

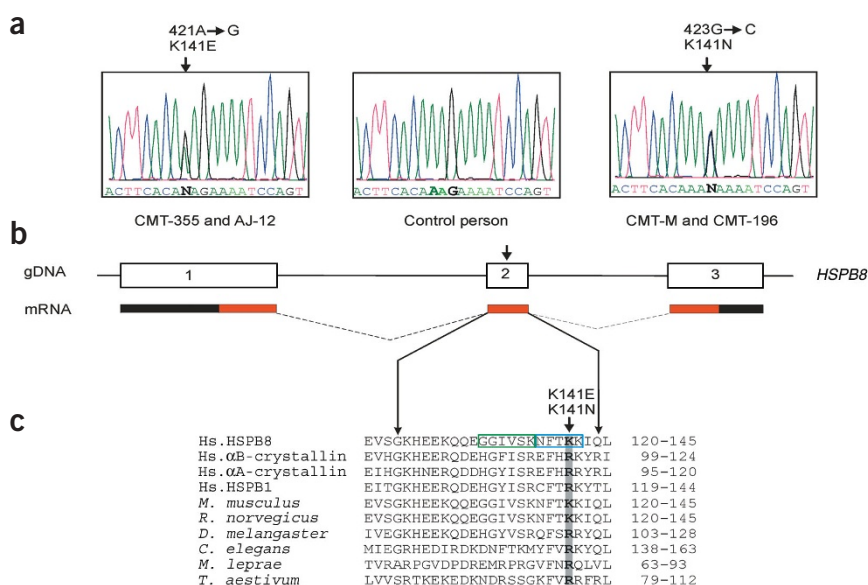
# Hot-spot residue in small heat-shock protein 22 causes distal motor neuropathy

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Distal hereditary motor neuropathies are pure motor disorders of the peripheral nervous system resulting in severe atrophy and wasting of distal limb muscles<sup>1</sup>. In two pedigrees with distal hereditary motor neuropathy type II linked to chromosome 12q24.3, we identified the same mutation (K141N) in small heat-shock 22-kDa protein 8 (encoded by *HSPB8*; also called HSP22). We found a second mutation (K141E) in two smaller families. Both mutations target the same amino acid, which is essential to the structural and

functional integrity of the small heat-shock protein  $\alpha$ A-crystallin<sup>2</sup>. This positively charged residue, when mutated in other small heat-shock proteins, results in various human disorders<sup>3,4</sup>. Coimmunoprecipitation experiments showed greater binding of both HSPB8 mutants to the interacting partner HSPB1. Expression of mutant HSPB8 in cultured cells promoted formation of intracellular aggregates. Our findings provide further evidence that mutations in heat-shock proteins have an important role in neurodegenerative disorders.

**Figure 1** DNA and protein sequence analysis of *HSPB8* and HSP22. (a) Electropherogram showing the 421A→G and 423G→C sequence variations in *HSPB8* exon 2, resulting in the K141E and K141N missense mutations in families AJ-12 and CMT-355 and families CMT-M and CMT-196, respectively. The corresponding genomic sequence of a control person is shown in the middle. (b) *HSPB8* spans ~16 kb and contains three exons. The  $\alpha$ -crystallin domain is indicated in red. (c) ClustalW multiple protein alignment of *HSPB8* orthologs. The protein sequences show the central  $\alpha$ -crystallin domain (Hs.HSPB8 protein fragment 120–145) surrounding the K141E and K141N mutations. Orthologs in *Homo sapiens* (Hs), *Mus musculus*, *Rattus norvegicus*, *Drosophila melanogaster*, *Mycobacterium leprae*, *Caenorhabditis elegans* and *Triticum aestivum* are shown. The putative N-myristoylation site is boxed in green; the N-glycosylation and protein kinase C motifs are boxed in blue. Both mutations are indicated by an arrow.

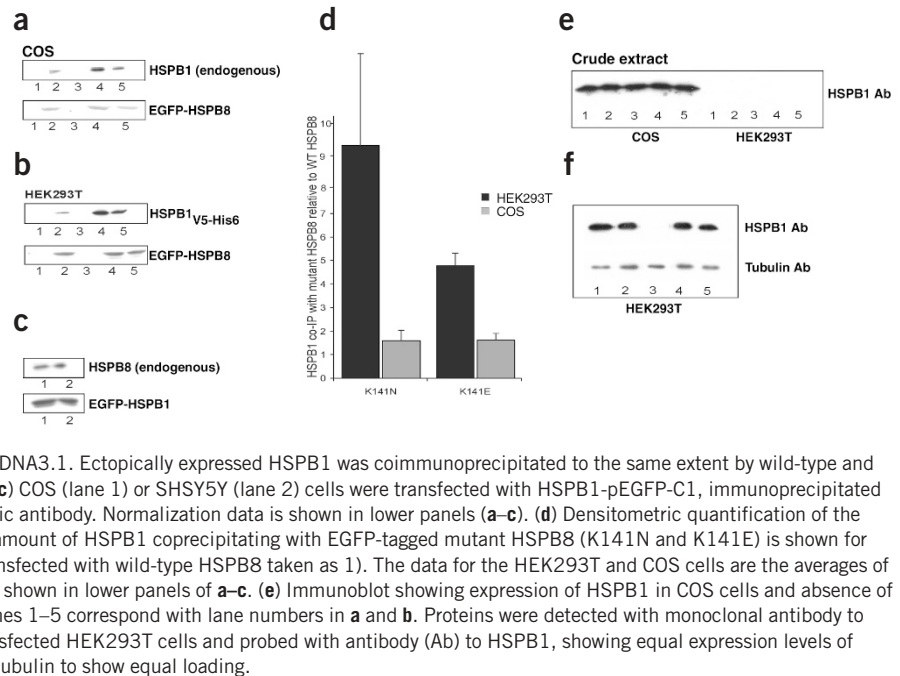


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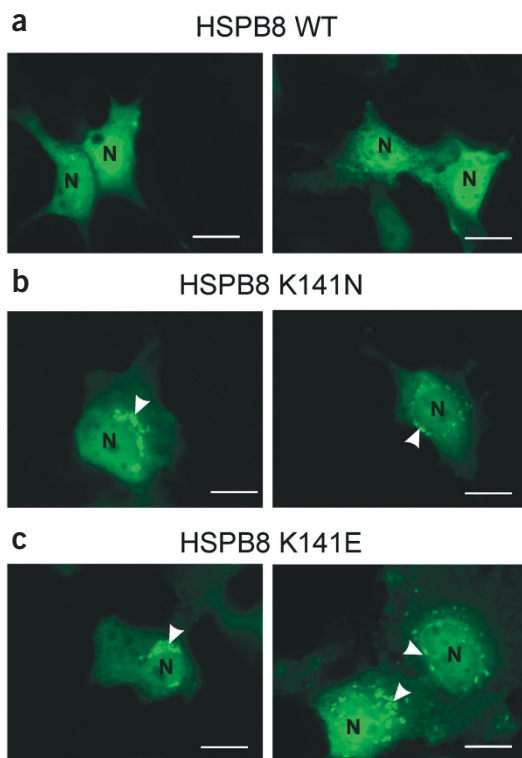
**Figure 2** HSPB8-HSPB1 coimmunoprecipitation experiments. **(a–c)** Expression of mutant HSPB8 does not disrupt interaction with HSPB1. **(a)** COS cells were transiently transfected with EGFP-tagged wild-type or mutant HSPB8, immunoprecipitated and analyzed for the presence of HSPB1 by immunoblotting with a monoclonal antibody. A stronger HSPB1 signal was observed for HSPB8-K141N (lane 4) and HSPB8-K141E (lane 5) than for wild-type HSPB8 (lane 2). Lane 1 (pEGFP-C1) and lane 3 (untransfected cell) represent controls. **(b)** The same experiment was done using HEK293T cells cotransfected with pcDNA3.1-HSPB1. HEK293T cells do not express HSPB1. Lane 1, pEGFP-C1 and pcDNA3.1; lane 2, HSPB8- pEGFP-C1 and HSPB1-pcDNA3.1; lane 3, untransfected cells; lane 4, K141N-pEGFP-C1 and HSPB1-pcDNA3.1; lane 5, K141E-pEGFP-C1 and HSPB1-pcDNA3.1. Ectopically expressed HSPB1 was coimmunoprecipitated to the same extent by wild-type and mutant HSPB8 as endogenous HSPB1 in COS cells. **(c)** COS (lane 1) or SHSY5Y (lane 2) cells were transfected with HSPB1-pEGFP-C1, immunoprecipitated and analyzed for the presence of HSPB8 with a specific antibody. Normalization data is shown in lower panels **(a–c)**. **(d)** Densitometric quantification of the immunoprecipitation experiments shown in **a–c**. The amount of HSPB1 coprecipitating with EGFP-tagged mutant HSPB8 (K141N and K141E) is shown for HEK293T and COS cells (normalized ratio in cells transfected with wild-type HSPB8 taken as 1). The data for the HEK293T and COS cells are the averages of three independent experiments. Normalization data is shown in lower panels of **a–c**. **(e)** Immunoblot showing expression of HSPB1 in COS cells and absence of expression in HEK293T cells. Cytosolic proteins in lanes 1–5 correspond with lane numbers in **a** and **b**. Proteins were detected with monoclonal antibody to HSPB1. **(f)** Western blot on lysates obtained from transfected HEK293T cells and probed with antibody (Ab) to HSPB1, showing equal expression levels of HSPB1. The blot was then reprobed with antibody to tubulin to show equal loading.



Distal hereditary motor neuropathy (HMN; OMIM 158590) is genetically heterogeneous and has been associated with eight loci and three genes so far<sup>5–7</sup>. We previously mapped autosomal dominant distal HMN type II to a 5-Mb candidate region at 12q24.3 in a Belgian family<sup>8,9</sup>. Distal HMN type II is an exclusive lower motor neuron disease without sensory loss, with an onset age of 15–25 years. The presenting symptoms are paresis of the extensor muscles of the big toe and later of

the extensor muscles of the feet. The disease progresses rapidly, and complete paralysis of all distal muscles of the lower extremities occurs within 5 years. Affected individuals have chronic neurogenic alterations in electromyography<sup>10</sup>.

Here, we identified a large Czech family with 34 affected individuals (**Supplementary Fig. 1** online) with a phenotype markedly similar to that of the previously reported Belgian family with distal HMN type II. In both families, we did a haplotype analysis using short tandem repeat (STR) markers and identified several recombinants, narrowing the candidate region from 5 Mb (ref. 8) to 1.7 Mb between *D12S349* and *PLA2G1B* (**Supplementary Fig. 2** online). This refined region contains 9 known and 14 predicted genes, of which 5 known genes have previously been excluded<sup>11,12</sup>. We selected four of the other genes for mutation analysis: *PRKAB1* (protein kinase, AMP-activated beta 1), *CIT* (citron; rho-interacting, serine-threonine kinase), *SIRT4* (sirtuin 4) and *HSPB8* (heat shock 22-kDa protein 8, also called *HSP22*, *H11* and *E2IG1*). We sequenced all known exons and intron-exon boundaries of each of these genes. In the Belgian and Czech families, we identified a heterozygous transversion, 423G→C, in exon 2 of *HSPB8* (resulting in the amino acid substitution K141N; **Fig. 1a,b**). In two other families (Bulgarian and English) with distal HMN, we found a heterozygous transition, 421A→G, involving the same lysine residue (amino acid substitution K141E; **Fig. 1a,b**). Both missense mutations cosegregated perfectly with the distal HMN type II phenotype in all families (**Supplementary Fig. 1** online), and both were absent in 400 Caucasian control chromosomes. Because the Belgian and Czech families and the Bulgarian and English families shared the



**Figure 3** Accumulation of cytoplasmic aggregates in COS cells transfected with K141N and K141E mutant HSPB8. The cells were processed for immunofluorescence analysis 48 h after transfection with pEGFP-C1 wild-type or mutant HSPB8. White arrows indicate HSPB8 aggregates (**b,c**), which were absent in cells transfected with wild-type (WT) HSPB8 (**a**). At least 200 cells were counted in three randomly chosen microscopic fields by two independent investigators. N, nucleus. Scale bars, 30  $\mu$ m.

mutations K141N and K141E, respectively, we carried out haplotype analysis with 11 STR markers and four intragenic *HSPB8* single-nucleotide polymorphisms (SNPs). The results suggest that the families are genetically not closely related (**Supplementary Fig. 2** online).

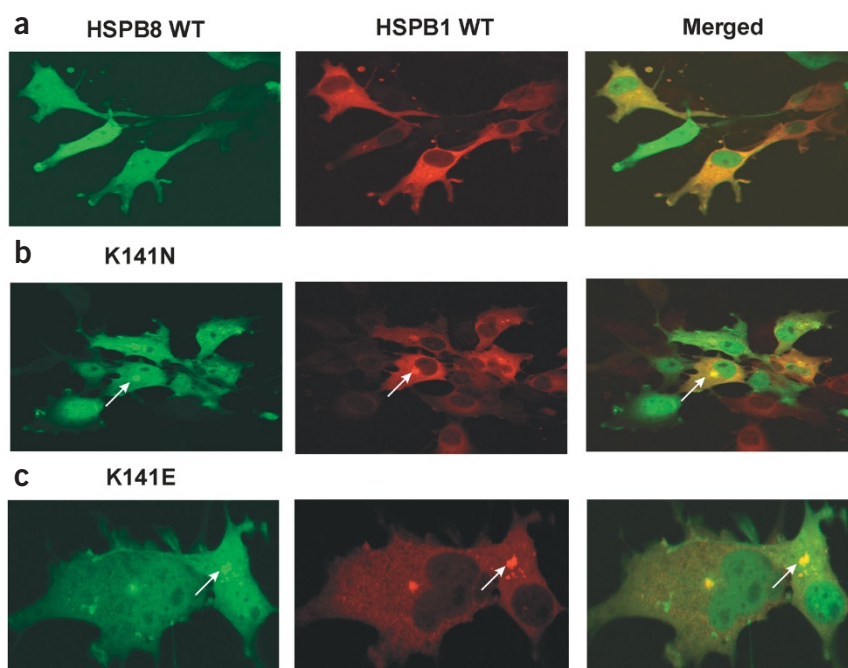
HSPB8 belongs to the superfamily of mammalian small heat-shock proteins or stress proteins<sup>13–16</sup>. Members of this superfamily share a conserved  $\alpha$ -crystallin domain in their C-terminal part and the WDPF motif in their N-terminal part, whereas other parts of the sequence (N-terminal halves and extreme C-terminal tails) are more variable<sup>17,18</sup>. The missense mutations K141N and K141E are located in the central  $\alpha$ -crystallin domain of HSPB8. Alignment of HSPB8 orthologs showed that the targeted lysine residue is highly conserved in the  $\alpha$ -crystallin domain (**Fig. 1c**). This positively charged amino acid is essential for the structural and functional integrity of  $\alpha$ A-crystallin<sup>2</sup>. We confirmed the HSPB8 expression profile as reported in the UniGene Database (Hs.111676 and Mm.21549) and in ref. 19 (**Supplementary Fig. 3** online). In the nervous system, we detected high expression in spinal cord, specifically in motor and sensory neurons (**Supplementary Fig. 3** online).

HSPB8 is a phosphoprotein that interacts with HSPB1 and might be involved in the modulation of HSPB1 activities<sup>13,19</sup>. We investigated the effect of the K141N and K141E mutations of HSPB8 on its interaction with HSPB1 by coimmunoprecipitation. We transiently expressed enhanced green fluorescent protein (EGFP)-tagged wild-type and mutant HSPB8 in simian fibroblasts (COS) and human embryonic kidney cells (HEK293T). We immunoprecipitated fusion proteins using a polyclonal antibody to EGFP and analyzed immunoprecipitates for the presence of HSPB1 by western blotting. In COS cells, endogenous HSPB1 coimmunoprecipitated with wild-type HSPB8 (**Fig. 2a**). Conversely, immunoprecipitation of EGFP-tagged HSPB1 pulled down endogenous wild-type HSPB8 from COS or human neuronal (SHSY5Y) cells (**Fig. 2c**). Immunoprecipitation of the HSPB8 mutants from COS lysates also showed association with endogenous HSPB1 (**Fig. 2a**), indicating that the missense mutations do not abolish interaction between HSPB8 and HSPB1.

Both HSPB8 mutants pulled down more HSPB1 protein than wild-type HSPB8 did. We quantified the immunoprecipitation data from three independent experiments by enhanced chemiluminescence detection densitometry (**Fig. 2d**). To verify the specificity of the interaction between HSPB8 and HSPB1, we cotransfected HEK293T cells that do not express HSPB1 (**Fig. 2e**) with EGFP-HSPB8 (wild-type or mutant) and HSPB1 carrying V<sub>5</sub> and His<sub>6</sub> epitope tags. The results were identical to those obtained with COS cells: both wild-type and mutant HSPB8 interacted with HSPB1, and both mutants pulled down more HSPB1 than wild-type HSPB8 did (**Fig. 2b,d**). These differences were not due to varying expression levels of HSPB8 or HSPB1. Western blots with antibodies to HSPB8 and HSPB1 showed equal expression levels of EGFP-tagged wild-type and mutant HSPB8 and HSPB1 in both COS and HEK293T cells (**Fig. 2a,b,f**). Rabbit and mouse IgGs did not immunoprecipitate the HSPB8-HSPB1 complex (data not shown).

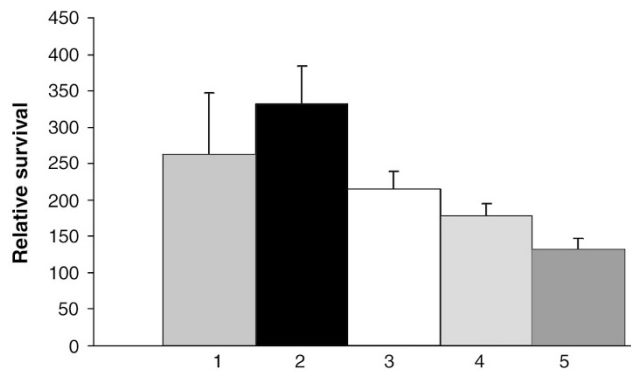
The residue Lys141 in HSPB8 that is mutated in families with distal HMN type II corresponds to the residue Arg120 that is mutated in  $\alpha$ B-crystallin, causing a familial desmin-related myopathy (DRM; OMIM 601419; ref. 3). In DRM, intracellular desmin aggregates form in cells with the R120G mutation<sup>20</sup>. To test whether K141N or K141E mutant HSPB8 could also lead to the formation of aggregates, we carried out immunofluorescence experiments in transfected COS cells (**Fig. 3**). After 48 h in COS cells, EGFP-tagged wild-type HSPB8 showed a homogeneous distribution throughout the cytoplasm and nucleus (**Fig. 3a**). In contrast, the mutant forms led to cytoplasmic or perinuclear HSPB8-labeled aggregates (**Fig. 3b,c**). The percentage of aggregate-positive cells was significantly higher in cells transfected with K141N HSPB8 ( $62\% \pm 2\%$ ) or with K141E HSPB8 ( $59\% \pm 4\%$ ) than in those transfected with wild-type HSPB8 ( $3\% \pm 0.3\%$ ;  $P = 1.5 \times 10^{-6}$  and  $2.8 \times 10^{-5}$ , respectively; data are from three independent experiments). Western-blot analysis 48 h after transfection showed similar levels of expression of HSPB8 in COS cells transfected with wild-type and mutant forms of HSPB8 (data not shown). Colocalization studies showed that wild-type HSPB8 and HSPB1 were located in the cytoplasm of COS cells. In cells transfected with HSPB8 mutants, we observed colocalization between HSPB8 aggregates and HSPB1 (**Fig. 4**). We investigated whether the aggregates reduced cell viability. When transfected into N2a neuronal cells, K141E mutant HSPB8 significantly reduced cell viability, as compared with wild-type HSPB8, measured 48 h after transfection ( $P = 0.007$ ). K141N mutant HSPB8 also reduced viability, but to a lesser extent ( $P = 0.043$ ) (**Fig. 5**).

We showed that mutations in *HSPB8* are closely associated with motor neuropathy, specifically distal HMN type II. The missense mutations in the HSPB8 protein do not disrupt interaction with HSPB1, but strengthen it, leading to the formation of aggregates.



**Figure 4** Confocal microscopic analysis of cells expressing HSPB8 and HSPB1. COS cells were cotransfected with pEGFP-C1 wild-type or mutant HSPB8 (green) and pcDNA3.1-HSPB1 (red). The cells were processed for immunofluorescence analysis 48 h after transfection and visualized with a laser scanner microscope. (a) Wild-type (WT) HSPB8, wild-type HSPB1 and merged confocal image. (b) K141N mutant HSPB8, wild-type HSPB1 and merged image. (c) K141E mutant HSPB8, wild-type HSPB1 and merged image. Arrows indicate the aggregates.





**Figure 5** Survival assay of N2a neuronal cells transfected with wild-type and mutant HSPB8 constructs. The relative survival was measured after 48 h. 1, control (empty pEGFP-C1 vector); 2, control (no vector); 3, pEGFP-C1 wild-type HSPB8; 4, pEGFP-C1 HSPB8 K141N; 5, pEGFP-C1 HSPB8 K141E. Data shown are mean  $\pm$  s.e.m. of five independent experiments. *P* values: HSPB8 K141N, 0.043; HSPB8 K141E, 0.007. Statistical significance was determined using a Student's *t*-test.

Missense mutations in other sHSPs, namely  $\alpha$ A-crystallin (R116C) and  $\alpha$ B-crystallin (R120G) cause an autosomal dominant congenital cataract (OMIM 123580; ref. 4) and DRM<sup>3</sup>, respectively. Arg116 and Arg120 correspond to Lys141 in HSPB8. The R120G mutation in  $\alpha$ B-crystallin associated with DRM causes the formation of large desmin aggregates<sup>3,20</sup>, and the R116C mutation in  $\alpha$ A-crystallin that causes congenital cataract shows a higher affinity for hetero-aggregation with  $\alpha$ B-crystallin<sup>21</sup>. Whether the aggregates of HSPB8 and HSPB1, observed as a short-term result in *in vitro* studies, also occur during the slow degeneration of motor neurons in individuals with distal HMN type II is an open question that can be addressed only by the generation of animal models carrying the HSPB8 mutations. Some sHSPs have crucial roles in neuronal apoptosis and muscle function<sup>3,22</sup>. R116C in  $\alpha$ A-crystallin diminishes its protective ability against stress-induced lens epithelial cell apoptosis<sup>23</sup>, and  $\alpha$ B-crystallin negatively regulates apoptosis by inhibiting caspase-3 activation<sup>24</sup>. HSPB2 associates specifically with and activates the myotonic dystrophy protein kinase, which causes myotonic dystrophy (OMIM 160900; ref. 25). Mutation of Arg116 to an acidic residue caused marked changes in protein structure, suggesting that a positive charge must be preserved at this position for the structural and functional integrity of  $\alpha$ A-crystallin<sup>2</sup>. The mutations of HSPB8 at Lys141 have a gain-of-function effect, which may lead to dysfunction of axonal transport and dysregulation of the cytoskeleton, causing motor neuron death in distal HMN. Other important unknown mechanisms may also be involved in the pathogenesis of distal HMN. Notably, transcriptional and post-translational regulation of HSPB1, the molecular partner of HSPB8, is necessary for sensory and motor neuron survival after peripheral nerve injury<sup>22</sup>. HSPB1 was upregulated in a transgenic model of amyotrophic lateral sclerosis<sup>26</sup>. Our finding that HSPB8 has a pivotal role in the biology of the motor axon suggests that upregulation of HSPB8 is a potential strategy to protect the motor neuron from degeneration.

## METHODS

**Affected individuals.** Our study included four families with distal HMN (Supplementary Fig. 1 online). In the Belgian family (CMT-M), we previously identified the locus on chromosome 12q24.3 linked with distal HMN type II (refs. 8–10). The Czech (CMT-196), English (CMT-355) and Bulgarian (AJ-12) families were recently identified. From affected family members and control persons, we isolated genomic DNA from total blood samples using a standard

extraction protocol. Informed consent was obtained from all family members, and the study was approved by the Institutional Review Boards of the Universities of Antwerp, Oxford, Prague and Sofia.

**Genotyping and genetic linkage analysis.** We carried out PCR amplification with dye-labeled STR primers on a DYAD thermocycler (MJ Research). We analyzed fragments on an ABI3700 DNA sequencer with the ABI GENESCAN 3.1 and GENOTYPER 2.1 software (Applied Biosystems). For analysis of linkage to HSPB8 and STR markers, we computed two-point lod scores using the MLINK program of the FASTLINK package. We assumed autosomal dominant inheritance, a disease frequency of 1 in 10,000, equal male and female recombination rates and complete penetrance. We calculated lod scores assuming equal allele frequencies for all markers. We established the order of markers on the genetic and physical maps by consulting the Genome Database and the National Center for Biotechnology Information genome database.

**Mutation analysis.** We used the National Center for Biotechnology Information Entrez Genome Map Viewer, the Ensembl Human Genome Server and the GenBank database to find known genes, expressed-sequence tags and putative new genes in the region linked with distal HMN type II. We determined the exon-intron boundaries of the candidate sequences by BLAST searches against the high-throughput genome sequences. All exons of the genes *PRKAB1*, *CIT*, *SIRT4* and *HSPB8* were amplified by PCR using intronic primers (sequences available on request). We sequenced PCR products using the DYEnamic ET Terminator Cycle Sequencing Kit (Amersham Pharmacia Biotech). Sequence reactions were loaded on the ABI3700 sequencer (Applied Biosystems). We collected and analyzed data using the ABI DNA sequencing analysis software, version 3.6.

**Expression analysis.** We obtained the plasmid clone IRALp962K0712, containing the complete human HSPB8 cDNA sequences, from RZPD (The Resource Center of the German Human Genome Project). We used gene-specific primers to construct a HSPB8 cDNA probe of 800 bp and used this probe to hybridize the Human 12- and 8-lane Multiple Tissue and Brain Northern blot (Clontech).

We extracted total RNA from mouse muscle (Navy Medical Research Institute) using the Totally RNA Kit (Ambion). We carried out RT-PCR using the Random Primer DNA Labeling System (Life Technologies). The full-length mouse HSPB8 cDNA was used as a probe to hybridize the Mouse Multiple Tissues and Embryos Northern blot (Clontech). We also hybridized northern blots with an ACTB cDNA probe (Clontech) as a control for RNA loading.

We isolated ventral horns and dorsal root ganglia from 13-d-old mouse embryos and extracted total RNA using the Totally RNA Kit (Ambion). We carried out RT-PCR using the SuperScript III First-Strand Synthesis System for RT-PCT (Invitrogen). We used mouse HSPB8 cDNA primers (sequences available on request) to amplify an HSPB8 cDNA fragment of 687 bp.

**cDNA cloning and mutagenesis.** We cloned cDNA encoding full-length wild-type human HSPB8 or HSPB1 (refs. 19,27) as a HindIII fragment into the pEGFP-C1 (Clontech) and pcDNA3.1/V5/His6 TOPO (Invitrogen) vectors by PCR using the RZPD plasmids IRALp962K0712 (HSPB8) and IRALp962H201 (HSPB1) as templates. For site-directed mutagenesis of the HSPB8 mutations 423C→G and 421A→G, we used the QuikChange site-directed mutagenesis kit (Stratagene). All constructs were verified by direct DNA sequencing.

**Immunoprecipitation.** We transfected HEK293T cells with calcium phosphate, COS cells with polyethylenimine and SHY5Y cells with lipofectamine 2000 according to standard procedures using 6–18  $\mu$ g of DNA construct. After 24 h, we washed cells twice with ice-cold phosphate-buffered saline (2.7 mM KCl, 1.47 mM KH<sub>2</sub>PO<sub>4</sub>, 137 mM NaCl, 8.1 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 7.4) and lysed them in phosphate-buffered saline with 0.5% Nonidet P-40, 5 mM levamisole, 10 mM sodium fluoride, 8 mM sodium- $\beta$ -glycerophosphate and a protease inhibitor cocktail mix (Roche Diagnostics). We disrupted cells by sonication and centrifuged the crude extract at 4  $^{\circ}$ C for 10 min (14,000 r.p.m.). We incubated 1 mg of proteins overnight with affinity-purified antibodies to EGFP and then for 4 h with protein G-Sepharose (Amersham Pharmacia Biotech). We washed the beads five times with lysis buffer, boiled them for 5 min in Laemmli sample buffer and fractionated the proteins by SDS-PAGE followed by western blotting. We visualized proteins by enhanced chemiluminescence detection (ECL kit; Amersham Pharmacia Biotech).

**Immunostaining and microscopy.** We viewed the cells directly for EGFP fluorescence after fixing them with 3% paraformaldehyde in phosphate-buffered saline for 20 min at room temperature. We examined cells using a Zeiss Axioplan II epifluorescence microscope equipped with a 63 objective. We captured images using a cooled CCD Axiocam Camera and KS100 software (Zeiss).

**Miscellaneous.** We affinity-purified antibodies to EGFP according to standard procedures<sup>28</sup>. We determined protein concentrations by the method of Bradford<sup>29</sup> using bovine serum albumin as a standard. We carried out SDS-PAGE according to Matsudaira and Burgess<sup>30</sup>.

**Cell viability test.** We cultured N2a neuroblastoma cells (CCL131; American Type Culture Collection) in a 1:1 mix of Dulbecco's modified Eagle medium and F12 medium with glutamax (Gibco Invitrogen) supplemented with nonessential amino acids, penicillin (100 U ml<sup>-1</sup>), streptomycin (100 g ml<sup>-1</sup>) and 10% fetal calf serum. We transfected cells with 5 g of mutated or wild-type *HSPB8* plasmid DNA using the Nucleofector technology (Amaxa). We collected exponentially growing N2a cells and resuspended them at a concentration of 2 × 10<sup>6</sup> cells per 100 l in a cuvette containing Nucleofector solution T. After transfection, we resuspended cells in serum-free medium and plated them in a 96-well plate. Using a fluorescence-activated cell sorter, we determined transfection efficiency after 24 h and 48 h (92% ± 4% and 88% ± 8%, respectively). We determined cell viability 3 h, 24 h and 48 h after transfection using the CellTiter 96 AQueous Cell Proliferation Assay (Promega). Western blots showed mutant and wild-type *HSPB8* proteins to be expressed at comparable levels.

**URLs.** The FASTLINK package is available at <http://www.cs.rice.edu/~schaffer/fastlink.html>. The Genome Database is available at <http://www.gdb.org/>, and the National Center for Biotechnology Information genome database is available at <http://www.ncbi.nlm.nih.gov/LocusLink/>. The National Center for Biotechnology Information Entrez Genome Map Viewer is available at [http://www.ncbi.nlm.nih.gov/mapview/map\\_search.cgi?chr=hum\\_chr.inf&query/](http://www.ncbi.nlm.nih.gov/mapview/map_search.cgi?chr=hum_chr.inf&query/). The Ensembl Human Genome Server is available at <http://www.ensembl.org/>, and the GenBank database is available at <http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=Nucleotide/>. The RZPD is available at <http://www.rzpd.de/>. ClustalW multiple protein alignment is available at [http://npsa-pbil.ibcp.fr/cgi-bin/npsa\\_automat.pl?page=npsa\\_clustalw.html](http://npsa-pbil.ibcp.fr/cgi-bin/npsa_automat.pl?page=npsa_clustalw.html).

**Accession numbers.** GenBank protein sequences: human *HSPB8*, NP\_055180; mouse *HSPB8*, NP\_109629; rat *HSPB8*, NP\_446064; *Drosophila melanogaster* *HSPB8*, NP\_729477; *Caenorhabditis elegans* *HSPB8*, NP\_509045; *Triticum aestivum* (bread wheat) GI 17942916, 1GMEA; *Mycobacterium leprae* Hsp16.7, P12809. The GenBank genomic sequence NT\_009775 includes the genes *PRKAB1* (NM\_006253), *CIT* (NM\_007174), *SIRT4* (NM\_012240) and *HSPB8* (NM\_014365). *HSPB8* SNPs in the National Center for Biotechnology Information Entrez SNP database: Rs2278181, Rs2278182, Rs11038 and Rs1133026.

*Note: Supplementary information is available on the Nature Genetics website.*

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#### COMPETING INTERESTS STATEMENT

The authors declare that they have no competing financial interests.

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