RFLPs OF FACTOR IX GENE IN JAPANESE HAEMOPHILIA B FAMILIES AND GENE DELETION IN TWO HIGH-RESPONDER-INHIBITOR PATIENTS

Sadaaki MIKAMI, Masato NISHINO, Takuya NISHIMURA, and Hiromu FUKUI

Department of Pediatrics, Nara Medical College, Kashihara, Nara 634, Japan

Summary The factor IX genes in four Japanese families with haemophilia B were analysed for the restriction fragment length polymorphisms (RFLPs) of *TaqI*, *XmnI* and *DdeI*, using subcloned intragenic DNA fragments as probes (probes VIII and XIII). The factor IX genes in 12 patients with haemophilia B and three high-responder-inhibitor cases showed no size difference using a cDNA probe (cVII) when restricted by *TaqI*, *Eco*RI and *HindIII*. Complete gene deletions were observed in two other highresponder-inhibitor cases.

INTRODUCTION

The gene for coagulation factor IX has been identified and completely sequenced by Choo et al. (1982), Anson et al. (1984), Kurachi and Davie (1982), Yoshitake et al. (1985) and Jaye et al. (1983). Also, the common polymorphic sites in the normal factor IX gene have been clarified (Yoshitake et al., 1985; Camerino et al., 1984; Giannelli et al., 1984; Winship et al., 1984), to which pathogenic genes for haemophilia B were detected to be linked in a number of families using subcloned DNA fragments as probes and a cDNA sequence. These restriction fragment length polymorphisms (RFLPs) are now used for the female carrier detection and the prenatal diagnosis (Camerino et al., 1984; Giannelli et al., 1984; Winship et al., 1984; Camerino et al., 1985; Peake et al., 1984; Hay et al., 1986; Grunebaum et al., 1984; Brocker-Vriends et al., 1985). Although the abnormal factor IX genes in haemophiliacs have been analysed by the several investigators, a partial or complete deletion of the genes was found mainly in the inhibitor patients (Giannelli et al., 1983; Hassan et al., 1985a, 1985b; Bernardi et al., 1985).

This paper described a study of the RFLPs in 10 Japanese families with haemophilia B, using subcloned intragenic DNA fragments of the factor IX gene as probes.

Received September 16, 1986; revised version received January 9, 1987; Accepted January 21, 1987

Also, the restriction fragment length in 14 patients with haemophilia B, including five inhibitor cases were observed.

MATERIALS AND METHODS

Subjects. Eleven haemophilia B patients from 10 unrelated families, all of whom were treated in our clinic, consisted of six patients with severe form, two with the moderate or mild form, one with Bm form and two with inhibitor (alloantibody) type. The two inhibitor cases were high responders with more than 50 Bethesda units, one of whom had a temporary inhibitor with a peak of 78 Bethesda units that subsequently disappeared even when factor IX preparation was administered. The family members tested included the propositus, other patients if they existed, and the propositus' parents. All mothers were detected as definite carriers according to their family pedigrees and/or their haemostatic findings. Their DNAs were used for RFLPs analysis using subcloned DNA probes as described below.

DNAs from the 11 patients and three other inhibitor cases, were studied for the detection of abnormal restriction sites in their factor IX genes using a cDNA probe. These three inhibitor cases were all high-responders.

Gene probes. Both the subcloned factor IX gene probes VIII and XIII and the cDNA probe cVII provided by Prof. Brownlee of Oxford University were described by Giannelli *et al.* (1984) and Winship *et al.* (1984). The probes VIII, XIII and cVII were cloned into the *PvuII* site of ampicillin-resistant plasmid vectors pATs, and were stabbed in *E. coli* MC1061 that were cultured in media with ampicillin. After chloramphenicol amplification, the plasmids were separated by sucrose centrifugation. Then the DNAs from the plasmids were separated by ethidium bromidecesium chloride centrifugation. After the excision of plasmids DNAs with the appropriate restriction enzymes described in Table 1, factor IX probes were separated by agarose gel electrophoresis. The probes VIII, XIII and cVII were 2.5, 0.46 and 2.0 kb in length, respectively. They were radiolabeled by nick translation (Rigby *et al.*, 1977), and were denatured by heating before hybridisation.

RFLPs analysis. Whole citrated-blood, 10 to 20 ml, were frozen at -70° C. After centrifugation, the precipitates were treated with SDS and proteinase K by the method of Grunebaum *et al.* (1984). Then DNA was obtained by repeated phenol-chloroform extractions.

 Table 1.
 Subcloned intragenic DNA probes (VIII and XIII) and cDNA probe (cVII) for factor IX gene.

Probes	Length	Plasmid vectors	Stabbers	Excision enzymes
VIII (subclone) XIII (subclone)	2.5 kb 0.46 kb	рАТ 153/ <i>Рvu</i> П/8 рАТХ	MC 1061 MC 1061	EcoRI/HindIII BamHI/HindIII
$cVII$ $\begin{pmatrix} cDNA \\ clone \end{pmatrix}$	2.0 kb	pAT 153/PvuII/8	MC 1061	BamHI/HindIII

22

Jpn. J. Human Genet.

DNA samples were then submitted to reaction with an appropriate restriction enzyme. *TaqI*, *XmnI* and *DdeI* were used for RFLP analysis in the families using probes VIII and XIII, and *TaqI*, *Eco*RI and *HindIII* were used for detection of the restriction fragment length in the patients' DNA using probe cVII. After ethanol



Fig. 1. Three common polymorphic sites of factor IX gene and hybridisation sites of the probes VIII and XIII.

Table 2.	Method of RFLP analysis of factor IX gene.	

(Whole blood)		
Freezing at -40° C		
Centrifuging		
SDS and proteinase K treatment		
Repeated phenol-chloroform extraction		
(DNA)		
Restriction enzyme treatment		
[TaqI, XmnI, DdeI for probe VIII, XIII		
TaqI, EcoRI, HindIII for probe cVII		
Ethanol extraction		
(Restricted DNA)		
Agarose gel electrophoresis with SDS		
Denaturation by 0.5 м NaOH		
Southern blotting to nitrocellulose filter		
Prehybridisation		
Hybridisation with ³² P probe VIII, XIII or cVII		
Autoradiography		

Vol. 32, No. 1, 1987

precipitation and SDS treatment, samples were electrophoresed in 0.7% agarose using a submarine apparatus. Then the DNA fragments in the gel were denatured by 0.5 M sodium hydroxide, and transferred onto a nitrocellulose membrane filter by Southern blotting (Southern, 1975). After prehybridisation, the filter was hybridised with each radiolabeled gene probe, and used for autoradiography (Table 2).

RESULTS

RFLPs in the carriers restricted by TaqI, XmnI or DdeI

TaqI, XmnI and DdeI polymorphisms in the two factor IX genes in the carriers of 10 haemophilia B families, were observed. All carriers were diagnosed by either their family pedigrees or by their haemostatic findings.

TaqI restriction is a typical way to detect a dimorphism of the gene, but only



Fig. 2. TaqI RFLP in a haemophilia B family.

Jpn, J. Human Genet.

one carrier out of 10 showed dimorphism of fragment length. In the XmnI restriction of the gene, the DNAs from two carriers out of the remaining nine, and in *Dde*I restriction, one carrier out of the remaining seven showed dimorphisms. Although Giannelli *et al.* (1984) and Winship *et al.* (1984) reported that approximately 60% of haemophilia B genes could be detected in the families by observation of these three polymorphic sites, only four families out of 10 were informative by RFLP analysis of their factor IX genes in our observations.

Taql RFLP in a haemophilia B family

A TaqI RFLP in a family is shown in Fig. 2. The two genes in the mother (who was the carrier having 40% IX:C plasma activity), showed a dimorphism of 5.3+1.8 and 5.3+1.3 kb, and the gene in the propositus (who has 1-3% plasma IX: C activity and 11% IX antigen), was 5.3+1.8 kb. Therefore the 5.3+1.8 kb gene was detected as the pathogenic gene. And since the gene in the father showed 5.3+1.3 kb, the sister of the propositus, whose genes consisted of 5.3+1.3 from the father and 5.3+1.8 which was the pathogenic gene from the mother, was considered to be a carrier. Her clotting activity and antigen of factor IX were both approximately 40%.



Fig. 3. XmnI RFLP in a haemophilia B family.

Vol. 32, No. 1, 1987

XmnI and DdeI RFLPs in haemophilia B families

In XmnI restriction of a family, DNA from the mother showed a dimorphism of 11.5 and 6.5+5.0 kb by probes VIII and XIII. Since the propositus and the brother who was also a patient of severe form, showed two small restricted bands of 6.5+5.0, this gene was determined as the pathogenic gene (Fig. 3).

In the *Dde*I restriction of another family, the genes from the mother showed 1.75 and 1.70 kb. The former had a 50 bp insertion. Since the propositus had 1.70 kb gene, it was the pathogenic gene. The genes from the sister who was already detected as a carrier according to her haemostatic findings, consisted of the abnormal gene of 1.70 kb from the mother and the normal gene of 1.75 kb from the father. The brother who was not a patient, had the normal gene of 1.75 kb from the mother (Fig. 4).

TaqI restriction of the patient genes

In order to observe either deletions of abnormalities of the restriction fragments of factor IX gene in the patients, DNAs from all patients were restricted by enzymes and their fragment length were detected using the cDNA probe cVII. This probe runs from residues 17 to 2,002 of the factor IX cDNA, so as to hybridise with the amino acid-coding exons. A total number of 14 patients including five inhibitor cases, was tested.

In the TaqI restriction, several positive bands consisting of three major bands of 3.5, 2.6 and 1.7 kb were observed in both normal and patient DNAs except two patients as is shown in Fig. 5. Two inhibitor cases with high titers and one temporary inhibitor case were contained in these band-positive patients. The length



Fig. 4. DdeI RFLP in a haemophilia B family.

Jpn. J. Human Genet.



Fig. 5. Fragment length of factor IX genes restricted by Taql in 14 haemophilia B patients.



Fig. 6. Fragment length of factor IX genes restricted by EcoRI in 14 haemophilia B patients.

of the positive fragment bands in the 12 patients was the same as that in normal subjects, and corresponded to the sizes reported by Hassan *et al.* (1985a). Though we can expect to find size differences if any partial deletion or mutation occurred in the *TaqI* TCGA site which includes a hot spot of mutation, no abnormality of the fragment length could be observed in comparison with those in the normal gene.

In the remaining two inhibitor cases who were both high responders, no positive bands were detected with probe cVII hybridisation.

EcoRI restriction of the patient genes

As the next restriction enzyme for patient genes, we used EcoRI which attacks G/AATTC. As can be seen in Fig. 6, three major fragments of 6.5, 5.2 and 4.5 kb nucleotides were observed except in two inhibitor cases. These fragment sizes of the gene were similar to those reported by Hassan *et al.* (1985b), and no discrepancy of length was observed between the 12 tested subjects. In two inhibitor cases, no bands were detected as was found after *TaqI* digestion.

S. MIKAMI et al.



Fig. 7. Fragment length of factor IX genes restricted by HindIII in 14 haemophilia B patients.

HindIII restriction of the patient genes

As a final treatment with restriction enzyme, HindIII was chosen to attack the A/AGCTT sequences. Several hot bands were observed on the film in both normal and patient genes except the previously mentioned two inhibitor cases, showing 15.3, 8.3, 6.0, 5.3, 4.3 and 4.2 kb from the cathodal side. No size difference was detectable between the band-positive subjects. In the two inhibitor cases, no hybridisation bands were seen in *HindIII* restricted genes (Fig. 7). Consequently, these two patients showed complete deletions of their factor IX genes.

DISCUSSION

The three common polymorphic sites of the factor IX gene are now thought to be very useful for the detection of the pathogenic gene in haemophilia B families (RFLP) (Yoshitake et al., 1985; Camerino et al., 1984; Winship et al., 1984). Many reports used for this purpose subcloned DNA and cDNA probes (Camerino et al., 1984: Giannelli et al., 1984; Winship et al., 1984; Camerino et al., 1985; Peake et al., 1984; Hay et al., 1986; Grunebaum et al., 1984; Brocker-Vriends et al., 1985). A TaqI site (T/CGA) in intron D is a typical polymorphic site (Yoshitake et al., 1985; Giannelli et al., 1984), and one of the hot spots of mutations (Barker et al., 1984). Though approximately 25% of the two factor IX genes in women are reported to have dimorphism in this site, only one carrier out of 10 showed it in our observation. All the remaining carriers showed a monomorphism of $2 \times (5.3 \text{ and}$ 1.8 kb). In this family, the pathogenic gene which showed 5.3 and 1.3 kb fragments must be inherited from the mother of the propositus. The sister of the propositus who was detected as a carrier according to her haemostatic findings, proved to have two factor IX genes consisting of one abnormal gene of 5.3 and 1.3 kb from the mother and the other normal gene of 5.3 and 1.8 kb from the father. Since the propositus showed 1-3% activity but 11% antigen, it seems difficult to perform prenatal diagnosis by the conventional method.

28

In XmnI and DdeI RFLP analysis, the rates of dimorphisms in the carriers were also lower in our observation than those reported by Winship et al. (1984), and the fragment length of the genes in the most of carriers showed $2 \times (11.5 \text{ kb})$ by XmnI and $2 \times (1.70 \text{ kb})$ by DdeI in contrast with the result reported by Winship et al. (1984). Since only a small number of carriers were tested in this observation, it will be necessary to examine the frequency of the polymorphic alleles in a much larger samples of the Japanese population before predicting the usefullness of these RFLP's. We must make efforts to search appropriate enzymes and DNA markers for RFLPs of Japanese factor IX genes.

In regard to the abnormalities of the factor IX genes in haemophilia B patients, either a partial or a total deletion of factor IX gene was reported in inhibitor cases (Giannelli *et al.*, 1983; Hassan *et al.*, 1985a, 1985b; Bernardi *et al.*, 1985) except in one family (Chen *et al.*, 1985). However all of the inhibitor cases do not show a detectable gene deletion. In our observations, the factor IX genes in two inhibitor cases had the total gene. They were all high responders, though one of them was a temporary inhibitor case.

Point mutations of the genes in haemophilia B patients were reported in the CRM-positive variant form, for example, factor IX Chapel Hill (Noyes *et al.*, 1983) and factor IX Alabama (Davis *et al.*, 1984). Restriction enzymes such as *TaqI* are a useful marker to search for a point mutation, and several point mutations were reported in haemophilia A patients detected by the method of RFLP search (Antonarakis *et al.*, 1985; Gitscher *et al.*, 1985; Gitscher *et al.*, 1986). In haemophilia B genes, a definitive mutation point has not been found by screening using restriction enzymes. However a mutation in a splice junction was determined by the analysis of DNA sequence (Rees *et al.*, 1985). In our research, *TaqI*, *Eco*RI and *HindIIII* were used for reaction with the genes of 12 patients, and no abnormal fragments were detected. It may be required to observe size difference of the fragments using other enzymes.

Acknowledgments We wish to thank Prof. Brownlee at Oxford University for providing gene probes for factor IX, and Dr. Inagaki and Dr. Iizuka for providing inhibitor plasma. We also acknowledge the valuable technical help of Dr. Murotsu in the Institute for Molecular and Cellular Biology, Osaka University.

REFERENCES

- Antonarakis, S.E., Waber, P.G., Kittur, S.D., Patel, B.S., Kazazian, H.H., Jr., Mellis, M.A., Counts, R.B., Stamatoyannopoulos, G., Bowie, E.J.W., Fass, D.N., Pittman, M.S., Wozney, J.M., and Toole, J.J. 1985. Hemophilia A. Detection of molecular defects and carriers by DNA analysis. *New Engl. J. Med.* 313: 842–848.
- Anson, D.S., Choo, K.H., Rees, D.J.G., Giannelli, F., Could, K., Huddleton, J.A., and Brownlee, G.G. 1984. The gene structure of anti-haemophilic factor IX. *EMBO* 3: 1053–1060.
- Barker, D., Schafer, M., and White, R. 1984. Restriction sites containing CpG show a higher frequency of polymorphism in human DNA. *Cell* **36**: 131–138.

Vol. 32, No. 1, 1987

- Bernardi, F., del Senno, F., Barbieri, R., Buzzoni, D., Gambari, R., Marchetti, G., Conconi, F., Panicucci, F., Positano, M., and Pitruzzello, S. 1985. Gene deletion in an Italian haemophilia B subject. J. Med. Genet. 22: 305–307.
- Brocker-Vriends, A.H.J.T., Briet, E., Quadt, R., Bertina, R.M., van der Linden, I.K., van de Kamp, J.J.P., Pearson, P.L., and Veltkamp, J.J. 1985. Carrier detection of haemophilia B by using an intragenic restriction-fragment length polymorphism. *Thromb. Haemostas.* 54: 506–509.
- Camerino, G., Grzeschik, K.H., Jaye, M., de la Salle, H., Tolstoshev, P., Lecocq, J.P., Heilig, R., and Mandel, J.L. 1984. Regional localization on the human X chromosome and polymorphism of the coagulation factor IX gene (hemophilia B locus). *Proc. Natl. Acad. Sci. U.S.A.* 81: 498–502.
- Camerino, G., Oberle, I., Drayna, D., and Mandel, J.L. 1985. A new *Msp1* restriction fragment length polymorphism in the hemophilia B locus. 1985. *Hum. Genet.* **71**: 79–81.
- Chen, S-H., Yoshitake, S., Chance, P.F., Bray, G.L., Thompson, A.R., Scott, C.R., and Kurachi, K. 1985. An intragenic deletion of the factor IX gene in a family with hemophilia B. J. Clin. Invest. 76: 2161-2164.
- Choo, K.H., Could, K.G., Rees, D.J.G., and Brownlee, G.G. 1982. Molecular cloning of the gene for human anti-haemophilic factor IX. *Nature* 299: 178-180.
- Davis, L.M., McGraw, R.A., Graham, J.B., Roberts, H.R., and Stafford, D.W. 1984. Identification of the genetic defect in factor IX Alabama: DNA sequence analysis reveals a Gly substitution for Asp⁴⁷. *Blood* 64: 262a (Suppl. 1) (Abst.).
- Giannelli, F., Choo, K.H., Rees, D.J.G., Boyd, Y., Rizza, C.R., and Brownlee, G.G. 1983. Gene deletions in patients with haemophilia B and anti-factor IX antibodies. *Nature* 303: 181–182.
- Giannelli, F., Anson, D.S., Choo, K.H., Rees, D.J.G., Winship, P.R., Ferrari, N., Rizza, C.R., and Brownlee, G.G. 1984. Characterisation and use of an intragenic polymorphic marker for detection of carriers of haemophilia B (factor IX deficiency). *Lancet* I: 239–241.
- Gitschier, J., Wood, W.I., Tuddenham, E.G.D., Shuman, M.A., Goralka, T.M., Chen, E.Y., and Lawn, R.M. 1985. Detection and sequence of mutations in the factor VIII gene of haemophiliacs. *Nature* 315: 427–430.
- Gitschier, J., Wood, W.I., Shuman, M.A., and Lawn, R.M. 1986. Identification of a missense mutation in the factor VIII gene of a mild hemophiliac. *Science* 232: 1415–1416.
- Grunebaum, L., Cazenave, J-P., Camerino, G., Kloepfer, C., Mandel, J-L., Tolstoshev, P., Jaye, M., de la Salle, H., and Lecocq, J-P. 1984. Carrier detection of hemophilia B by using a restriction site polymorphism associated with the coagulation factor IX gene. J. Clin. Invest. 73: 1491– 1495.
- Hassan, H.J., Orland, M., Leonardi, A., Chelucci, C., Guerriero, R., Mannucci, P.M., and Peschle, C. 1985a. Intragenic factor IX restriction site polymorphism in hemophilia B variants. *Blood* 65: 441–443.
- Hassan, H.J., Leonardi, A., Guerriero, R., Chelucci, C., Cianetti, L., Ciavarella, N., Ranieri, P., Pilolli, D., and Peschle, C. 1985b. Hemophilia B with inhibitor: Molecular analysis of the subtotal deletion of the factor IX gene. *Blood* 66: 728–730.
- Hay, C.W., Robertson, K.A., Yong, S-L., Thompson, A.R., Growe, G.H., and MacGillivray, T.A. 1986. Use of a *Bam*HI polymorphism in the factor IX gene for the determination of hemophilia B carrier status. *Blood* 67: 1508–1511.
- Jaye, M., dela Salle, H., Schamber, F., Balland, A., Kohli, V., Findeli, A., Tolstoshev, P., and Lecocq, J-P. 1983. Isolation of a human anti-haemophilic factor IX cDNA clone using a unique 52base synthetic oligonucleotide probe deduced from the amino acid sequence of bovine factor IX. *Nucleic Acids Res.* 11: 2325–2335.
- Kurachi, K. and Davie, E.W. 1982. Isolation of a cDNA for human factor IX. *Proc. Natl. Acad.* Sci. U.S.A. **79**: 6461–6464.
- Noyes, C.M., Griffith, M.J., Roberts, H.R., and Lundblad, R.L. 1983. Identification of the molecu-

lar defect in factor IX Chapel Hill: Substitution of histidine for arginine at position 145. *Proc. Natl. Acad. Sci. U.S.A.* **80**: 4200–4202.

- Peake, I.R., Furlong, B.L., and Bloom, A.L. 1984. Carrier detection by direct gene analysis in a family with haemophilia B (factor IX deficiency). *Lancet* I: 242-243.
- Rees, D.J.G., Rizza, C.R., and Brownlee, G.G. 1985. Haemophilia B caused by a point mutation in a donor splice junction of the human factor IX gene. *Nature* **316**: 643–645.
- Rigby, P.W.J., Dieckmann, M., Rhodes, C., and Berg, P. 1977. Labeling deoxyribonucleic acid to high specific activity in vitro by nick translation with polymerase I. J. Mol. Biol. 113: 237–251.
- Southern, E.M. 1975. Detection of specific sequences among DNA fragments separated by gel electrophoresis. J. Mol. Biol. 98: 503-517.
- Winship, P.R., Anson, D.S., Rizza, C.R., and Brownlee, G.G. 1984. Carrier detection in haemophilia B using two further intragenic restriction fragment length polymorphism. *Nucleic Acids Res.* 12: 8861–8872.
- Yoshitake, S., Schach, B.G., Foster, D.C., Davie, E.W., and Kurachi, K. 1985. Nucleotide sequence of the gene for human factor IX (antihemophilic factor B). *Biochemistry* 24: 3736–3750.