

Dir Ins(9)(q34.3q22.1q31.3) or Inv Ins(9)(q34.3q22.3q21.2)?

To the Editor:

Dr. Kajii and his colleagues have studied a large kindred, in which apparently the same chromosome rearrangement as in our report (Narahara *et al.*, 1986) is segregating, suggesting that a karyotype of inversion carriers is *inv ins*(9)(q34.3q22.3q21.2) instead of *dir ins*(9)(q34.3q22.1q31.3). Our reply to them is as follows: First, the two kindreds are probably related to each other, although their common ancestral origin is yet to be detected. Second, it seems very difficult to identify the precise breakpoints of such a complex rearrangement, because the short segment involved in the insertional translocation is in the region showing mirror-image banding patterns. Results of conventional G- and R-banding were compatible with either *inv ins*(9)(q34.3q22.3q21.2) or *dir ins*(9)(q34.3q22.1q32), but analysis of chromosomes at the level of near 850 bands per haploid set suggested that *dir ins*(9)(q34.3q22.1q31.3) is more likely than *inv ins*(9)(q34.3q22.31q21.2). The conclusion of which interpretation is correct has to await a study of dosage effect of a gene whose locus is mapped to the region in question. Third, intrachromosomal shift is not absolutely rare, eleven cases having been described (Table 1). Of these, five cases had inverted insertion, one had direct insertion and the remaining five had insertion of unknown direction owing to the shortness of the inserted segments. In all but one (Grass *et al.*, 1981), the three-breakpoint-rearrangements were ascertained through recombinant products. Intrachromosomal shift, inverted or direct, would yield two loops during meiosis I, one involving the inserted segment and the other the interposing (non-insertional) segment. An odd number of crossing-over in the latter loop would result in duplication or deficiency of the inserted segment, while that in the former loop would produce various types of recombinants, depending upon

Table 1. Reported cases with intrachromosomal shifts.

Reference	Karyotype	Reason for ascertainment
Therkelsen <i>et al.</i> (1973)	<i>dir ins</i> (2)(q34p13p24)	Recombinant (Rec) dup 2p
Palmer <i>et al.</i> (1977)	<i>inv ins</i> (1)(p22q41q25)	Rec dup 1q
Pan <i>et al.</i> (1977)	<i>ins</i> (1)(p32q25q31)	Rec dup 1q and del 1q
Miller <i>et al.</i> (1979)	<i>ins</i> (7)(p15p21q22)	Rec del 7p and dup 7p
Strobel <i>et al.</i> (1980)	<i>inv ins</i> (11)(q14.5p14.2p11.2)	Rec del 11p and dup 11p
Wyandt <i>et al.</i> (1980)	<i>inv ins</i> (3)(p25.5p21.1p13.5)	Rec del 3p
Grass <i>et al.</i> (1981)	<i>ins</i> (X)(p11q22q24)	Infertility
Allderdice <i>et al.</i> (1983)	<i>inv ins</i> (9)(q22.1q34.3q34.1)	Rec dup 9q and del 9q
Cohen <i>et al.</i> (1983)	<i>ins</i> (16)(q13p11p13)	Rec dup 16p
Pai <i>et al.</i> (1983)	<i>ins</i> (2)(p13q31q33)	Rec del 2q
Martin <i>et al.</i> (1985)	<i>inv ins</i> (5)(p13q22q33)	Rec dup 5q

the direction of insertion. The exclusive occurrence of recombinants with pure deficiency or duplication of the inserted segments among the kindreds so far reported (Table 1) may indicate another possibility that the inserted segment and its homologue are omitted from meiotic pairing without forming a loop. Unfortunately, there has been no meiotic study of carriers with intrachromosomal shifts.

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Kouji NARAHARA

*Department of Pediatrics, Okayama University  
School of Medicine, Okayama 700, Japan*

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