

field, they are thus able to exercise total power over what is and is not published. Their power is also extensive in the two reviewer system if their eminence is used to justify rejection based on their single negative review. Dr. Goodstein's comments should be taken seriously by journal editors, who should return to a three reviewer system. This system may take a bit more time and effort, but the cost is minimal when one considers that "eminent" scientists can no longer be expected not to use their eminence to keep new people from competing for the limited resources currently available to fund science.

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A missing category

To the editor:

In the review article by Lisa Evans and Mark Burns on focusing techniques, entitled "Solute Focusing Techniques for Bioseparations," (Bio/ Technology 13:46-52, January) we have missed a highly cost-effective category for isolation, purification, and concentration of biological preparations. The Evans/Burns' review reported on various techniques based on the principle of isoelectrofocusing, but did not mention the existence of a novel purification technology called "autofocusing," which was originally described in various publications and patent applications from the Institute of Animal Physiology, Kosice, Slovak Republic (e.g., J. Chromatography 320:15-22, 1985) entitled "Autofocusing—A method for isoelectric focusing without carrier ampholytes," and another one entitled "Industrial autofocusing—A new technology for large-scale isolectrofocusing" (J. Chromatography 320:2113-218, 1985). The serendipity of this observation/ finding was based on the omission of carrier ampholytes in an isoelectric-focusing run by a technician before the weekend. The term "autofocusing" was coined from the combination "automatic isolectrofocusing," and has demonstrated its applicability particularly for enzymes [see J. Chromatography 358:274-278, 1986 for uricase; J. Chromatography 411:486-489, 1987 for alpha-amylase; and J. Chromatography 474:430-434, 1989 for peroxidase].

Therefore, autofocusing provides for various biological preparations adequate resolution and higher concentration than the best categories reported in Evans' review article, and at the same time preserves the biological activities. The technique requires no ampholyte buffers or resins, as opposed to a number of modifications described, and uses very little electrical energy. This autofocusing method is currently in use on an industrial scale in a small number of enterprises on both the European and American continents, particularly to (semi)purify thousands of liters per day of nonbacterial enzymes for environmental friendly cleaning or rehabilitation of oil-contaminated soils, subsoil waters, waste waters, and slaughterhouses, etc., using mixtures of proteolytic, amylolytic, and lipolytic enzymes from earthworms.

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German gene therapy *To the editor:*

Your enjoyable and informative article entitled "Germany: A Dominant Force by the Year 2000" omitted to make mention of Germany's first gene therapy company, Orthogen, GmbH. Orthogen was founded in 1992 by Dr. Peter Wehling, an orthopedic surgeon at the University of Düsseldorf, with startup funds provided by the Ministry of Economic Department of the State of North Rhineland-Westphalia. Based in Düsseldorf, it specializes in the development of gene therapies for treating arthritis and other disorders of the musculoskeletal system. Its success provides further support for the main theme of your article.

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Erratum

Due to a printing error, Figure 4 in Collins-Racie, L.A., McColgan, J.M., Grant, K.L., DiBlasio-Smith, E.A., McCoy, J.M., and LaVallie, E.R. 1995. Production of recombinant bovine enterokinase catalytic subunit in Escherichia coli using the novel secretory fusion partner DsbA. Bio/Technology 13:982-987 appeared incorrectly. The corrected figure and legend are reproduced below.

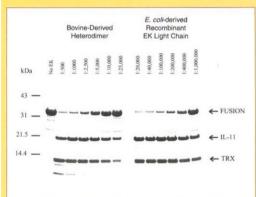


FIGURE 4.

Comparison of Trx/IL-11 fusion protein digestion with native bovine-derived enterokinase heterodimer (EKn) and E. coli-derived recombinant EKL (rEKL). Fusion protein at a concentration of approximately1 mg/ml in 50 mM Hepes, pH 8.0 was incubated with varying amounts of enterokinase (ratios expressed as w/w) at 37/C for 20 hrs. The mobilities of the uncleaved fusion protein and the cleaved constituents thioredoxin (Trx) and interleukin 11 (IL-11) are indicated with arrows to the right of the figure. The resulting digests were lyophilized and electrophoresed on a 10% SDS-tricine gel prior to staining with Coomassie blue.