Cannabidiol Modulates Fear Memory Formation Through Interactions with Serotonergic Transmission in the Mesolimbic System

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Emerging evidence suggests that the largest phytochemical component of cannabis, cannabidiol (CBD), may possess pharmacotherapeutic properties in the treatment of neuropsychiatric disorders. CBD has been reported to functionally interact with both the mesolimbic dopamine (DA) and serotonergic (5-HT) receptor systems. However, the underlying mechanisms by which CBD may modulate emotional processing are not currently understood. Using a combination of *in vivo* electrophysiological recording and fear conditioning in rats, the present study aimed to characterize the behavioral, neuroanatomical, and pharmacological effects of CBD within the mesolimbic pathway, and its possible functional interactions with 5-HT and DAergic transmission. Using targeted microinfusions of CBD into the shell region of the mesolimbic nucleus accumbens (NASH), we report that intra-NASH CBD potently blocks the formation of conditioned freezing behaviors. These effects were challenged with DAergic, cannabinoid CB1 receptor, and serotonergic (5-HT_{1A}) transmission blockade, but only 5-HT_{1A} blockade restored associative conditioned freezing behaviors. *In vivo* intra-ventral tegmental area (VTA) electrophysiological recordings revealed that behaviorally effective doses of intra-NASH CBD elicited a predominant decrease in spontaneous DAergic neuronal frequency and bursting activity. These neuronal effects were reversed by simultaneous blockade of 5-HT_{1A} receptor transmission. Finally, using a functional contralateral disconnection procedure, we demonstrated that the ability of intra-NASH CBD to block the formation of conditioned freezing behaviors was dependent on intra-VTA GABAergic transmission substrates. Our findings demonstrate a novel NAc → VTA circuit responsible for the behavioral and neuronal effects of CBD within the mesolimbic system via functional interactions with serotonergic 5-HT_{1A} receptor signaling.

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INTRODUCTION

There is ongoing debate regarding the potential therapeutic potential of cannabis-derived phytochemicals in the treatment of neuropsychiatric conditions. Central to this debate is emerging evidence demonstrating that separate phytochemical constituents of cannabis may possess differential pharmacological and psychotropic effects. Although Δ -9-tetrahydrocannabinol (THC) is considered the primary psychoactive component of marijuana, cannabis contains over 70 distinct compounds (ElSohly and Slade, 2005; Radwan *et al*, 2009). Of these, cannabidiol (CBD), which is

considered non-psychoactive, is the most abundant. In contrast to THC, CBD is an agonist at the 5HT_{1A} receptor (Russo *et al*, 2005) and decreases cellular reuptake and hydrolysis of the endocannabinoid anandamide, potentiating its central effects (Bisogno *et al*, 2001).

In terms of psychotropic profiles, THC is associated with transient and long-term psychotomimetic effects (Bhattacharyya *et al*, 2012; Murray *et al*, 2007), whereas clinical and preclinical research has shown that CBD can produce antipsychotic and anxiolytic effects (Crippa *et al*, 2011; Campos *et al*, 2012; Fogaca *et al*, 2014; Zuardi *et al*, 2006, 2012; Mechoulam *et al*, 2007; Leweke *et al*, 2012; Renard *et al*, 2016a; Schubart *et al*, 2014; Gomes *et al*, 2015). In addition, CBD may modulate emotional memory processing and decrease symptoms associated with emotional memory disorders such as post-traumatic stress disorder (PTSD; Blessing *et al*, 2015; Betthausen *et al*, 2015).

While the precise neuroanatomical regions responsible for CBD's actions are not known, considerable evidence implicates the nucleus accumbens (NAc) as an important site

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for CBD's modulatory effects on various cognitive and behavioral phenomena (Guimarães *et al*, 2004; Bhattacharyya *et al*, 2009; Valvassori *et al*, 2011; Mijangos-Moreno *et al*, 2014; Pedrazzi *et al*, 2015). For example, CBD attenuates THC-induced dysregulation of the ventral striatum during verbal recall tasks (Bhattacharyya *et al*, 2009) and increases c-fos and adenosine levels in rodent NAc (Guimarães *et al*, 2004; Mijangos-Moreno *et al*, 2014). In addition, CBD blocks amphetamine-induced oxidative stress in the NAc (Valvassori *et al*, 2011) and intra-NAc CBD attenuates amphetamine-induced deficits in prepulse inhibition (Pedrazzi *et al*, 2015; Renard *et al*, 2016b). Nevertheless, the precise functional and pharmacological mechanisms by which CBD may produce these effects in the mesolimbic system are not currently understood.

In the present study, we investigated the potential effects of CBD on fear-related memory formation and how CBD may modulate neuronal activity states within the mesolimbic circuitry. Using a combination of behavioral conditioning and *in vivo* neuronal electrophysiological recordings, we report that intra-NAc CBD dose-dependently blocks the formation of associative fear memories measured by conditioned freezing behaviors and blunts the activity of DAergic neurons in the ventral tegmental area (VTA). Furthermore, these effects were dependent on intra-shell region of the mesolimbic nucleus accumbens (NASh) serotonergic (5-HT_{1A}) receptor transmission and functional interactions between the NASh and GABAergic transmission in the VTA.

MATERIALS AND METHODS

Rats and Surgery

Male Sprague-Dawley rats (300–350 g; Charles River, Senneville, QC, Canada) were anesthetized with ketamine (80 mg/ml)–xylazine (6 mg/kg; intraperitoneally) and placed in a stereotaxic apparatus. An incision was made to expose the skull, and stainless steel guide cannulae (22 G; Plastics1) were implanted into the NASh using the following stereotaxic coordinates (12° angle, mm from bregma): anteroposterior (AP) +1.8, lateral (LAT) ± 2.6, and ventral (V) – 7.4 from the dural surface and the following coordinates for the VTA (10° angle, mm from bregma): AP – 5.0, LAT ± 2.6, and V – 8.0 from the dural surface. Coordinates were based on the Atlas of Paxinos and Watson (2005). Rats in the NASh-VTA disconnection groups received single unilateral NASh cannulation and contralateral VTA cannulation. All experimental procedures were performed in accordance with the regulations of the Canadian Council on animal care (CCAC) and the University of Western Ontario.

Drug Administration

The broad-spectrum DA receptor antagonist, α -flupenthixol (α -flu; Tocris), the GABA_A antagonist, bicuculline methiodide (Tocris), and the GABA_B antagonist, hydroxysaclofen (Tocris), were dissolved in physiological saline (pH adjusted to 7.4). The 5-HT_{1A} receptor antagonist, NAD 299 hydrochloride (Tocris), CBD (Tocris), and CB1R antagonist, SR141716A (rimonabant, RIM), were first dissolved in

dimethyl sulfoxide (DMSO) and then diluted in PBS for a final 1% DMSO in PBS vehicle (VEH) solution. Microinfusions were performed over 1 min through a Hamilton microsyringe. To ensure adequate diffusion, injector cannulae were left in place for an additional 1 min.

Olfactory Fear Conditioning

We used a previously described olfactory fear conditioning paradigm to measure fear memory formation (Lauzon *et al*, 2009; Draycott *et al*, 2014). Two distinct environments were used. Environment A was a 30 in × 30 in Plexiglass box with black spots on a white background and environment B was a 30 in × 30 in Plexiglass box with black and white stripes. The designated shock environment had a metallic grid shock floor, while the designated test environment had a smooth gray Plexiglas floor. The environments were assigned as 'shock' and 'test' in a counterbalanced manner such that all rats with 'shock' environment A were tested in environment B and all rats with 'shock' environment B were tested in environment A. On day 1 (habituation), rats were given sham microinfusions into the NASh and habituated to both conditioning environments for 30 min. Footshocks were delivered at an intensity of 0.8 mA (for 1 s), which we have previously reported to produce highly robust conditioned freezing behaviors (Lauzon *et al*, 2009; Draycott *et al*, 2014). On day 2 (conditioning phase), rats were returned to the room and, immediately before being placed in previously assigned 'shock' environment, rats received intra-NASh microinfusions of CBD (1, 10, and 100 ng/0.5 μ l); the DA antagonist α -flu (100 ng–1.0 μ g/0.5 μ l), alone or in combination with CBD; the 5-HT_{1A} antagonist NAD 299 hydrochloride (10–100 ng/0.5 μ l), alone or in combination with CBD (100 ng). Two odors were delivered during conditioning, almond and peppermint. One odor was presented with a footshock (CS+) and the other was presented in the absence of a footshock (CS–). After 1 min in the 'shock' environment, the CS– odor was presented for 20 s. After 2 min, the CS+ odor was presented for 19 s followed by a 1 s footshock delivered through the shock floor. This cycle was repeated five times. On day 3 (test phase), rats were placed in the previously assigned test environment. They were given 1 min to explore the environment before odor presentations began, during which time baseline levels of freezing behavior were recorded. Both CS+ and CS– odors were presented in a fully counterbalanced order for 5 min and amounts of time rat's spent freezing (lack of movement except for respiration) was recorded and analyzed with ANY-MAZE video software for offline analysis.

Footshock Sensitivity Tests

To ensure that intra-NASh infusions of CBD, NAD 299, or α -flu were having no effect on baseline sensitivity to footshock stimulation during fear memory acquisition, separate control groups received either bilateral intra-NASh microinfusions of VEH, CBD (100 ng/0.5 μ l), NAD 299 (500 ng/0.5 μ l), or α -flu (1000 ng/0.5 μ l) alone immediately before a sensitivity testing phase wherein rats received five test shock administrations (0.8 mA, 1 s, in the absence of any cue delivery), once per minute over a total of 5 min. During

this time, levels of freezing behavior, total distance traveled, and the number of jumping events were recorded with ANY-MAZE video software and analyzed offline.

In Vivo Electrophysiological Recordings

In vivo single-cell extracellular VTA recordings were performed as described previously (Loureiro *et al*, 2015; Tan *et al*, 2009). Briefly, rats were anesthetized with urethane (1.4 g/kg, intraperitoneally) and placed in a stereotaxic frame with body temperature maintained at 37 °C. A scalp incision was made and holes were drilled in the skull above the NASH and VTA. For intra-NASH microinfusion of CBD (100 ng/0.5 µl), a 1 µl Hamilton syringe was slowly lowered at the same coordinates used for behavioral studies. For intra-VTA extracellular recording, glass microelectrodes (with an average impedance of 6–8 MΩ) filled with a 2% Pontamine Sky Blue solution were lowered using a hydraulic micro-positioner (Kopf 640) at the following flat skull stereotaxic coordinates (in mm from bregma): AP – 5.3, LAT ± 0.7 from midline, ventral (V) – 7.0 to – 8.5 from the dural surface. Extracellular signals were amplified using a MultiClamp 700B amplifier (Molecular Devices) and recorded through a Digidata 1440A acquisition system (Molecular Devices) using the pClamp 10 software. Extracellular recordings were filtered at 1 kHz and sampled at 5 kHz. VTA DA neurons were identified according to well-established electrophysiological features (Ungless *et al*, 2004; Jalabert *et al*, 2011): (1) a relatively long action potential width (> 2.5 ms); (2) a slow spontaneous firing rate (2–5 Hz); (3) a biphasic waveform consisting of a notch on the rising phase followed by a delayed after potential; and (4) a single irregular or bursting firing pattern. VTA GABA interneurons were also characterized based on previously reported criteria: (1) a narrow action potential width (< 1 ms); (2) relatively fast firing rates (typically between 10 and 20 Hz); and (3) the absence of bursting activity. Neurons that failed to clearly meet the aforementioned criteria for VTA DA or VTA non-DA electrophysiological neuronal properties were excluded from postexperimental analyses. Recordings analyses were accomplished using the Clampfit 10 software. Response patterns of isolated VTA neurons following intra-NASH CBD microinfusions were determined by comparing neuronal frequency rates between the 5 min preinfusion *vs* postinfusion epochs. Classification of drug-infusion effects used a criterion of a ≥ 10% increase in firing frequency postinfusion to be classified as an increase; a ≤ 10% decrease to be classified as a decrease. Neurons showing firing frequency parameters within these cutoff points were classified as ‘no change’. For VTA DA neurons, we also analyzed the bursting rate and number of spikes within each burst. The onset of a burst was defined as the occurrence of two consecutive spikes with an interspike interval of < 80 ms. For histological analysis of extracellular VTA neuronal recording sites, recording electrode positions were marked with iontophoretic deposit of Pontamine Sky Blue dye (–20 µA, continuous current for 12–15 min). Rats were then perfused transcardially with isotonic saline followed by 10% formalin. Brains were removed and stored in a 25% sucrose-formalin solution before sectioning (40 µm sections) on a freezing cryostat. Following this, sections were stained with neutral red and

infusion and/or neuronal recording sites were confirmed with light microscopy.

NASH–VTA Functional Disconnection Studies

Previous research has established the functional connectivity between the NAc and the VTA, whereby GABAergic projections from the NAc are capable of modulating VTA DAergic and non-DAergic neuronal activity states (Nauta *et al*, 1978; Kalivas *et al*, 1993). To determine if the behavioral effects of intra-NASH CBD were due to modulatory influences from the NASH → VTA, we performed a functional disconnection procedure, similar to one described previously (Rosen *et al*, 2015). In this procedure, to disconnect the effects of intra-NASH CBD from intra-VTA GABAergic transmission, experimental groups had microinjector cannulae placed unilaterally in the NASH of one hemisphere, and the VTA of the contralateral hemisphere. Group 1 received intra-VTA VEH followed by intra-NASH VEH. Group 2 received intra-VTA VEH followed by intra-NASH CBD (100 ng/0.5 µl). Group 3 received intra-VTA microinfusion of the GABA_A receptor antagonist bicuculline methiodide (50 ng/0.5 µl) with the selective GABA_B antagonist hydroxysaclofen (100 ng/0.5 µl), followed by intra-NASH CBD (100 ng/0.5 µl). Group 4 received the intra-VTA bicuculline/hydroxysaclofen mixture followed by intra-NASH VEH. All groups received the intra-VTA treatment 2 min before intra-NASH treatments. The hemispheres for unilateral/contralateral cannulations were counter-balanced within groups to control for laterality. All groups were then subjected to the same olfactory fear conditioning paradigm described above.

Data Analysis

Data were analyzed with one- or two-way ANOVA, followed by *post hoc* paired-samples *t*-tests or Newman–Keuls tests, where appropriate.

RESULTS

Histological Analyses

Histological analysis revealed injector placements localized within the anatomical boundaries of the shell subdivision of the NAc or VTA, according to Paxinos and Watson (2005). Figure 1a presents a microphotograph displaying a typical injector placement within the NASH, relative to the ‘core’ subdivision. Figure 1b displays a schematic illustration showing representative intra-NASH cannulae placements along the rostral–caudal axis of the NAc. Black circles represent VEH control rats receiving for shows a microphotograph displaying typical bilateral intra-NASH injector placements.

Intra-NASH CBD Dose-Dependently Blocks the Formation of Associative Fear Memory

We first examined the potential effects of intra-NASH CBD on the acquisition of associative fear memory (see Materials and Methods). Using a wide dose range of bilateral intra-NASH CBD (1 ng–100 ng/0.5 µl), we challenged the

Figure 1 Histological analysis of intra-shell region of the mesolimbic nucleus accumbens (NASh) microinjection sites. (a) Microphotograph of a representative injector placement within the shell subdivision of the nucleus accumbens (NAc). (b) Schematic representation of select intra-NASh injector locations. ● = 100 ng cannabidiol (CBD) groups, ◆ = 100 ng CBD + 100 ng NAD299 group. (c) Microphotograph of representative intra-NASh bilateral cannulae placements (black arrows).

acquisition of fear memory by administering CBD immediately before the fear conditioning session. ANOVA comparing percentages of time spent freezing during testing revealed a significant main effect of group ($F_{(7,287)} = 5.75$, $p = 0.0001$). *Post hoc* analyses revealed that whereas rats treated with intra-NASh VEH ($n = 6$) and rats treated with a lower dose of CBD (1 ng/0.5 μ l, $n = 7$) spent significantly greater amounts of time freezing in the presence of the CS+ (p 's $< 0.05, 0.01$, respectively), rats receiving either 10 ng ($n = 8$) or 100 ng ($n = 6$) of intra-NASh CBD showed no associative freezing in response to CS- vs CS+ cue presentations (p 's > 0.05 ; Figure 2a). In addition, associative freezing in response to CS+ presentations were significantly lower relative to VEH controls for both the 10 ng and 100 ng CBD groups (p 's < 0.01 ; Figure 2a). Thus, bilateral intra-NASh CBD dose-dependently blocks the formation of conditioned fear memories measured with conditioned freezing behaviors. Accordingly, we selected the highest behaviorally effective dose of 100 ng/0.5 μ l as the challenge dose for subsequent behavioral and electrophysiological experiments.

Intra-NASh CBD Modulates Fear Memory Formation Through a 5-HT_{1A}-Dependent Mechanism Memory

Given previous evidence demonstrating that CBD produces its pharmacological effects via the 5-HT_{1A} receptor (Russo *et al*, 2005), we next challenged the effects of intra-NASh CBD (100 ng) with coadministration of the selective 5-HT_{1A} receptor antagonist, NAD 299 (10–100 ng/0.5 μ l). ANOVA comparing percentages of time spent freezing during testing revealed a significant main effect of treatment ($F_{(1,37)} = 13.9$; $p < 0.001$). *Post hoc* analyses revealed that rats receiving either the lower (10 ng/0.5 μ l) or higher (100 ng/0.5 μ l) dose

of NAD 299 (10 ng, $n = 7$; 100 ng, $n = 6$) with CBD, demonstrated robust associative freezing behaviors in response to CS+ presentations, relative to VEH controls ($n = 7$, p 's < 0.01 ; Figure 2b). To control for the potential effects of NAD 299 alone, a separate control group received bilateral intra-NASh NAD 299 (100 ng/0.5 μ l; $n = 8$) alone, before training. These rats displayed normal conditioned freezing behavior, freezing significantly more to CS+ presentations ($p < 0.05$). Next, to examine the possible involvement of DA receptor transmission in the effects of intra-NASh CBD, we challenged the effects of intra-NASh CBD with the broad spectrum DA receptor antagonist, α -flu (100–1000 ng/0.5 μ l). ANOVA revealed a significant main effect of group ($F_{(3,50)} = 5.65$; $p < 0.01$) on times spent freezing to CS+ vs CS- presentations. *Post hoc* analyses revealed that rats receiving coadministration of CBD (100 ng/0.5 μ l) with either the lower dose of α -flu (100 ng/0.5 μ l; $n = 6$) or the higher dose of α -flu (1000 ng/0.5 μ l; $n = 8$) showed no associative freezing during CS+ presentations (p 's > 0.05 ; Figure 2b). To control for any potential effects of α -flu alone on fear memory formation, a separate control group ($n = 8$) received intra-NASh α -flu alone (1000 ng/0.5 μ l). This group demonstrated significant associative freezing behaviors ($p < 0.05$; Figure 2b). Thus, whereas 5-HT_{1A} receptor blockade reversed the effects of CBD-induced block of fear memory acquisition, DA receptor blockade had no effect. In addition, neither the highest behaviorally effective doses of NAD 299 nor α -flu produced any effects on fear memory acquisition in and of themselves.

Beyond the 5-HT_{1A} receptor system, previous studies have suggested that CBD may produce pharmacological actions on the CB1 receptor system (McPartland *et al*, 2015). To examine if the effects of CBD on fear memory acquisition may be mediated through a CB1 receptor substrate, separate groups of rats received intra-NASh CBD (100 ng/0.5 μ l) coadministered with the selective CB1 antagonist, RIM (50–500 ng/0.5 μ l; $n = 9$, $n = 10$, respectively). Comparing the effects of RIM/CBD coadministration on freezing behaviors revealed a significant effect of group ($F_{(2,47)} = 10.53$; $p < 0.01$), with *post hoc* analyses revealing that neither dose of RIM reversed the effects of CBD on fear memory blockade, relative to VEH controls, with both groups displaying no associative freezing in response to CS+ vs CS- presentations (p 's > 0.05 ; Figure 2b).

To ensure that the highest doses of CBD, NAD 299, or α -flu were not producing any unconditioned effects on footshock sensitivity during the fear memory acquisition phase, separate control groups received footshock sensitivity tests (see Materials and Methods) following bilateral intra-NASh microinfusions of VEH ($n = 8$), CBD (100 ng/0.5 μ l, $n = 7$), NAD 299 (500 ng/0.5 μ l, $n = 8$), or α -flu (1000 ng/0.5 μ l, $n = 8$). Group comparisons with ANOVA revealed no significant group differences in the amount of freezing behavior following footshock (0.8 mA) administration ($F_{(3,29)} = 1.10$; $p > 0.05$; Figure 2c); total distance traveled during the sensitivity testing ($F_{(3,29)} = 0.92$; $p > 0.05$; Figure 2d) or total average number of jumping events during footshock administrations ($F_{(3,29)} = 0.25$; $p > 0.05$; Figure 2e). Thus, neither CBD, NAD 299 nor α -flu produced any observable alterations in footshock sensitivity during fear memory acquisition training.

Figure 2 Effects of intra-shell region of the mesolimbic nucleus accumbens (NASH) cannabidiol (CBD) on conditioned freezing behaviors: modulation by serotonin 1A receptor (5-HT_{1A}) receptor transmission. (a) Relative to vehicle (VEH) control rats ($n=6$), bilateral administration of intra-NASH CBD (1 ng, $n=7$; 10 ng, $n=8$ or 100 ng/0.5 μ l, $n=6$) dose-dependently blocks the acquisition of suprathreshold (0.8 mA) olfactory associative fear memory. (b) The effects of intra-NASH CBD on associative fear memory formation are dose-dependently reversed by coadministration of a selective 5-HT_{1A} receptor antagonist (NAD 299, 10 ng, $n=7$; 100 ng/0.5 μ l, $n=6$), but not by the broad-spectrum dopamine (DA) receptor antagonist, α -flu (100 ng ($n=6$), 1000 ng/0.5 μ l ($n=8$)), or the CB1 receptor-selective antagonist, RIM (50 ng ($n=9$), 500 ng/0.5 μ l, $n=10$). Neither intra-NASH NAD 299 (500 ng, $n=8$) nor α -flu (1000 ng, $n=8$) administered alone produce any effects on fear memory formation. Separate footshock sensitivity control groups (see Materials and Methods) receiving VEH ($n=8$) or the effective doses of intra-NASH CBD (100 ng, $n=7$), α -flu (1000 ng, $n=8$), or NAD 299 (100 ng, $n=8$) revealed no significant differences in measures of (c), freezing behavior; (d), locomotor activity or (e), jumping behaviors. * $p < 0.05$; ** $p < 0.01$. Bars represent means $\pm$ S.E.M. for this and all subsequent figures. CS, conditioned stimulus (positive or negative).

Intra-NASH CBD Modulates DAergic vs Non-DAergic Neuronal Activity States in the VTA

We next performed *in vivo*, single-unit neuronal recordings in the VTA to determine the potential effects of intra-NASH CBD administration on spontaneous neuronal activity patterns in isolated populations of DAergic vs non-DAergic VTA neurons (see Materials and Methods; Figure 3a). Accordingly, our previously determined behaviorally effective dose of CBD (100 ng) was microinfused into the NASH while simultaneously recording single-cell extracellular VTA

neuronal units. A total of $n=15$ VTA-DA neurons were recorded and analyzed following intra-NASH CBD micro-infusions. A cell was considered to have changed its firing rate if there was a minimum of 10% difference in frequency rate from baseline. Using this previously established and reported criterion (Draycott *et al*, 2014; Tan *et al*, 2009), qualitative analysis revealed that 60% of DA neurons showed decreased activity, 7% increased, and 33% were unchanged (Figure 3b). Subgroup analyses of average frequency recorded during 5 min pre- vs post-intra-NASH microinfusions revealed that CBD significantly decreased the firing

Figure 3 Effects of intra-shell region of the mesolimbic nucleus accumbens (NASH) cannabidiol (CBD) on DAergic and non-DAergic neuronal activity patterns in the ventral tegmental area (VTA). (a) Representative microphotograph showing typical intra-VTA *in vivo* dopamine (DA) neuron recording location (indicated by arrow). (b) Group summary of all VTA DA neuronal effects ($n = 15$) showing distribution of neuronal response patterns (no change in frequency, decreased activity, increased activity) across experimental conditions. (c) Intra-NASH CBD significantly decreased spontaneous VTA DA neuronal firing frequency and (d) bursting rates. (e) Sample rastergram showing typical VTA DAergic neuronal response pattern following intra-NASH CBD (100 ng/0.5 µl) administration. (f) Summary of non-DA neuronal group response patterns following intra-NASH CBD administration ($n = 15$). (g) A sub-population of non-DAergic VTA neurons show a significant increase in spontaneous firing frequency following intra-NASH CBD (100 ng/0.5 µl). (h) Sample rastergram showing a typical VTA non-DA neuronal response pattern following intra-NASH CBD administration. * $p < 0.05$; ** $p < 0.01$. CS, conditioned stimulus (positive or negative).

rate ($t_{(14)} = 3.13$, $p < 0.01$; Figure 3c) and bursting rate ($t_{(14)} = 2.76$, $p < 0.05$) without significantly decreasing the number of spikes per burst ($t_{(14)} = 2.06$, $p = 0.06$) (Figure 3d). Thus, intra-NASH CBD (100 ng) causes a predominantly inhibitory effect on VTA DAergic neuronal frequency and bursting rates. In Figure 3e, we present a rastergram of a single VTA DA neuron showing a typical decreased activity pattern following intra-NASH CBD microinfusion.

We next recorded and analyzed presumptive VTA GABA neurons ($n = 15$) during intra-NASH CBD (100 ng) administration. Qualitative analysis of overall population activity revealed that 27% of non-DA neurons decreased activity, 40% increased, and 33% demonstrated no significant change in frequency (Figure 3f). Statistical analyses of neuronal sub-populations recorded 5 min pre- vs post-NASH microinfusions showed that while CBD did not significantly modify the collective (group) firing frequency of VTA GABA interneurons, $t_{(14)} = 0.04$; $p = 0.97$, separate sub-population analyses revealed that neurons showing increased activity were significantly elevated in firing frequency relative to baseline ($t_{(5)} = 3.8$; $p < 0.01$; Figure 3g), whereas neurons showing either decreased ($t_{(3)} = 2.9$; $p > 0.05$) or no change in activity ($t_{(4)} = 0.55$; $p > 0.05$) were not significantly changed from preinfusion baseline levels (data not shown). Collectively, these data demonstrate that intra-NASH CBD induces heterogeneous effects on presumptive VTA GABAergic neurons. However, a plurality of these neurons (40%) significantly increased their spontaneous firing rates. In Figure 3h, we present a rastergram showing a typical non-DAergic neuronal response pattern following intra-NASH CBD microinfusion.

CBD Modulates DAergic Neuronal Activity in the VTA Selectively Through NASH 5-HT_{1A} Transmission

We next sought to determine how the effects of coadministration of selective 5-HT_{1A} or DA receptor antagonists on CBD-mediated behaviors might influence VTA DAergic neuronal activity. Accordingly, we performed *in vivo*, single-unit intra-VTA neuronal recordings to determine the potential effects of intra-NASH CBD, NAD 299/CBD, or α -flu/CBD coadministration on VTA DA neuron firing frequency and bursting levels (see Materials and Methods), using our behaviorally effective doses of CBD (100 ng/0.5 µl), NAD 299 (100 ng/0.5 µl), or α -flu (1 µg/0.5 µl). We sampled a total of $n = 44$ VTA DA neurons (VEH group, $n = 10$ cells; CBD (100 ng) group, $n = 15$ cells; CBD (100 ng)+NAD 299 (100 ng) group, $n = 10$ cells; CBD (100 ng)+ α -flu (1000 ng), $n = 9$ cells). A summary of VTA DAergic neuronal activity profiles after intra-NASH microinfusions are represented in Figures 4a. For rats receiving intra-NASH VEH, 50% of neurons showed no change, 30% increased, and 20% showed decreased activity. For rats receiving intra-NASH CBD, 60% of neurons decreased firing frequency, 33% showed no change, and 7% increased frequency. In rats receiving intra-NASH CBD+NAD 299, neurons decreasing, increasing, or showing no change in activity levels were 60%, 20%, and 20%, respectively. In rats microinfused with CBD+ α -flu, 78% of the recorded cells decreased their firing frequency, 22% showed no change, and no cells demonstrated increased frequency. Analyses of pre- vs postinfusion activity rates for VTA DA neurons revealed average changes from baseline of

Figure 4 Effects of intra-shell region of the mesolimbic nucleus accumbens (NASH) cannabidiol (CBD) and serotonergic/dopaminergic antagonists on VTA DAergic neuronal activity patterns in the ventral tegmental area (VTA). (a) Summary of experimental neuronal groups showing relative changes (no change, increase or decrease) firing frequencies following intra-NASH pharmacological treatments (total $n = 44$ neurons sampled). (b) Intra-NASH CBD alone (100 ng/0.5 μ l) caused a significant decrease in spontaneous VTA dopamine (DA) neuronal firing frequency rates. This inhibitory effect was reversed by coadministration of the effective dose of NAD 299 (500 ng/0.5 μ l). In contrast, coadministration with the DA receptor antagonist, α -flu (1 μ g/0.5 μ l), had no effect on CBD's inhibition of VTA DA neuron activity. (c) Comparing VTA DA neuron bursting rates revealed that intra-NASH CBD (100 ng/0.5 μ l) significantly decreases VTA DA neuron bursting rates. Again, this effect was significantly reversed by coadministration of NAD 299 (500 ng/0.5 μ l). In contrast, coadministration with the DA receptor antagonist, α -flu (1 μ g/0.5 μ l), had no effect on CBD's inhibition of VTA DA neuron bursting rates. (d) Intra-NASH NAD 299 (500 ng) or α -flu (1000 ng) produced heterogeneous effects on spontaneous VTA DA neuron activity patterns. However, neither drug alone produced any significant effects on firing frequency (e) or bursting rates (f). (g) Sample rastergram showing a typical VTA DA neuron inhibitory response pattern following intra-NASH CBD. (h) Sample rastergram showing a typical VTA DA neuron response following coadministration of CBD with NAD 299 and (i) sample rastergram showing a typical response pattern following coadministration of CBD with α -flu. * $p < 0.05$; ** $p < 0.01$. CS, conditioned stimulus (positive or negative).

+7% for rats treated with VEH, -27% for rats treated with CBD, +3% with CBD+NAD 299, and -23% for rats microinfused with CBD+ α -flu (Figure 4b). ANOVA comparing groups revealed a significant main effect of treatment on VTA DA neuron firing frequency rates ($F_{(3,43)} = 3.57$, $p < 0.05$). *Post hoc* analysis revealed that rats treated with intra-NASH CBD alone or with CBD+ α -flu showed

significantly decreased firing frequencies relative to baseline (p 's < 0.05). However, rats treated with CBD+NAD 299 showed significantly increased firing frequency relative to these groups, and no significant difference from baseline levels ($p < 0.05$; Figure 4b). Thus, cotreatment with NAD 299, but not α -flu, reversed CBD-induced reductions in spontaneous DA neuron frequency rates.

Analysis of VTA DA neuron bursting rates revealed average changes from baseline of -4% , -35% , $+29\%$, and -40% for rats treated with intra-NASH VEH, CBD, CBD+NAD 299, and CBD+ α -flu, respectively (Figure 4c). ANOVA comparing groups revealed a significant effect of treatment ($F_{(3,43)} = 4.24$, $p < 0.01$) and *post hoc* comparisons showed that bursting rates were significantly different between rats treated with CBD vs CBD+NAD 299 ($p < 0.05$) and between CBD+ α -flu and CBD+NAD 299 groups ($p < 0.05$). Thus, cotreatment with NAD 299, but not α -flu, reversed CBD-induced reductions in spontaneous DA neuron bursting rates. Next, to examine if intra-NASH α -flu or NAD 299 had any effects on VTA DA neuron frequency or bursting rates in and of themselves, separate control groups received either intra-NASH NAD 299 (500 ng/0.5 μ l; $n = 10$ neurons) or α -flu (1000 ng/0.5 μ l; $n = 9$ neurons). For intra-NASH NAD 299, qualitative analysis revealed that 10% of neurons showed increased, 30% decreased, and 60% no change in frequency (Figure 4d). For intra-NASH α -flu, qualitative analysis revealed that 20% of neurons increased, 35% decreased, and 45% showed no changes (Figure 4d). ANOVA revealed no significant treatment effect on VTA DA neuron firing rates across groups ($F_{(2,28)} = 0.63$; $p > 0.05$; Figure 4e). ANOVA analysis of bursting rates similarly revealed no significant treatment effect on DA neuron burst rates across groups ($F_{(2,26)} = 1.33$; $p > 0.05$; Figure 4f). Thus, neither intra-NASH NAD 299 nor α -flu had any effects on VTA DA neuron activity parameters in and of themselves.

Representative neuronal rastergrams showing single VTA DA neuronal response patterns following intra-NASH CBD (100 ng alone; Figure 4g); CBD (100 ng)+NAD 299 (100 ng; Figure 4h); and CBD (100 ng)+ α -flu (1000 ng; Figure 4i) demonstrate typically observed neuronal activity patterns following these treatments. Taken together, these results show that intra-NASH CBD decreases VTA DA neuronal frequency and bursting rates. These effects were blocked by coinfusion of a 5-HT_{1A} antagonist, but remained unchanged by the coinfusion of a DA receptor antagonist.

Functional Disconnection of the NASH $\rightarrow$ VTA Pathway Reverses the Effects of Intra-NASH CBD on Fear Memory Formation

Medium spiny neurons (MSNs) in the NAc send GABAergic terminals to VTA neurons (Nauta *et al*, 1978; Kalivas *et al*, 1993). Given our previously described finding (Figures 3f–h) that intra-NASH CBD caused significant increases in non-DA, presumptive VTA GABAergic neurons, we hypothesized that the behavioral effects of intra-NASH CBD may depend on NAc $\rightarrow$ VTA GABAergic projections and therefore depend on GABA receptor transmission directly in the VTA. Accordingly, using a functional cerebral disconnection procedure (see Materials and Methods), we examined whether intra-VTA blockade of GABAergic transmission may modulate the ability of intra-NASH CBD to block associative fear memory in rats with unilateral cannulae implantations into the NASH and VTA. Rats received a combination of intra-VTA bicuculline (50 ng) and saclofen (100 ng) to block intra-VTA GABA_{A/B} receptor transmission immediately before a unilateral microinfusion of intra-NASH CBD (100 ng) in the contralateral hemisphere (Figure 5a). Histological analysis revealed intra-VTA injector locations within the anatomical boundaries of

the VTA or NASH as defined by Paxinos and Watson (2005) (Figures 5b and c). ANOVA comparing behavioral associative freezing scores across groups revealed a significant main effect of group ($F_{(3,38)} = 3.261$; $p \leq 0.05$; Figure 5d) on percentages of time spent freezing to either CS+ or CS– presentations. *Post hoc* analyses revealed that VEH control rats (NASH/VEH vs VTA/VEH; $n = 6$) demonstrated robust associative freezing behaviors in response to CS+ presentations ($p < 0.01$). In contrast, rats receiving the intra-NASH CBD ($n = 6$; 100 ng/0.5 μ l) with intra-VTA VEH showed no associative fear memory ($p > 0.05$), consistent with previous results. However, rats receiving intra-VTA GABA_{A/B} antagonists before intra-NASH CBD (100 ng/0.5 μ l) ($n = 6$) demonstrated robust associative freezing behaviors in response to CS+ presentations ($p < 0.05$), demonstrating that contralateral blockade of VTA GABA_{A/B} transmission is sufficient to reverse the behavioral effects of intra-NASH CBD on blockade of associative fear memory formation. Finally, to ensure that intra-VTA GABA_{A/B} receptor blockade did not in itself influence fear memory formation, a separate group received intra-VTA GABA_{A/B} antagonist treatment before intra-NASH VEH. Similar to VEH/VEH-treated controls, these rats displayed significant associative fear responses during CS+ presentations ($n = 6$; $p < 0.05$).

DISCUSSION

In the present study, we report that intra-NASH CBD blocks the formation of fear-related memory via a 5-HT_{1A} receptor-dependent mechanism. Furthermore, we found that intra-NASH CBD decreased spontaneous VTA DAergic neuronal activity, both in terms of frequency and bursting levels, and conversely, increased the spontaneous activity of non-DAergic, presumptive VTA GABAergic neurons. CBD's modulation of VTA DAergic neuronal activity was dependent on intra-NASH 5-HT_{1A} transmission, but not DAergic or CB1 receptor transmission, as only blockade of 5HT_{1A} transmission reversed the effects of CBD on VTA DAergic activity states and restored associative fear memory formation. Finally, consistent with our electrophysiological findings, functional disconnection of GABAergic transmission between the contralateral NASH and VTA reversed the effects of intra-NASH CBD on associative fear memory formation. Taken together, these findings demonstrate several new insights into the potential role for CBD in the modulation of emotional memory processing within the mesolimbic pathway, implicating serotonergic transmission as an important mediator of these effects.

Our findings are consistent with previous research demonstrating a role for CBD in modulating emotional memory. For example, Stern *et al* (2012) reported that systemically administered CBD could block reconsolidation of associative fear memories in rats in a temporally specific manner. Das *et al* (2013) reported that systemic CBD could enhance the consolidation of fear-related extinction learning in human subjects. Marinho *et al* (2015) recently reported that CBD microinfusions into the prelimbic or infralimbic regions of rat prefrontal cortex could induce either anxiolytic or anxiogenic effects through a 5-HT_{1A}-dependent mechanism. The present findings are the first report that CBD can produce effects on fear-memory acquisition directly in the NAc, via 5-HT_{1A}-dependent modulation of VTA neuronal activity.

Figure 5 Disconnection of shell region of the mesolimbic nucleus accumbens/ventral tegmental area (NASH/VTA) γ -aminobutyric acid, type A/B ($GABA_{A/B}$) transmission reverses the effects of intra-NASH cannabidiol (CBD) on conditioned freezing. (a) Summary of experimental design for NASH-VTA disconnection procedure with sample microphotographs showing representative intra-VTA and intra-nucleus accumbens (NAc) microinjector placements. (b and c) Schematic summaries showing unilateral placements for intra-VTA microinfusions of the $GABA_{A/B}$ antagonist mixture and intra-NASH CBD microinfusion locations (see Materials and Methods). ● = 100 ng CBD+50 ng saclofen/50 ng bicuculline group, ◆ = 100 ng CBD+saline group. (d) Schematic representation of experimental groups with description of contralateral intra-VTA vs intra-NASH treatments. (e) Consistent with previous results (Figure 2a), intra-NASH CBD effectively blocks formation of associative fear memory in rats receiving intra-NASH CBD vs contralateral intra-VTA vehicle (VEH) ($n=6$), relative to VEH/VEH controls ($n=6$). In contrast, rats receiving intra-NASH CBD vs intra-VTA $GABA_{A/B}$ receptor blockade in the contralateral hemisphere ($n=6$) show no blockade of fear memory formation. In addition, rats receiving intra-NASH CBD vs contralateral VTA $GABA_{A/B}$ receptor blockade ($n=6$) demonstrate normal fear memory formation. * $p < 0.05$. CS, conditioned stimulus (positive or negative).

CBD Modulates Conditioned Freezing Behavior Through a 5-HT_{1A}-Dependent Mechanism in the NAc

Considerable evidence links the pharmacological and behavioral effects of CBD to the serotonergic system. For example, CBD is a 5-HT_{1A} receptor agonist (Russo *et al*, 2005) and systemic CBD modulates the reward-facilitating

effects of morphine via 5-HT_{1A} receptors in the dorsal raphe nucleus (Katsidoni *et al*, 2013). In addition, CBD has been shown to modulate anxiety-like behaviors through actions on the 5-HT_{1A} receptor (Fogaca *et al*, 2014; Marinho *et al*, 2015). Consistent with the current findings, previous evidence has demonstrated that CBD's ability to interact with the mesolimbic system involves 5-HT_{1A} receptor

transmission. For example, 5-HT_{1A} activation in the NASH blocks apomorphine-induced behaviors in rodents (Fujita *et al*, 2008), inhibits amphetamine-induced DA release in the rat NAc (Ichikawa *et al*, 1995), and reduces striatal DA synthesis (Johnson *et al*, 1993). Given these effects of 5-HT_{1A} activation, blockade of 5-HT_{1A} transmission would be expected to counteract the effects of CBD, removing the inhibitory effects of intra-NASH CBD on VTA DAergic activity. This prediction is consistent with our electrophysiological findings showing that blockade of 5-HT_{1A} transmission restores VTA DAergic tone in the presence of CBD and reverses the behavioral effects of CBD on fear memory formation. In contrast, coadministration of a DA receptor antagonist or selective CB1 receptor antagonist failed to reverse the behavioral effects of CBD. Previous studies have reported that CBD can act as an antagonist or inverse agonist of the CB1 receptor (Thomas *et al*, 1998; Pertwee, 2008). However, other reports have found that CBD has relatively low binding affinity and/or activity at the CB1 receptor (Bisogno *et al*, 2001; Thomas *et al*, 2007). The present findings would suggest that, at least within the NASH, CBD produces its effects through non-CB1 receptor-dependent mechanisms.

CBD Modulates Neuronal Activity Patterns in the VTA Through a 5-HT_{1A}-Dependent Mechanism

Interestingly, previous studies using systemic CBD administration have reported both effects of CBD on spontaneous activity patterns in VTA neuronal populations (French *et al*, 1997) or increases in mesolimbic DA release (Murillo-Rodríguez *et al*, 2006). These discrepancies are likely due to differences in administration routes, as the present study exclusively used targeted microinfusions into the NASH *vs* systemic administration. However, it is of interest that systemically applied CBD may be interacting with neural circuits extrinsic to the mesocorticolimbic system, which may in turn produce different effects on DAergic transmission patterns.

Within the VTA, we found that behaviorally effective doses of intra-NASH CBD were sufficient to inhibit VTA DAergic activity levels while simultaneously increasing the activity states of presumptive inhibitory VTA GABAergic neurons. This effect is consistent with previous observations demonstrating that blunting or potentiating VTA DAergic neuronal activity can potently modulate the formation of associative fear memories. For example, we have reported that activation of CB1 receptors in the rat prefrontal cortex can simultaneously block the formation of normally suprathreshold conditioned freezing behaviors and inhibit subcortical DAergic activity (Draycott *et al*, 2014). In contrast, overstimulation of DAergic transmission in the PFC can amplify normally non-salient associative fear conditioning cues (Lauzon *et al*, 2009). The present findings suggest that intra-accumbal modulation of VTA DAergic activity via CBD/5-HT_{1A}-dependent mechanisms may similarly regulate emotional salience and the acquisition of conditioned freezing behavior via functional interactions with select VTA neuronal populations. For example, by blunting VTA DAergic neuronal activity during the acquisition of associative fear memory, CBD may serve to block conditioned freezing behaviors. Future studies are required

to more precisely characterize the effects of CBD across these distinct learning phases of emotional memory processing.

How might accumbal 5HT_{1A} transmission modulate upstream DAergic mesolimbic transmission? Anatomical evidence demonstrates that NAc GABAergic MSNs send functional GABAergic projections to the VTA (Nauta *et al*, 1978; Kalivas *et al*, 1993). In the present study, we found that intra-NASH CBD increased the activity rates of presumptive, non-DA GABAergic neurons in the VTA. Accordingly, we hypothesized that intra-NASH CBD may regulate VTA DAergic activity (and corresponding fear-related associative memory formation) via indirect modulation of GABAergic transmission substrates in the VTA. To test this, we performed a disconnection study wherein intra-NASH CBD effects were disconnected from the contralateral hemisphere with combined intra-VTA microinfusions of GABA_{A/B} receptor antagonists. Consistent with this hypothesis, disconnection blockade of VTA GABAergic transmission was capable of reversing the effects of intra-NASH CBD on fear memory formation. While beyond the scope of the present study, one interesting implication from the present results is that 5-HT_{1A} receptor activation within the NASH might similarly modulate downstream VTA neuronal activity states and fear memory formation. Future studies are required to investigate these possibilities.

Implications of CBD's Modulatory Role on Mesolimbic Activity

The present study adds to a growing body of preclinical and clinical evidence demonstrating that CBD can modulate activity states of the mesolimbic DA system, and, in turn, regulate emotional processing. Importantly, the ability of CBD to inhibit mesolimbic DAergic activity and blunt fear-related memory formation is particularly intriguing given the large body of evidence demonstrating that the primary psychoactive phytochemical in cannabis, THC, produces propsychotic effects (Kuepper *et al*, 2011). Furthermore, in direct contrast to the effects of CBD, THC has been shown to induce a state of hyperactive mesolimbic DAergic activity following adolescent neurodevelopmental exposure (Renard *et al*, 2016a). In addition, we have recently reported that intra-NASH CBD blocks hyper-DAergic activity and psychomotor sensitization induced by amphetamine exposure (Renard *et al*, 2016b).

Several clinical and preclinical reports have suggested that CBD can reduce anxiety and may serve to ameliorate symptoms associated with emotional memory disorders such as PTSD. For example, CBD has been reported to facilitate the extinction of previously acquired traumatic or associative fear memories (Bitencourt *et al*, 2008; Stern *et al*, 2012; Das *et al*, 2013). While currently no clinical evidence has reported specific long-term therapeutic effects of CBD in PTSD patient populations, PTSD has been associated with dysregulation of 5-HT_{1A} expression levels in brain regions responsible for emotional memory and processing, such as the amygdala (Sullivan *et al*, 2013). Furthermore, preclinical evidence has shown also that genetic deletion of the 5-HT_{1A} receptor in mice leads to potentiated reactivity to fear-related conditioning cues (Klemenhagen *et al*, 2006). Future research is required to more precisely characterize the mechanisms by which CBD and 5-HT_{1A} receptor substrates

may interact and how these processes may, in turn, regulate emotional memory formation.

In summary, the present study reveals several novel insights regarding the functional effects of CBD on emotional memory processing and identifies a 5-HT_{1A} receptor mechanism within the NASH as an underlying mechanism by which CBD may regulate mesolimbic activity and modulate the formation of associative emotional memories. Furthermore, these findings underscore growing evidence demonstrating that differential phytochemical constituents of cannabis (CBD vs THC) can produce divergent and opposing effects on DAergic activity states and emotional memory behaviors.

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The authors declare no conflict of interest.

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