

Recurrent *BCOR* internal tandem duplication and *BCOR* or *BCL6* expression distinguish primitive myxoid mesenchymal tumor of infancy from congenital infantile fibrosarcoma

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Primitive myxoid mesenchymal tumor of infancy is a rare sarcoma that preferentially affects infants. It can be locally aggressive and rarely metastasizes, but the long-term outcome of children with this tumor is mostly unknown. Histologically, it is characterized by primitive cells with abundant myxoid stroma. Internal tandem duplication of B-cell CLL/lymphoma 6 (*BCL6*)-interacting co-repressor (*BCOR*) exon 15 has recently been described in clear cell sarcoma of kidney, central nervous system high-grade neuroepithelial tumor with *BCOR* alteration, and primitive myxoid mesenchymal tumor of infancy. Herein, we report five cases of primitive myxoid mesenchymal tumor of infancy: three girls and two boys with mean age of 6.5 months. The tumors were located in the paraspinal region ($n=3$), back ($n=1$), or foot ($n=1$) and ranged in size from 2.5 to 10.2 cm. *BCOR* internal tandem duplication was confirmed by PCR and sequencing in all five cases. The minimally duplicated region consisted of nine residues, which is shorter than was previously reported in other *BCOR*-associated tumors. To assess the clinical value and specificity of the *BCOR* internal tandem duplication, a group of 11 *ETV6*-rearranged congenital infantile fibrosarcomas were evaluated and no *BCOR* internal tandem duplication was identified in any case. Though not detected in congenital infantile fibrosarcomas, *BCOR* and *BCL6* immunoreactivity was present in >90% of the nuclei of tumor cells in each of the five primitive myxoid mesenchymal tumor of infancy. The presence of *BCOR* internal tandem duplication in all five primitive myxoid mesenchymal tumors of infancy provides evidence that it is a recurrent somatic abnormality and substantiates the concept that this tumor is a unique sarcoma of infancy. Our findings indicate that identification of *BCOR* internal tandem duplication and/or nuclear immunoreactivity for *BCOR* or *BCL6* can aid in the diagnosis of primitive myxoid mesenchymal tumor of infancy and help to differentiate it from congenital infantile fibrosarcoma.

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Primitive myxoid mesenchymal tumor of infancy, first described in 2006 by Allagio *et al*,¹ is a primitive mesenchymal soft tissue sarcoma with no distinctive lineage of differentiation. This tumor typically affects young children, particularly infants, frequently presenting as a painless mass. Until recently, there were no known specific immunohistochemical markers or molecular alterations to distinguish primitive myxoid mesenchymal tumor of infancy from its histologic mimics, and the diagnosis was based solely on morphologic features and the

absence of the *ETV6*–*NTRK3* gene fusion that is typically present in congenital infantile fibrosarcoma.² However, Kao *et al*³ have recently reported internal tandem duplication of the X-linked *BCL6* co-repressor (*BCOR*) gene (*BCOR* internal tandem duplication) in six of the seven primitive myxoid mesenchymal tumor of infancy cases examined. *BCOR* internal tandem duplication has been previously identified as a recurrent somatic abnormality in approximately 85% of clear cell sarcomas of kidney^{4–7} and in a subset of central nervous system high-grade neuroepithelial tumors with *BCOR* alterations.⁸

With only a limited number of cases reported in the literature, primitive myxoid mesenchymal tumor of infancy was not included in the most recent World Health Organization classification of tumors of the soft tissue (4th edition, February 2013).⁹ Based on

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available current data, the biological behavior of this mesenchymal soft tissue sarcoma is considered to be intermediate due to the fact that it can be locally aggressive and rarely metastasizes, similar to congenital infantile fibrosarcoma. Nevertheless, the outcome and long-term survival in children with primitive myxoid mesenchymal tumor of infancy is mostly unknown.^{10–18} This study aimed to evaluate the clinical utility of testing for the *BCOR* internal tandem duplication and the use of *BCOR* and B-cell CLL/lymphoma 6 (*BCL6*) immunohistochemistry as a diagnostic marker for this rare soft tissue sarcoma. Specifically, we wished to evaluate the effectiveness of these tests to differentiate primitive myxoid mesenchymal tumor of infancy from congenital infantile fibrosarcoma.

Materials and methods

Patients

This study was approved by our Institutional Review Board. A retrospective review of our institutional records was performed. Five children with low-grade primitive sarcoma diagnosed as primitive myxoid mesenchymal tumor of infancy were identified and all available clinicopathologic data, hematoxylin and eosin (H&E)-stained slides, immunohistochemistry-stained slides, and interphase fluorescent *in situ* hybridization results were reviewed. All five cases of primitive myxoid mesenchymal tumor of infancy identified in our files were pathology consultations, with treatment and follow-up performed at an outside institution.

DNA Extraction and PCR Analysis

Genomic DNA was extracted from formalin-fixed paraffin-embedded tissue by using a Maxwell 16 formalin-fixed paraffin-embedded Plus LEV DNA purification kit (Promega) in accordance with the manufacturer's instructions. Targeted PCR amplification of the duplicated segment of the *BCOR* gene (exon 15) was performed on genomic DNA by using GoTaq DNA polymerase (Promega) and *BCOR* internal tandem duplication primers previously described.⁶ PCR products were sequenced using BigDye Terminator v3.1 Sequencing Kit (Applied Biosystems). The sequence results were analyzed using 3730xl DNA Analyzer (Life Technologies) in conjunction with CLC Workbench 6.0 (CLCBio). All tandem duplications detected were confirmed by independent replicate analyses. Negative (normal tissue) and positive (10 cases of clear cell sarcomas of kidney) controls were used to validate the detection assay. In addition to the primitive myxoid mesenchymal tumor of infancy cases, 11 cases of *ETV6*-rearranged congenital infantile fibrosarcoma were also evaluated for *BCOR* internal tandem duplication.

Immunohistochemical Staining

Both primitive myxoid mesenchymal tumors of infancy and congenital infantile fibrosarcoma cases were assessed for expression of *BCOR* and *BCL6* by immunohistochemistry. Formalin-fixed paraffin-embedded tissue (4 μ m in thickness) from primitive myxoid mesenchymal tumors of infancy, congenital infantile fibrosarcomas, and appropriated negative and positive controls were subjected to immunohistochemistry staining for *BCL6* using a BenchMark ULTRA automated slide stainer (Ventana Medical Systems) and immunohistochemistry staining for *BCOR* using a Dako Omnis automated staining platform in accordance with the manufacturers' recommended procedure. In brief, immunohistochemistry for *BCL6* was performed after deparaffinization and antigen retrieval using cell conditioning solution (CC1) for 64 min, followed by 32 min incubation at 37 °C with mouse monoclonal anti-*BCL6* antibody (Bond ready-to-use primary antibody BCL-6 [LN22]; Leica Biosystems, catalog number PA0204). Immunohistochemistry for *BCOR* was also performed following deparaffinization and antigen retrieval using EnVision FLEX high pH Target Retrieval Solution (Dako) for 30 min. The slides were incubated with mouse monoclonal IgG1 *BCOR* antibody, which targets the N terminus of *BCOR* protein (1:100 dilution, incubated for 20 min at 36 °C; clone C-10, catalog number SC-514576, Santa Cruz Biotechnology). An OptiView DAB immunohistochemistry detection kit (OptiView, Ventana Medical Systems) was used as an indirect biotin-free system for signal detection. Before light microscopy examination, the slides were counterstained with hematoxylin.

Results

Clinicopathologic Characteristics of Primitive Myxoid Mesenchymal Tumor of Infancy

We identified five cases of primitive myxoid mesenchymal tumor of infancy in our records. The mean age at diagnosis of the three girls and two boys was 6.5 months (range, 1 week to 13 months). The tumors were located in the paraspinal region (three cases), back (one case), or foot (one case). The tumor size was available for three patients and ranged from 2.5 to 10.2 cm (mean, 5.3 cm) in the largest dimension. At initial diagnosis, all patients had localized disease with no evidence of metastasis. However, one patient in whom primitive myxoid mesenchymal tumor of infancy was initially diagnosed at 1 week of age (primitive myxoid mesenchymal tumor of infancy—case 1) experienced local recurrence after 6 months. A concise summary of the clinicopathologic features is presented in Table 1.

Histologically, all five tumors displayed a diffuse growth pattern, a vaguely multinodular appearance, and variable degrees of cellularity. Round-to-oval or

Table 1 Clinical features, pathological and molecular characteristics of the primitive myxoid mesenchymal tumor of infancy (PMMTI) and congenital infantile fibrosarcoma (CIF) cases

Case	Age at initial diagnosis	Gender	Tumor location	Tumor size (largest dimension) (cm)	BCL6 IHC staining	BCOR IHC staining	ETV6 rearrangement (by FISH)	BCOR-ITD	ITD length (bp)
PMMTI 1	1 week	Male	Paraspinal (occipital)	10.2	Positive	Positive	Negative	Positive	90
PMMTI 2	9 months	Female	Back	2.5	Positive	Positive	Negative	Positive	66
PMMTI 3	8 months	Female	Paraspinal (T9 to L3)	NA	Positive	Positive	Negative	Positive	NA
PMMTI 4	13 months	Female	Paraspinal (L2 to S2)	NA	Positive	Positive	Negative	Positive	96
PMMTI 5	2 months	Male	Left foot	3.4	Positive	Positive	Negative	Positive	96
CIF 1	4 months	Male	Left upper arm	7.0	Negative	Negative	Positive	Negative	—
CIF 2	6 months	Male	Left leg	NA	Negative	Negative	Positive	Negative	—
CIF 3	12 months	Male	Right arm	9.0	Negative	NP	Positive	Negative	—
CIF 4	7 months	Female	Face and neck	9.0	Negative	Negative	Positive	Negative	—
CIF 5	5 weeks	Male	Neck	NA	Negative	NP	Positive	Negative	—
CIF 6	8 months	Female	Right arm	5.0	Negative	Negative	Positive	Negative	—
CIF 7	3 months	Female	Left forearm	6.0	Negative	NP	Positive	Negative	—
CIF 8	6 months	Female	Right thigh	10.0	Negative	Negative	Positive	Negative	—
CIF 9	7 weeks	Male	Left foot	NA	Negative	Negative	Positive	Negative	—
CIF 10	5 weeks	Male	Right hand	4.0	Negative	NP	Positive	Negative	—
CIF 11	9 months	Female	Left forearm	8.4	Negative	Negative	Positive	Negative	—

Abbreviations: BCL6, B-cell CLL/lymphoma 6; BCOR, BCL6 interacting co-repressor; bp, base pair; ETV6, ETS variant gene 6 (TEL oncogene); iFISH, interphase fluorescence *in situ* hybridization; IHC, immunohistochemical; ITD, internal tandem duplication; NA, not available; NP, not performed.

slightly spindle-shaped primitive cells were distributed in a loose myxoid matrix. A well-developed vasculature was noted. The nuclei were mostly ovoid, with fine chromatin and discreet nucleoli (Figures 1a and b). The mean mitotic rate was three mitoses per 10 high-power fields. Only typical mitoses were identified. Areas with hemangiopericytoma-like vascular pattern (Case 3, Figure 1c), prominent myxoid background (Case 4, Figure 1d), and focal mild inflammatory infiltrate were also noted (Case 5).

A summary of all the different immunohistochemistry stains performed on the primitive myxoid mesenchymal tumor of infancy cases are presented in Figure 2a. In four cases in which immunohistochemistry staining for CD99 was performed, all were diffusely positive, and all five primitive myxoid mesenchymal tumor of infancy were negative for S-100. Interphase fluorescence *in situ* hybridization analysis to detect ETV6 rearrangement, as seen in congenital infantile fibrosarcoma, was negative in all five primitive myxoid mesenchymal tumors of infancy cases (Table 1).

Recurrent Somatic BCOR Internal Tandem Duplication in Primitive Myxoid Mesenchymal Tumor of Infancy

To determine whether the BCOR internal tandem duplication could be used clinically to distinguish primitive myxoid mesenchymal tumor of infancy from congenital infantile fibrosarcoma, we compared a cohort of 11 ETV6-rearranged congenital infantile fibrosarcomas to our cohort of primitive myxoid mesenchymal tumors of infancy for the presence of the BCOR internal tandem duplication using targeted polymerase chain reaction for the duplicated region of BCOR (exon 15) on DNA extracted from formalin-fixed paraffin-embedded tumor tissue. Clear cell sarcomas of kidney and normal brain were used as positive and negative controls of the assay, respectively. All five cases of primitive myxoid mesenchymal tumor of infancy demonstrated the BCOR internal tandem duplication, which was confirmed by sequencing. In contrast, the BCOR internal tandem duplication was not detected in any of the 11 congenital infantile fibrosarcoma cases (Table 1).

BCOR and BCL6 Expression in Primitive Myxoid Mesenchymal Tumor of Infancy and Congenital Infantile Fibrosarcoma

To determine whether BCL6 or BCOR immunoreactivity could be used as a surrogate of the BCOR internal tandem duplication and help distinguish primitive myxoid mesenchymal tumor of infancy and congenital infantile fibrosarcoma, we performed immunohistochemistry staining for both BCL6 and BCOR on our cohort of tumors. All 10 congenital infantile fibrosarcoma cases tested for BCL6

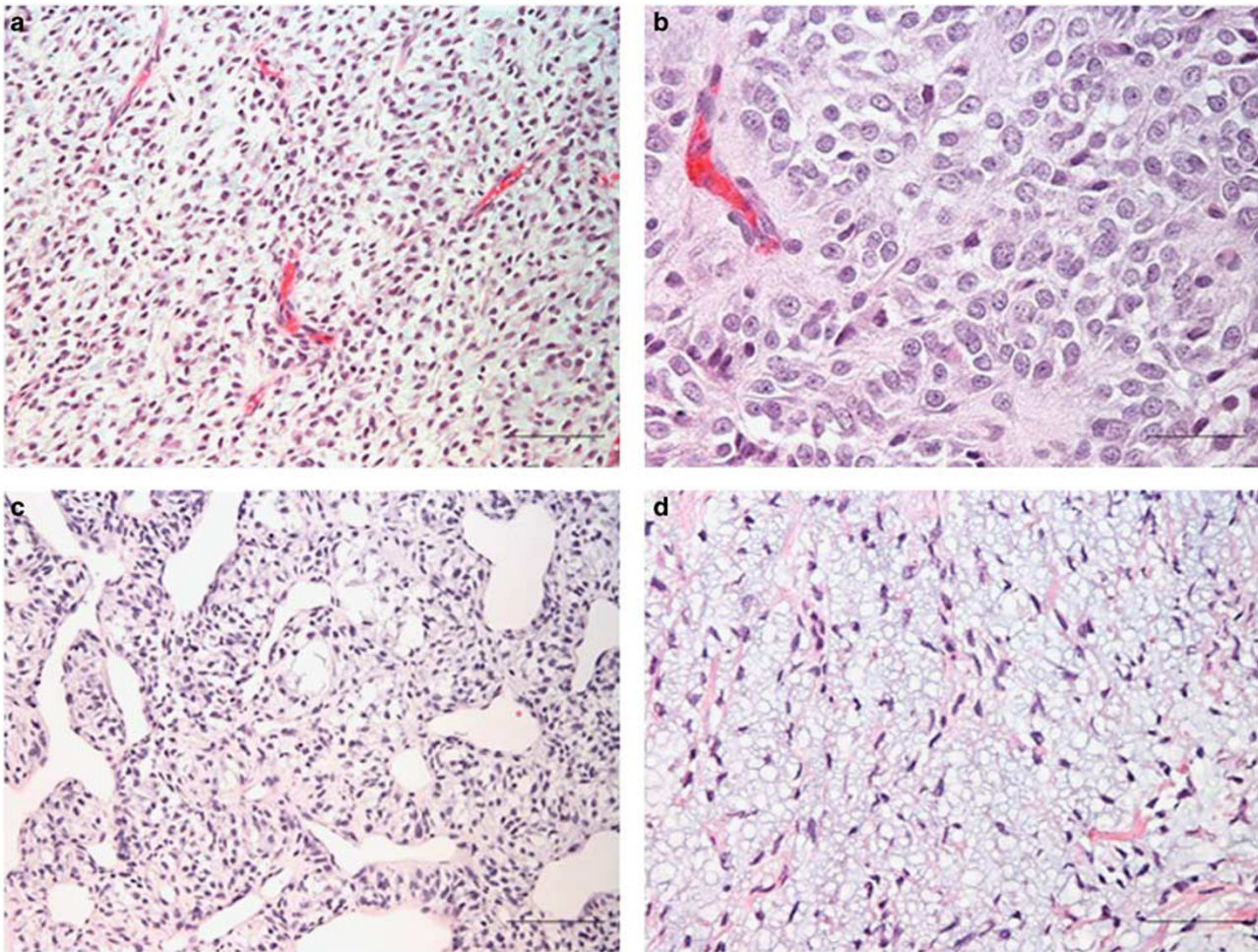


Figure 1 Histomorphology of primitive myxoid mesenchymal tumor of infancy. (a) Primitive cell morphology with delicate vessels and myxoid stroma (primitive myxoid mesenchymal tumor of infancy—case 1, $\times 200$); (b) tumor cells show uniform round-to-oval nuclei with inconspicuous nucleoli (primitive myxoid mesenchymal tumor of infancy—case 1, $\times 400$); (c) areas with hemangiopericytoma-like vascular pattern (primitive myxoid mesenchymal tumor of infancy—case 3, $\times 200$) and (d) prominent myxoid background were also noted (primitive myxoid mesenchymal tumor of infancy—case 4, $\times 400$). Hematoxylin and eosin.

expression were negative, and only rare intratumoral inflammatory cells were immunoreactive for BCL6. BCOR immunostaining was completely negative in all seven congenital infantile fibrosarcomas examined. In contrast, each of the primitive myxoid mesenchymal tumors of infancy examined demonstrated nuclear positivity for both BCOR and BCL6 in over 90% of the tumor cells (Figure 3).

Discussion

The main differential diagnosis of primitive myxoid mesenchymal tumor of infancy is congenital infantile fibrosarcoma, which also presents as a soft tissue mass and most commonly affects infants. The presence of recurrent *ETV6-NTRK3* fusion is a diagnostic feature of congenital infantile fibrosarcoma that is absent in cases of primitive myxoid mesenchymal tumor of infancy. Various primary sites have been reported, but primitive myxoid

mesenchymal tumor of infancy has a slight predilection for involving deep soft tissue, especially around the spinal cord.^{12,13} Indeed, three of our five cases involved a paraspinal location. Both primitive myxoid mesenchymal tumor of infancy and congenital infantile fibrosarcoma frequently recur locally if not completely excised and rarely metastasize.^{1,2} Nevertheless, based on the currently available data, primitive myxoid mesenchymal tumor of infancy appears to be more aggressive than congenital infantile fibrosarcoma, exhibiting frequent local recurrence, poor response to chemotherapy, and the potential to transform into an undifferentiated sarcoma with marked cellular atypia.¹⁷

Before the identification of *BCOR* internal tandem duplication, ultrastructural, molecular, and cytogenetic analysis of published cases of primitive myxoid mesenchymal tumor of infancy failed to demonstrate any specific alteration. Electron microscopic

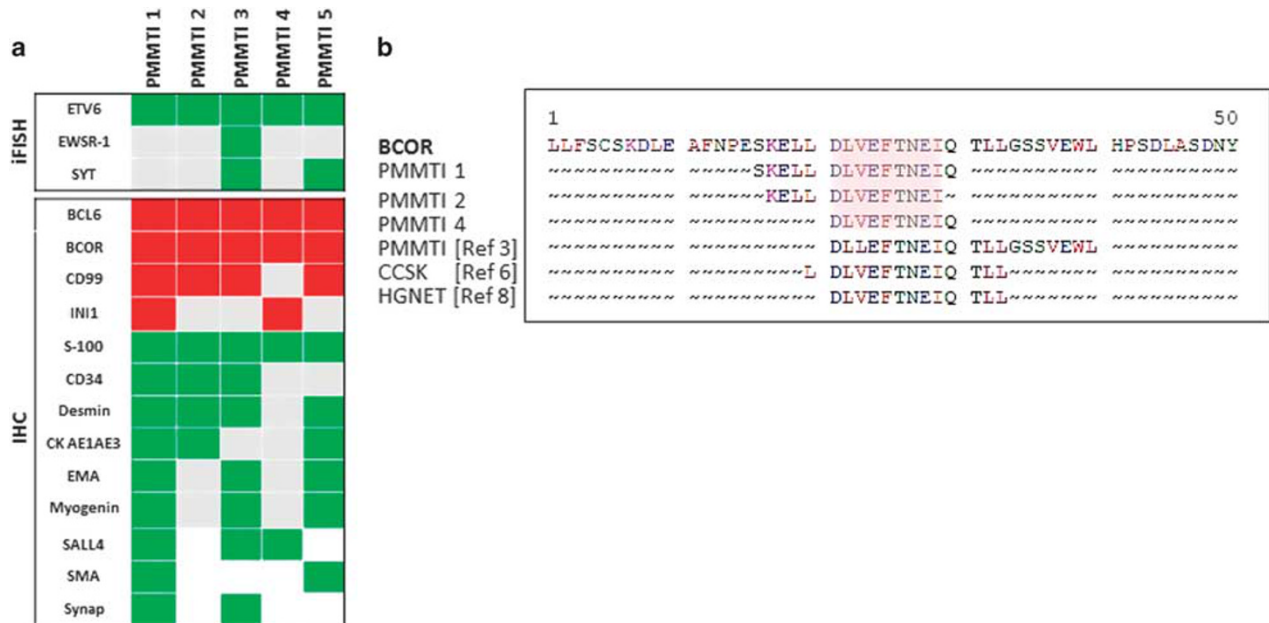


Figure 2 (a) Immunohistochemical staining and interphase fluorescence *in situ* hybridization results for five cases of primitive myxoid mesenchymal tumor of infancy. Green, negative; red, positive; gray, not tested. (b) Diagram of predicted BCOR protein sequences from BCOR internal tandem duplication-associated tumors. BCOR wild-type protein sequence is presented on top followed by three cases from our cohort (primitive myxoid mesenchymal tumor of infancy—cases 1, 2, and 4). A homologous sequence of nine residues is seen (red box). The common duplicated region identified in the primitive myxoid mesenchymal tumors of infancy by Kao *et al*,³ clear cell sarcomas of kidney by Roy *et al*,⁶ and central nervous system high-grade neuroepithelial tumors with BCOR alterations by Sturm *et al*,⁸ includes 20, 14, and 13 residues, respectively. BCL6, B-cell CLL/lymphoma 6; CD, cluster of differentiation; CK, cytokeratin; EMA, epithelial membrane antigen; ETV6, ETS variant gene 6 (TEL oncogene); EWSR-1, Ewing sarcoma breakpoint region 1; INI1, SMARCB1 SWI/SNF-related, matrix-associated, actin-dependent; SALL4, spalt-like transcription factor 4; SMA, smooth muscle actin; Synap, synaptophysin; SYT, synovial sarcoma translocation.

analysis revealed primitive mesenchymal features and the presence of abundant dilated rough endoplasmic reticulum in the cytoplasm, but found no distinctive features other than an apparent fibroblastic line of differentiation.^{1,16,18}

The BCOR gene, located on the short arm of chromosome X (ch Xp11.4), encodes a protein that selectively interacts with BCL6 as a co-repressor. Although somatic alterations in the BCOR gene have been reported in different types of human neoplasm, particularly undifferentiated round cell sarcomas of the bone that harbor a BCOR-CCNB3 translocation^{19,20} and, most recently, clear cell sarcomas of kidney, in which BCOR internal tandem duplication was found in 85% of the cases analyzed,^{4–7} BCOR germline mutation recognized in patients with X-linked oculofaciocardiodental syndrome is not considered a cancer predisposing condition.^{21–24}

Until recently, the diagnosis of primitive myxoid mesenchymal tumor of infancy was based exclusively on morphologic features and the absence of ETV6-NTRK3 gene fusion; hence, the identification of BCOR internal tandem duplication in six of the seven cases studied by Kao *et al*³ and in all five of our cases of primitive myxoid mesenchymal tumor of infancy indicates that BCOR internal tandem duplication has a diagnostic utility in this tumor.

Kao *et al* have also shown that BCOR immunohistochemistry stain was diffuse and strongly positive in 11 out of 14 BCOR internal tandem duplication positive tumors (78%), moderately positive in two cases (14%) and negative in one patient (7%).²⁵ Even though they have shown that a large number of variable sarcomas are negative for BCOR immunostaining, congenital infantile fibrosarcomas were not among those tested.²⁵ In our hands, both immunohistochemistry staining for BCL6 and BCOR demonstrated diffuse nuclear positivity in more than 90% of tumor cells in all five of the primitive myxoid mesenchymal tumors of infancy examined, whereas it was negative in all congenital infantile fibrosarcomas tested (Fisher's exact test, $P < 0.01$). Therefore, the diffuse nuclear expression of BCOR and BCL6 observed in primitive myxoid mesenchymal tumor of infancy appears useful in differentiating it from congenital infantile fibrosarcoma, particularly when molecular testing for BCOR internal tandem duplication and/or ETV6-NTRK3 is unavailable.

Five distinct types of in-frame BCOR internal tandem duplication mutants with varying lengths of internal tandem duplication (in base pairs, bp) were identified in a cohort of clear cell sarcomas of kidney: type I, 96 bp (five cases); type II, 93 bp (three cases); type III, 90 bp (one case); type IV, 87 bp (one case); and type V, 114 bp (one case), all located

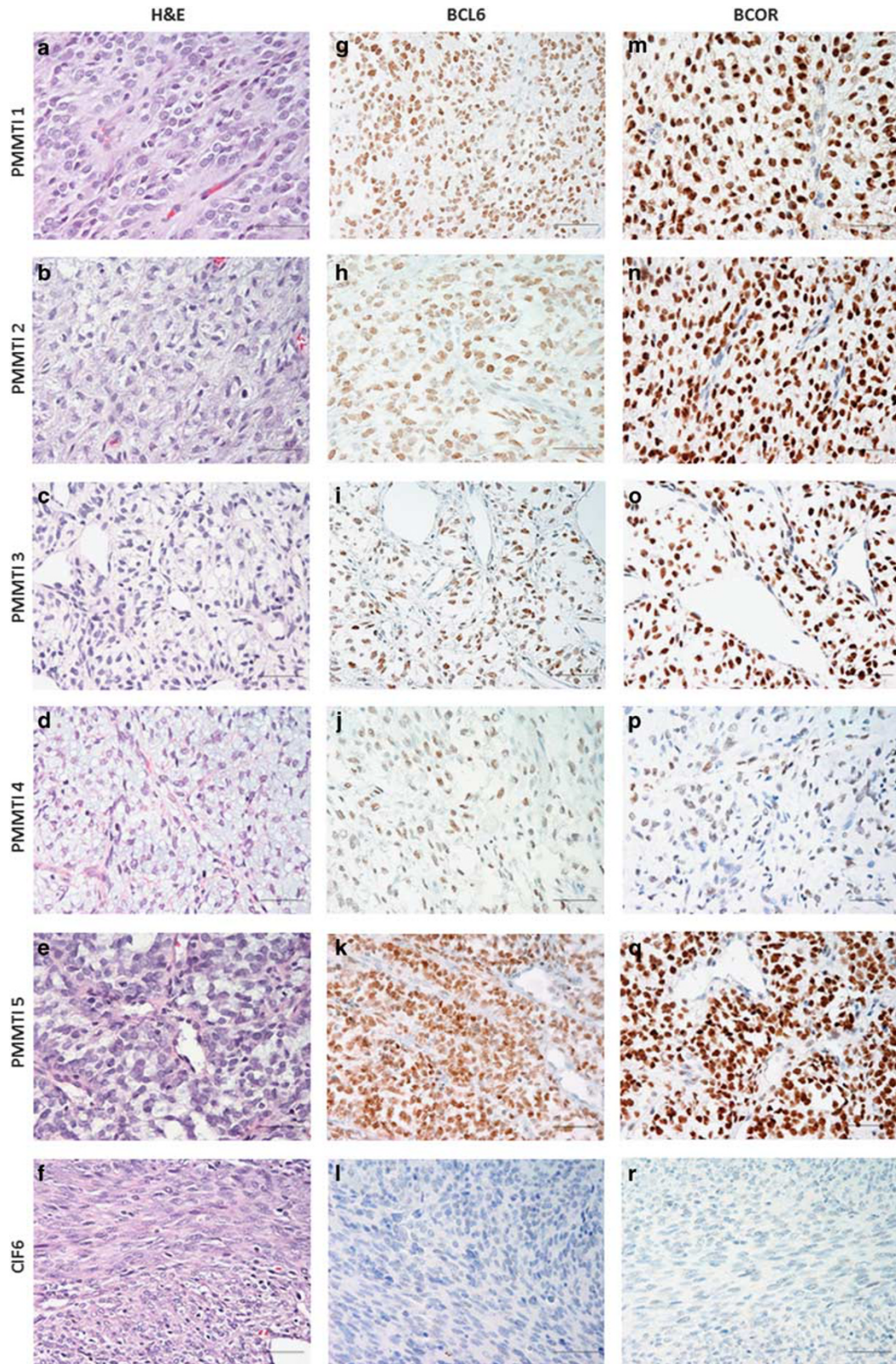


Figure 3 Primitive myxoid mesenchymal tumor of infancy in contrast with congenital infantile fibrosarcoma. (a–r) Five cases of primitive myxoid mesenchymal tumor of infancy, all negative for *ETV6* rearrangement by interphase fluorescent *in situ* hybridization and positive for *BCOR* internal tandem duplication (first column, H&E, × 400); immunohistochemistry staining for BCL6 (second column) and BCOR (third column) show diffuse nuclear positivity in all primitive myxoid mesenchymal tumor of infancy, but negative staining in an *ETV6*-rearranged congenital infantile fibrosarcoma (congenital infantile fibrosarcoma—case 6), immunohistochemistry stains, × 400. *ETV6*, ETS variant gene 6 (TEL oncogene); BCOR, B-cell CLL/lymphoma 6 (BCL6)-interacting co-repressor.

within the C-terminal coding region.⁶ In our five cases of primitive myxoid mesenchymal tumor of infancy, the internal tandem duplication ranged from 66 to 96 bp (Table 1). The length of the minimally duplicated region was nine amino acid residues, which is shorter than what was previously reported in *BCOR* internal tandem duplication positive primitive myxoid mesenchymal tumor of infancy,³ clear cell sarcomas of kidney,⁶ and central nervous system high-grade neuroepithelial tumors with *BCOR* alterations⁸ (Figure 2b). The significance of the insert size and/or length of the homologous region in positive *BCOR* internal tandem duplication tumors are still unknown. The molecular characterization of a larger number of primitive myxoid mesenchymal tumors of infancy is required to establish the clinical, biological, and prognostic implications of *BCOR* gene alteration, and *BCOR* and *BCL6* protein overexpression. Supplementary molecular analysis and methylation studies would be beneficial for characterizing this distinctive tumor.

Central nervous system high-grade neuroepithelial tumors with *BCOR* alterations⁸ also share some histologic similarities with primitive myxoid mesenchymal tumor of infancy and clear cell sarcomas of kidney. Although the phenotypic resemblance among these three *BCOR* internal tandem duplication positive tumors does not necessarily indicate that they have similar histogenesis, it does cause us to speculate whether these three embryonal tumors constitute another 'triad of tumors' involving soft tissue, kidney, and CNS. This is similar to the triad of extrarenal rhabdoid tumor of soft tissue, rhabdoid tumor of the kidney, and atypical teratoid rhabdoid tumor of the brain, which share similar morphologic features and, in almost all cases, a common genetic alteration affecting the *SMARCB1* locus on the long arm of chromosome 22 that results in the loss of *SMARCB1* (*INI1*) protein expression.^{26–28} Moreover, primitive myxoid mesenchymal tumor of infancy and clear cell sarcoma of kidney share morphological features and similar molecular aberration analogous to congenital infantile fibrosarcoma of soft tissue and the cellular variant of mesoblastic nephroma of the kidney, which have identical histology and share a common t(12;15)(p13;q25) translocation that results in an *ETV6–NTRK3* gene fusion.

In summary, we have confirmed the association between *BCOR* internal tandem duplication and primitive myxoid mesenchymal tumor of infancy and provided supporting evidence that *BCOR* internal tandem duplication is indeed a recurrent somatic abnormality in primitive myxoid mesenchymal tumor of infancy. Furthermore, we have demonstrated that *BCOR* internal tandem duplication identification and/or nuclear immunoreactivity for *BCOR* and/or *BCL6* can have diagnostic utility and can help to differentiate primitive myxoid mesenchymal tumor of infancy from congenital infantile fibrosarcoma. Future studies are needed to elucidate

the biologic role of the *BCOR* internal tandem duplication and *BCOR/BCL6* protein overexpression in these rare mesenchymal tumors. Additional research into this area is necessary and may provide insight into the use of targeted therapeutics in such patients.

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Disclosure/conflict of interest

The authors declare no conflict of interest.

References

- 1 Alaggio R, Ninfo V, Rosolen A, *et al*. Primitive myxoid mesenchymal tumor of infancy: a clinicopathologic report of 6 cases. *Am J Surg Pathol* 2006;30:388–394.
- 2 Knezevich SR, McFadden DE, Tao W, *et al*. A novel *ETV6–NTRK3* gene fusion in congenital fibrosarcoma. *Nat Genet* 1998;18:184–187.
- 3 Kao YC, Sung YS, Zhang L, *et al*. Recurrent *BCOR* internal tandem duplication and *YWHAE–NUTM2B* fusions in soft tissue undifferentiated round cell sarcoma of infancy: overlapping genetic features with clear cell sarcoma of kidney. *Am J Surg Pathol* 2016;40:1009–1020.
- 4 Astolfi A, Melchionda F, Perotti D, *et al*. Whole transcriptome sequencing identifies *BCOR* internal tandem duplication as a common feature of clear cell sarcoma of the kidney. *Oncotarget* 2015;6:40934–40939.
- 5 Karlsson J, Valind A, Gisselsson D. *BCOR* internal tandem duplication and *YWHAE–NUTM2B/E* fusion are mutually exclusive events in clear cell sarcoma of the kidney. *Genes Chromosomes Cancer* 2015;55:120–123.
- 6 Roy A, Kumar V, Zorman B, *et al*. Recurrent internal tandem duplications of *BCOR* in clear cell sarcoma of the kidney. *Nat Commun* 2015;6:8891.
- 7 Ueno-Yokohata H, Okita H, Nakasato K, *et al*. Consistent in-frame internal tandem duplications of *BCOR* characterize clear cell sarcoma of the kidney. *Nat Genet* 2015;47:861–863.
- 8 Sturm D, Orr BA, Toprak UH, *et al*. New brain tumor entities emerge from molecular classification of CNS-PNETs. *Cell* 2016;164:1060–1072.
- 9 Fletcher CDM, Bridge JA, Hogendoorn P, Mertens F. WHO Classification of Tumours of Soft Tissue and Bone, Fourth Edition. IARC: Lyon, 2013.
- 10 Lam J, Lara-Corrales I, Cammisuli S, *et al*. Primitive myxoid mesenchymal tumor of infancy in a preterm infant. *Pediatr Dermatol* 2010;27:635–637.
- 11 Mulligan L, O'Meara A, Orr D, *et al*. Primitive myxoid mesenchymal tumor of infancy: a report of a further

- case with locally aggressive behavior. *Pediatr Dev Pathol* 2011;14:75–79.
- 12 Gong Q, Wang Z, Li X, *et al*. Primitive myxoid mesenchymal tumor of infancy: report of two cases and review of the literature. *Pathol Int* 2012;62:549–553.
 - 13 Saito A, Taketani T, Kanai R, *et al*. A case with sacrococcygeal primitive myxoid mesenchymal tumor of infancy: a case report and review of the literature. *J Pediatr Hematol Oncol* 2013;35:e280–e282.
 - 14 Su TC, Hwang MJ, Li CF, *et al*. A rare malignant tumor of scalp in a 3-month-old Taiwanese infancy: case report of primitive myxoid mesenchymal tumor of infancy with molecular study. *Med Mol Morphol* 2013;46:109–113.
 - 15 Cuthbertson DW, Caceres K, Hicks J, *et al*. A cooperative approach to diagnosis of rare diseases: primitive myxoid mesenchymal tumor of infancy. *Ann Clin Lab Sci* 2014;44:310–316.
 - 16 Cipriani NA, Ryan DP, Nielsen GP. Primitive myxoid mesenchymal tumor of infancy with rosettes: a new finding and literature review. *Int J Surg Pathol* 2014;22:647–651.
 - 17 Guilbert MC, Rougemont AL, Samson Y, *et al*. Transformation of a primitive myxoid mesenchymal tumor of infancy to an undifferentiated sarcoma: a first reported case. *J Pediatr Hematol Oncol* 2015;37:e118–e120.
 - 18 Foster JH, Hicks J, Vasudevan S, *et al*. Primitive myxoid mesenchymal tumor of infancy involving chest wall in an infant: a case report and clinicopathologic correlation. *Pediatr Dev Pathol* 2015;19:244–248.
 - 19 Pierron G, Tirode F, Lucchesi C, *et al*. A new subtype of bone sarcoma defined by BCOR-CCNB3 gene fusion. *Nat Genet* 2012;44:461–466.
 - 20 Puls F, Niblett A, Marland G, *et al*. BCOR-CCNB3 (Ewing-like) sarcoma: a clinicopathologic analysis of 10 cases, in comparison with conventional Ewing sarcoma. *Am J Surg Pathol* 2014;38:1307–1318.
 - 21 Horn D, Chyrek M, Kleier S, *et al*. Novel mutations in BCOR in three patients with oculo-facio-cardio-dental syndrome, but none in Lenz microphthalmia syndrome. *Eur J Hum Genet* 2005;13:563–569.
 - 22 Davoody A, Chen IP, Nanda R, *et al*. Oculofaciocardio-dental syndrome: a rare case and review of the literature. *Cleft Palate Craniofac J* 2012;49:e55–e60.
 - 23 Oberoi S, Winder AE, Johnston J, *et al*. Case reports of oculofaciocardiodental syndrome with unusual dental findings. *Am J Med Genet A* 2005;136:275–277.
 - 24 Jiang YH, Fang P, Adesina AM, *et al*. Molecular characterization of co-occurring Duchenne muscular dystrophy and X-linked oculo-facio-cardio-dental syndrome in a girl. *Am J Med Genet A* 2009;149A:1249–1252.
 - 25 Kao YC, Sung YS, Zhang L, *et al*. BCOR overexpression is a highly sensitive marker in round cell sarcomas with BCOR genetic abnormalities. *Am J Surg Pathol* 2016;40:1670–1678.
 - 26 Weeks DA, Beckwith JB, Mierau GW, *et al*. Rhabdoid tumor of kidney. A report of 111 cases from the National Wilms' Tumor Study Pathology Center. *Am J Surg Pathol* 1989;13:439–458.
 - 27 Kodet R, Newton WA Jr., Sachs N, *et al*. Rhabdoid tumors of soft tissues: a clinicopathologic study of 26 cases enrolled on the Intergroup Rhabdomyosarcoma Study. *Hum Pathol* 1991;22:674–684.
 - 28 Rorke LB, Packer R, Biegel J. Central nervous system atypical teratoid/rhabdoid tumors of infancy and childhood. *J Neurooncol* 1995;24:21–28.