describes the most important aspects of marker gene contextual data,

258 and that would encourage users to deposit this contextual data in a standardized fashion.

259

260 ***Results of community-led surveys***

261 Community surveys are an excellent way to determine researcher preferences for core  
262 descriptors. To date, there have been four online surveys about descriptors for marker  
263 genes. In the same manner as the Department of Energy Joint Genome Institute's (DOE-  
264 JGI) user survey focusing on general descriptor contextual data for marker genes in 2005,  
265 the Ribosomal Database Project (RDP) <sup>56,57</sup>, SILVA <sup>58</sup> and the Terragenome Consortium  
266 (<http://www.terragenome.org>) conducted three more user surveys focusing on prevalent  
267 habitats for rRNA gene surveys, general descriptor contextual data for rRNA gene  
268 sequences and soil metagenome project contextual data, respectively (supplementary  
269 information 1). Additionally, following a special session during the 2005 International  
270 Census of Marine Microbes (ICoMM), an extensive set of contextual data items were  
271 selected, and were analyzed along with survey results.

272 These results of these user surveys provided valuable insights into community requests  
273 for contextual data items to be included in the MIENS specification and main habitats  
274 constituting the environmental packages.

275

276 ***Survey of published parameters***

277 We reviewed published rRNA gene studies, retrieved via SILVA and the ICoMM  
278 database MICROBIS (<http://icomm.mbl.edu/microbis>) to further supplement contextual  
279 data items that are included in the respective environmental packages. In total, thirty-nine  
280 publications from SILVA; including twenty-three publications with more than 500

281 sequences, and thirteen others retrieved with habitat-specific study queries; and over 40  
282 ICoMM projects were scanned for contextual data items to constitute the core of the  
283 environmental package sub-tables (supplementary information 1).

284

#### 285 *Survey of INSDC source feature qualifiers*

286 As a final analysis step, we surveyed usage statistics of INSDC source feature key  
287 qualifier values of rRNA gene sequences contained in SILVA (supplementary  
288 information 1). Most striking of these results is that <10% of the 1.2 million 16S rRNA  
289 gene sequences (SILVA release 100) were associated with even basic information such as  
290 latitude/longitude, collection date or PCR primers.

291

#### 292 *The MIENS checklist in full*

293 The MIENS specification provides users with an ‘electronic laboratory notebook’  
294 containing core contextual data items required for consistent reporting of marker gene  
295 investigations. A number of experts in a wide array of topics, guided by a solid  
296 rationalization procedure at each step along the way, led the development of these  
297 contextual data items.

298 Project details are hosted in the ‘Investigation’ section of MIENS, facilitating access to  
299 the outline of contextual data of a marker gene survey. The ‘Environment’ section  
300 provides the geospatial, temporal and environmental context. Fourteen ‘environmental-  
301 packages’ were developed, with the assistance from user surveys, publication reviews and  
302 expert communities working on their respective environments, and were integrated into  
303 the ‘MIMS/MIENS extension’ section. These packages provide a wealth of

304 environmental and epidemiological contextual data fields for a complete description of  
305 sampling environments (supplementary information 2). Researchers within The Human  
306 Microbiome Project <sup>59</sup> contributed the host associated and all human packages. The  
307 Terragenome Consortium contributed sediment and soil packages. Finally, ICoMM,  
308 Microbial Inventory Research Across Diverse Aquatic Long Term Ecological Research  
309 Sites (MIRADA-LTERS), and the Max Planck Institute for Marine Microbiology  
310 contributed the water package. The MIENS working group developed the remaining  
311 packages (air, microbial mat/biofilm, miscellaneous natural or artificial environment,  
312 plant-associated, and wastewater/sludge). The package names describe high-level habitat  
313 terms in order to be exhaustive. The miscellaneous natural or artificial environment  
314 package contains a generic set of parameters, and is included for any other habitat that  
315 does not fall into the other thirteen categories. Whenever needed, multiple packages may  
316 be used for the description of the environment.

317 The MIGS/MIMS specifications are applicable to MIENS with respect to the nucleic acid  
318 sequence source and sequencing contextual data, but have been complemented with  
319 further experimental contextual data such as PCR primers and conditions, or target  
320 gene/locus.

321 For clarity and ease of use, all items within the MIENS specification are presented with a  
322 value syntax description, as well as a clear definition of the item. Whenever terms from a  
323 specific ontology are required as the value of an item, these terms can be readily found in  
324 the respective ontology browsers, which are linked by URLs in the item definition.  
325 Although this version of the MIENS specification does not contain unit specifications, we  
326 recommend all units to be chosen from and follow the International System of Units (SI)

327 recommendations. In addition, we strongly urge the community to provide feedback  
328 regarding the best unit recommendations for given parameters. To facilitate comparative  
329 studies, unit standardization across data sets will be vital in future versions of MIENS.

330

### 331 *Accessing the MIGS/MIMS/MIENS checklists*

332 The MIGS/MIMS/MIENS checklists are maintained in a relational database system on  
333 behalf of the GSC community. This provides a secure and stable mechanism for updating  
334 the checklist suite and versioning. An excel version of the checklist is also provided to  
335 the community on the GSC web site at: [http://gensc.org/gc\\_wiki/index.php/MIENS](http://gensc.org/gc_wiki/index.php/MIENS). The  
336 checklist is updated on the GSC web site as development work is carried out on the  
337 database end. In the future, we plan to develop programmatic access to this database in  
338 order to allow automatic retrieval of the latest version of each checklist for INSDC  
339 databases and for GSC community resources.

340

### 341 **MIENS Adoption by Major Database and Informatics Resources**

342 A variety of efforts are under way to aid sequence submitters in compliance. In the past,  
343 the INSDC has issued a reserved 'BARCODE' keyword for the Consortium for the  
344 Barcoding of Life (CBOL) <sup>60,61</sup>. Following this model, the INSDC has recently  
345 recognized the GSC as an authority for the MIGS/MIMS/MIENS standards and issued it  
346 with an official keyword within INSDC nucleotide sequence records <sup>62</sup>. This greatly  
347 facilitates automatic validation of the submitted contextual data and provides support for  
348 datasets compliant with previous versions by including the checklist version in the  
349 keyword.



350 GenBank accepts MIENS metadata in tabular format using the sequin and tbl2asn  
351 submission tools, validates MIENS compliance, and reports the MIENS fields in the  
352 structured comment block. The ENA Webin submission system provides prepared web  
353 forms for the submission of MIENS compliant data; it presents all of the appropriate  
354 fields with descriptions, explanations and examples, in addition to validation of the data  
355 entered in the forms. An example which can aid in submission via Sequin or Webin  
356 systems is MetaBar<sup>63</sup>; a spreadsheet and web-based software tool designed to assist users  
357 in the consistent acquisition, electronic storage and submission of contextual data  
358 associated with their samples in compliance with the MIGS/MIMS/MIENS  
359 specifications.

360 The next-generation Sequence Read Archives (SRA) collects and displays MIENS  
361 compliant metadata in the sample and experiment objects. There are several tools that are  
362 already available or under development to assist users in SRA and ERA submissions. The  
363 myRDP SRA PrepKit, allows users to prepare and edit their submissions of reads  
364 generated from ultra-high-throughput sequencing technologies. A set of suggested  
365 attributes in the data forms assist researchers in providing metadata conforming to the  
366 MIMS and MIENS specifications. The Investigation/Study/Assay (ISA) Infrastructure is  
367 a flexible, freely available software suite that assists in the curation, reporting, and local  
368 management of experimental metadata from studies employing one or a combination of  
369 technologies, including high-throughput sequencing. Specific ISA configurations  
370 (available from [http://gensc.org/gc\\_wiki/index.php/Adopters#ISA\\_infrastructure](http://gensc.org/gc_wiki/index.php/Adopters#ISA_infrastructure)) have  
371 been developed to ensure MIENS compliance by providing templates and validation  
372 capability while another tool, ISAconverter, produces SRA.xml documents, thereby

373 facilitating submission to ERA and SRA repositories<sup>64</sup>.  
374 The SILVA, RDP, Greengenes and the ICoMM resources have participated in the  
375 development of MIENS, and are now taking the standardization one step further by  
376 establishing tools and resources to aid in compliance.  
377 Further detailed guidance for submission processes can be found under the respective  
378 wiki pages ([http://gensc.org/gc\\_wiki/index.php/MIENS](http://gensc.org/gc_wiki/index.php/MIENS)) of the MIENS standard.

379

### 380 *Examples of MIENS compliant datasets*

381 Several MIENS compliant reports are included in the supplementary information 3.  
382 These include; a 16S rRNA gene survey from samples obtained in the North Atlantic, an  
383 18S pyrotag study of anaerobic protists in permanently anoxic basin of the North Sea, a  
384 *pmoA* survey from desert soils of Negev Desert, Israel, a *dsrAB* survey from marine  
385 sediments from the Gulf of Mexico, and finally a 16S pyrotag study of bacterial diversity  
386 in the Western English Channel (publicly accessible via SRA study accession number  
387 SRP001108). Two further MIENS compliant 16S submissions are available in INSDC  
388 under the accession numbers GU949561.1 and GU949562.1.

389

### 390 *MIENS – a ‘living standard’*

391 MIENS, as well as MIGS/MIMS, are ‘living checklists’ and not final specifications.  
392 Therefore, further developments, extensions, and enhancements will be recognized, and  
393 improved versions of the checklists, if necessitated, will be released annually, while  
394 preserving the validity of former versions. A public ticketing system will be set up to  
395 track changes and feature requests. The final decisions about their implementation will be

396 done by the MIENS working group.

### 397 **Conclusions and Call for Action**

398 The GSC ([www.gensc.org](http://www.gensc.org)) is an international working body with a stated mission of  
399 working towards richer descriptions of our complete collection of genomes and  
400 metagenomes. With the development of the MIENS specification, this mission has been  
401 extended to marker gene sequences as well. The GSC is an open initiative that welcomes  
402 the participation of the wider community. This includes an open call to contribute to  
403 refinements of the MIENS specification or its implementation.

404 The adoption of the MIENS standard by major data providers and organizations as well  
405 as the three primary public sequence data repositories (INSDC) with an active poll for  
406 MIENS compliant data underlines and seconds the efforts to contextually enrich our  
407 marker gene collection, and complements the recent efforts to contextually enrich other  
408 (meta) omics data. The MIENS checklist has been developed to the point that it is ready  
409 to be used in the publication of sequences. A defined procedure for requesting new  
410 features and the stable release cycles will facilitate implementation of the standard across  
411 the community. Widespread compliance among authors, adoption by journals and use by  
412 informatics resources will vastly improve our collective ability to mine and integrate  
413 invaluable sequence data collections for knowledge and application driven research. In  
414 particular, the ability to combine microbial community samples collected from any  
415 source, using the universal Tree of Life as a yardstick to compare even the most diverse  
416 communities, should provide new insights into the dynamic spatial and temporal  
417 distribution of microbial life on our planet and even on our own bodies.

418

419 **References**

- 420 1 Community cleverness required. *Nature* **455**, 1-1 (2008).
- 421 2 Field, D. *et al.*, 'Omics Data Sharing. *Science* **326**, 234-236 (2009).
- 422 3 Taylor, C. F. *et al.*, Promoting coherent minimum reporting guidelines for  
423 biological and biomedical investigations: the MIBBI project. *Nat Biotechnol* **26**, 889-896  
424 (2008).
- 425 4 Woese, C. R. and Fox, E., Phylogenetic structure of the prokaryotic domain: the  
426 primary kingdoms. *Proc Nat Acad Sci USA* **74**, 5088-5090 (1977).
- 427 5 Woese, C. R., Kandler, O., and Wheelis, M. L., Towards a natural system of  
428 organisms: proposal for the domains *Archaea*, *Bacteria*, and *Eucarya*. *Proc Nat Acad Sci*  
429 *USA* **87**, 4576-4579 (1990).
- 430 6 Amann, R. I., Ludwig, W., and Schleifer, K. H., Phylogenetic identification and  
431 in-situ detection of individual microbial cells without cultivation. *Microbiol Rev* **59**, 143-  
432 169 (1995).
- 433 7 Curtis, T. P., Sloan, W. T., and Scannell, J. W., Estimating prokaryotic diversity  
434 and its limits. *Proc Nat Acad Sci USA* **99**, 10494-10499 (2002).
- 435 8 Turrioni, F. *et al.*, Human gut microbiota and bifidobacteria: from composition to  
436 functionality. *Antonie van Leeuwenhoek* **94**, 35-50 (2008).
- 437 9 Ludwig, W. *et al.*, Bacterial phylogeny based on comparative sequence analysis.  
438 *Electrophoresis* **19**, 554-568 (1998).
- 439 10 Ludwig, W. and Schleifer, K. H., in *Microbial phylogeny and evolution, concepts*  
440 *and controversies*, edited by J. Sapp (Oxford university press, New York, 2005), pp. 70-  
441 98.
- 442 11 Ciccarelli, F. D. *et al.*, Toward automatic reconstruction of a highly resolved tree  
443 of life. *Science* **311**, 1283-1287 (2006).
- 444 12 Teeling, H. and Glöckner, F. O., RibAlign: a software tool and database for  
445 eubacterial phylogeny based on concatenated ribosomal protein subunits. *BMC*  
446 *Bioinformatics* **7** (2006).
- 447 13 Stahl, D. A., Analysis of hydrothermal vent associated symbionts by ribosomal  
448 RNA sequences. *Science* **224**, 409-411 (1984).
- 449 14 Pace, N. R., Stahl, D. A., Olsen, G. J., and Lane, D. J., Analyzing natural

450 microbial populations by rRNA sequences. *ASM News* **51**, 4-12 (1985).

451 15 Olsen, G. J. *et al.*, Microbial ecology and evolution: a ribosomal RNA approach.  
452 *Annu Rev Microbiol* **40**, 337-365 (1986).

453 16 Giovannoni, S. J., Britschgi, T. B., Moyer, C. L., and Field, K. G., Genetic  
454 diversity in Sargasso Sea bacterioplankton. *Nature* **345**, 60-63 (1990).

455 17 Ward, D. M., Weller, R., and Bateson, M. M., 16S rRNA sequences reveal  
456 numerous uncultured microorganisms in a natural community. *Nature* **345**, 63-65 (1990).

457 18 López-García, P., López-López, A., Moreira, D., and Rodríguez-Valera, F.,  
458 Diversity of free-living prokaryotes from a deep-sea site at the Antarctic Polar Front.  
459 *Fems Microbiol Ecol* **36**, 193-202 (2001).

460 19 Moon-van der Staay, S. Y., De Wachter, R., and Vaultot, D., Oceanic 18S rDNA  
461 sequences from picoplankton reveal unsuspected eukaryotic diversity. *Nature* **409**, 607-  
462 610 (2001).

463 20 DeLong, E. F., *Archaea* in coastal marine environments. *Proc Nat Acad Sci USA*  
464 **89**, 5685-5689 (1992).

465 21 Pace, N. R., A molecular view of microbial diversity and the biosphere. *Science*  
466 **276**, 734-740 (1997).

467 22 Huber, J. A., Butterfield, D. A., and Baross, J. A., Temporal changes in archaeal  
468 diversity and chemistry in a mid-ocean ridge seafloor habitat. *Appl Environ Microbiol*  
469 **68**, 1585-1594 (2002).

470 23 Rappe, M. S. and Giovannoni, S. J., The uncultured microbial majority. *Annu Rev*  
471 *Microbiol* **57**, 369-394 (2003).

472 24 Hewson, I. and Fuhrman, J. A., Richness and diversity of bacterioplankton species  
473 along an estuarine gradient in Moreton Bay, Australia. *Appl Environ Microbiol* **70**, 3425-  
474 3433 (2004).

475 25 Doolittle, W. F., Fun With Genealogy. *Proc Nat Acad Sci USA* **94**, 12751-12753  
476 (1997).

477 26 Huynen, M. A. and Bork, P., Measuring genome evolution. *Proc Nat Acad Sci*  
478 *USA* **95**, 5849-5856 (1998).

479 27 Ivars-Martinez, E. *et al.*, Biogeography of the ubiquitous marine bacterium  
480 *Alteromonas macleodii* determined by multilocus sequence analysis. *Mol Ecol* **17**, 4092-

481 4106 (2008).

482 28 Cole, J. R., Konstantinidis, K., Farris, R. J., and Tiedje, J. M., in *Environmental*  
483 *Molecular Microbiology*, edited by W.-T. Liu and J.K. Jansson (Caister Academic Press  
484 UK, 2010), pp. 1-19.

485 29 Zehr, J. P., Mellon, M. T., and Zani, S., New nitrogen-fixing microorganisms  
486 detected in oligotrophic oceans by amplification of nitrogenase (nifH) genes. *Appl*  
487 *Environ Microbiol* **64**, 3444-3450 (1998).

488 30 Francis, C. A., Beman, J. M., and Kuypers, M. M. M., New processes and players  
489 in the nitrogen cycle: the microbial ecology of anaerobic and archaeal ammonia  
490 oxidation. *Isme J* **1**, 19-27 (2007).

491 31 Minz, D. *et al.*, Diversity of sulfate-reducing bacteria in oxic and anoxic regions  
492 of a microbial mat characterized by comparative analysis of dissimilatory sulfite  
493 reductase genes. *Appl Environ Microbiol* **65**, 4666-4671 (1999).

494 32 Thomas, S. *et al.*, Evidence for phosphonate usage in the coral holobiont. *Isme J*  
495 (2009).

496 33 Gilbert, J. A. *et al.*, Potential for phosphonoacetate utilization by marine bacteria  
497 in temperate coastal waters. *Environ Microbiol* **11**, 111-125 (2009).

498 34 Martinez, A., W. Tyson, G., and DeLong, E., F., Widespread known and novel  
499 phosphonate utilization pathways in marine bacteria revealed by functional screening and  
500 metagenomic analyses. *Environ Microbiol* **9999** (2009).

501 35 Hebert, P. D. N., Cywinska, A., Ball, S. L., and Dewaard, J. R., Biological  
502 identifications through DNA barcodes. *Proc R Soc Lond B Biol Sci* **270**, 313-321 (2003).

503 36 von Mering, C. *et al.*, Quantitative phylogenetic assessment of microbial  
504 communities in diverse environments. *Science* **315**, 1126-1130 (2007).

505 37 Pommier, T. *et al.*, Global patterns of diversity and community structure in  
506 marine bacterioplankton. *Mol Ecol* **16**, 867-880 (2007).

507 38 Fuhrman, J. A. *et al.*, A latitudinal diversity gradient in planktonic marine  
508 bacteria. *Proc Nat Acad Sci USA* **105**, 7774-7778 (2008).

509 39 Gilbert, J., A. *et al.*, The seasonal structure of microbial communities in the  
510 Western English Channel. *Environ Microbiol* **11**, 3132-3139 (2009).

511 40 Lozupone, C. A. and Knight, R., Global patterns in bacterial diversity. *Proc Nat*

512 *Acad Sci USA* **104**, 11436-11440 (2007).

513 41 Auguet, J.-C., Barberan, A., and Casamayor, E. O., Global ecological patterns in  
514 uncultured Archaea. *Isme J* **4**, 182-190 (2010).

515 42 Costello, E. K. *et al.*, Bacterial community variation in human body habitats  
516 across space and time. *Science* **326**, 1694-1697 (2009).

517 43 Kaschner, K. *et al.*, AquaMaps: Predicted range maps for aquatic species,  
518 Available at <http://www.aquamaps.org/>, (2008).

519 44 Workshops Report and Recommendations DNA Barcoding of Marine  
520 Biodiversity (MarBOL) presented at the MarBOL Workshops, 2009 (unpublished).

521 45 Tamames, J. *et al.*, Environmental distribution of prokaryotic taxa. *BMC*  
522 *Microbiology* **10**, 85.

523 46 Schriml, L. M. *et al.*, GeMInA, Genomic Metadata for Infectious Agents, a  
524 geospatial surveillance pathogen database. *Nucl Acids Res* **38**, D754-D764 (2010).

525 47 Palmer, C. *et al.*, Development of the Human Infant Intestinal Microbiota. *PLoS*  
526 *Biol* **5**, e177 (2007).

527 48 Ravel, J. *et al.*, Vaginal microbiome of reproductive-age women. *Proc Nat Acad*  
528 *Sci USA e-pub ahead of print* (2010).

529 49 Qin, J. *et al.*, A human gut microbial gene catalogue established by metagenomic  
530 sequencing. *Nature* **464**, 59-65 (2010).

531 50 DeLong, E. F. *et al.*, Community genomics among stratified microbial  
532 assemblages in the ocean's interior. *Science* **311**, 496-503 (2006).

533 51 Moreira, D., Rodriguez-Valera, F., and Lopez-Garcia, P., Metagenomic analysis  
534 of mesopelagic Antarctic plankton reveals a novel deltaproteobacterial group.  
535 *Microbiology* **152**, 505-517 (2006).

536 52 Lauber, C. L., Hamady, M., Knight, R., and Fierer, N., Soil pH as a predictor of  
537 soil bacterial community structure at the continental scale: a pyrosequencing-based  
538 assessment. *Appl Environ Microbiol* **75**, 5111-5120 (2009).

539 53 Hirschman, L. *et al.*, Habitat-Lite: a GSC case study based on free text terms for  
540 environmental metadata. *OMICS* **12**, 129-136 (2008).

541 54 Field, D. *et al.*, The minimum information about a genome sequence (MIGS)  
542 specification. *Nat Biotechnol* **26**, 541-547 (2008).

543 55 Field, D. *et al.*, Meeting reports from the Genomic Standards Consortium (GSC)  
544 Workshops 6 and 7. *SIGS* **1**, 68-71 (2009).

545 56 Cole, J. R. *et al.*, The ribosomal database project (RDP-II): introducing myRDP  
546 space and quality controlled public data. *Nucl Acids Res* **35**, D169-172 (2007).

547 57 Cole, J. R. *et al.*, The Ribosomal Database Project: improved alignments and new  
548 tools for rRNA analysis. *Nucl Acids Res* **37**, D141-145 (2009).

549 58 Pruesse, E. *et al.*, SILVA: a comprehensive online resource for quality checked  
550 and aligned ribosomal RNA sequence data compatible with ARB. *Nucl Acids Res* **35**,  
551 7188-7196 (2007).

552 59 Turnbaugh, P. J. *et al.*, The Human Microbiome Project. *Nature* **449**, 804-810  
553 (2007).

554 60 Benson, D. A. *et al.*, GenBank. *Nucl. Acids Res.* **35**, D21-25 (2007).

555 61 Benson, D. A. *et al.*, GenBank. *Nucl. Acids Res.* **36**, D25-30 (2008).

556 62 Lynette, H. *et al.*, Meeting report: Metagenomics, Metadata and Meta-analysis”  
557 (M3) Workshop at the Pacific Symposium on Biocomputing 2010. *SIGS* **2**, 357-360  
558 (2010).

559 63 Hankeln, W. *et al.*, MetaBar - a tool for consistent contextual data acquisition and  
560 standards compliant submission. *BMC Bioinformatics* **11**, 358 (2010).

561 64 Rocca-Serra, P. *et al.*, ISA infrastructure: supporting standards-compliant  
562 experimental reporting and enabling curation at the community level. *Bioinformatics* **in**  
563 **press** (2010).

564



		Report type	
		MIENS survey	MIENS culture
<b>Investigation</b>			
Submitted to INSDC <sup>[boolean]</sup>	Depending on the study (large-scale e.g. done with next generation sequencing technology, or small-scale) sequences have to be submitted to SRA (Short Read Archives), ENA (European Nucleotide Archive), DRA (DDBJ Read Archive) or via the classical Webin/Sequin systems to Genbank, ENA and DDBJ	M	M
Investigation type <sup>[survey or culture]</sup>	Nucleic Acid Sequence Report is the root element of all MIENS compliant reports as standardized by Genomic Standards Consortium (GSC). This field is either MIENS survey or MIENS culture	M	M
Project name	Name of the project within which the sequencing was organized	M	M
<b>Environment</b>			
Geographic location (latitude and longitude <sup>[float, point, transect and region]</sup> )	The geographical origin of the sample as defined by latitude and longitude. The values should be reported in decimal degrees and in WGS84 system	M	M
Geographic location (depth <sup>[integer, point, interval, unit]</sup> )	Please refer to the definitions of depth in the environmental packages	E	E
Geographic location (elevation of site <sup>[integer, unit]</sup> , altitude of sample <sup>[integer, unit]</sup> )	Please refer to the definitions of either altitude or elevation in the environmental packages	E	E
Geographic location (country and/or sea <sup>[INSDC or GAZ]</sup> ; region <sup>[GAZ]</sup> )	The geographical origin of the sample as defined by the country or sea name. Country, sea, or region names should be chosen from the INSDC list ( <a href="http://insdc.org/country.html">http://insdc.org/country.html</a> ), or the GAZ ontology ( <a href="http://biportal.bioontology.org/visualize/40651">http://biportal.bioontology.org/visualize/40651</a> )	M	M
Collection date <sup>[ISO8601]</sup>	The time of sampling, either as an instance (single point in time) or interval. In case no exact time is available, the date/time can be right truncated i.e. all of these are valid times: 2008-01-23T19:23:10+00:00; 2008-01-23T19:23:10; 2008-01-23; 2008-01; 2008; Except: 2008-01; 2008 all are ISO6801 compliant	M	M
Environment (biome <sup>[EnvO]</sup> )	In environmental biome level are the major classes of ecologically similar communities of plants, animals, and other organisms. Biomes are defined based on factors such as plant structures, leaf types, plant spacing, and other factors like climate. Examples include: desert, taiga, deciduous woodland, or coral reef. EnvO terms listed under environmental biome can be found from the link: <a href="http://www.ebi.ac.uk/ontology-lookup/browse.do?ontName=ENVO&amp;termId=ENVO%3A00000428&amp;termName=biome">http://www.ebi.ac.uk/ontology-lookup/browse.do?ontName=ENVO&amp;termId=ENVO%3A00000428&amp;termName=biome</a>	M	M
Environment (feature <sup>[EnvO]</sup> )	Environmental feature level includes geographic environmental features. Examples include: harbor, cliff, or lake. EnvO terms listed under environmental feature can be found from the link: <a href="http://www.ebi.ac.uk/ontology-lookup/browse.do?ontName=ENVO&amp;termId=ENVO%3A00002297&amp;termName=environmental%20feature">http://www.ebi.ac.uk/ontology-lookup/browse.do?ontName=ENVO&amp;termId=ENVO%3A00002297&amp;termName=environmental%20feature</a>	M	M

Environment (material <sup>[EnvO]</sup> )	The environmental material level refers to the matter that was displaced by the sample, prior to the sampling event. Environmental matter terms are generally mass nouns. Examples include: air, soil, or water. EnvO terms listed under environmental matter can be found from the link: <a href="http://www.ebi.ac.uk/ontology-lookup/browse.do?ontName=ENVO&amp;termId=ENVO%3A00010483&amp;termName=environmental%20matter">http://www.ebi.ac.uk/ontology-lookup/browse.do?ontName=ENVO&amp;termId=ENVO%3A00010483&amp;termName=environmental%20matter</a>	M	M
<b>MIGS/MIMS/MIENS Extension</b>			
Environmental package <sup>[air, host-associated, human-associated, human-skin, human-oral, human-gut, human-vaginal, microbial mat/biofilm, miscellaneous natural or artificial environment, plant-associated, sediment, soil, wastewater/sludge, water]</sup>	MIGS/MIMS/MIENS extension for reporting of measurements and observations obtained from one or more of the environments where the sample was obtained. All environmental packages listed here are further defined in separate subtables. By giving the name of the environmental package, a selection of fields can be made from the subtables and can be reported	M	M
<b>Nucleic acid sequence source</b>			
Isolation and growth conditions <sup>[PMID, DOI, or url]</sup>	Publication reference in the form of pubmed ID (PMID), digital object identifier (DOI), or url for Isolation and growth condition specifications of the organism/material	-	M
<b>Sequencing</b>			
Target gene or locus (e.g. 16S rRNA, 18S rRNA, nif, amoA, rpo, V6, ITS)	Targeted gene, locus or gene region name for marker gene study	M	M
Sequencing method (e.g. dideoxysequencing, pyrosequencing, polony) <sup>[OBI]</sup>	Sequencing method used; e.g. Sanger, pyrosequencing, ABI-solid. This field accepts OBI terms, for a browser of OBI terms please see <a href="http://biportal.bioontology.org/visualize/40547">http://biportal.bioontology.org/visualize/40547</a>	M	M

**Table 1. Items for the MIENS specification and their mandatory (M), conditionally mandatory (C) (the item is mandatory only when applicable to the study) or recommended (X) status for both MIENS-survey and MIENS-culture checklists. ‘-‘ denotes that an item is not applicable for a given checklist. ‘E’ denotes that a field has environment-specific requirements. For example, while ‘depth’ is mandatory for environments water, sediment or soil; it is optional for human-associated environments. Item names are followed by a short description of the value of the item in parentheses and/or value type in brackets as a superscript. Whenever applicable, value types are chosen from a controlled vocabulary (CV) from the OBO foundry**

(<http://www.obofoundry.org>), EFO (ArrayExpress Environmental Factor Ontology), EnvO (Environment Ontology), CABRI (Common Access to Biological Resources and Information), OBI (Ontology for Biomedical Investigations). Items that are mandatory or conditionally mandatory for both MIENS report types are indicated by shaded rows and constitute the core of the MIENS specification. This table only presents the very core of MIENS checklists, i.e. only mandatory items for each checklist. Supplementary information 2 in spreadsheet format contains all MIENS items, the tables for environmental packages in the MIMS/MIENS extension, and GenBank structured comment name that should be used for submitting MIENS data to GenBank.