COMPLEX CHROMOSOME REARRANGEMENTS: REPORT OF THREE PATIENTS

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Three unrelated patients are described, each with a complex, Summarv de novo chromosome rearrangement involving four or more break points. One of the patients had few clinical abnormalities and an apparently balanced karyotype with seven break points (1q32, 2q37, 3q26.2, 5q11.2, 5q15, 6q25, 10p13) in six derivative chromosomes. Another patient had multiple congenital anomalies and an apparently balanced complex chromosome rearrangement (CCR) involving four break points (5q13, 5q35, 8p11, 11p15) in three derivatives. The other patient showed multiple anomalies and an unbalanced CCR with seven break points (4g21, 4g25, 6q15, 6q21, 10p13, 10q22, 10q25) in three derivatives including [del(4) (q21q25)]. Each person's parents had normal karyotypes and showed no spontaneous chromosome instability. The fragile sites induced with the FrdU method in two of the three pairs of parents did not correspond to the break points in the CCRs in their offspring. In the last patient, a QFQheteromorphism study revealed that del(4) is of paternal origin. The cause of the CCRs in the three patients is unknown. None of their parents had a history of exposure to teratogenic agents or of radiation of the gonads. None of the parents was an atomic bomb victim although four of them lived in Nagasaki.

Received January 8, 1988; revised version received February 25, 1988; Accepted March 12, 1988 This study was supported in part by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science and Culture of Japan (No. 60480468).

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INTRODUCTION

Congenital complex chromosome rearrangement (CCR) is defined as a congenital chromosome aberration involving three or more break points and with reciprocal exchange of segments between two or more chromosomes (Kousseff *et al.*, 1987). It has been classified into two groups: familial and *de novo*. Among some 60 CCRs reported, about half are familial and half are *de novo* (Pai *et al.*, 1980). *De novo* CCRs involving five or more break points are infrequent, having been reported in only 17 cases (Fukushima *et al.*, 1986; Kousseff *et al.*, 1987).

We report here two cases of *de novo* CCR involving seven break points and one case with four break points, together with the data on the fragile sites in their parents.

OBSERVATIONS

Clinical observations. Patient 1, a girl, was born with a weight of 3,200 g to healthy parents after a 38 weeks' uneventful pregnancy. When examined by us at the age of 4 years, she had generalized hypotonicity, joint hyperextensibility and severe mental retardation (I.Q.=25). Bilateral epicanthus was noted. Computerized tomography of the brain revealed mild dilatation of the lateral ventricles.

Patient 2, a female infant, was born at 39 weeks of gestation with a weight of 2,040 g to healthy parents. She was admitted to our hospital because of neonatal hypoglycemia. On re-examination at the age of 14 months, the following clinical features were noted: developmental retardation (D.Q.=45), an open sagittal suture of the skull with widely spaced frontal bones, a hairy forehead, low-set and malformed ears, preauricular dimples, wide-set nipples, pigmented external genitalia, an enlarged clitoris, bilateral partial syndactyly of the second and third fingers, pes valgus, fibularly flexed toes, large fifth toes, hypoplastic toe nails, pulmonary stenosis, and generalized hypotonia.

Patient 3 is a male newborn baby whose birth weight and length were appropriate for his gestational age of 41 weeks. He was hospitalized immediately after birth because of neonatal respiratory distress. Abnormal findings included porencephaly, brain atrophy, agenesis of corpus callosum, low-set ears, cleft palate, micrognathia, overlapping fingers, a sacral dimple, deformed feet and developmental retardation.

Cytogenetic observations. Chromosome preparations from the three patients and their parents were obtained from three-day cultures of peripheral blood lymphocytes. Standard GTG-banding revealed complex chromosome rearrangements (CCRs) in all three patients (Figs. 1 and 2): Patient 1: 46,XX,t(1;2)(1pter \rightarrow 1q32:: 2q37 \rightarrow 2qter; 2pter \rightarrow 2q37:: 1q32 \rightarrow 1qter),t(6; 10)(6pter \rightarrow 6q25:: 10p13 \rightarrow 10pter; 6qter \rightarrow 6q25:: 10p13 \rightarrow 10qter),dir ins(3;5)(3pter \rightarrow 3q26.2:: 5q11.2 \rightarrow 5q15:: 3q26.2 \rightarrow



Fig. 1. Partial karyotypes of Patients 1 (A), 2 (B) and 3 (C). Wedges on intact chromosomes (left) show break points for derivative chromosomes (right).



Fig. 2. Diagrams of CCRs in Patients 1 (a), 2 (b) and 3 (c).

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3qter; 5pter \rightarrow 5q11.2:: 5q15 \rightarrow 5qter). Patient 2: 46, XX, t(8; 11) (11pter \rightarrow 11p15:: 8p11 \rightarrow 8qter; 8pter \rightarrow 8p11:: 11p15 \rightarrow 11qter), inv(5)(pter \rightarrow q13:: q35 \rightarrow q13:: q35 \rightarrow qter). Patient 3: 46, XY, t(6; 10; 10)(6pter \rightarrow 6q15:: 10q22 \rightarrow 10q25:: 10p13 \rightarrow 10pter; 10qter \rightarrow 10q25:: 10p13 \rightarrow 10q22:: 6q21 \rightarrow 6q15:: 6q21 \rightarrow 6qter), del(4)(pter \rightarrow q21:: q25 \rightarrow qter). Of a total of 18 break points pooled in the three patients, 2q37, 5q15 and 6q25 in Patient 1, 11p15 in Patient 2, and 10q22 and 10q25 in Patient 3 corresponded to the rare or common fragile sites (HGM8, 1985). GTG-banded chromosomes in the parents of these patients were all normal and without spontaneous chromosome breaks. Thus, the CCRs in the three patients were all of *de novo* origin. The deleted chromosome 4 in Patient 3 was of paternal origin (Fig. 3) with the method described previously (Niikawa and Kajii, 1984).

In order to know whether or not the *de novo* rearrangements of the patients are related to fragile sites in their parents, the chromosomes from the parents of Patients 2 and 3 were analyzed in the following two ways: a culture for 72 hr in a folic acid-free medium (MEM-FA, Nissui, Tokyo) supplemented with 5% fetal bovine serum (FBS); another culture for 72 hr in conventional MEM with 10% FBS and by



Fig. 3. Q-banded chromosomes 4 of Patient 3 (C), his mother (M) and his father (P). The del(4) in the patient originated from a chromosome 4 in the father.

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Fragile sites	Number of metaphases with fragile site(s)				
	Patient 1		Patient 2		
	Mother	Father	Mother	Father	
1p36	2	1		4	
1p22				1	
1p 21.2		1		1	
1q21.3	1				
1q25.1				2	
1q42		1		2	
1q44.1				1	
2q13 ^a				1	
2q31		1		· 4	
2q33		3			
2q37.3				1	
3p14.2	3	9	20	13	
3p24.2	1	1		1	
3р25 ь			2		
4p16.1		1		1	
4q12	1				
4q31.1	3		3	1	
5q15			1		
5q31.1			1		
6p22.2			1		
6p23 ª				2	
6q25.1	1	3	2	1	
7p13		1		3	
7p14.2			1		
7q22		3			
7q31.2				1	
7q32.3	1	1	4	7	
8q22.1			2	1	
9q22.1				1	
9q32	2		2	1	
11p13		1			
12q24	1			2	
13q13.2		1		2	
14q21.2		1		1	
14q23		1			

Table 1. Fragile sites observed in the parents of Patients 2 and 3.

Fragile sites	Number of metaphases with fragile site(s)				
	Patient 1		Patient 2		
	Mother	Father	Mother	Father	
14q24.11				4	
15q22		3	2		
16q22.1		2			
16q23.2	2	3	6	6	
18q21.3		1			
18q23 b			2		
20p12.2				1	
Xp22.31		1		4	
Xq22.1		1	1		
Xq27.3 ª			1		
Others ^c	9	10	3	9	
Total number of fragile sites	27	51	54	79	
Number of cells observed	100	100	135	150	

Table 1. (Cont'd)

^a Listed as rare fragile sites in HGM8. Their heredity was not tested in the present families. ^b Not listed in HGM8, but observed in two or more cells. ^c Not listed in HGM8, and observed only in one cell.

adding FrdU (10^{-7} M) 24 hr prior to harvest. Metaphase plates from each culture were collected by adding Colcemid ($0.02 \ \mu g/ml$) 60 min prior to harvest. Chromosomes were first stained with Giemsa, photographed, destained, GTG-banded and then re-photographed. In every parent examined, 50 to 100 mitotic cells were analyzed from each culture. None of the revealed fragile sites corresponded to the break points in the rearrangements in the respective patient (Table 1).

DISCUSSION

The chromosome abnormality in Patient 1 was composed of six derivative chromosomes with seven break points, while that in Patient 2 consisted of three derivatives with at least four break points, and that in Patient 3 involved three derivatives with seven break points. The rearrangements in the three patients were all apparently balanced except for a small interstitial deletion at 4q22-24 in Patient 3. The absence of grave clinical abnormalities in Patient 1 could thus be attributable to her apparently balanced karyotype. The presence of various anomalies in Patient

2, on the other hand, did not reflect her apparently balanced karyotype. There is a possibility, however, of a submicroscopic deletion or position effect of a gene(s) on her derivative chromosomes. The multiple anomalies observed in Patient 3 are most likely due to the loss of genetic material from chromosome 4. He shared del(4)(q21q25) and most of his clinical manifestations with a patient reported by Loughman *et al.* (1979).

The causes that lead to CCRs have not been identified. The importance of extrinsic factors has repeatedly been stressed, such as viral infection and exposure to radiation or chemicals in the parents, particularly the mothers, of the CCR carriers (Kousseff *et al.*, 1987). However, there was no history of such exposure in any of the parents of our patients. Neither the parents nor the patients were exposed to the atomic bomb, although four of the six parents lived in Nagasaki after the Second World War.

Unbalanced CCRs are often *de novo* in origin. Of 10 such instances, seven, including Patient 3 in this series, had a deletion, while the remaining three had excess chromosome material (Prieto *et al.*, 1978; Pai *et al.*, 1980; McGavran *et al.*, 1982). None of them was a mosaic.

Kleczkowska *et al.* (1982), in reviewing CCRs in the literature, identified 20 break points preferentially involved in CCR, out of a total of 100 break points observed. Of the 18 break points observed in our three patients, 10q22 was one of the 20 preferential break points, while another two were among the additional 80 break points. Hecht and Hecht (1984a; 1984b) proposed a hypothesis that certain fragile sites may be more susceptible to chromosome breakages in meiosis leading to a chromosome rearrangement in the offspring. None of the fragile sites observed in two pairs of parents in the present study, however, corresponded to the break points in the CCRs in their offspring.

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