

Overexpression of fatty acid synthase is associated with palmitoylation of Wnt1 and cytoplasmic stabilization of β -catenin in prostate cancer

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Fatty acid synthase (FASN), a key metabolic enzyme for liponeogenesis highly expressed in several human cancers, displays oncogenic properties such as resistance to apoptosis and induction of proliferation when overexpressed. To date, no mechanism has been identified to explain the oncogenicity of FASN in prostate cancer. We generated immortalized prostate epithelial cells (iPrECs) overexpressing FASN, and found that ¹⁴C-acetate incorporation into palmitate synthesized *de novo* by FASN was significantly elevated in immunoprecipitated Wnt-1 when compared to isogenic cells not overexpressing FASN. Overexpression of FASN caused membranous and cytoplasmic β -catenin protein accumulation and activation, whereas FASN knockdown by short-hairpin RNA resulted in a reduction in the extent of β -catenin activation. Orthotopic transplantation of iPrECs overexpressing FASN in nude mice resulted in invasive tumors that overexpressed β -catenin. A strong significant association between FASN and cytoplasmic (stabilized) β -catenin immunostaining was found in 862 cases of human prostate cancer after computerized subtraction of the membranous β -catenin signal ($P < 0.001$, Spearman's $\rho = 0.33$). We propose that cytoplasmic stabilization of β -catenin through palmitoylation of Wnt-1 and subsequent activation of the pathway is a potential mechanism of FASN oncogenicity in prostate cancer.

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Prostate cancer is the most prevalent noncutaneous malignancy in men in the United States. Prostate cancer has also been associated with obesity, high-fat intake and the so-called metabolic syndrome.^{1–3} Fatty acid synthase (FASN) is a key enzyme for liponeogenesis in the metabolic syndrome.⁴

Cancer cells display increased anaerobic glycolysis even in the presence of oxygen (the so-called Warburg effect), as well as significantly increased *de novo* synthesis of fatty acids through FASN.⁵ In fact, FASN is minimally expressed in most normal human tissues with the exception of the liver and adipose tissue whereas upregulation of FASN is a common feature of many tumor cell lines and tissues. In addition,

FASN overexpression has been correlated to poor prognosis or shorter disease-free survival in patients with several human malignancies.^{6–12} In the prostate, FASN overexpression has been associated with both early and late phases of carcinogenesis. *In vitro*, the expression of FASN in the androgen-dependent prostate cancer cells, LNCaP is enhanced by androgens whereas silencing of the FASN gene by RNA interference induces apoptosis.^{13,14} *In vivo*, prostate adenocarcinomas with FASN upregulation display a more aggressive biologic behavior.^{15,16} The overexpression and copy number gain of the FASN gene were also demonstrated in tissues from prostate cancer patients and associated with

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different molecular tumor signatures.^{8,17} In addition, high cellular levels of FASN in prostate cancer samples were inversely correlated with the expression of PTEN and were associated with the activation and nuclear localization of Akt.^{18,19} Taken together these observations suggest that FASN overexpression confers a selective growth advantage to prostate cancer cells *in vitro* and *in vivo*. Nevertheless, no specific mechanism has been identified so far to explain the oncogenicity of FASN in prostate cancer.

The principal enzymatic product of FASN is palmitic acid.²⁰ Palmitoylation, as well as other lipid modifications such as myristoylation, represents key regulatory switches in most signal transduction pathways including the Wnt/ β -catenin pathway.^{21,22} β -Catenin is a dual function member of the Wnt signaling pathway that exists in three cellular pools: at the membrane where it is involved in cell–cell adhesion and in the cytoplasm or in the nucleus where it acts as a transcription factor activator.²³ In the absence of Wnt signal, β -catenin is bound to the adenomatous polyposis coli (APC)/axin/GSK3 β complex, phosphorylated and then degraded through the ubiquitin/proteasome system.²⁴ Under activated Wnt signaling or in case of β -catenin gene mutations, the protein does not complex to APC, it stabilizes in the cytoplasm, it is carried into the nucleus where it activates members of the Tcf/Lef transcription factor family and can be in equilibrium with the cytoplasmic pool.²³ In the prostate, β -catenin promotes androgen signaling through its ability to bind to the androgen receptor (AR) and activate transcription of androgen-regulated genes.^{25,26} *In vitro*, β -catenin is overexpressed in several prostate cancer cell lines, whereas, *in vivo*, activation of the APC gene and stabilization of β -catenin in the murine prostate result in prostatic intraepithelial neoplasia and prostate cancer.^{27–29} Overexpression of β -catenin in prostate cancer tissues also correlates with advanced stage and hormone refractory progression of the disease.^{29,30} β -catenin is thus involved in prostate carcinogenesis despite the evidence that less than 5% of human prostate cancer harbor mutations of this gene.^{31,32} Subcellular localization of the β -catenin protein is also peculiar in prostate cancer compared to other epithelial tumors: it has been described as membranous and cytoplasmic whereas its nuclear expression is seen rarely if at all.^{29,33,34}

Here we hypothesized that FASN overexpression in immortalized human prostate epithelial cells (iPrECs) would lead to a specific pattern of palmitoylation involving the activation of the Wnt/ β -catenin pathway. We then analyzed the relationship between FASN and β -catenin in orthotopic xenografts of AR-expressing iPrEC cells (AR-iPrECs) overexpressing FASN and in human prostate cancer cell lines and tissue samples.

MATERIALS AND METHODS

Generation of FASN-Overexpressing Immortalized Prostate Epithelial Cells and Culture Conditions

Immortalized human prostate epithelial cell lines and AR-iPrEC were engineered. The human FASN cDNA (7536 bp)

was generated, sequenced and subcloned into the pBabe retroviral vector containing a puromycin resistance gene (Addgene, Cambridge, MA, USA). To obtain stable FASN-expressing cell lines, 293T-packaging cells plated in 60 mm dishes were cotransfected with the retroviral vector pBabe-FASN (2 μ g) and the packaging plasmid pCL (1 μ g), using the Fugene 6 transfection agent (Roche, Mannheim, Germany). Cells were transduced through infection using 10 mg/ml polybrene (Sigma, St Louis, MO, USA); at 48 h after infection, 2 μ g/ml puromycin (Sigma) was added to the culture medium to select for puromycin-resistant cells. Cells infected with pBabe vector alone (iPrEC-vector or AR-iPrEC-vector) were used as control. iPrEC and AR-iPrEC cell lines were grown in PrEBM medium obtained from Cambrex Bioscience (Walkersville, MD, USA).

Protein Labeling, Immunoprecipitation, and Immunoblotting

Immortalized prostate epithelial cell-vector and iPrEC-FASN cells were starved for 3 h and then pulsed for 2–3 h in medium containing 0.3 mCi/ml of [³H]-palmitate (Perkin-Elmer Life Sciences, Boston, MA, USA). Labeled cells were lysed in RIPA buffer (50 mM Tris, 1 mM EDTA, 150 mM NaCl, 1% NP-40, 0.5% deoxycholate, and 0.1% SDS) containing complete protease inhibitor mix with EDTA (Roche), 1 mM Na₃VO₄, 1 mM NaF, and the whole-cell lysates were subjected to SDS–PAGE under nonreducing conditions. For immunoprecipitation, the lysates were cleared with protein A- or G-Sepharose. Anti-Wnt1 antibody (Santa Cruz Biotechnology, Santa Cruz, CA, USA) was incubated for 1 h at 4°C, followed by overnight incubation with protein A- or G-Sepharose. Finally, the beads were washed with lysis buffer and immune complexes were subjected to SDS–PAGE under nonreducing conditions. Gels were transferred to PVDF membrane and then exposed to BioMax MS Film using intensifying screens for 2–7 days at –80°C.

Wnt-1 Palmitoylation Assay

Immortalized prostate epithelial cell-vector and iPrEC-FASN cells (at 70% confluence) were washed twice with PBS, then pulsed for 24 h in medium containing 2 μ Ci/ml of ^{2–14}C acetate sodium (Amersham Biosciences, Piscataway, NJ, USA). Labeled cells were lysed in RIPA buffer. After 30 min at 4°C, insoluble material was removed by centrifugation at 13 000 r.p.m. Protein quantification was performed and 500 μ g of proteins were used for immunoprecipitation. After 1 h preclearing with protein G-Sepharose, anti-Wnt 1 polyclonal antibody (Zymed, Invitrogen, Rockville, MD, USA) was incubated overnight at 4°C, followed by 1 h incubation with protein G-Sepharose.

Purified rabbit IgG were used as negative control of immunoprecipitation. Immune complexes were collected by centrifugation, washed five times in lysis buffer, and resuspended in 300 μ l of lysis buffer. Each sample was placed in a vial filled with liquid scintillation mixture and counted

in a β -counter (Beckman 5801 LS). Mean c.p.m. in immune complex from lysates of iPrEC-vector and iPrEC-FASN subtracted the background count (c.p.m. in the negative control sample) were measured.

Western Blotting

Immortalized human prostate epithelial cell and AR-iPrEC were lysed in RIPA buffer. Cellular debris was then pelleted by centrifugation, and supernatants were quantitated using a Bradford protein assay (Bio-Rad, Hercules, CA, USA). Equal amounts of protein were resolved on a precast 4–12% Tris-glycine SDS-polyacrylamide gels (Invitrogen), and transferred to Hybond nitrocellulose membrane (Amersham Biosciences). The membranes were blocked for 1 h in TBS containing 0.05% Tween-20 and then incubated overnight with the primary antibody. The following antibodies were used: anti-FASN, anti- β -catenin, (both from BD Transduction laboratories, San Diego, CA, USA) at 1:1000 dilution, anti-active β -catenin (Millipore, Billerica, MA, USA) at 1:2000 dilution, and anti- β actin antibody (Sigma) at 1:5000 dilution. After incubation with peroxidase-coupled secondary antibodies (Bio-Rad), blots were developed using enhanced chemiluminescence (Amersham) and exposed to X-ray films (Kodak, Rochester, NY, USA).

RNAi

To stably suppress FASN, we used pLKO.1 lentiviral constructs containing three different short-hairpin RNA (shRNA) sequences generated by the RNAi Consortium (shFASN no. 1: CATGGAGCGTATCTGTGAGAA; shFASN no. 2: CGAGAGCACCTTTGATGACAT; shFASN no. 3: CCTCCACAATTAAACCGCAT). Lentiviral infections were performed as previously described with some modifications.³⁵ Briefly, 0.5×10^6 293T cells were seeded in 60 mm dishes. The following day the 293T cells were transfected using Fugene 6 (Roche) with pLKO.1 plasmids simultaneously with packaging plasmids gag-pol and VSV-g. The media containing the progeny virus released from the 293T cells was collected and used to infect target cell lines (LNCaP, CL1, PC3 and DU145) for 3–6 h in the presence of 8 μ g/ml polybrene (Sigma). The target cells were selected with 2 μ g/ml of puromycin (Sigma-Aldrich) for 48 h, incubated for additional 48 h without puromycin and then lysed for western blotting analysis. Three different shRNAs were utilized (data not shown) and the shFASN no. 2 was selected as the optimal, as it resulted in the most profound knockdown of the target. The experiments were performed in triplicate, showing comparable results.

Xenografts

AR-iPrEC-FASN (5×10^6) were injected orthotopically in 6-week-old immunodeficient mice into the anterior lobe of prostate tissue as previously described.³⁶ Tumor size was monitored every 3 days and mice were killed when the tumor size reached a diameter of ~ 1.5 cm.

Histological Samples

A series of 862 men who underwent radical prostatectomy was used to assess the correlation between the expression of FASN and β -catenin. At least three cores from each prostate cancer sample were included in nine high-density tissue microarrays (TMAs). Benign prostate tissue cores ($n = 40$) were also included in the TMAs. The men were participants in the Health Professionals Follow-up Study (<http://www.hsph.harvard.edu/hpfs/>) and the Physicians' Health Study (<http://phs.bwh.harvard.edu/>), and were diagnosed with prostate cancer between 1982 and 2004. The median age (s.d.) at cancer diagnosis was 66 (3.7 years). Briefly, 61% of the cases were Gleason 7, and 15% Gleason 8–10. Quality control specimens were included in each array to assess possible drift or batch variability in staining.

Immunofluorescence

For immunofluorescence experiments, 5×10^4 cells were seeded in four-well chamber slides (BD Biosciences, Bedford, MA, USA) for 24 h. Cells were washed twice with PBS, fixed with 4% paraformaldehyde for 10 min at RT, and permeabilized with PBS/0.1% Triton X 100 for 5 min at RT. After two washes with PBS, cells were blocked with PBS/1% BSA for 1 h, followed by incubation with the primary antibody diluted in PBS/1% BSA for 1 h at RT. The antibodies used were anti- β -catenin, (BD Transduction laboratories, 1:100 dilution) and anti-active β -catenin (Millipore, 1:250 dilution). Cells were rinsed three times with PBS and then incubated with Texas-Red anti mouse IgG (Vector laboratories, Burlingame, CA, USA; 1:200 dilution) for 1 h at RT. Cells were washed three times, followed by incubation with 1 μ M Hoechst 33342 (Invitrogen) for 5 min and three additional washes. Finally, the chamber slides were mounted with Vectashield mounting medium (Vector laboratories). Cells were imaged at $\times 63$ with a Plan-Apochromat oil immersion lens on an Axioplan 2 Apoptome epifluorescence microscope (Zeiss, Germany).

Immunohistochemistry

Sections 4 μ m thick were cut from each TMA and from formalin-fixed paraffin-embedded xenograft tumor samples and used for subsequent immunohistochemical analysis. Sections were incubated using a rabbit polyclonal antibody for FASN (Assay Designs, Ann Arbor, MI, USA; diluted 1:400) and a mouse monoclonal antibody for β -catenin (Dako, Hamburg, Germany; diluted 1:200). Antigen retrieval for FASN was performed in microwave for 15 min in citrate buffer. Antigen retrieval for β -catenin was performed in pressure cooker at 120° for 5 min in citrate buffer. Antigen-antibody reactions were revealed with standardized development times using the streptavidin method with diaminobenzidine as substrate on an automated Biogenex i6000 immunostainer (Biogenex, San Ramon, CA, USA).

QD-labeled anti-rabbit (Qdot 605 nm, yellow) and anti-mouse (Qdot 655 nm, red) IgG antibodies (Invitrogen) were

used for QDot multiplex immunostaining. Signal was detected using a Leitz Diaplan fluorescence microscope and a CRI Nuance spectral analyzer (CRI Inc., Woburn, MA, USA). Image files were collected at 5-nm wavelength intervals from 450 to 700 nm. The composite image file was deconvoluted using the spectra of autofluorescence and the QD emission spectrum, according to manufacturer's instructions.

Computer-Based Image Analysis System for Immunohistochemical Scoring of TMAs

Tissue microarray slides were scanned using the Ariol instrument SL-50 (Applied Imaging, Grand Rapids, MI, USA) and tumor areas of each core were selected for quantitative analysis. FASN and β -catenin immunostainings were quantitatively scored by Ariol as the area of staining. Linear progressive values were obtained from each core and finally normalized using a logarithmic scale. Total and membrane β -catenin stainings were then analyzed separately by the Ariol and the cytoplasmic area was obtained by subtracting the membrane from the total staining. Finally, the area of cytoplasmic β -catenin was normalized using a logarithmic scale.

Statistical Analyses

FASN and β -catenin expression scores were summarized as mean (s.d.) and median (range) of values for the normal and tumor samples. Comparisons between the paired normal and tumor samples were conducted using Wilcoxon signed-rank test. The association between FASN and cytoplasmic β -catenin was statistically evaluated using a two-sided Spearman's correlation coefficient. Two-tailed P -value < 0.05 was used to define statistical significance. The statistical analysis was undertaken using SAS version 9 (SAS Institute Inc., Cary, NC, USA).

Statistical significance of the results of the palmitoylation assay was determined by using one-tail t -test and GraphPad Prism 4.0 software.

RESULTS

Stable Overexpression of FASN in Immortalized Human Prostate Cells Results in a Unique Pattern of Protein Palmitoylation

We first subcloned the human FASN cDNA (7536 bp) into a pBabe retroviral vector and used it to infect iPrEC-FASN cells. iPrEC-FASN and iPrEC-vector cells were supplemented with exogenous ^3H -palmitate, the main product of FASN activity. Cell lysates of iPrEC-FASN cells showed different levels of palmitoylation of a variety of as yet unidentified proteins when compared to iPrEC-vector (Figure 1a left). A high level of palmitoylation of a band of approximately 45 kDa, corresponding to Wnt1, was found in iPrEC-FASN cells after immunoprecipitation (Figure 1a right).

We thus hypothesized that FASN overexpression would lead to a specific pattern of palmitoylation of Wnt 1, an upstream key regulator of β -catenin, in tumor cells.

FASN-Dependent Activation of Wnt1 Occurs via its Palmitoylation

To assess whether FASN was responsible for the increase in Wnt1 palmitoylation, we quantified the incorporation of labeled acetate, the required precursor of palmitate in *de novo* fatty acid synthesis. iPrEC cells stably overexpressing FASN showed significantly increased incorporation of ^{14}C -acetate in immunoprecipitated Wnt-1 (Figure 1b and c).

The result of the Wnt1 palmitoylation assay suggests that a portion of the palmitate synthesized *de novo* by FASN in iPrEC-FASN cells is utilized for the palmitoylation of Wnt-1 that may affect, at least in part, the activation of β -catenin.

FASN Overexpression in Immortalized Prostate Epithelial Cells Results in Activation and Stabilization of β -Catenin

We then assessed whether, palmitoylation of Wnt1 was associated with activation of β -catenin. Significantly, higher levels of β -catenin protein were detected in iPrEC-FASN and AR-iPrEC-FASN cells compared to the control cells (Figure 2a). In addition, using a specific antibody for the active form of the protein, ie an antibody raised against Ser37- and Thr41-dephosphorylated β -catenin, we also found that β -catenin was activated in iPrEC-FASN and AR-iPrEC cells compared to the control cells (Figure 2a).

Immunofluorescence staining for total β -catenin showed a mixed cytoplasmic and membrane positivity in iPrEC-FASN compared with a predominant membrane staining in iPrEC-vector cells (Figure 2b left). Accordingly, an increased cytoplasmic staining for the active form of β -catenin was detected in iPrEC-FASN compared to iPrEC-vector cells (Figure 2b right).

Taken together, these observations demonstrate that β -catenin is upregulated and stabilized in immortalized human prostate cells overexpressing FASN *in vitro*.

FASN Knockdown Causes Inactivation of β -Catenin in Androgen-Dependent and -Independent Prostate Cancer Cells

To assess the effect of FASN downregulation on β -catenin activity, we checked the expression of the active form of β -catenin after knockdown of FASN by shRNA in androgen-dependent (LNCaP) and androgen-independent (CL-1, PC3, DU145) prostate cancer cell lines.

Figure 3 shows that downregulation of FASN by shRNA is consistently associated with decreased expression of the active form of β -catenin in all the cell lines examined. In contrast, the expression of total β -catenin did not change after FASN knockdown. Four days after the infection with FASN shRNA, cells were still viable but showed a slower growth rate than the control cells infected with GFP-shRNA (data not shown).

This experiment strongly suggests that the activation of β -catenin is modulated by FASN levels in prostate cancer cell lines.

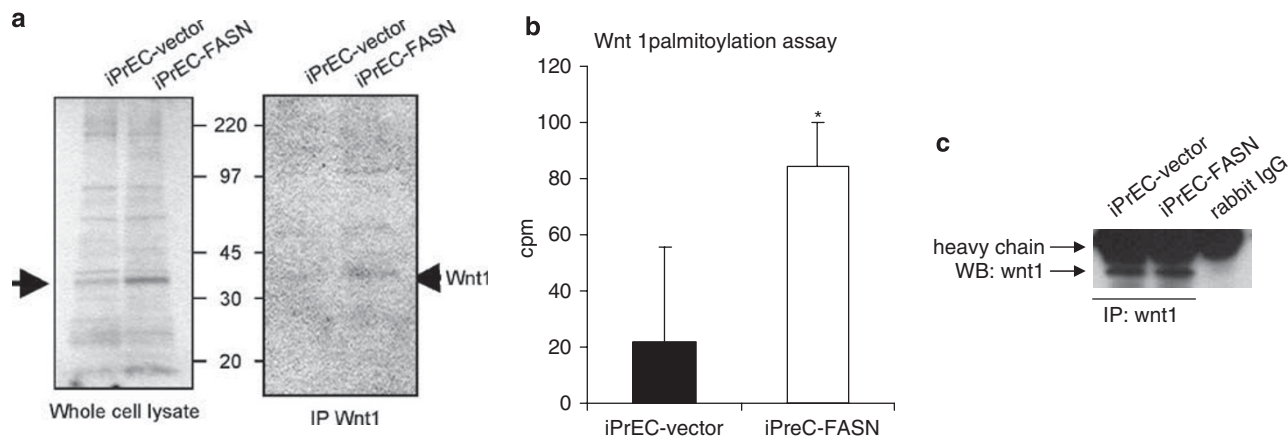


Figure 1 Substantiation of the hypothesis that Wnt1 is palmitoylated in iPReC cells stably overexpressing FASN. **(a)** Overexpression of FASN led to a specific pattern of palmitoylation in lysates of iPReC-FASN compared to iPReC-vector cells pulsed in medium containing ^3H -palmitate (left panel). A band of approximately 45 kDa corresponding to Wnt1 was revealed after immunoprecipitation (right panel). **(b)** The Wnt 1 palmitoylation assay showed increased incorporation of labeled acetate in immunoprecipitated Wnt-1 in iPReC-FASN compared to iPReC-vector cells. C.p.m. in Wnt 1 immune complex samples after subtraction of the background (c.p.m. in the negative control sample: rabbit purified IgG immunoprecipitation) are shown in the graph. Error bars represent mean standard error ($P < 0.05$). **(c)** Representative western blotting of immunoprecipitated Wnt-1 in the same samples used for the palmitoylation assay.

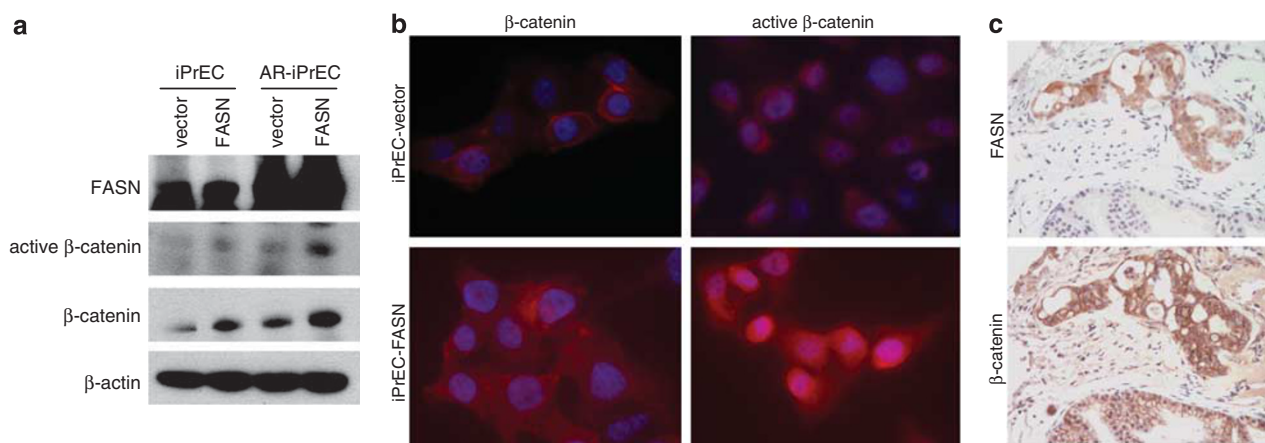


Figure 2 *In vitro* and *in vivo* stabilization and activation of β -catenin in immortalized prostate cells stably overexpressing FASN. **(a)** Immunoblot analysis of FASN and β -catenin in iPReC-FASN and AR-iPReC-FASN cell lines and their respective controls. An increase of β -catenin total expression and a slight increase of its activated form were observed in correspondence to FASN overexpression. **(b)** A predominant cytoplasmic staining for total and active β -catenin was detectable by immunofluorescence in iPReC-FASN cells (bottom pictures) whereas the signal in iPReC-vector (top pictures) was mainly at the membrane (DAPI-Texas Red staining, $\times 63$ magnification). **(c)** Colocalization between mixed membrane/cytoplasmic staining for β -catenin and FASN was found by immunohistochemistry in AR-iPReC-FASN xenograft tumors. The upper panel shows intense immunostaining for FASN in the tumor cells and no staining in the normal surrounding mouse prostate tissue. The lower panel shows mixed membrane/cytoplasmic staining for β -catenin in the tumor cells and exclusively membrane staining in the surrounding normal mouse prostate glands (DAB, $\times 200$ magnification).

Association of FASN and Cytoplasmic β -Catenin in AR-iPReC-FASN Cells Orthotopically Transplanted *In Vivo*

To confirm the association between FASN and β -catenin *in vivo*, we created an orthotopic (anterior lobe) xenograft prostate tumor model by injecting the AR-iPReC-FASN cells in nude mice. AR-iPReC-FASN cells formed invasive adenocarcinoma in mouse prostate tissue with 90% efficiency, whereas AR-iPReC-vector cells formed tumors in about 30% of injected mice (data not shown). In addition, immunohistochemical analysis of FASN and β -catenin in these tumors showed colocalization of the two proteins

(Figure 2c). Both membrane and cytoplasmic expression of β -catenin was observed in these tumors whereas nuclear immunostaining was never detected.

Thus, the stabilized cytoplasmic fraction of β -catenin colocalizes with FASN in the cytosol of the AR-iPReC-FASN xenograft tumors.

FASN and Stabilized β -Catenin are Coexpressed in the Cytoplasm of Human Prostate Cancer Specimens

We next determined whether high-FASN levels correlate with stabilized β -catenin by immunohistochemical staining in

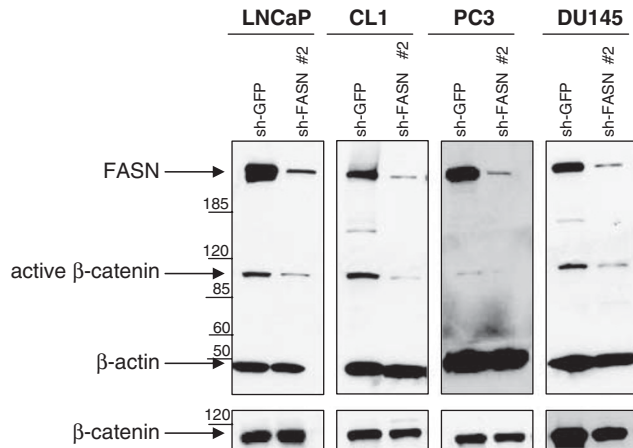


Figure 3 Activation and expression of β -catenin in FASN-knockdown human prostate tumor cell lines. Treatment of LNCaP, CL1, PC3 and DU145 prostate cancer cell lines with a shRNA against FASN resulted in marked downregulation of FASN. A significant decrease of the active form of β -catenin was found in all the prostate cancer cell lines after 4 days as the infection with the pKLO.1 plasmid encoding the sh-FASN no. 2. No significant difference in total β -catenin expression levels was observed after FASN inhibition.

TMA's containing 862 cases of prostate cancer and representative cores of normal prostate tissue. FASN expression in normal prostate tissue was weak and limited to occasional cells in the secretory cell layer. β -Catenin immunostaining in normal prostate was always membranous in both basal and secretory cells. Increased FASN expression and increased cytoplasmic stabilization of β -catenin were significantly associated with prostatic adenocarcinoma compared to paired normal samples ($P < 0.0001$ Wilcoxon test). All the prostate cancer cores showed cytoplasmic staining for FASN and either membranous or cytoplasmic staining for β -catenin, with a considerable variability in the localization of the staining (Figure 4a and b). The colocalization of FASN and β -catenin in the cytoplasm of the same prostate cancer cells was demonstrated using dual quantum dot immunohistochemistry followed by deconvolution by multispectral imaging (Figure 4c). Nuclear localization of β -catenin was never seen by immunohistochemistry in our series of prostate cancers.

Digital Quantitative Assessment of FASN and Cytoplasmic β -Catenin Expression

To avoid the bias derived from human scoring both FASN and β -catenin immunostainings were acquired and automatically scored by image analysis. Figure 5a–c depict how we achieved and quantitated the cytoplasmic immunostaining for β -catenin after digital subtraction of the membrane area from the overall area of immunostaining. A strong significant association was found between FASN immunostaining and cytoplasmic β -catenin ($P < 0.001$, Spearman's $\rho = 0.33$, $n = 862$; Figure 5d).

Overexpression of FASN and cytoplasmic stabilization of β -catenin are thus strongly associated in human prostate cancer tissues.

DISCUSSION

Prostate cancer is currently the second cause of tumor-related death after lung cancer in US males.¹ A diet rich in fat and carbohydrate as well as high body-mass index have been correlated to a higher risk of dying from prostate cancer.³⁷ FASN plays a key role in the metabolic conversion of excess dietary carbohydrates into fatty acids and it is overexpressed in prostate cancer. Although several mechanisms of FASN-mediated oncogenicity have been hypothesized, clear pathways linking the activity of this enzyme and cancer have not been elucidated to date.^{38,39}

Here we hypothesized that the Wnt/ β -catenin pathway is a potential mediator of FASN oncogenicity in the prostate. The Wnt/ β -catenin pathway has previously been implicated as an oncogenic factor in prostate cancer progression. Wnt proteins are hydrophobic and are post-transcriptionally modified by palmitoylation, thus explaining their poor solubility.⁴⁰ Furthermore, palmitoylation is required for receptor activation by Wnt proteins.²¹ Our data demonstrated that Wnt-1 palmitoylation is increased in prostate epithelial cells stably overexpressing FASN when compared to their isogenic counterpart, as determined by increased incorporation into Wnt-1 of *de novo* synthesized ¹⁴C-acetate, the precursor of palmitate. FASN-mediated palmitoylation could also affect other transduction pathways apart from Wnt/ β -catenin. In addition, as the pattern of palmitoylation in the lysates of FASN overexpressing cells differs from that obtained from untransfected cells, the effects of increased synthesis of palmitate in prostate tumor cells, may have additional pleiotropic effects as a result of palmitoylation and subsequent activation or altered subcellular localization of target proteins.

Our results also show that β -catenin is stabilized in FASN-overexpressing cells. The cell lysates of iPrEC-FASN and AR-iPrEC-FASN cells contained higher amounts of total and active β -catenin compared to non-FASN-overexpressing cells. The specific antibody for the active form of β -catenin is directed against an epitope containing two of the four non-phosphorylated residues of the protein, which are dephosphorylated when β -catenin is activated.⁴¹ Its detection is thus informative for the stabilized, nonphosphorylated form of β -catenin. The evidence of a link between the overexpression of FASN and the activation of β -catenin was in fact supported by a decrease in the active form of β -catenin after FASN knockdown in both androgen-dependent and -independent prostate cancer cell lines. Taken together, these data suggest that the Wnt/ β -catenin pathway is activated in FASN-overexpressing prostate cells and that this activation is associated with an increase in the cytoplasmic fraction of β -catenin.

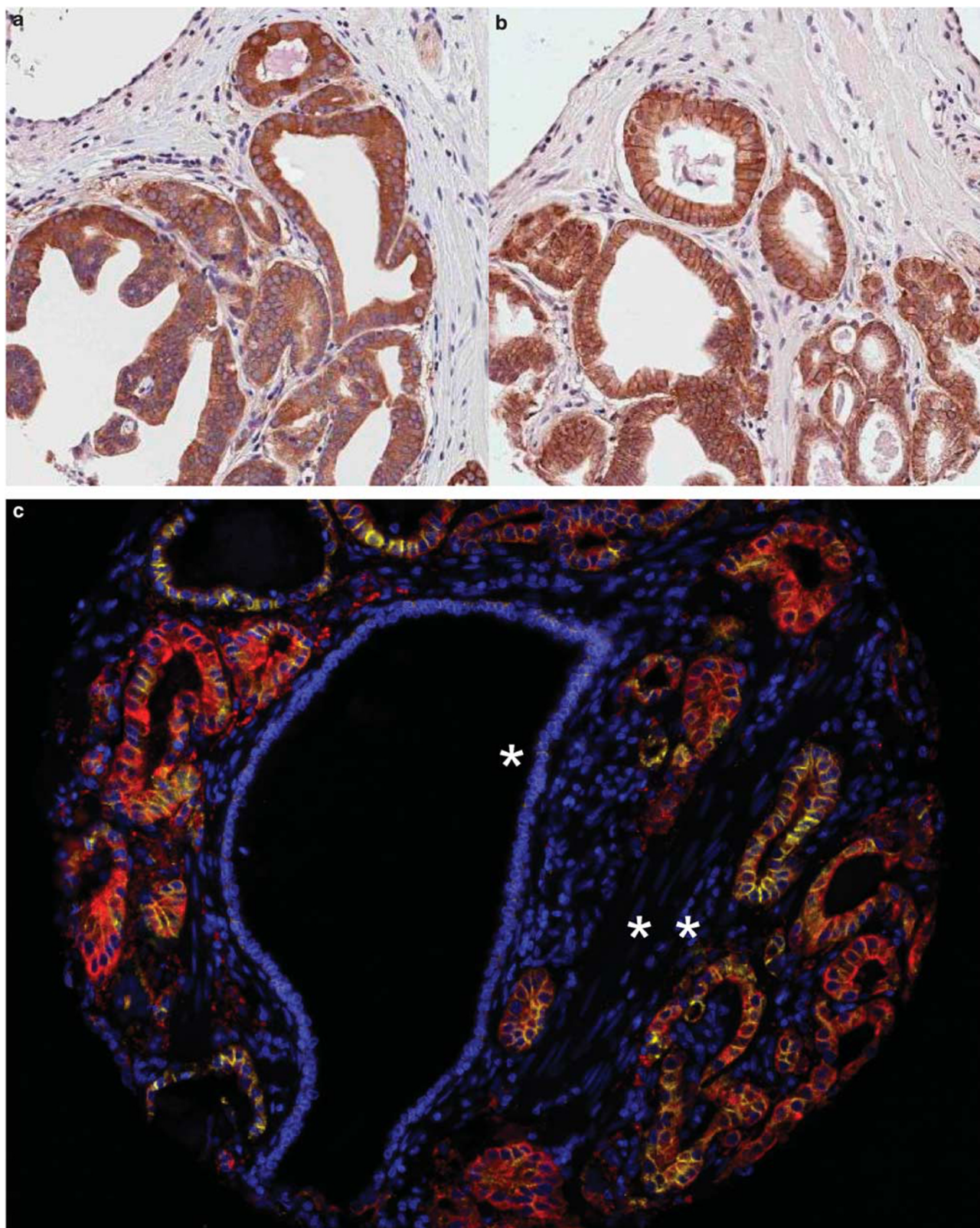


Figure 4 Representative images of immunohistochemistry showing overexpression of FASN (a) and mixed membrane/cytoplasmic expression of β -catenin (b) (DAB, $\times 100$ magnification). (c) Pseudo-colored dual quantum dot imaging of benign prostate epithelium (one star) and prostate adenocarcinoma (two stars). FASN (Qdot 605 nm, yellow) and β -catenin (Qdot 655 nm, red) were seen to colocalize in the cytoplasm of the prostate adenocarcinoma glands with minimal colocalization and expression in the adjacent benign epithelium (QDots, $\times 100$ magnification).

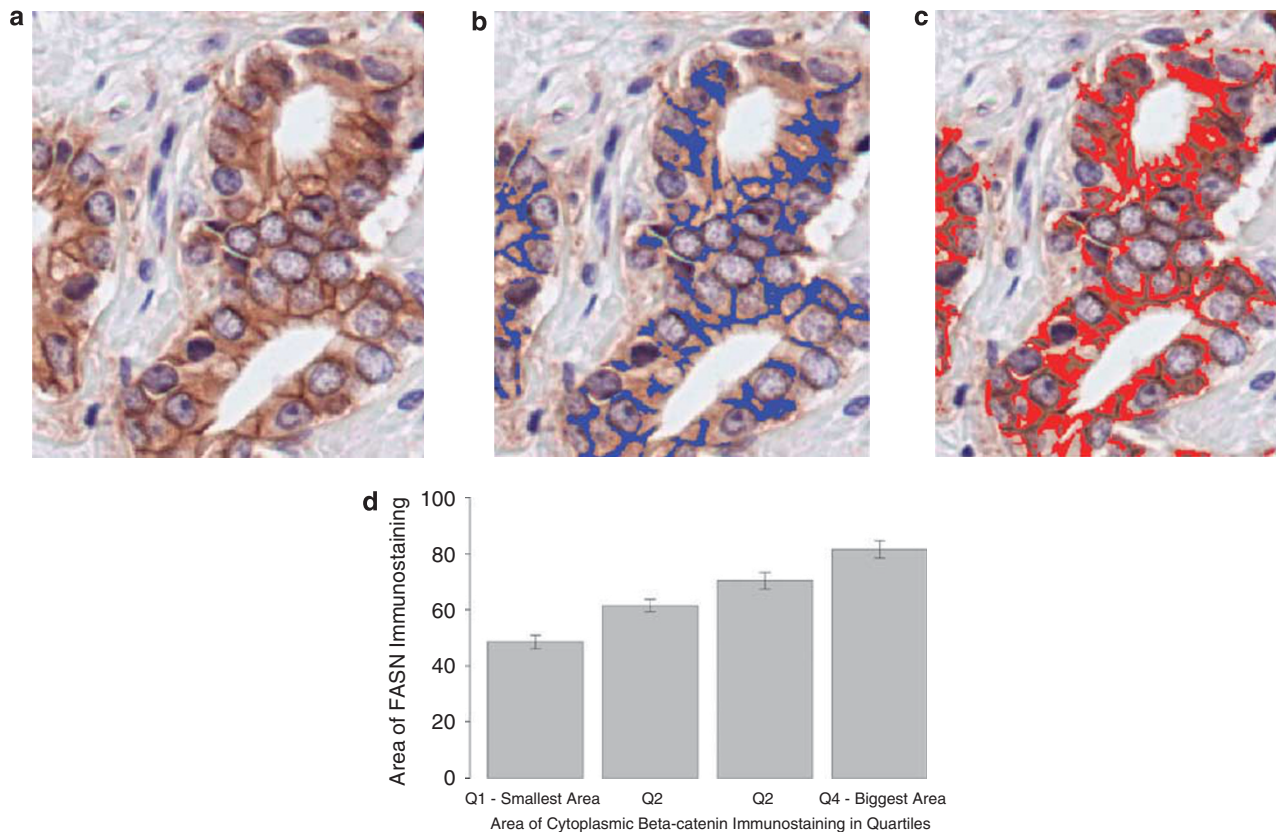


Figure 5 Quantitative assessment of cytoplasmic β -catenin immunostaining by computerized digital analysis in a case of prostate cancer Gleason grade 3. (a) The section stained for β -catenin is digitally acquired. (b) The membranous immunostaining is pseudo-colored in blue. (c) The cytoplasmic immunostaining is pseudo-colored in red and then it can be quantified by digital subtraction of the membranous from the total staining. Finally a score based on the area of staining is generated. (d) Correlation between the area of cytoplasmic β -catenin, divided into quartiles, and the area of FASN immunostaining in serial sections of the same cores ($\times 100$ magnification).

To quantitate the amount of cytoplasmic, ie stabilized and activated β -catenin immunostaining in prostate cancer, we developed a computer-based quantitative method. We were able to quantify by image analysis the area of cytoplasmic β -catenin after digital subtraction of the membrane staining from total immunostaining. In normal conditions, cytoplasmic β -catenin is bound to the APC/GS3K/axin complex and degraded through the ubiquitin-proteasome pathway. Under Wnt signaling, the destruction complex disassembles. β -Catenin is not phosphorylated and can either accumulate in the cytoplasm or shift into the nucleus to bind transcription factors of the TCF family.²³ Once in the nucleus the fate of β -catenin is activation of transcription whereas a nuclear-cytoplasmic shuttle mediated by chaperone proteins occurs.²⁴ Therefore, cytoplasmic β -catenin can be considered a *bona fide* surrogate of nuclear immunostaining.

Importantly, we show that FASN and the cytoplasmic stabilized form of β -catenin colocalize in human prostate cancer samples. It might be argued that the stabilization of β -catenin in the cytoplasm might result in an increase in the nuclear fraction with subsequent activation of downstream effectors of the Wnt pathway. Nuclear localization of β -catenin has been described in many human solid tumors

such as colorectal cancer and hepatocellular carcinoma in association with mutations of the *CTNNB1* gene (very rare in prostate cancer).^{32,33,42} Interestingly, nuclear localization of β -catenin in prostate cancer is a rare event.^{30,33,34} In fact, we also failed to identify any nuclear staining for β -catenin in our 862 prostate cancer samples. Further, cytoplasmic stabilization of β -catenin has been previously described in apoptosis-resistant, hormone-refractory prostate cancer.^{29,30} Thus, cytoplasmic stabilization of β -catenin can be considered a surrogate of activated β -catenin in prostate cancer.

The AR and β -catenin functionally interact and increased AR expression is consistently associated with the development of androgen-independent prostate cancers.^{25,43} Therefore, β -catenin may play an important role in the maintenance/progression of castration-resistant prostate cancer.⁴⁴ Interestingly, high levels of FASN, Wnt-1 and β -catenin expression have been previously found to be associated with advanced, metastatic, hormone refractory prostate cancer.^{8,29} It is tempting to speculate that FASN overexpression may result in β -catenin activation through Wnt1 palmitoylation with subsequent ligand-independent AR activation.

On the basis of our findings, we hypothesized a new model for the mechanism of action of FASN in prostate oncogen-

esis. We propose that stabilization of β -catenin through the palmitoylation of Wnt1 is a potential effector of FASN oncogenicity in prostate cancer.

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- Albertsen PC, Hanley JA, Fine J. 20-year outcomes following conservative management of clinically localized prostate cancer. *JAMA* 2005;293:2095–2101.
- Andersson SO, Wolk A, Bergström R, *et al*. Body size and prostate cancer: a 20-year follow-up study among 135006 Swedish construction workers. *J Natl Cancer Inst* 1997;89:385–389.
- Calle EE, Rodriguez C, Walker-Thurmond K, *et al*. Overweight, obesity, and mortality from cancer in a prospectively studied cohort of U.S. adults. *N Engl J Med* 2003;348:1625–1638.
- Kuhajda FP, Pizer ES, Li JN, *et al*. Synthesis and antitumor activity of an inhibitor of fatty acid synthase. *Proc Natl Acad Sci USA* 2000;97:3450–3454.
- Warburg O. On the origin of cancer cells. *Science* 1956;123:309–314.
- Kusakabe T, Maeda M, Hoshi N, *et al*. Fatty acid synthase is expressed mainly in adult hormone-sensitive cells or cells with high lipid metabolism and in proliferating fetal cells. *J Histochem Cytochem* 2000;48:613–622.
- Kuhajda FP, Jenner K, Wood FD, *et al*. Fatty acid synthesis: a potential selective target for antineoplastic therapy. *Proc Natl Acad Sci USA* 1994;91:6379–6383.
- Rossi S, Graner E, Febbo P, *et al*. Fatty acid synthase expression defines distinct molecular signatures in prostate cancer. *Mol Cancer Res* 2003;10:707–715.
- Visca P, Sebastiani V, Botti C, *et al*. Fatty acid synthase (FAS) is a marker of increased risk of recurrence in lung carcinoma. *Anticancer Res* 2004;24:4169–4173.
- Takahiro T, Shinichi K, Toshimitsu S. Expression of fatty acid synthase as a prognostic indicator in soft tissue sarcomas. *Clin Cancer Res* 2003;9:2204–2212.
- Shurbaji MS, Kalbfleisch JH, Thurmond TS. Immunohistochemical detection of a fatty acid synthase (OA-519) as a predictor of progression of prostate cancer. *Hum Pathol* 1996;27:917–921.
- Menendez JA, Lupu R. Fatty acid synthase and the lipogenic phenotype in cancer pathogenesis. *Nat Rev Cancer* 2007;7:763–777.
- Swinnen JV, Esquenet M, Goossens K, *et al*. Androgens stimulate fatty acid synthase in the human prostate cancer cell line LNCaP. *Cancer Res* 1997;57:1086–1090.
- De Schrijver E, Brusselmans K, Heyns W, *et al*. RNA interference-mediated silencing of the fatty acid synthase gene attenuates growth and induces morphological changes and apoptosis of LNCaP prostate cancer cells. *Cancer Res* 2003;63:3799–3804.
- Swinnen JV, Roskams T, Joniau S, *et al*. Overexpression of fatty acid synthase is an early and common event in the development of prostate cancer. *Int J Cancer* 2002;98:19–22.
- Epstein JI, Carmichael M, Partin AW. OA-519 (fatty acid synthase) as an independent predictor of pathologic state in adenocarcinoma of the prostate. *Urology* 1995;45:81–86.
- Shah US, Dhir R, Gollin SM, *et al*. Fatty acid synthase gene overexpression and copy number gain in prostate adenocarcinoma. *Hum Pathol* 2006;37:401–409.
- Van de Sande T, Roskams T, Lerut E, *et al*. High-level expression of fatty acid synthase in human prostate cancer tissues is linked to activation and nuclear localization of Akt/PKB. *J Pathol* 2005;206:214–219.
- Bandyopadhyay S, Pai SK, Watabe M, *et al*. FAS expression inversely correlates with PTEN level in prostate cancer and a PI3-kinase inhibitor synergizes with FAS siRNA to induce apoptosis. *Oncogene* 2005;24:5389–5395.
- Kuhajda FP. Fatty acid synthase and cancer: new application of an old pathway. *Cancer Res* 2006;66:5977–5980.
- Miura GI, Treisman JE. Lipid modification of secreted signaling proteins. *Cell Cycle* 2006;5:1184–1188.
- Kurayoshi M, Yamamoto H, Izumi S, *et al*. Post-translational palmitoylation and glycosylation of Wnt-5a are necessary for its signaling. *Biochem J* 2007;402:515–523.
- Daugherty RL, Gottardi CJ. Phospho-regulation of β -catenin adhesion and signaling functions. *Physiology* 2007;22:303–309.
- Cong F, Varmus H. Nuclear-cytoplasmic shuttling of axin regulates subcellular localization of β -catenin. *Proc Natl Acad Sci USA* 2004;101:2882–2887.
- Truica CI, Byers S, Gelmann EP. Beta-catenin affects androgen receptor transcriptional activity and ligand specificity. *Cancer Res* 2000;60:4709–4713.
- Yardy GW, Brewster SF. Wnt signaling and prostate cancer. *Prostate Cancer Prostatic Dis* 2005;8:119–126.
- Gounari F, Signoretti S, Bronson R, *et al*. Stabilization of beta-catenin induces lesions reminiscent of prostatic intraepithelial neoplasia, but terminal squamous transdifferentiation of other secretory epithelia. *Oncogene* 2002;21:4099–4107.
- Bruxvoort KJ, Charbonneau HM, Giambernardi TA, *et al*. Inactivation of Apc in the mouse prostate causes prostate carcinoma. *Cancer Res* 2007;67:2490–2496.
- Chen G, Shukeir N, Potti A, *et al*. Up-regulation of Wnt-1 and beta-catenin production in patients with advanced metastatic prostate carcinoma: potential pathogenetic and prognostic implications. *Cancer* 2004;101:1345–1356.
- de la Taille A, Rubin M, Chen WM, *et al*. β -catenin-related anomalies in apoptosis-resistant and hormone-refractory prostate cancer cells. *Clin Cancer Res* 2003;9:1801–1807.
- Voeller HJ, Truica CI, Gelmann EP. Beta-catenin mutations in human prostate cancer. *Cancer Res* 1998;58:2520–2523.
- Chesire DR, Ewing CM, Gage WR, *et al*. *In vitro* evidence for complex modes of nuclear beta-catenin signaling during prostate growth and tumorigenesis. *Oncogene* 2002;21:2679–2694.
- Bismar TA, Humphrey PA, Grignon DJ, *et al*. Expression of beta-catenin in prostatic adenocarcinomas: a comparison with colorectal adenocarcinomas. *Am J Clin Pathol* 2004;121:557–563.
- Horvath LG, Henshall SM, Lee CS, *et al*. Lower levels of nuclear beta-catenin predict for a poorer prognosis in localized prostate cancer. *Int J Cancer* 2005;113:415–422.
- Moffat DA, Gruenemberg X, Yang SY, *et al*. A lentiviral RNAi library for human and mouse genes applied to an arrayed viral high-content screen. *Cell* 2006;124:1283–1298.
- Berger R, Febbo PG, Majumder PK, *et al*. Androgen-induced differentiation and tumorigenicity of human prostate epithelial cells. *Cancer Res* 2004;64:8867–8875.
- Wright ME, Chang SC, Schatzkin A, *et al*. Prospective study of adiposity and weight change in relation to prostate cancer incidence and mortality. *Cancer* 2007;109:675–684.
- Baron A, Migita T, Tang D, *et al*. Fatty acid synthase: a metabolic oncogene in prostate cancer? *J Cell Biochem* 2004;91:47–53.
- Menendez JA, Lupu R. Fatty acid synthase and the lipogenic phenotype in cancer pathogenesis. *Nat Rev Cancer* 2007;7:763–777.
- Willert K, Brown JD, Danenberg E, *et al*. Wnt proteins are lipid-modified and can act as stem cell growth factors. *Nature* 2003;423:448–452.
- van Noort M, Meeldijk J, van der Zee R, *et al*. Wnt signaling controls the phosphorylation status of beta-catenin. *J Biol Chem* 2002;277:17901–17905.
- Fiorentino M, Altimari A, Ravaioli M, *et al*. Predictive value of biological markers for hepatocellular carcinoma patients treated with orthotopic liver transplantation. *Clin Cancer Res* 2004;10:1789–1795.
- DeMarzo A, Nelson WG, Isaacs WB, *et al*. Pathological and molecular aspects of prostate cancer. *Lancet* 2003;361:955–964.
- Mulholland DJ, Dedhar S, Coetzee GA, *et al*. Interaction of nuclear receptors with the Wnt/beta-catenin/Tcf signaling axis: Wnt you like to know? *Endocr Rev* 2005;26:898–915.