Agrobacterium tumefaciens-mediated transformation of filamentous fungi

Because of a production error, the experimental protocol in "*Agrobacterium tumefaciens*-mediated transformation of filamentous fungi," by Marcel J.A. de Groot et al., which appeared in *Nature Biotechnology* **16:**839 (September 1998), was incorrect. The third section of the protocol should read as follows:

T-DNA transfer. Protoplasts of *A. awamori* were prepared as described by Punt and van den Hondel[27]. *A. awamori* conidia were obtained by growing the strain on a nitrocellulose filter placed on a potato dextrose agar (PDA) plate for several days and washing the filter with a physiologic salt solution. Cocultivations between *A. tumefaciens* and *A. awamori* was performed as follows. For transformation of protoplasts, 100 µl of protoplasts at a concentration of 3×10^7 or 10^8 protoplasts/ml were mixed with 100 µl of the *Agrobacterium* culture prepared as described[11]. When conidia were transformed, 100 µl conidia at a concentration of 10^6, 10^7, or 10^8 conidia/ml were used. Subsequently, these mixtures and dilutions thereof were plated on nitrocellulose filters placed on induction medium (IM) plates[11] containing 5 mM glucose with or without AS. The plates were incubated at room temperature for 2 days. The filters were transferred to *Aspergillus* minimal medium plates[28] containing 200 µM cefotaxime to kill the *Agrobacterium* cells and 100 µg/ml hygromycin to select for transformants. Conidia from *A. niger*, *F. venenatum*, *T. reesei*, *C. gloeosporioides*, and *N. crassa* were obtained as described above. Conidia from *A. bisporus* were purchased from Proefstation voor de Champignoncultuur (Horst, The Netherlands). Transformations were performed as described for *A. awamori*. The numbers of conidia used for transformation are listed in Table 2. Also, a rehydrated freeze-dried American Type Culture Collection (ATCC) culture of *F. venenatum* was used for transformation. The freeze-dried material had been rehydrated and stored at 4°C for approximately 2 weeks. An aliquot of 100 µl ATCC material was mixed with 200 µl *A. tumefaciens*. The IM plates were incubated at room temperature for 2 or 5 days. To germinate the conidia of *A. bisporus* malt extract agar (2% malt extract, 10 mM MOPS, 1.5% agar, pH 7.0, with KOH) was used. About 1.2×10^7 conidia were plated on a nitrocellulose filter placed on malt extract. An *A. bisporus* breeding-granule was added to facilitate germination of the conidia. The plates were incubated for 5 or 7 days at room temperature. For transformation, the filters were submerged in 25 ml of *A. tumefaciens* culture. The IM plates were incubated at room temperature for 5 days. To select for transformants the following hygromycin concentrations were used: *A. niger* 200 µg/ml, *F. venenatum* 150 µg/ml, *T. reesei* 100 µg/ml, *C. gloeosporioides* 100 µg/ml, *N. crassa* 200 µg/ml, and *A. bisporus* 25 µg/ml.

Identification of a calcium channel modulator using a high-throughput yeast two-hybrid screen

Because of a production error, Figure 5A of "Identification of a calcium channel modulator using a high-throughput yeast two-hybrid screen," by Kathleen Young et al., which appeared in *Nature Biotechnology* **16:**946 (October 1998), was incorrect. The correct figure appears below: