gression analysis (Enzfitter, Elsevier-Biosoft, Cambridge, UK). Linear regression analysis of an indirect Hill plot transformation of the dose–response curve for NBMPR inhibition of hENT1-mediated uridine influx (Fig. 2*d*) gave a Hill coefficient of 0.98 ± 0.03 (indicating interaction of NBMPR with a single population of binding sites) and an IC_{50} value of 3.4 ± 0.03 nM. Correction of the latter value for inhibitor depletion caused by partitioning into oocyte lipids (determined using HPLC-purified [^{3}H]NBMPR (Moravek Biochemicals) to be 26%) gave an apparent K_i value for NBMPR inhibition of uridine influx of 2.4 nM (calculated using competitive inhibition[1,13] and a uridine K_m of 0.24 mM). Gemcitabine was a generous gift from Eli Lilly Company.

Acknowledgments

The laboratories of S.A.B., C.E.C. & J.D.Y. contributed equally to this project. Supported by the Medical Research Council of the United Kingdom, the Cancer Research Campaign of the United Kingdom, the Yorkshire Cancer Research Campaign, the National Cancer Institute of Canada, the Medical Research Council of Canada, and the Alberta Heritage Foundation for Medical Research. J.D.Y. is a Heritage Medical Scientist of the AHFMR; C.E.C. is a Terry Fox Cancer Research Scientist of the NCIC. We thank H.E. Fincham, J. Ingram and D. Mowles for technical assistance.

RECEIVED 15 JULY; ACCEPTED 15 NOVEMBER 1996

1. Cass, C.E. Nucleoside transport. in *Drug Transport in Antimicrobial and Anticancer Chemotherapy* (ed. Georgopapadakou, N.H.) 403–451 (Marcel Dekker, New York, 1995).
2. Kwong, F.Y.P. *et al.* Purification of the human erythrocyte nucleoside transporter by immunoaffinity chromatography. *Biochem. J.* **255**, 243–249 (1988).
3. Kwong, F.Y.P. *et al.* Mammalian nitrobenzylthioinosine-sensitive nucleoside transport proteins: Immunological evidence that transporters differing in size and inhibitor-sensitivity share sequence homology. *J. Biol. Chem.* **267**, 21954–21960 (1992).
4. Barros, L.F. *et al.* Immunolocalisation of nucleoside transporters in human placental trophoblast and endothelial cells: Evidence for multiple transporter isoforms. *Pfluegers Arch. Eur. J. Physiol.* **429**, 394–399 (1995).
5. Kwong, F.Y.P. *et al.* Enzymic cleavage as a probe of the molecular structures of mammalian equilibrative nucleoside transporters. *J. Biol. Chem.* **268**, 22127–22134 (1993).
6. Huang, Q.Q. *et al.* Cloning and functional expression of a complementary DNA encoding a mammalian nucleoside transport protein. *J. Biol. Chem.* **269**, 17757–17760 (1994).
7. Che, M., Ortiz, D.F. & Arias, I.M. Primary structure and functional expression of a cDNA encoding the bile canalicular, purine-specific Na$^+$-nucleoside co-transporter. *J. Biol. Chem.* **270**, 13596–13599 (1995).
8. Craig, J.E., Zhang, Y. & Gallagher, M.P. Cloning of the *nupC* gene of *Escherichia coli* encoding a nucleoside transport system, and identification of an adjacent insertion element, IS *186*. *Mol. Microbiol.* **11**, 1159–1168 (1994).
9. Westh-Hansen, S.E., Jensen, N. & Munch-Petersen, A. Studies on the sequence and structure of the *Escherichia coli* K-12 *nupG* gene, encoding a nucleoside-transport system. *Eur. J. Biochem.* **168**, 385–391 (1987).
10. Ouellette, B.F.F. *et al.* Sequencing of chromosome I from *Saccharomyces cerevisiae*: Analysis of a 32 kb region between the *LTE1* and *SPO7* genes. *Genome* **36**, 32–42 (1993).
11. Williams, J.B. & Lanahan, A.A. A mammalian delayed-early response gene encodes HNP36, a novel, conserved nucleolar protein. *Biochem. Biophys. Res. Commun.* **213**, 325–333 (1995).
12. Huang, Q.Q., Harvey, C.M., Paterson, A.R., Cass, C.E. & Young, J.D. Functional expression of Na$^+$-dependent nucleoside transport systems of rat intestine in isolated oocytes of *Xenopus laevis*. *J. Biol. Chem.* **268**, 20613–20619 (1993).
13. Young, J.D. & Jarvis, S.M. Nucleoside transport in animal cells. *Biosci. Rep.* **3**, 309–322 (1983).
14. Bryson, H.M. & Sorkin, E.M. Cladribine. A review of its pharmacodynamic and pharmacokinetic properties and therapeutic potential in haematological malignancies. *Drugs* **46**, 872–894 (1993).
15. Rustum, Y.M. & Raymakers, R.A.P. 1-β-Arabinofuranosylcytosine in therapy of leukaemia: Preclinical and clinical overview. *Pharmacol. Ther.* **56**, 307–321 (1992).
16. Ross, S.R., McTavish, D. & Faulds, D. Fludarabine: A review of its pharmacological properties and therapeutic potential in malignancy. *Drugs* **45**, 737–759 (1993).
17. Kaye, S.B. Gemcitabine: Current status of phase I and II trials. *J. Clin. Oncol.* **12**, 1527–1531 (1994).
18. Mizushima, S. & Nagata, S. pEF-BOS, a powerful mammalian expression vector. *Nucleic Acids Res.* **18**, 5322 (1990).
19. Kozak, M. The scanning model for translation: An update. *J. Cell Biol.* **108**, 229–241 (1989).
20. Hofmann, K. & Stoffel, W. TMbase — A database of membrane spanning protein segments. *Biol. Chem. Hoppe-Seyler* **347**, 166 (1993).

ERRATA

Tethered epidermal growth factor as a paradigm for growth factor-induced stimulation from the solid phase

PHILIP R. KUHL & LINDA G. GRIFFITH-CIMA

Nature Medicine **2**, 1022–1027 (1996)

On page 1025 of the September issue, Figure 4 was incorrect. The correct version is displayed below.

Identification of primitive human hematopoietic cells capable of repopulating NOD/SCID mouse bone marrow: Implications for gene therapy

A. LAROCHELLE, J. VORMOOR, H. HANENBERG, J.C.Y. WANG, M. BHATIA, T. LAPIDOT, T. MORITA, B. MURDOCH, X.L. XIAO, I. KATO, D.A. WILLIAMS & J.E. DICK

Nature Medicine **2**, 1329–1337 (1996)

The abstract incorrectly stated that the SCID-repopulating cells were exclusively present in the $CD4^+CD8^-$ fraction; they were exclusively in the $CD34^+CD38^-$ fraction. We regret the error.

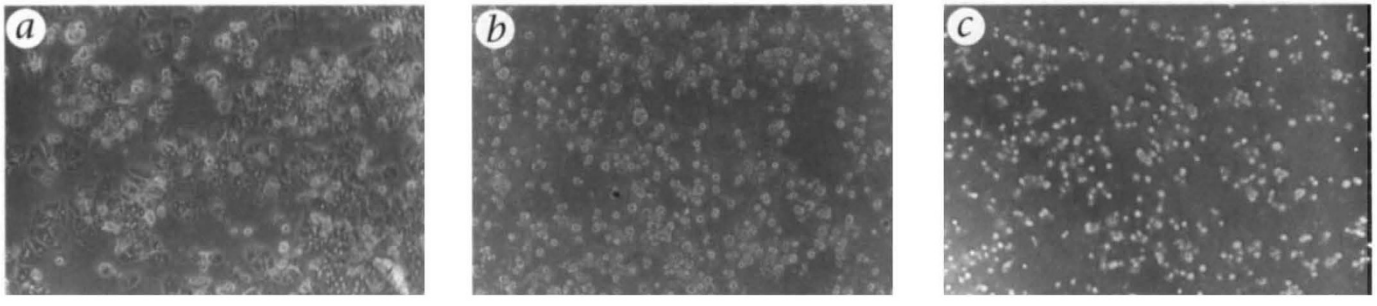

Fig. 4 Attenuation of cell spreading by small PEO stars and soluble or tethered EGF. Morphology of cells (×10 magnification, phase contrast) after 1 day in culture. Under all conditions, cell morphologies were invariant during the culture period after initial spreading had occurred. Two-thirds of the medium was replaced daily. *a*, Moderate inhibition of spreading of hepatocytes cultured on aminated glass slides grafted with star PEO (f = 70, M_a = 5200) from a 1% solution; *b*, complete inhibition of hepatocyte spreading by the addition of 10 ng/ml soluble EGF to hepatocytes cultured on slides grafted with star PEO (f = 70, M_a = 5200) from a 1% solution; *c*, complete inhibition of hepatocyte spreading by 4 ng/cm^2 tethered EGF, where EGF is linked to star PEO (f = 70, M_a = 5200) grafted to the surface from a 1% solution.