

## A DRAM-atic end for T cells

Infection of CD4<sup>+</sup> T cells with human immunodeficiency virus (HIV) can result in a caspase-independent cell-death pathway, but the details of this process remain unclear. In *PLoS Pathogens*, Estaquier and colleagues link the autophagy-regulatory protein DRAM to HIV-triggered death of CD4<sup>+</sup> T cells. Infection of T cells with HIV induces DRAM expression in a manner dependent on the tumor suppressor p53 and subsequent activation of autophagy and cell death. However, autophagy is not critical for cell death; instead, after infection with HIV, DRAM associates with lysosomes and initiates destabilization of their membranes—a classic cell-intrinsic death initiator. How DRAM actually disrupts lysosomal membranes is unclear, but the cell death that results from DRAM activity substantially impairs viral infection. DRAM-mediated killing may therefore represent a cell-intrinsic mechanism for eliminating virus-infected cells. **ZF**

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## Peripheral fate

A pool of T cells responding to a given antigen differentiates into a mixture of qualitatively distinct kinds of effector T cells. In *Cell*, Jenkins and colleagues track the progeny of individual T cells to determine the signals that give rise to three different effector fates: T<sub>H1</sub>, T<sub>FH</sub> or germinal center T<sub>FH</sub> (GC-T<sub>FH</sub>). A pool of responding T cells produces a characteristic proportion of T<sub>H1</sub> cells, T<sub>FH</sub> cells and GC-T<sub>FH</sub> cells that depends on the type of experimental bacterial challenge and dose of the challenging antigen. Single naive T cells give rise to a distinct pattern of effector cells; however, averaging the response of all the potential responder cells results in a ratio of effector cells characteristic of the particular antigenic challenge. The fate of a given cell depends at least in part on the strength of signaling via the T cell antigen receptor, which thus has a key instructive role in determining the fate of effector cells. **ZF**

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## Affinity testing

The generation of high-affinity antibodies depends on the ability of B cells to acquire antigens from antigen-presenting cells. In *Science*, Tolar and colleagues show that B cells acquire antigens by dynamic contractions that pull out and engulf the presenting membranes. By comparing plasma membrane sheets and planar lipid bilayers, the authors show that flexibility of the presenting membrane is important. B cells use actomyosin contractility to pull out and invaginate the presenting membranes and internalize the antigen through clathrin- and actin-dependent processes. Diminishing the strength of myosin IIa contraction improves the internalization of low-affinity antigens. This suggests that although bonds between the B cell receptor (BCR) and antigen in small microclusters might break under the pulling forces, larger microclusters resist the contractile forces for long enough (20–30 seconds) to allow the association of clathrin and internalization. Thus, contractile forces mechanically test the strength of BCR binding and provide a mechanism for affinity discrimination. **IV**

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## Becoming pandemic

Mutant-selection experiments with the highly pathogenic avian influenza virus H5N1 have identified a virus that is transmissible to ferrets. In *Nature*, Xiong *et al.* report that the transmissible mutant virus has slightly higher binding affinity of hemagglutinin (HA) for its human receptor (sialic acid in  $\alpha$ 2,6-linkage to galactose on sugar side chains) and much less affinity for the avian receptor (sialic acid in  $\alpha$ 2,3-linkage). The crystal structure of the transmissible mutant HA in complex with receptor analogs shows that the mutant HA binds human receptors in the same folded-back conformation as did HA from viruses in the pandemics of 1918 (H1), 1957 (H2) and 1968 (H3), although the binding is weaker. This binding mode is different from that of wild-type H5 HA and arises from conformational changes due to a Q226L point substitution. The structural consequences of this substitution are similar to those of the pandemic viruses H2 and H3. **IV**

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## Mast cell–DC axis

Malaria continues to pose a considerable global health threat; hence, understanding the immune response to plasmodia is of utmost importance. In *Nature Medicine*, Guermonprez *et al.* describe a mast cell–dendritic cell (DC) axis associated with plasmodia infection. Infection of mice with *Plasmodium chabaudi* leads to release of the cytokine Flt3L from mast cells, which substantially increases the number of CD8 $\alpha^+$  DCs and, in turn, activates CD8<sup>+</sup> T cells. The authors trace those effects to higher expression of xanthine dehydrogenase in a yet-to-be identified radiosensitive cell population. That population responds to type I interferons produced during infection. Plasmodia-infected erythrocytes release hypoxanthine, which then can be converted to uric acid by xanthine dehydrogenase. Uric acid triggers mast cell degranulation and cleavage of membrane-tethered Flt3L. A similar response occurs in human patients with malaria, which suggests that this innate response to plasmodia is conserved. **LAD**

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## Systemic alarmin

Cochlin, a glycosylated extracellular matrix protein found in the inner ear, is also expressed in the spleen, but its physiological role there remains unknown. In *Immunity*, Py *et al.* show cochlin is secreted by follicular DCs. After bacterial infection, cochlin is proteolytically cleaved by aggrecanase 1 (whose expression is induced by tumor-necrosis factor), which results in the release of a bioactive amino-terminal fragment that functions systemically to activate innate immune responses. *Cochl*<sup>-/-</sup> mice have normal splenic and lymph node structure and adaptive immunity but fail to control infection with *Pseudomonas* or *Staphylococcus*. A defect in the recruitment of monocytes and neutrophils leads to greater bacterial burdens, which ultimately kills the *Cochl*<sup>-/-</sup> mice. Thus, cleavage of cochlin releases a systemic alarmin that contributes to innate immunity. **LAD**

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Written by Laurie A. Dempsey, Zoltan Fehervari & Ioana Visan