0–15 μ M. For the competition EMSA, the corresponding unlabelled 26-nucleotide RNA or DNA of 0–30 μ M was used in addition to the 32 P-labelled RNA and PAZ domain protein.

Site-directed mutagenesis

Expression constructs for mutant proteins were generated using the QuikChange site-directed mutagenesis kit (Stratagene). The presence of appropriate mutations was confirmed by DNA sequencing. Mutant proteins were purified by affinity chromatography using nickel-NTA columns (Qiagen). Proper folding of each PAZ domain mutant was confirmed by NMR. Samples for the NMR competition assay were prepared by mixing 15 N-labelled wild-type PAZ domain (0.2 mM), ssRNA oligonucleotide (5'-pUGACA) and unlabelled mutant PAZ domain with a molar ratio of 1:1:5.

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Correspondence and requests for materials should be addressed to M.-M.Z. (Ming-Ming.Zhou@mssm.edu). Coordinates for the NMR structure of the *D. melanogaster* Ago1 PAZ domain are deposited in the Protein Data Bank under ID code 1R4K.

addendum

Specific cytotoxic T cells eliminate cells producing neutralizing antibodies

Oliver Planz, Peter Seiler, Hans Hengartner & Rolf M. Zinkernagel

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The experiments in BALB/c mice using hybridoma technology reported in this Letter indicated that neutralizing-antibody-producing B cells and hybridomas could have been preferentially infected by lymphocytic choriomeningitis virus (LCMV), whereas B cells specific for the internal viral components were not. We concluded that during LCMV infection, B cells specific for the viral-surface antigens may be selectively and productively infected by LCMV through the membrane-anchored neutralizing-antibody receptor and are subsequently eliminated by virus-specific cytotoxic T lymphocytes. These experiments were performed in normal or CD8⁺ T-cell-depleted BALB/c H-2^d mice. In later experiments designed to generalize these findings, we studied the infection patterns of hybridomas using C57BL/6 mice, but the results were variable and not conclusive. We also failed to observe a preferential infection of B cells or of hybridomas generated from IgM or isotype-switchable transgenic mice with a high percentage of neutralizing-antibody-expressing B cells^{1,2}. In a third series of experiments with a recombinant virus that combines the envelope glycoprotein from vesicular stomatitis virus (VSV) with an LCMV replicon, we did not detect a preferential CD8⁺ T-cell-dependent depletion of the anti-VSV antibody response (D. Pinschewer *et al.*, unpublished observations). We have also not so far been able to reproduce our original findings in new experiments with LCMV and BALB/c (SPF) mice. We therefore wish to alert the community to these problems until further experiments have been performed. □

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