

# LETTERS TO NATURE

## Untranslated T7 Phage mRNA is stabilized in *suA* Host

It has been shown<sup>1</sup> that relief from the polar effect of nonsense mutations in the tryptophan operon by the *suA* allele<sup>2</sup> is associated with increased survival of mRNA operator distal to the nonsense mutation. Experiments described here confirm the effect of *suA* on mRNA distal to amber mutations and directly demonstrate the complete mRNA molecule only in the presence of the *suA* mutation.

Phage-specific RNA from T7-infected cells has been resolved into multiple discrete species by gel electrophoresis<sup>3</sup> and one of these species has been identified as the transcription product of gene 1 (ref. 4). This gene is required for transcription of the majority of the T7 genome, so the RNA complement is quite simple in cells infected with T7 carrying mutations in this gene<sup>4</sup>. For this reason, the metabolism of this mRNA was examined in *su<sup>-</sup>*, *su<sub>II</sub>* and *suA* hosts. The protein product of this gene was also analysed in order to check if *suA* permitted translation past the amber codon (that is, insertion of mis-sense).

Infection of the *su<sup>-</sup>* host (*E. coli* B<sub>s-1</sub>/1) with T7<sup>+</sup> and T7 *am* 342 (gene 1, near operator distal end of the gene<sup>5</sup>) resulted in RNA profiles on gel electrophoresis which were similar to those already published<sup>4</sup>. The gene 1 mRNA is not seen as a discrete band on the gel in the case of infection with T7 *am* 342, whereas a sharp peak at about  $1.1 \times 10^6$  Daltons is characteristic of the complete mRNA species observed after infection with T7<sup>+</sup>. Similar results were obtained with T7 *am* 23 (operator proximal end of gene<sup>5</sup>) and T7 *am* 94 (middle of gene<sup>5</sup>). The fact that in no case was a discrete band of lower molecular weight observed suggests that this RNA was either not present at all or was terminated in some non-uniform way.

The electrophoretic analysis of T7 RNA made after infection of *E. coli* XA7007 (*suA*) with T7 *am* 342 is shown in Fig. 1 along with the analysis of RNA from the *su<sup>-</sup>* host also infected with T7 *am* 342. Whereas no pool of full length gene 1 mRNA molecules accumulates in the *su<sup>-</sup>* host, a significant amount of complete gene 1 mRNA is present in the *suA* host.

Analysis of the T7 proteins by SDS-gel electrophoresis<sup>6</sup> showed that a short peptide fragment is made in both the *su<sup>-</sup>* and *suA* hosts, which indicated that translation is terminated at the amber codon in the *suA* host as well as in the *su<sup>-</sup>* strain. This implies, then, that the T7 message distal to the amber codon is stabilized in the *suA* strain, but not translated. Similar conclusions were drawn by Morse and Primakoff<sup>1</sup>. It was suggested earlier<sup>1</sup> that the *suA* mutation may affect an endonuclease which normally attacks the untranslated portion of mRNAs. Such an interpretation is compatible with the results presented here. In T7-infected cells the mRNA appears to be stable<sup>7,8</sup>. This stability, however, may reflect a deficiency in RNAase V activity<sup>9</sup> (5' exonucleolytic activity associated with ribosomes), while the hypothetical endonuclease is still free to make internal cuts in untranslated T7 mRNA as in the case of the amber mutant in an *su<sup>-</sup>* host.

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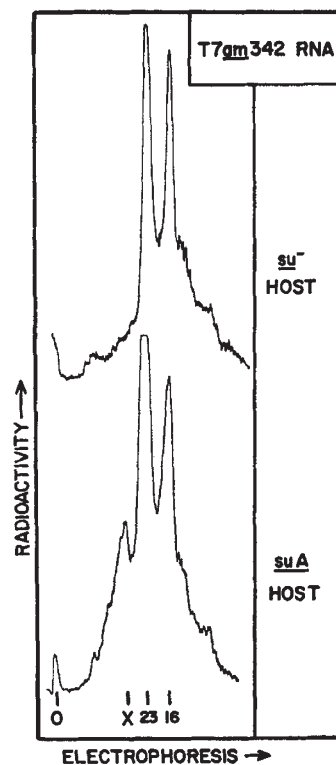


Fig. 1 Gel electrophoresis of <sup>14</sup>C-RNA from cells infected with T7 *am* 342. Upper trace shows RNA from *su<sup>-</sup>* cells; lower trace shows RNA from *suA* cells. Microdensitometer tracing is of a contact autoradiograph of a longitudinally sliced, dried 2.5% polyacrylamide-0.5% agarose gel. Electrophoresis, labelling, and sample preparation were carried out as described previously<sup>3,4</sup>. The band marked (x) corresponds to the mRNA for the T7 gene 1 protein. The 23s and 16s host species are indicated.

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