β -catenin/Wnt signalling pathway in fibromatosis, metaplastic carcinomas and phyllodes tumours of the breast

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Wnt signalling pathway is known to have a critical role in carcinogenesis and in epithelial-to-mesenchymal transition. Upon Wnt activation, β -catenin is translocated from the membrane to the cytoplasm and nucleus, where it interacts with transcriptional activators. It has been suggested that various spindle cell lesions of the breast may harbour Wnt pathway activation. Given that β -catenin nuclear localization constitutes a good surrogate marker of Wnt canonical pathway activation, we have investigated the distribution of β -catenin in spindle cell lesions of the breast and whether it could be employed in the differential diagnosis of these lesions. A total of 52 metaplastic breast carcinomas, eight fibromatoses and 23 phyllodes tumours were retrieved from our institutions' archives. We performed immunohistochemistry using two anti- β -catenin antibodies. In all, three fibromatoses and 21 metaplastic breast carcinomas were subjected to CTNNB1 (β -catenin encoding gene) mutation analysis by direct gene sequencing. A good correlation between the two antibodies was observed (Spearman's r > 0.82, P < 0.001). All fibromatoses and 23% of metaplastic breast carcinomas expressed nuclear β -catenin. In fibromatosis, β-catenin was more often diffusely expressed, whereas in metaplastic breast carcinomas, expression was more frequently focal. Membranous β -catenin expression was significantly lower in spindle cell carcinomas than in other subtypes of metaplastic breast carcinomas. In phyllodes tumours, stromal cells of benign and malignant subtypes displayed nuclear β-catenin expression in 94 and 57% of cases, respectively. No CTNNB1 mutation was identified in any of the 21 metaplastic carcinomas analysed, whereas the mutations 45S>S/P and 41T>T/A were found in samples of fibromatosis. In conclusion, β -catenin nuclear expression is a common feature in fibromatoses and in the stromal component of phyllodes tumours, but may also be observed in metaplastic breast carcinomas. β-catenin nuclear expression should not be used as a single marker to differentiate fibromatosis from other spindle cell tumours of the breast.

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 β -catenin is a multifunctional protein located to the intracellular side of the cytoplasmic membrane, which has a critical role in cell-to-cell adhesion by linking cadherins to the actin cytoskeleton.¹ Upon Wnt activation, β -catenin is translocated from the mem-

brane to the cytoplasm and nucleus, where it interacts with transcriptional activators to modulate genes such as c-MYC,² cyclin D1^{3,4} or NF- κ B target genes.⁵

Cadherin–catenin complexes participate in the normal development and stabilization of the mammary gland architecture, affecting cell fate, polarity and motility.¹ There are numerous lines of evidence to suggest that a dysfunctional Wnt/ β -catenin signal-ling pathway has a critical role in carcinogenesis and in epithelial-to-mesenchymal transition.^{1,3–7} Furthermore, there is evidence that β -cateninactivating mutations have oncogenic properties.^{8,9}

Several studies have investigated β -catenin expression in spindle cell lesions of the breast,

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including fibromatoses, phyllodes tumours and metaplastic carcinomas.^{7,10–13} Åberrant β -catenin expression, that is, cytoplasmic and/or nuclear, is typically found in fibromatosis,11 however, it has also been described in metaplastic breast carcinomas¹⁰ and phyllodes tumours.^{12–15} Aberrant β -catenin expression, and particularly the nuclear localization, which can be detected by immunohistochemistry, constitutes a good surrogate marker of Wnt canonical pathway activation.^{1,3,10} Therefore, and on the basis of its pattern of expression and cellular localization, β -catenin has been proposed as a useful immunohistochemical marker for the differential diagnosis of spindle cell lesions;^{16–18} however, its use has not been addressed specifically in breast tumours.

 β -catenin gene (CTNNB1) mutations have been shown to be frequently found in fibromatosis of the breast and other anatomical sites (up to 45.5% of the cases in breast, up to 85% in sporadic desmoid tumours); these mutations lead to β -catenin stabilization, nuclear accumulation and Wnt pathway activation.^{11,19} β -catenin nuclear accumulation, however, does not equate with CTNNB1 mutation. Indeed, in desmoid tumours, either sporadic or associated with familial adenomatous polyposis syndrome, APC mutations may be the mechanism leading to β -catenin nuclear expression.^{11,20} In phyllodes tumours, despite evidences of β -catenin nuclear accumulation, no CTNNB1 mutation has been so far identified in both benign and malignant subtypes,¹² suggesting that β -catenin nuclear accumulation in these lesions is driven by mechanisms other than CTNNB1 gene mutations (eg, APC mutation, Wnt5a expression).¹²

In breast cancers, *CTNNB1* gene mutations and β -catenin nuclear expression appear to be less frequent than in other types of carcinomas.^{6,9} Interestingly, Hayes *et al*¹⁰ have recently described the presence of *CTNNB1* gene mutations in ~26% of metaplastic breast carcinomas; however, in an independent study, Hennessy *et al*⁷ could not confirm this observation.

The aims of this study were threefold: (1) to address the distribution of β -catenin expression in metaplastic carcinomas, fibromatosis and phyllodes tumours using two distinct commercially available antibodies; (2) to determine whether this marker could be employed in the differential diagnosis of these lesions; and (3) to investigate whether metaplastic carcinomas harbour *CTNNB1* mutations.

Materials and methods

Case Selection

A total of 52 metaplastic breast carcinomas, eight fibromatosis and 23 phyllodes tumours of the breast were retrieved from our institutions' archives. Cases were selected consecutively, based on the availability of a formalin-fixed and paraffin-embedded block for the study. All cases were centrally reviewed at M Lacroix-Triki *et al*

least by two pathologists (MLT, FCG, AHL and/or JRF) and classified according to previously published criteria.^{12,13,21,22} Out of the 52 metaplastic breast carcinomas, 22 were classified as spindle cell carcinomas, 17 as carcinomas with squamous metaplasia, and 13 as carcinomas with heterologous elements as previously described.^{21,23,24} Out of the 23 phyllodes tumours of the breast, 16 were classified as benign, four as borderline and three as malignant using previously defined criteria.^{12,13,25,26} This study was approved by the local ethical committees.

Immunohistochemistry

Immunohistochemistry was performed on $3-\mu$ m-thick representative whole tissue sections from 83 cases, which were mounted on polylysine-coated slides. Two commercially available monoclonal antibodies raised against the C-terminal domain of β -catenin were used in this study: clone $14/\beta$ -catenin (BD) Transduction Laboratories, San Jose, CA, USA) and 17C2 (Novocastra/Leica, Newcastle Upon Tyne, UK), which were used in 1:6000 and 1:100 dilutions, respectively. Heat-induced antigen retrieval was employed for both antibodies: 30 min in buffer pH 6.0 in a pre-treatment module (Labvision, Fremont, CA, USA) for $14/\beta$ -catenin antibody and 2 min in citrate buffer pH 6.0 in a pressure cooker for clone 17C2. Sections were blocked with 1.5% H₂O₂ in methanol for 10 min and incubated with β -catenin antibodies for 60 min at room temperature. Detection was achieved with the Vector avidin-biotin complex (ABC) system (Vector Laboratories, Burlingame, CA, USA) according to the manufacturer's recommendations, using 3,3'-diaminobenzidine (Dako, Glostrup, Denmark) as a chromogenic substrate. Slides were lightly counterstained with haematoxylin. Positive controls (normal breast section) and negative controls (omission of the primary antibody and substitution of the primary antibody by IgG-matched control) were included in each experiment.

 β -catenin immunostains obtained with the two antibodies were analysed independently by two of the authors (MLT and FCG) using the Allred scoring system that combines the staining intensity and the percentage of stained cells (intensity score 0-3 + %score 0–5).²⁷ For each case, the score was assessed separately for the membranous, cytoplasmic and nuclear reactivity. In phyllodes tumours, the epithelial and spindle cell components were evaluated separately and in metaplastic breast carcinomas with clearly distinguishable morphologically distinct components (ie, carcinomatous and sarcomatous), the two components were also evaluated separately. An Allred score of >2 was considered as positive. For membranous staining, cases were further categorized according to the Allred score obtained as displaying a normal expression of β -catenin (Allred score 7 to 8), reduced expression (Allred score 3-6) or no expression (Allred score 0–2). Immunohistochemical analysis with the $14/\beta$ -catenin clone was performed with the observers blinded to the results of the analysis of 17C2 clone.

For a comparison with fibromatosis, we considered the expression patterns and levels of β -catenin in the stromal cells of phyllodes tumours and in the most abundant component of metaplastic carcinomas. Data analysis was performed with the results obtained with each antibody and also with combined results (ie, for nuclear/cytoplasmic staining, positivity is defined as positive for at least one antibody; for membranous staining, the highest Allred score was taken into account).

Microdissection and DNA Extraction

Metaplastic carcinomas, one breast fibromatosis and two desmoid tumours were subjected to CTNNB1 sequencing. All cases were microdissected to ensure >75% of purity of neoplastic cells. Microdissection was performed with a sterile needle under a stereomicroscope (Olympus SZ61, Tokyo, Japan) from ten consecutive 8- μ m-thick sections stained with nuclear fast red as previously described.²⁸ DNA was extracted using the DNeasy Kit (Qiagen Ltd, Crawley, UK) according to the manufacturer's recommendations. DNA concentration was measured with the Pico-Green assay as per the manufacturer's instructions (Invitrogen, Paisley, UK).²⁸ Out of the 52 metaplastic breast carcinomas, microdissection yielded sufficient DNA of optimal quality in 21 samples. One breast fibromatosis and two desmoid tumours vielded sufficient DNA of optimal quality.

CTNNB1 Mutation Analysis

Sequencing of known mutation hotspots of CTNNB1 on exon $3^{6,9-11,19}$ was performed in 21 metaplastic carcinomas (8 cases with β -catenin nuclear expression and 13 cases without nuclear expression), one breast fibromatosis and two sporadic desmoid tumours. As a positive control, DNA samples of the HCT116 colon cancer cell line, which is known to harbour CTNNB1 mutation in exon 3,²⁹ were included in each experiment. The primers used for *CTNNB1* sequencing have been previously described.¹⁰ In all, 50 ng tumour DNA was amplified and sequencing reactions were carried out using the DNA Sequencing Kit BigDye Terminator v 1.1 Cycle Sequencing Ready Reaction Mix (Applied Biosystems, Warrington, UK) as previously described.²³ Sequences of the forward and reverse strands were analysed with Mutation Surveyor software (Softgenetics, PA, USA). All reactions were carried out in duplicate from the original DNA sample.

Statistical Analysis

The SPSS statistical software package was used for all statistical analysis. Spearman's correlation coefficient and κ agreement value were assessed to analyse the concordance between results from the two β -catenin antibodies used in parallel in this study. For each antibody, correlations between categorical variables were performed using the χ^2 test and Fisher's exact test. A *P*-value of 0.05 was considered as statistically significant.

Results

Comparison of the Two Anti-β-Catenin Antibodies

Two commercially available antibodies raised against β -catenin, clones 14/ β -catenin and 17C2, were analysed independently on 83 breast lesions. We observed statistically significant correlations between the two antibodies for the semiquantitative assessment of β -catenin nuclear, cytoplasmic and membranous expression (Spearman's r > 0.82, P < 0.0001 for all comparisons, Table 1). Analysis

Table 1 Comparison of β -catenin immunostainings in 83 spindle cell lesions of the breast using two commercially available antibodies (clones 14/ β -catenin and 17C2)

	Clone 14/β-catenin (n = 83)	Clone 17C2 (n = 83)
β-catenin membranous Allred score	s expression	
0	38 (45.8%)	32 (38.6%)
2	0	4 (4.8%)
3	2 (2.4%)	8 (9.6%)
4	3 (3.6%)	5 (6%)
5	5 (6%)	3 (3.6%)
6	10 (12%)	7 (8.4%)
7	8 (9.6%)	13 (15.7%)
8	17 (20.6%)	11 (13.3%)
Spearman's <i>r,</i> two-tailed	0.90 (P<0.	0001)
β-catenin cytoplasmic Allred score	expression	
0	15 (18.1%)	13 (15.7%)
2	0	1 (1.2%)
3	5 (6%)	6 (7.2%)
4	13 (15.7%)	22 (26.6%)
5	16 (19.2%)	14 (16.8%)
6	22 (26.6%)	23 (27.7%)
7	12 (14.4%)	4 (4.8%)
8	0	0
Spearman's <i>r</i> , two-tailed	$0.84 \ (P < 0.0)$	0001)
β-catenin nuclear exp Allred score	ression	
0	45 (54.3%)	39 (47.1%)
2	3 (3.6%)	9 (10.9%)
3	9 (10.9%)	10 (12%)
4	8 (9.6%)	10 (12%)
5	4 (4.8%)	12 (14.4%)
6	12 (14.4%)	3 (3.6%)
7	2 (2.4%)	0
8	0	0
Spearman's <i>r</i> , two-tailed	0.83 (P<0.	0001)

		Clone 14/β-catenin β-catenin nuclear		Clone 17C2			$Combined^{a}$			
				r	β-catenin nuclear			β-catenin nuclear		
n		Positive (Allred >2)	Nego (Allre	ative d 0–2)	Positive (Allred >2)	Neg (Allre	ative ed 0–2)	Positive (Allred >2)	Neg (Allre	ative d 0–2)
Squamous	17	3 (17.6%)	14 (83	2.4%)	3 (17.6%)	14 (8	2.4%)	5 (29.4%)	12 (7	0.6%)
Heterologous	13	2 (15.4%)	11 (84	4.6%)	1 (7.7%)	12 (9	2.3%)	2 (15.4%)	11 (8	4.6%)
elements Spindle cell Total Fisher's test, 3×2	22 52	4 (18.2%) 9 (17.3%) P = 0.999 β-catenin cytoplasmic		$\begin{array}{cccc} 4 & (18.2\%) & 18 & (81.8\%) \\ 8 & (15.4\%) & 44 & (84.6\%) \\ & P \!=\! 0.788 \end{array}$		5 (22.7%) 17 (77.3%) 12 (23%) 40 (77%) P = 0.651				
				β-catenin cytoplasmic		β-catenin cytoplasmic				
	n	Positive (Allred > 2)	Nego (Allre	ative d 0–2)	Positive (Allred >2)	Neg (Allre	ative ed 0–2)	Positive (Allred >2)	Neg (Allre	ative d 0–2)
Squamous	17	17 (100%)	0		17 (100%)	0		17 (100%) 0		
metaplasia Heterologous elements Spindle cell Total Fisher's test, 3×2	13	13 (100%)	0		13 (100%)	0		13 (100%)	0	
	22 52	22 (100%) 52 (100%)	0 0 NP		22 (100%) 0 52 (100%) 0 NP		22 (100%) 52 (100%)	00%) 0 .00%) 0 NP		
		β-catenin membranous			β-catenin membranous			β-catenin membranous		
		Normal (Allred 7–8)	Reduced (Allred 3–6)	Negative (Allred 0–2)	Normal (Allred 7–8)	Reduced (Allred 3–6)	Negative (Allred 0–2)	Normal (Allred 7–8)	Reduced (Allred 3–6)	Negative (Allred 0–2)
Squamous metaplasia Heterologous elements Spindle cell Total Fisher's test, 3×3	17	12 (70.6%)	5 (29.4%)	0	12 (70.6%)	5 (29.4%)	0	14 (82.3%)	3 (17.7%)	0
	13	7 (53.8 %)	6 (46.2%)	0	7 (53.8 %)	6 (46.2%)	0	8 (61.5%)	5 (38.5%)	0
	22 52	6 (27.3%) 25 (48.0%)	9 (40.9%) 20 (38.5%) P=0.009	7 (31.8%) 7 (13.5%)	5 (22.7%) 24 (46.2%)	12 (54.6%) 23 (44.2%) P = 0.011	5 (22.7%) 5 (9.6%)	6 (27.3%) 28 (53.8%)	13 (59.1%) 21 (40.4%) P = 0.005	3 (13.6%) 3 (5.8%)

Table 2 β -catenin expression in metaplastic breast carcinomas (n = 52)

NP, not performed (not computed as the data are constant).

^aCombines the data obtained with both antibodies (ie, for nuclear/cytoplasmic staining, positivity is defined as positive for at least 1 antibody; for membranous staining, the highest Allred score was taken into account).

of agreement between the two antibodies for nuclear, cytoplasmic and membranous expression revealed nearly perfect agreement for all comparisons,³⁰ with *K* scores of 0.88 (95% confidence intervals 0.78–0.98), 0.89 (95% confidence intervals 0.73–1.01) and 0.85 (95% confidence intervals 0.74–0.97) when positive (Allred score > 2) vs negative (Allred score \leq 2) categories were used.

β-Catenin Expression in Metaplastic Carcinomas

By combining the results of both antibodies, 49 out of 52 metaplastic breast carcinomas displayed β catenin membranous expression (Table 2). β -catenin membranous expression was significantly lower in spindle cell carcinomas than in carcinomas with squamous metaplasia and in carcinomas with heterologous elements (Fisher's exact test 3×3 , two-tailed P = 0.005 when combining both antibodies; Table 2 and Figure 1). In seven cases exhibiting at least two morphologically distinct components in the same sections, the sarcomatous elements consistently displayed loss or reduction of membranous expression when compared with the epithelial component (Fisher's exact test, two-tailed, P = 0.0005 and P = 0.004 for clones $14/\beta$ -catenin and 17C2, respectively; Table 3).

Analysis of β -catenin nuclear expression revealed 17% (9/52) and 15% (8/52) of cases displaying nuclear expression with the 14/ β -catenin and 17C2 clones, respectively. When results from both antibodies were combined (ie, a case scored as positive for only one antibody was considered positive), 23% (12/52 cases) of metaplastic breast carcinomas displayed β -catenin nuclear expression (Table 2 and Figure 2). Regardless of the antibody used, no 1441



Figure 1 β -catenin membranous expression in metaplastic breast carcinomas. A carcinoma with squamous metaplasia (a) displaying strong β -catenin reactivity at the membrane of the majority of the neoplastic cells (b). A spindle cell carcinoma (c) displaying lack/ reduction of β -catenin membranous expression (d). Both cases lacked β -catenin nuclear staining.

association between the subtypes of metaplastic breast carcinomas and β -catenin nuclear expression was found (Table 2 and Figure 2). In addition, in the seven cases where two morphologically distinct components were evaluated, β -catenin nuclear accumulation was not significantly associated with any of the components (Table 3). For both antibodies, all positive cases for nuclear expression displayed <33% of positive neoplastic cells, with variable intensity, never exceeding an Allred score of 5 (Table 4).

β-Catenin Expression in Fibromatosis

With both antibodies, all fibromatoses analysed (n=8) expressed nuclear β -catenin in at least 33% of neoplastic cells, with variable intensity, and lacked any membranous expression. Cytoplasmic staining was observed in 100 and 87% of cases with 14/ β -catenin and 17C2 clones, respectively (Table 5 and Figure 3).

β-Catenin Expression in Phyllodes Tumours

In all phyllodes tumours, membranous staining was observed with both antibodies only in the epithelial component (Table 5 and Figure 4). Nuclear expression was found in epithelial and stromal cell components in up to 75 and 94%, respectively, of benign phyllodes tumours (Figure 4 and Table 5), with a preferential localization of the nuclear-positive stromal cells around the epithelial structures. In accord with the results described by Sawyer *et al*,¹² a trend for a lower prevalence of nuclear expression in stromal cells of borderline/malignant phyllodes tumours than in benign phyllodes tumours was found when the 14/ β -catenin clone was employed (Fisher's exact test, two-tailed P = 0.067; Figure 4 and Table 5).

CTNNB1 Mutation Analysis in Metaplastic Carcinomas

To investigate the presence of *CTNNB1* mutations in metaplastic breast cancers, we sequenced eight

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		Clone 14/	β-catenin	Clone	17C2
	MBC subtype	Carcinomatous component (Allred score)	Sarcomatous component (Allred score)	Carcinomatous component (Allred score)	Sarcomatous component (Allred score)
Membranous expression					
Case 1	Spindle cell	7	0	5	3
Case 6	Squamous metaplasia	8	0	8	3
Case 16	Squamous metaplasia	8	3	8	3
Case 31	Heterologous elements	8	3	8	3
Case 33	Spindle cell	7	0	7	0
Case 45	Spindle cell	8	0	7	0
Case 46	Spindle cell	7	0	7	3
Fisher's exact test	1	P=0.	0005	P = 0	.004
(normal <i>vs</i> reduced/negative) ^a					
Cytoplasmic expression					
Case 1	Spindle cell	6	4	6	6
Case 6	Squamous metaplasia	6	5	6	5
Case 16	Squamous metaplasia	7	5	6	4
Case 31	Heterologous elements	6	5	6	4
Case 33	Spindle cell	7	6	6	5
Case 45	Spindle cell	5	5	5	5
Case 46	Spindle cell	5	5	5	4
Fisher's exact test		N	Р	N	Р
(positive <i>vs</i> negative) ^b					
Nuclear expression					
Case 1	Spindle cell	0	0	0	0
Case 6	Squamous metaplasia	0	0	0	4
Case 16	Squamous metaplasia	0	0	0	0
Case 31	Heterologous elements	0	0	0	0
Case 33	Spindle cell	0	2	0	2
Case 45	Spindle cell	0	2	0	0
Case 46	Spindle cell	0	3	0	0
Fisher's exact test	-	P = 0	.999	P = 0	.999
(positive <i>vs</i> negative) ^b					

Table 3	β -catenin ex	pression in met	aplastic breas	t carcinomas	(n = 7)	displaying tw	o morphologically	y distinct tumour comp	onents
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MBC, metaplastic breast carcinoma; NP, not performed (not computed as the data are constant).

^aNormal: Allred score 7–8; reduced: Allred score 3–6; negative: Allred score 0–2.

^bPositive: Allred score 3–8; negative: Allred score 0–2.

metaplastic carcinomas with β -catenin nuclear expression and 13 cases without β -catenin nuclear expression. Positive controls, included in each experiment, showed the expected presence of the previously reported CTNNB1 mutation in exon 3 (deletion of codon 45) in the HCT116 colo-rectal cell line.²⁹ In addition, to test the accuracy of the CTNNB1 mutation analysis on formalin-fixed and paraffin-embedded tissues, a case of breast fibromatosis from this cohort and two sporadic desmoid tumours were included in the analysis and were found to harbour activating CTNNB1 mutations on exon 3 (25195T>TC:45S>S/P in breast fibromatosis, Figure 3b; 25183A>AG:41T>T/A in both desmoid tumours), which have been previously described in sporadic desmoid tumours.¹⁹ None of the 21 metaplastic carcinomas analysed here harboured any exon 3 CTNNB1 gene mutation. In two cases, the distinct components (ductal and spindle in one case; spindle, rhabdoid and chondroid in another case) were separately microdissected and

subjected to *CTNNB1* sequencing analysis, but no mutation was found.

Discussion

 β -catenin nuclear expression in neoplastic lesions constitutes a surrogate marker of Wnt pathway activation and may also suggest the presence of mutations of the *CTNNB1* gene and other genes in the Wnt pathway (eg, *APC*). Given the high prevalence of *CTNNB1* and *APC* gene mutations in fibromatoses from various anatomical sites including breast,^{11,19,20} β -catenin immunohistochemical assessment has been proposed as a useful tool in the differential diagnosis between fibromatosis and other spindle cell lesions of the breast.^{16–18} Here, we demonstrate that although fibromatosis consistently express β -catenin nuclear expression, 23% of metaplastic breast carcinomas and the stromal cells of 93% of phyllodes tumours also display β -catenin nuclear expression.

β -catenin expression in breast cancers

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Figure 2 β -catenin nuclear expression in metaplastic breast carcinomas. A carcinoma with squamous metaplasia (a) displaying focal strong nuclear β -catenin staining (b). A carcinoma with heterologous elements (c) displaying focal weak-to-moderate nuclear expression of β -catenin (d). A spindle cell carcinoma (e) displaying strong β -catenin nuclear reactivity (f). Note that the carcinoma with squamous metaplasia and the carcinomatous component of the carcinoma with heterologous elements (d, asterisk) and the spindle cell carcinoma showed reduction/lack of β -catenin membranous expression.

Fibromatosis, phyllodes tumours and a subset of metaplastic breast carcinomas may in some instances give rise to a diagnostic difficulty, especially in core-

Table 4 Summary of Allred scores for nuclear β -catenin expression in 52 metaplastic carcinomas according to the antibody clones $14/\beta$ -catenin and 17C2

Allred score	Clone 14/β-catenin Number of cases	Clone 17C2 Number of cases	
Negative			
0	42	36	
2	1	8	
Total	43	44	
Positive			
3	6	2	
4	2	5	
5	1	1	
6	0	0	
7	0	0	
8	0	0	
Total	9	8	

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needle biopsies that may not be representative of the whole lesion.¹⁶ In our series, all fibromatosis displayed diffuse β -catenin nuclear expression and lack of membrane expression. It should be pointed out that a previous study on a larger cohort of 33 breast fibromatoses reported that up to 18% of the cases failed to display nuclear expression.¹¹ β -catenin cytoplasmic expression is often observed in fibromatosis; however, fibromatosis neoplastic cells consistently lack membranous expression (our results and those of Abraham $et al^{11}$). Likewise, the stromal component of phyllodes tumours do not express β -catenin at the membrane and may harbour nuclear reactivity in up to 93% of the cases described in this study. Therefore, given that the patterns of β -catenin expression in fibromatosis and a subset of phyllodes tumours and metaplastic breast carcinomas clearly overlap, the differential diagnosis between these three spindle cell lesions should not rely solely on the use of β -catenin immunohistochemistry.

Å previous study¹⁰ described aberrant β -catenin expression in 92% of metaplastic carcinomas and



Figure 3 β -catenin expression and *CTNNB1* exon 3 mutation in breast fibromatosis. A breast fibromatosis (a) adjacent to an entrapped normal breast lobule, which harboured an activating 25195T > TC:45S > S/P *CTNNB1* exon 3 mutation (b) and cytoplasmic and nuclear β -catenin expression with clones 14/ β -catenin (c) and 17C2 (d). No membranous staining was detected with any of the antibodies tested.







Figure 4 β -catenin expression in phyllodes tumours of the breast. A benign (**a**, **b**) and a malignant (**c**, **d**) phyllodes tumours. Membranous and nuclear β -catenin expression was observed, respectively, in epithelial and stromal cells of both lesions. Note that nuclear expression was predominantly found in stromal cells surrounding the epithelial cells. In addition, less stromal cells displayed nuclear expression in the malignant phyllodes tumour.

identified exon 3 CTNNB1 gene mutations in 26% of cases. Using two antibodies against β -catenin, in this study we demonstrate that more than a third (46%) of metaplastic breast carcinomas displayed reduction or lack of β -catenin membrane staining and that abnormal cytoplasmic and/ or nuclear staining was found in 100% of cases. Although these results suggest that Wnt pathway may be dysfunctional in the vast majority of metaplastic breast cancers, it should be noted that the mechanisms leading to β -catenin activation in these lesions remain contentious. In contrast with the results from Hayes $et al^{10}$ and in agreement with Hennessy et al,⁷ no exon 3 CTNNB1 gene mutations were identified in our analysis of microdissected metaplastic breast carcinomas. The stark contrast between the results by Hayes *et al*¹⁰ and the combined results of Hennessy et al^7 and those presented in this study may stem from different case selections, different methods for gene sequencing and different approaches for

sample accrual. Importantly, in this study, all cases were microdissected and sequenced in duplicate, minimizing the artefacts induced by formalin fixation. Furthermore, in two cases, morphologically distinct components of metaplastic breast carcinomas were microdissected separately and no mutations were identified.

In this study we demonstrate that spindle cell metaplastic breast carcinomas display significantly lower levels of β -catenin membrane staining than other subtypes of metaplastic cancers. We^{31,32} and others^{7,33,34} have previously demonstrated that metaplastic cancers display transcriptomic features suggestive of an epithelial-to-mesenchymal transition phenotype. Given the role of the Wnt pathway in epithelial-to-mesenchymal transition, our results are consistent with the hypothesis that at least a subgroup of metaplastic breast cancers have features of epithelial-to-mesenchymal transition and that Wnt pathway activation may have a role in this

Table 5 β -catenin expression in fibromatosis ($n = 8$) and in the
distinct components of phyllodes tumours ($n = 23$, benign $n = 16$,
borderline/malignant $n = 7$).

	Clone 14/ β-catenin	Clone 17C2
Fibromatosis (n = 8) Nuclear		
Positive	8 (100%)	8 (100%)
Negative	$\begin{pmatrix} 0 \\ 0 \end{pmatrix}$	$\begin{bmatrix} 0 \\ 5 \\ (1 \\ 0) \end{bmatrix}$
Median Alfred score (range)	b (4-7)	5 (4-6)
Cytoplasmic		
Positive	8 (100%)	7 (87.5%)
Median Allred score (range)	0 4 (3–7)	4.5 (2–5)
Membranous		
Normal	0	0
Reduced	0	0
Negative	8 (100%)	8 (100%)
Phyllodes tumours $(n = 23)$ Nuclear		
Benign PTs		
Positive	15 (93.7%)	15 (93.7%)
Negative	1 (6.3%)	1 (6.3%)
Borderline/malignant PTs		- (
Positive	4(57.1%)	5(71.4%)
Fisher's exact test two-tailed	3(42.9%) P=0.0671	2(20.0%) P=0.209
Epithelial component	1 = 0.0071	1 - 0.205
Benign PTs	(750/)	0 (50 00/)
Positive	12 (75%)	9 (56.2%)
Borderline/malignant PTs	4 (20 /0)	7 (10.070)
Positive	3 (42.9%)	3 (42.9%)
Negative	4 (57.1%)	4 (57.1%)
Fisher's exact test, two-tailed	P = 0.182	P = 0.667
Cytoplasmic		
Spindle cell component		
Benign P1s Positivo	5 (21.2%)	6 (37 5%)
Negative	11(68.8%)	10(62.5%)
Borderline/malignant PTs		
Positive	3 (42.9%)	4 (57.1%)
Negative Fisher's exact test, two tailed	4(57.1%)	3 (42.9%)
Enithelial component	P = 0.057	P = 0.050
Positive	23 (100%)	23 (100%)
Negative	0	0
Membranous		
Spindle cell component	0	0
Reduced	0	0
Negative	23 (100%)	23 (100%)
Epithelial component		
Benign PTs	40 (750/)	40 (00 50/)
Normal Reduced	12(75%) 4(25%)	10 (62.5%)
Negative	4 (25 /0) 0	0 (37.370)
Borderline/malignant PTs	-	-
Normal	4 (57.1%)	2 (28.6%)
Keduced	3 (42.9%)	5 (71.4%)
Fisher's exact test. 2×3	P = 0.625	P = 0.193

PTs, phyllodes tumours.

process. Given the apparent lack of mutations in the exon 3 hotspot in this series and in the study by Hennessy *et al*,⁷ our results suggest that β -catenin aberrant expression in metaplastic breast carcinomas may stem from mutations affecting other exons of the *CTNNB1* gene or other genes in the Wnt pathway (eg, the *APC* gene). Alternatively, β -catenin aberrant expression and Wnt pathway activation in these tumours may be the consequence of a global epithelial-to-mesenchymal transition programme.^{33,34}

Consistent with the hypothesis of β -catenin aberrant expression and epithelial-to-mesenchymal transition, in seven biphasic metaplastic lesions, we observed a lack/reduction of membranous β -catenin expression to be significantly associated with the sarcomatous component (Table 3). Furthermore, nuclear β -catenin expression was restricted to the sarcomatous counterpart in two of these cases (one with each antibody); however, no statistically significant difference was observed in the prevalence of β -catenin nuclear expression in the sarcomatous and carcinomatous components. Therefore, in some cases, activation of the β -catenin/Wnt pathway is coincidental with the morphological characteristics of epithelial-to-mesenchymal transition, which may indicate some degree of clonal divergence between the morphologically distinct components of metaplastic breast carcinomas. Likewise, we have previously shown that intratumour morphological heterogeneity in some metaplastic carcinomas of the breast may be underpinned by distinct patterns of genetic aberrations.²³ Further studies investigating the prevalence of intratumour genetic heterogeneity in relation to the presence of mutations of Wnt pathway genes in distinct components of metaplastic breast cancers are warranted.

In conclusion, β -catenin nuclear expression is a common feature in fibromatosis and in the stromal component of phyllodes tumours, but it may also be observed in metaplastic breast carcinomas. Therefore, β -catenin nuclear expression should not be used as a single marker to differentiate fibromatosis from other spindle cell tumours of the breast. Despite the presence of β -catenin nuclear expression in a subgroup of metaplastic breast carcinomas, no exon 3 *CTNNB1* gene mutations were found in 21 microdissected metaplastic breast cancers, suggesting that Wnt pathway activation in these tumours may be driven either by mutations in other exons of the *CTNNB1* gene or activation/inactivation of other components of the Wnt pathway.

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Disclosure/conflict of interest

The authors declare no conflict of interest.

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