Widespread expression of the testis-determining gene *SRY* in a marsupial

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Correspondence should be addressed to J.L.H. There is compelling evidence from mutation analysis¹ and transgenesis² that the SRY gene isolated from human³ and mouse⁴ encodes the testisdetermining factor on the mammalian Y chromosome. However, how SRY achieves this function is unclear. Although marsupials have been separated from eutherian mammals for ~100 million years, homologues of SRY have been localised to the Y chromosome of two unrelated marsupial species. the tammar wallaby and the Darling Downs dunnart⁵. Gonadal development is fundamentally similar in eutherian and marsupial mammals, but the timing of morphological events is different. Fetal Sry transcripts are confined to somatic cells of the male mouse genital ridge between 10.5-12.5 days post coitum⁶, corresponding with the onset of testis differentiation. Analysis of Sry gene expression in the genital ridge of normal and germ celldeficient fetal mice has established that this gene acts in the somatic cell lineage⁶, and is presumed to induce the formation of Sertoli cells7,8. This assumption can be tested more critically in the tammar, where the equivalent stages of testis differentiation are observed over a 7-day period⁹. We have examined the relationship of SRY expression to testis differentiation in the tammar wallaby. We show the the marsupial SRY gene cannot be exclusively coupled to Sertoli cell differentiation, as this gene is expressed in the male fetus from several days before genital ridge formation until 40 days after birth. SRY transcripts are also present in a variety of extra-gonadal tissues in the developing young and adult male, a pattern of SRY expression similar to that observed in humans. These data indicate that, in addition to a role in testis determination, SRY may have other functions.

Total RNA was extracted from pooled tammar fetuses and neonatal gonads and analysed for SRY expression using reverse transcription and polymerase chain reaction (RT-PCR). In the tammar, the gonadal primordium is first apparent at day 21 of the 26.5 day gestation. By day 25 the cytoplasmic:nuclear ratio in male pre-Sertoli

cells is greater than in female cells, but it is not until two days post-partum that seminiferous cords are obvious with putative Leydig cells appearing in the interstitium⁹. We initially tested for the presence of SRY transcripts from the day after the genital ridge is established (day 22 of gestation) until two days after the development of seminiferous tubules (day 4 post partum). Surprisingly, SRY expression was detected throughout this entire developmental period in male fetal tissue and in gonads from male pouch young (Fig. 1a). We therefore extended our sampling period from day 19 of gestation (early head-fold; ~20 somites) to 40 days after birth and established that in the tammar SRY is expressed from at least 6 days before the appearance of Sertoli cells until long after the immature testis has fully differentiated (Fig. 1b). Thus, expression of the tammar SRY gene does not result in immediate Sertoli cell differentiation.

Examination of the distribution of tammar SRY transcripts showed that SRY mRNA is expressed in all male fetal and pouch young tissues assayed, with the exception of liver. Female tissues also were tested to ensure that the SRY primers did not amplify sequences of SRY-related genes common to both sexes and, as expected, no bands were seen (Figs 1 and 2). SRY transcripts were observed not only in fetal gonad with attached mesonephros, but also in heart and brain (Fig. 2a), and skin, limb bud, phallus, adrenal, kidney and lung tissues (data not shown). In male pouch young, SRY transcripts were found in gonad and brain, at a lower level in heart and mesonephros (Fig. 2a), and in phallus, kidney and lung (data not shown). In adult males, SRY mRNA was detected in testis, epididymis and brain, at lower levels in kidney and heart, and was undetectable in prostate or liver (Fig. 2b). RNase A treatment of RT(+/-) products, as described¹⁰, confirmed that the RT(+) bands in Figs 1 and 2 correspond to transcripts and that the presence of a faint band for some RT(-) samples represents genomic DNA contamination (data not shown). The pattern

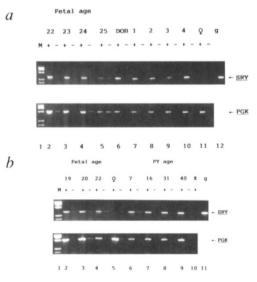


Fig. 1 Time course of tammar *SRY* expression. Total RNA preparations from fetal tissue (posterior to the forelimb bud) or pouch young (PY) gonads were used in a reverse transcription reaction, with (+) and without (-) reverse transcriptase (RT). Separate PCR

reactions were performed, using the RT products or tammar genomic DNA, with primer pairs for SRY (upper panel) or PGK (lower panel). A 405-bp control PGK band was present in all RT(+) samples and a 296-bp SRY band was found for all male RT(+) and genomic DNA samples. *a*, Lanes 1, molecular size markers (bp) 577, 500, 404, 242 and 190; 2, one male fetus at day 22; 3, three male fetuses at day 23; 4, two male fetuses at day 24; 5, one male fetus at day 25; 6–10, gonads from one male PY at, day of birth (DOB) and days 1–4 after birth; 11, gonads from a single female 1 day after birth; 12, male genomic DNA. *b*, Lanes 1, molecular size markers; 2, two male fetuses at day 29, 4, one male fetuses at day 22; 6-9, gonads from a single male PY at 7, 16, 31 and 40 days, respectively, 11, male genomic DNA. A PGK band (lower panel) is present for all samples except when template was omitted (lane 10).