

Casting nets

Neutrophils are among the first responders recruited to sites of microbial infection. After activation, neutrophils release proteolytic enzymes and reactive oxygen species that can kill invading microbes. In *Science*, Brinkman *et al.* identify extracellular fibers that are rapidly released by neutrophils in response to bacteria or inflammatory mediators. These NETs (neutrophil extracellular traps) comprise chromatin fibers and granule proteins, such as neutrophil elastase and myeloperoxidase. Efficient microbial killing and toxin inactivation occurred within NETs and required chromatin to provide the essential structure for antimicrobial activity. NET formation is independent of apoptosis and, instead, seems to be an innate mechanism for the trapping and exposure of pathogens to high concentrations of toxic antimicrobial products.

LAD

Science **303**, 1532–1535 (2004)

Pick me

Data from cellular experiments and computer modeling show that ligation by peptide bound to major histocompatibility complex proteins (pMHCs) marks T cell receptors (TCRs) for internalization and degradation. However, the molecular events that drive this process are unclear. In the *Journal of Immunology*, La Gruta *et al.* suggest that an architectural change in the TCR-CD3 complex may be the indicator for eventual TCR downregulation. Ligation of pMHC exposes the TCR-associated CD3 ζ N terminus that is normally buried in the complex. This process requires the Src family protein tyrosine kinase and is a late event, presumably to allow signaling events to occur. The change in CD3 ζ and its likely dissociation from the TCR-CD3 complex may further reveal buried lysosomal targeting motifs on other CD3 subunits, which subsequently direct ligated TCR-CD3 complexes for degradation.

PTL

J. Immunol. **172**, 3662–3669 (2004)

The eleventh bell Tolls

Mammals express ten Toll-like receptors (TLRs) involved in the recognition of pathogen-associated molecular patterns (PAMPs). In *Science*, Ghosh and colleagues have cloned an additional TLR in mice. TLR11 expression was restricted to macrophages and epithelial cells of the liver, kidney and bladder. This TLR induced NF- κ B activation and a signaling pathway involving MyD88, IRAK and TRAF6. TLR11-expressing cells could not respond to known TLR ligands but could respond specifically to uropathogenic bacteria such as *Escherichia coli* 8NU. Consistently, TLR11-deficient mice were highly susceptible to kidney infection with *E. coli* 8NU. Thus, mouse TLR11 probably mediates protection against infection of the urogenital system. The exact ligand remains to be identified and it is unclear so far if humans express TLR11.

JDKW

Science **303**, 1522–1526 (2004)

CHIPS in staph arsenal

Staphylococcus aureus escapes host defense by producing an unknown factor that inhibits the migration of neutrophils toward chemo-attractants such as fMetLeuPhe and the C5a fragment of complement protein C5. In the *Journal of Experimental Medicine*, de Haas *et al.* identify and name this factor 'chemotaxis inhibitory protein of

S. aureus' (CHIPS). CHIPS specifically bound to neutrophils and monocytes but not lymphocytes. It inhibited fMetLeuPhe- or C5a-induced calcium mobilization. Despite having a greater potency in human than in mouse, CHIPS was able to prevent neutrophil influx in a mouse peritonitis model. The prevalence of the gene encoding CHIPS in clinical isolates of *S. aureus* suggests that it may be a useful target for anti-inflammatory therapy.

PTL

J. Exp. Med. **199**, 687–695 (2004)

Microbial assassins using TLR4

Recognition of microbial pathogens by TLR4 usually triggers activation of immune responses that lead to clearance of the infecting pathogen. In *Nature*, Hsu *et al.* report that certain virulent strains of bacteria evade immune responses by rerouting TLR4 signaling to induce apoptosis of responding macrophages. *Bacillus anthracis*, the causal organism of anthrax, as well as several *Salmonella* and *Yersinia* species, can induce TLR4 activation of protein kinase R (PKR), which blocks protein synthesis by inactivating the translation initiation factor eIF2 α . Bacterial-specific virulence factors blocked the normal TLR4-mediated activation of NF- κ B (which induces expression of host survival genes). Macrophages that lacked expression of TLR4, PKR or a mutated eIF2 α (S51A) that could not be phosphorylated survived bacterial infections. These results help to explain why these bacteria are such successful pathogens.

LAD

Nature **428**, 341–345 (2004)

MyD88 in cross-priming

Dendritic cells (DCs) can internalize exogenous antigen and present processed MHC class I epitopes to CD8⁺ cytolytic T lymphocytes (CTLs) by the still ill-defined process of cross-presentation. In the *Journal of Immunology*, Palliser *et al.* show that the adaptor protein MyD88 is required for DCs to induce CTL effector function in response to exogenous antigen, both *in vivo* and *in vitro*. MyD88-deficient DCs were less efficient than wild-type or TLR4-deficient DCs in processing internalized protein for MHC class I presentation. CD8⁺ T cells could proliferate in response to MyD88-deficient DCs that had been given exogenous protein, but had much diminished cytolytic activity and interferon- γ production. Thus, the efficiency of processing internalized antigen for CTL cross-priming is sensitive to MyD88 signals.

LAD

J. Immunol. **172**, 3415–3421 (2004)

Enzyme HINTs for regulation

MITF is a transcription factor that is essential for mast cell function. HINT, a member of the histidine triad family that can bind nucleotidyl substrates, associates with and suppresses MITF activity. In *Immunity*, Lee *et al.* showed that the enzyme lysyl-tRNA (LysRS) is a key regulator of MITF transcriptional activity. LysRS associates with MITF to form a multicomplex that includes HINT. Ap₄A, a nucleotidyl compound synthesized by LysRS, can bind HINT and induce HINT-MITF dissociation. In immunoglobulin E-activated mast cells, intracellular Ap₄A concentrations rapidly increase, therefore reducing the association of HINT with MITF. This renders MITF transcriptionally active. Thus, both LysRS and Ap₄A are key regulators of MITF activity in mast cells.

JDKW

Immunity **20**, 145–151 (2004)

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