#### ORIGINAL ARTICLE

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# Catalog of 434 single-nucleotide polymorphisms (SNPs) in genes of the alcohol dehydrogenase, glutathione S-transferase, and nicotinamide adenine dinucleotide. reduced (NADH) ubiquinone oxidoreductase families

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Abstract An approach based on development of a large archive of single-nucleotide polymorphisms (SNPs) throughout the human genome is expected to facilitate large-scale studies to identify genes associated with drug efficacy and side effects, or susceptibility to common diseases. We have already described collections of SNPs present among various genes encoding drug-metabolizing enzymes. Here we report SNPs for such enzymes at additional loci, including 8 alcohol dehydrogenases, 12 glutathione S-transferases, and 18 belonging to the NADHubiquinone oxidoreductase family. Among DNA samples from 48 Japanese volunteers, we identified a total of 434 SNPs at these 38 loci: 27 within coding elements, 52 in 5' flanking regions, five in 5' untranslated regions, 293 in introns, 20 in 3' untranslated regions, and 37 in 3' flanking regions. The ratio of transitions to transversions was approximately 2.1 to 1. Among the 27 coding SNPs, 13 were nonsynonymous changes that resulted in amino acid substitutions. Our collection of SNPs derived from this study should prove useful for investigations designed to detect associations between genetic variations and common diseases or responsiveness to drug therapy.

**Key words** Single-nucleotide polymorphism (SNP) · Alcohol dehydrogenase · Glutathione S-transferase · NADH ubiquinone oxidoreductase · High-dense SNP map Nonsynonymous substitution · Japanese population

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## Introduction

Human genetic variations result from a dynamic process over time that includes sudden mutations, random genetic drift, and, in some cases, founder effects. Variations at a single gene locus are useful as markers of individual risk for adverse drug reactions or susceptibility to complex diseases (for reviews, see Risch and Merikangas 1996; Kruglyak 1997; McCarthy and Hilfiker 2000). Common types of sequence variation in the human genome include singlenucleotide polymorphisms (SNPs), insertion/deletion polymorphisms, and variations in the number of repeats of certain motifs (e.g., microsatellites and variable number of tandem repeat loci). Among those variations, SNPs are the most abundant, stable, and widely distributed across the genome; moreover, they lend themselves to automated analysis on a very large scale with high-throughput typing technologies (for reviews, see Wang et al. 1998; Halushka et al. 1999).

Projects to establish large collections of SNPs at candidate loci and to develop detailed SNP maps across the human genome have been under way for some time in various laboratories (Sherry et al. 2000, 2001). Construction of an SNP database for the Japanese population began in 2000, and already around 34,000 SNPs have been archived (http://snps.ims.u-tokyo.ac.jp). Simultaneously, we have been focusing on identifying SNPs within genes encoding drug-metabolizing enzymes, drug receptors, and transporters. Local, highly dense SNP maps have been completed for four gene families related to drug metabolism, including 17 members of the S-transferase family, three quinone oxidoreductases, two epoxide hydrolases, and five N-acetyltransferases (Iida et al. 2001; Saito et al. 2001; Sekine et al. 2001). Such genes are likely to have significant roles in responsiveness to drugs and/or in susceptibility to common diseases such as arteriosclerosis, diabetes mellitus, and cancer.

In this study we report a total of 434 SNPs that we identified in a Japanese population sample by direct sequencing of 38 gene loci encoding alcohol

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dehydrogenases, glutathione S-transferases, and nicotinamide adenine dinucleotide, reduced (NADH) uniquinone oxidoreductases.

### **Subjects and methods**

DNA samples and genes encoding drug-metabolizing enzymes

Blood samples were obtained, with informed consent, from 48 healthy Japanese individuals for this study, which was approved by the SNP Research Center, RIKEN. Genomic DNA was extracted from each sample according to standard protocols. On the basis of DNA sequences released from the GenBank database, we predicted structures and designed primers to amplify each gene in its entirety as well as up to 2kb upstream from the first exon (putative promoter region) and downstream from the last exon. Accession numbers are shown at the top of each gene map in Fig. 1. Regions corresponding to repetitive sequences were predicted by the RepeatMasker program with the option "Do not mask simple repeats and lowcomplexity DNA" selected (http://ftp.genome.washington. edu/cgibin/RepeatMasker).

Polymerase chain reaction (PCR), direct sequencing, and detection of SNPs

PCR experiments and DNA sequencing were performed according to methods described previously (Ohnishi et al. 2000). Each PCR experiment was carried out using 20 ng of DNA pooled from three individuals. All SNPs detected by the PolyPhred computer program (Nickerson et al. 1997) were confirmed by sequencing both strands of each PCR product.

# **Results and discussion**

We present here a series of highly dense SNP maps that include a total of 434 SNPs among 38 genes belonging to three families of enzymes associated with drug metabolism. The genomic structures and locations of identified SNPs are illustrated schematically in Fig. 1. Eight of the genes studied here belong to the alcohol dehydrogenase family, 12 to the glutathione S-transferase family, and 18 to the NADH ubiquinone oxidoreductase family. Of the SNPs identified, 334 (77%) were not previously included in the National Center for Biotechnology Information (U.S.) dbSNP database.

Alcohol dehydrogenase family

The family of alcohol dehydrogenases (EC1.1.1.1) in vertebrates consists of several enzymes that are able to catalyze reversible oxidation of a wide variety of endogenous and xenobiotic primary and secondary alcohols, to produce the corresponding aldehydes and ketones. Several distinct classes of vertebrate alcohol dehydrogenase, encoded by nonorthologous genes, have been defined on the basis of sequence homologies and/or unique catalytic properties or gene expression patterns (for reviews, see Duester et al. 1999; Ashmarin et al. 2000). We analyzed here six of the seven known ADH gene loci and two genes that had shown sequence similarities to ADH by direct sequencing (Fig. 1, a1-a8). A total of 66 SNPs were identified from these eight loci: eight were located in coding elements, eight in 5' flanking regions, one in a 5' untranslated region, 37 in introns, eight in 3' untranslated regions, and four in 3' flanking regions. Among these 66 SNPs, 43 (65%) were novel and seven were located within regions corresponding to repetitive sequences predicted by the RepeatMasker program. Among the eight SNPs identified within coding elements, three were nonsynonymous and one, an Ala97Thr substitution (G289A) within the HEP27 gene, had not been reported before, while the remaining two (an Arg48His within ADH2 gene and an Arg272Gln with ADH3 gene) were identical to SNPs deposited in the dbSNP database (ID: rs1789884 and ID: rs1789912, respectively). With regard to the distribution of SNPs in each gene, we found 27 within the 27-kb genomic region containing the ADH7 gene but detected only three in the 20-kb ADH5 genomic region.

Glutathione S-transferase family

Glutathione S-transferases (EC2.5.1.18) are a unique group of multifunctional isozymes that play important roles in the conjugation and detoxification of various xenobiotics such as aflatoxins and polycyclic aromatic hydrocarbons. On the basis of similarities in amino acid sequences and crossreactivity of antibodies, mammalian cryptozoic glutathione S-transferases are divided into six classes: alpha, mu, kappa, theta, pi, and sigma. In addition, there is a class of microsomal glutathione S-transferases (for reviews, see Hayes and Pulford 1995; Strange et al. 2000). We analyzed a total of 12 glutathione S-transferase genes by direct sequencing (Fig. 1, g1-g12), and identified 127 SNPs: eight in coding elements, 22 in 5' flanking regions, none in 5' untranslated regions, 86 in introns, seven in 3' untranslated regions, and four in 3' flanking regions. Among these 127 SNPs, 104 (82%) were considered to be novel because they did not overlap with SNPs in the dbSNP database, and 15 were located within regions corresponding to repetitive sequences predicted by the RepeatMasker program. Of the eight SNPs identified within coding regions, six were nonsynonymous and two, a Thr163Ala substitution (A487G) in the GSTA4 gene and a Val224Ile substitution (G670A) in the GSTM3 gene, were novel. Regarding distribution, SNPs were most frequent at the GSTM1 gene locus (21 SNPs were present in the 12-kb region sequenced; i.e., one in every 480bp), while only three SNPs were detected at the 36.5-kb MGST2 locus (one in every 3000bp).





Fig. 1. Genomic organizations and locations of single-nucleotide polymorphisms (SNPs) in 8 alcohol dehydrogenase genes (a1-a8), 12 glutathione S-transferase genes (g1-g12), and 18 NADH ubiquinone oxidoreductase genes (n1-n18). Exons are represented by *open rectangles* and introns by *horizontal lines*. SNPs are indicated above each gene, and microsatellite sequences, and insertion–deletion polymorphisms are listed below. For cDNA and coding sequences in genomic DNA, the adenine of the initiator Met codon is denoted nucleotide +1. The first nucleotide of the 5' untranslated region is

numbered -1. Likewise, intronic DNA positive numbers start from the guanine of the donor-site invariant GT, and negative numbers start from the guanine of the acceptor-site invariant AG. *Italic type* indicates SNPs located within repetitive sequences predicted by the RepeatMasker program. The complete genomic structure of the *NDUFA2* gene, determined for the first time in this study, has been submitted to the DNA Data bank of Japan under accession number AB054976.1































Fig. 1. Continued



















Fig. 1. Continued

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tttaatctcctttaaatttc G/A caatttcacaacctagggta tttttttttttttgagac G/A aagtctcactcttgtcccct

ctgtagcctctgcctcccag G/A ttcaggcgattcgcgtacct cagattcaagtggttctcct G/C cctcagcctcccaagtagct

23

20 (intron 1 -1945)

(3' flanking region 150)

21 (intron 2 75) 22 (intron 2 148)







2 (5' flanking region -1436)

ttaaaagttgacttttttct G/A cc G/A ggcacggtggctcacgcctg









NADH ubiquinone oxidoreductase family

Respiratory NADH ubiquinone oxidoreductases (also known as complex I, which is embedded in the inner mitochondrial membrane; EC1.6.5.3) serve to dehydrogenate NADH and to shuttle electrons to coenzyme Q. This electron transport generates a proton gradient across the inner mitochondrial membrane, to provide the proton-motive force for ATP synthesis (Guenebaut et al. 1998). The total human complex consists of 42 or more subunits, seven of which are encoded by the mitochondrial genome and the remainder by the nuclear genome. So far, complete gene structures have been reported for six of the 35 nuclear members of this family, and chromosomal localization is known for 26 of the 35 (for reviews see Ton et al. 1997; Loeffen et al. 1998; Smeitink and van den Heuvel 1999). We analyzed 18 complex I genes by direct sequencing, based on their cDNA sequences (Fig. 1. n1-n18). A total of 241 SNPs were identified from the 18 loci: 11 in coding elements, 22 in 5' flanking regions, four in a 5' untranslated region, 170 in introns, five in 3' untranslated regions, and 29 in 3' flanking regions. Of these, 187 (78%) were novel and 38 were located within regions corresponding to repetitive sequences predicted by the RepeatMasker program. Of the 11 SNPs identified within coding elements, four (Leu81Val in the NDUFA3 gene, Ala9Val in the NDUFA6 gene, Pro66Ala in the NDUFA7 gene, and Tyr133His in the NDUFB5 gene) were nonsynonymous and had not been reported previously. With regard to distribution, we found 11 SNPs within the 4.4-kb NDUFA2 genomic locus, while only two were detected in the 18-kb genomic sequence containing the NDUFB3 gene.

The SNP collection reported here provides genetic data that should be helpful for personalized medical service and also for identifying genes associated with drug efficacy and/ or side effects. We believe that, in the near future, using an SNP-based approach to predict individual differences in drug efficacy or toxicity on the basis of genetic factors will be a realistic approach to treatment of human diseases or toxic conditions. Finally, we hope that the virtual experiments made possible by our catalog will accelerate certain aspects of human genetic research.

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