



## DNA assembly and cloning in an overnight run with the BioXp™ 3200 system

The BioXp™ 3200 system is an automated personal genomic workstation that builds and clones DNA fragments in a process that is virtually hands-free. In an overnight run, the instrument generates cloned DNA from custom-designed oligonucleotide pools and reagents engineered from sequence information. Here we discuss highlights and advantages of the BioXp system and the two modules currently available, the assembly module and the assembly-and-cloning module.

As the scale of genetic analysis has trended away from single-gene studies and toward gene-family, genomic and metagenomic studies, the demand for large-scale DNA-synthesis services and technologies has grown. Moreover, with newer cloning and sequencing technologies, the pace of molecular biology research is accelerating. Synthetic Genomics, Inc., has developed the BioXp system (**Fig. 1**), available through SGI-DNA, as an in-house laboratory tool to address the increased interest and demand for rapid DNA synthesis. With an assembly module, the BioXp system generates high-quality, linear DNA fragments from custom-designed oligonucleotide pools and reagents across a meaningful section of the complexity continuum in an overnight run. Now, with the introduction of the assembly-and-cloning module, the BioXp system has the additional capability to deliver cloned DNA from a custom DNA sequence in an overnight run (**Fig. 2**).



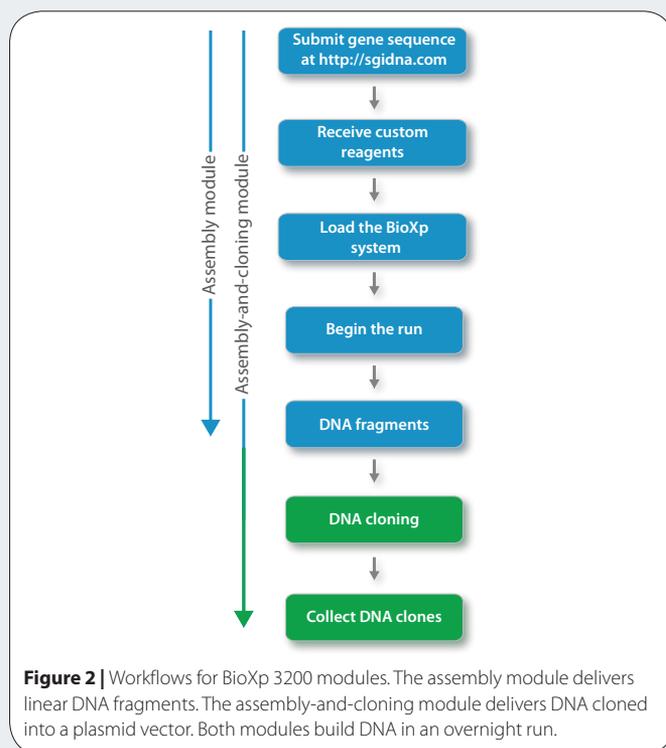
**Figure 1** | The BioXp 3200 system, a genomic workstation.

Since its launch in early 2015, commercial and research laboratories have been reaping the advantages of the BioXp system, whether

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in individual laboratories or in core facilities. On-site access to the automated BioXp system liberates researchers from the time-consuming, tedious steps needed to obtain DNA fragments, allowing them to instead focus on new discoveries and DNA analytics.



**Figure 2** | Workflows for BioXp 3200 modules. The assembly module delivers linear DNA fragments. The assembly-and-cloning module delivers DNA cloned into a plasmid vector. Both modules build DNA in an overnight run.

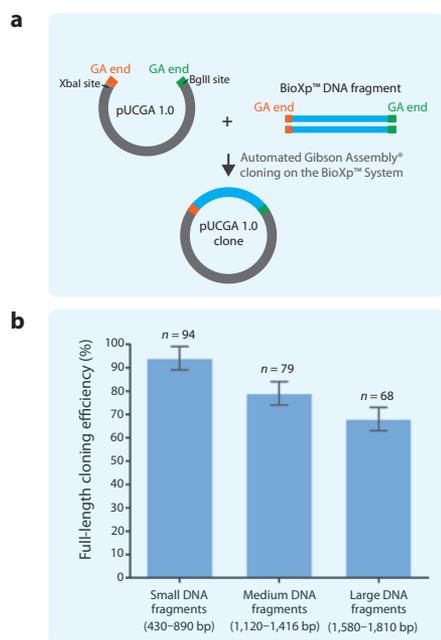
### The BioXp assembly-and-cloning module

With the assembly module, the BioXp system builds linear, blunt-end, double-stranded DNA fragments. Now, with the assembly-and-cloning module, the instrument has the added capability to build and clone DNA fragments of interest into the SGI-DNA pUCGA 1.0 vector. The pUCGA 1.0 clones generated by the BioXp system are immediately ready for transformation and further downstream analysis.

## APPLICATION NOTES

## BioXp system pUCGA 1.0 clones

DNA clones obtained from the BioXp system consist of a DNA fragment of interest (400–1,800 bp, 40–60% GC content) cloned into the 2.7-kb pUCGA 1.0 vector. The pUCGA 1.0 vector map and additional information are available at <http://sgidna.com>. With Gibson Assembly® technology, homologous overlap regions are automatically designed into the termini of the DNA fragments and are present in the pUCGA 1.0 vector to facilitate cloning. These homologous regions, referred to as GA ends, consist of 30 bases that have minimal sequence homology to naturally occurring genes. Highly accurate and robust automated cloning on the BioXp system is performed using the Gibson Assembly method (Fig. 3a). Clones obtained from the BioXp system are ready for transformation and subsequent downstream analysis.



**Figure 3** | The BioXp 3200 system utilizes high-efficiency Gibson Assembly cloning to deliver DNA in the pUCGA 1.0 vector. **(a)** Overview of Gibson Assembly cloning on the BioXp 3200 system with a BioXp fragment and the pUCGA 1.0 vector. **(b)** High cloning efficiency from the BioXp 3200 system. As expected, the highest cloning efficiencies were achieved with the smallest DNA fragments. The total cloning efficiency (CE) for full-length inserts was calculated using the following formula:  $CE (\%) = (\text{Number of white colonies} / \text{Total number of colonies}) \times (\text{Number of full insert colonies} / \text{Total number of colonies}) \times 100$ . The sample number ( $n$ ) is shown above each respective bar; error bars represent  $\pm$ s.d.

## Transforming pUCGA 1.0 clones

Once clones have been collected from the BioXp system deck after an assembly-and-cloning run, transformation is the next downstream step required for error-free clone identification and further analysis. The pUCGA 1.0 vector contains *lacZ* (the gene encoding the N-terminal fragment of  $\beta$ -galactosidase), which is disrupted by insertion of the BioXp DNA fragment, allowing for blue-white screening of recombinant clones. To assess clone quality, we performed transformation and calculated the cloning efficiency of pUCGA 1.0 clones. Briefly, an assembly-and-cloning run was repeated 13 times

on five different BioXp instruments. Twenty clones were randomly collected from the instruments after each run, transformed into TransforMax™ EPI300™ electrocompetent *Escherichia coli*, and plated onto LB plates containing  $100 \mu\text{g ml}^{-1}$  carbenicillin with  $40 \mu\text{g ml}^{-1}$  X-Gal and 0.1 mM isopropyl  $\beta$ -D-1-thiogalactopyranoside. After an overnight incubation, colonies were counted, picked and grown overnight, and plasmid DNA was prepared.

## High cloning efficiency of pUCGA 1.0 clones

The cloning efficiency of constructs generated with the BioXp assembly-and-cloning module is shown in Figure 3b. The overall cloning efficiency of full-length pUCGA 1.0 clones was 83%. Additionally, we grouped clones according to fragment size for further cloning-efficiency analysis. As expected, the highest cloning efficiency (>90%) was observed for the smallest DNA fragments (<900 bp).

Full-length inserts were identified from double enzyme digestion with XbaI and BglIII (Fig. 3a), which leaves a partial or full GA end sequence intact at the terminus of the BioXp fragment. For excision of only the fragment of interest from pUCGA 1.0, the BioXp fragment may be pre-engineered with restriction enzyme sites internal to the GA ends. Alternatively, PCR amplification with a high-fidelity DNA polymerase may be used to isolate the fragment of interest or to subclone the fragment in an alternate vector (e.g., an expression vector).

## Conclusion

The BioXp system brings a new, rapid, automated method of DNA synthesis and cloning directly to the laboratory benchtop. The instrument can currently assemble and clone 24 DNA fragments of interest simultaneously in an overnight run. The pUCGA 1.0 DNA clones obtained from the BioXp system exhibit high cloning efficiencies—greater than 90% for DNA fragments less than 900 bp in length, and 83% overall. Laboratories using the BioXp system have the capability for virtually hands-free assembly and cloning of genes into vectors in an overnight run.

## Additional information

Complete product information and additional resources are available at <http://www.sgidna.com/bxp3200>. BioXp™ is a trademark and Synthetic Genomics® and Gibson Assembly® are registered trademarks of Synthetic Genomics, Inc. TransforMax™ and EPI300™ are trademarks of Epicentre Technologies Corporation. Gibson Assembly US Patent Nos. 7,776,532, 8,435,736 and 8,968,999.

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