### ORIGINAL ARTICLE

# Naphthacemycins, novel circumventors of $\beta$ -lactam resistance in MRSA, produced by *Streptomyces* sp. KB-3346-5. II. Structure elucidation

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Seventeen new compounds, naphthacemycins  $A_1$ - $A_{11}$ ,  $B_1$ - $B_4$  and  $C_1$ - $C_2$ , were isolated from a cultured broth of *Streptomyces* sp. KB-3346-5 during screening for circumventors of  $\beta$ -lactam resistance in methicillin-resistant *Staphylococcus aureus*. Their structures were elucidated by spectroscopic studies, including NMR and X-ray crystallographic analysis. The naphthacemycin A series has a new skeleton displaying a 7-phenylnaphthacene-5,6,11(12*H*)-trione. In contrast, the quinone moiety of the A series is changed to dehydroxyquinol in the B series and to a semiquinone-like structure in the C series. *The Journal of Antibiotics* (2017) **70**, 568–573; doi:10.1038/ja.2017.29; published online 15 March 2017

#### INTRODUCTION

Methicillin-resistant Staphylococcus aureus (MRSA) is a major cause of untreatable and potentially fatal hospital-associated infections. Community-acquired MRSA has also become a serious public health issue.<sup>1</sup> MRSA is resistant to  $\beta$ -lactam antibiotics and usually resistant to most other classes of antibiotics. There are a few antibiotics used for MRSA, for example, vancomycin, teicoplanin, arbekacin, linezolid, daptomycin and tigecycline, but microorganism strains resistant to these compounds are increasingly being reported. In the course of screening for new anti-MRSA compounds, we found cyslabdan, which enhances the activity of imipenem (a carbapenem type β-lactam antibiotic) against MRSA.<sup>2,3</sup> Further screening for microbial metabolites that circumvent the β-lactam resistance of MRSA led us to find the naphthacemycins  $A_1-A_{11}$  (1-11),  $B_1-B_4$  (12-15) and  $C_1-C_2$ (16-17) (Scheme 1), reported as KB-3346-5 substances in the patent by our group,<sup>4</sup> from a cultured broth of Streptomyces sp. KB-3346-5.<sup>5</sup> Here we describe the structure elucidation of the naphthacemycins, based on NMR study and X-ray crystallographic analysis.

#### **RESULTS AND DISCUSSION**

#### Physicochemical properties

Naphthacemycins (1–17) were purified from a cultured broth of *Streptomyces* sp. KB-3346-5, isolated from a soil sample collected in Okinawa Prefecture, Japan.<sup>5</sup> They are red powders. Compounds **2**, **4-6**, **8**, **11** and **14–17** have one chlorine atom, while 7 and **10** have two chlorine atoms. The IR spectra showed they all have carbonyl (1600–1720 cm<sup>-1</sup>) and hydroxyl (3350–3440 cm<sup>-1</sup>) groups. UV spectra were observed at 274–280, 302–306 and 362–408 nm in

the naphthacemycin A series (1-11), at 246–249, 287–288, 352–354 and 414–417 nm in the naphthacemycin B series (12-15), and 248 and 410 nm in the naphthacemycin C series (16-17). Naphthacemycins are soluble in chloroform, ethyl acetate and methanol and insoluble in *n*-hexane and H<sub>2</sub>O.

#### Structure elucidation of the naphthacemycin A series

The molecular formula of naphthacemycin  $A_9$  (9) was established as C30H26O8 by HR-FAB-MS. 1H and 13C NMR spectra and HSQC analysis revealed the presence of 30 carbons, including six methyl, six  $sp^2$  methine, one  $sp^3$  quaternary and 17  $sp^2$  quaternary carbons (Table 1). The long-range couplings of HMBC correlation revealed fragments I and II (Figure 1) as follows; the cross peaks from H-1  $(\delta_{\rm H} 6.55)$  to C-2  $(\delta_{\rm C} 163.9)$ , C-3  $(\delta_{\rm C} 101.6)$ , C-4a  $(\delta_{\rm C} 110.0)$  and C-12a ( $\delta_{C}$  154.7), from H-3 ( $\delta_{H}$  6.25) to C-1 ( $\delta_{C}$  105.5), C-2, C-4  $(\delta_{C} 165.0)$  and C-4a, and from 4-OH  $(\delta_{H} 12.86)$  to C-3, C-4 and C-4a indicated a 2,3,5-trisubstituted phenol (ring A). The cross peaks from H<sub>3</sub>-13 ( $\delta_H$  1.77) and H<sub>3</sub>-14 ( $\delta_H$  1.83) to C-11a ( $\delta_C$  155.4), C-12 ( $\delta_{C}$  39.2) and C-12a indicated a dimethyl moiety being connected to C-12a via C-12. This was confirmed by the coupling between H-1 and C-12, and thus the fragment I was established. An oxygen atom is suggested to be attached to C-2 by its chemical shift. The cross peaks from H-8 ( $\delta_{\rm H}$  6.99) to C-6a ( $\delta_{\rm C}$  125.8), C-7 ( $\delta_C$  140.1), C-9 ( $\delta_C$  162.6) and C-10 ( $\delta_C$  109.5), from H-10  $(\delta_{\rm H}$  7.52) to C-6a, C-8  $(\delta_{\rm C}$  124.6), C-9, C-10a  $(\delta_{\rm C}$  135.8) and C-11 ( $\delta_C$  185.7), and from 9-OCH3 ( $\delta_H$  3.94) to C-9 indicated a 3,4,5-trisubstituted anisole (ring D). The cross peaks from H-17  $(\delta_H 6.39)$  to C-15  $(\delta_C 120.9)$ , C-16  $(\delta_C 156.7)$ , C-18  $(\delta_C 159.9)$  and

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Scheme 1 Structures of naphthacemycins  $A_1-A_{11}$  (1–11),  $B_1-B_4$  (12–15) and  $C_1-C_2$  (16–17).

C-19 ( $\delta_{\rm C}$  106.9), from H-19 ( $\delta_{\rm H}$  6.44) to C-15, C-17 ( $\delta_{\rm C}$  96.4) and C-18, from H\_3-21 ( $\delta_{\rm H}$  2.06) to C-15, C-19 and C-20 ( $\delta_{\rm C}$  137.0), from 16-OCH<sub>3</sub> ( $\delta_H$  3.65) to C-16, and from 18-OCH<sub>3</sub> ( $\delta_H$  3.80) to C-18 indicated a 2-monosubstituted 3,5-dimethoxytoluene (ring E). The coupling between H-8 and C-15 suggested a connection of rings D and E at C-7 and C-15, respectively, and thus the fragment II was established. Among the remaining three carbons, C-5 ( $\delta_{C}$  185.0) had couplings with H-1 and H-3 and C-6 ( $\delta_{\rm C}$  182.6) had a coupling with H-10. They are considered to be  ${}^{4}J_{CH}$  W-couplings, and C-5 and C-6 are suggested to bond to C-4a and C-6a, respectively (Figure 2). The chemical shifts of C-6 ( $\delta_C$  182.6) and C-11 ( $\delta_C$  185.7) indicated that they form a quinone. If the remaining C-5a ( $\delta_{C}$  135.5) bonds to C-5 and C-11a to form ring B, and C-5a-C-6 and C-11-C-11a bondings form ring C, the naphthacenequinone structure is constructed. C-2 is suggested to be hydroxylated by the molecular formula. The carbon connections unrevealed by HMBC were clarified by INADEQUATE analysis (Table 1). Thus the structure of 9 was elucidated as shown in Scheme 1. It has a 7-phenylnaphthacene-5,6,11 (12H)-trione skeleton. Most structurally related compounds, tetarimycin A<sup>6</sup> and fasamycins<sup>7</sup>, were recently reported by Brady and co-workers as antibacterial agents. The structure was confirmed by X-ray crystallographic analysis as shown in Figure 3. The naphthacenequinone forms a planar skeleton.

The molecular formula of naphthacemycin  $A_8$  (8) was established as  $C_{30}H_{25}ClO_8$  by HR-FAB-MS, and this suggested that 8 was chlorinated analog of 9. <sup>1</sup>H and <sup>13</sup>C NMR spectra of 8 were similar to those of 9, except for ring D (Table 1). The HMBC correlations from H-8 ( $\delta_H$  6.93) to C-6a ( $\delta_C$  127.1), C-7 ( $\delta_C$  137.4), C-9 ( $\delta_C$  158.4) and C-10 ( $\delta_C$  120.4) and from 9-OCH<sub>3</sub> ( $\delta_H$  3.95) to C-9 were observed, and C-10 was a quaternary carbon (Figure 4). Thus, the structure of 8 was elucidated as 10-chloro-9, which was confirmed by X-ray crystallographic analysis (Figure 5). In contrast to 9, the naphthacenequinone of 8 bent about 40° at two quinone carbonyl atoms of ring C.

The structures of the other naphthacemycin A series were elucidated by comparison with NMR data of **8** or **9** or other A series compounds (Supplementary Tables S1 and S2) and analyses of HMBC correlations (Supplementary Figure S1).

#### Structure elucidation of the naphthacemycin B series

The molecular formula of naphthacemycin B<sub>1</sub> (12) was established as  $C_{27}H_{22}O_7$  by HR-FAB-MS, which indicated 12 was 28 mass units (CO) less than 1. <sup>1</sup>H and <sup>13</sup>C NMR spectra and HSQC analysis revealed the presence of 27 carbons, including three methyl, seven  $sp^2$  methine, one  $sp^3$  quaternary and sixteen  $sp^2$  quaternary carbons (Table 1). The structure was elucidated by long-range couplings of HMBC correlation (Figure 4). The cross peaks from H-1 ( $\delta_H$  6.65) to

570

#### Table 1 NMR spectroscopic data for 8, 9, 12 and 17<sup>a</sup>

	<b>8</b> b		<b>9</b> b			12°		17	
Position	$\delta_{C}$	$\delta_H$	δ <sub>C</sub>	$\delta_H$	INADEQUATE	$\delta_{C}$	$\delta_H$	$\delta_{C}$	$\delta_H$
1	105.4	6.54 (1H, s)	105.5	6.55 (1H, s)	2, 12a	107.1	6.65 (1H, s)	105.4	6.46 (1H, s)
2	164.3		163.9		1,3	166.7		162.6	
3	101.3	6.27 (1H, s)	101.6	6.25 (1H, s)	2, 4	102.1	6.20 (1H, s)	102.1	6.26 (1H, s)
4	165.0		165.0		3, 4a	166.6		165.4	
4-0H		13.02 (1H, s)		12.86 (1H, s)			12.87 (1H, s)		12.52 (1H, s)
4a	109.2		110.0		4, 5, 12a	108.6		108.6	
5	183.8		185.0		4a, 5a	191.6		188.5	
5a	134.6		135.5		5, 6, 11a	107.6		107.6	
6	181.4		182.6		5a, 6a	160.1		168.8	
6a	127.1		125.8		6, 7, 10a	118.2		123.8	
6-0H							14.53 (1H, s)		14.74 (1H, s)
7	137.4		140.1		6a, 8, 15	140.8		139.3	
8	119.2	6.93 (1H, s)	124.6	6.99 (1H, s)	7, 9	122.5	6.72 (1H, s)	124.6	6.98 (1H, s)
9	158.4		162.6		8,10	166.9		161.9	
9-0Me	56.8	3.95 (3H, s)	55.8	3.94 (3H, s)				55.8	3.94 (3H, s)
10	120.4		109.5	7.52 (1H, s)	9, 10a	110.1	7.05 (1H, s)	110.1	7.73 (1H, s)
10a	133.0		135.8		6a, 10, 11	142.9		137.3	
11	186.5		185.7		10a, 11a	116.1	7.36 (1H, s)	197.8	
11a	158.1		155.4		5a, 11, 12	146.5		53.2	
12	39.4		39.2		11a, 12a, 13, 14	39.7		43.3	
12a	154.3		154.7		1, 4a, 12	155.5		152.9	
13	28.5	1.76 (3H, s)	29.6	1.77 (3H, s)	12	34.5	1.91 (3H, s)	21.0	1.59 (3H, s)
14	30.8	1.85 (3H, s)	30.0	1.83 (3H, s)	12	34.7	1.70 (3H, s)	26.5	1.01 (3H, s)
15	120.3		120.9		7, 16, 20	124.5		130.2	
16	156.4		156.7		15, 17	155.4		153.1	
16-0Me	55.7	3.62 (3H, s)	55.7	3.65 (3H, s)				60.5	3.63 (3H, s)
17	96.3	6.39 (1H, s)	96.4	6.39 (1H, s)	16, 18	100.8	6.23 (1H, s)	113.5	
18	160.2		159.9		17, 19	157.5		154.7	
18-0Me	55.2	3.83 (3H, s)	55.2	3.80 (3H, s)				56.3	3.94 (3H, s)
19	106.9	6.46 (1H, s)	106.9	6.44 (1H, s)	18, 20	108.8	6.26 (1H, s)	108.6	6.61 (1H, s)
20	137.1		137.0		15, 19, 21	138.3		134.9	
21	20.7	2.08 (3H, s)	20.6	2.06 (3H, s)	20	20.7	1.88 (3H, s)	20.3	1.96 (3H, s)
22							- , ,	52.4	2.89 (1H, d, 17.7), 3.52 (1H, d, 17.7)
23								205.4	,, . , , , ,
24								29.6	1.89 (3H, s)

<sup>a</sup>300 MHz for <sup>1</sup>H NMR and 75 MHz for <sup>13</sup>C NMR.

<sup>b</sup>Solvent: CDCl<sub>3</sub>.

<sup>c</sup>Solvent: acetone-d<sub>6</sub>.  $\delta_{H}$  (Int, mult, J in Hz).



Figure 1 Partial structures of 9.

C-2 ( $\delta_{C}$  166.7), C-3 ( $\delta_{C}$  102.1) and C-4a ( $\delta_{C}$  108.6), from H-3 ( $\delta_{H}$  6.20) to C-1 ( $\delta_{C}$  107.1), C-2, C-4 ( $\delta_{C}$  166.6) and C-4a, and from 4-OH ( $\delta_{H}$  12.87) to C-3, C-4 and C-4a indicated a 2,3,5-trisubstituted phenol (ring A). The cross peaks from H<sub>3</sub>-13 ( $\delta_{H}$  1.91) and H<sub>3</sub>-14 ( $\delta_{H}$  1.70) to C-11a ( $\delta_{C}$  146.5), C-12 ( $\delta_{C}$  39.7) and C-12a ( $\delta_{C}$  155.5)

indicated a dimethyl moiety connected to C-12a via C-12. This was confirmed by the coupling between H-1 and C-12. The cross peaks from H-8 ( $\delta_{\rm H}$  6.72) to C-6a ( $\delta_{\rm C}$  118.2), C-9 ( $\delta_{\rm C}$  166.9) and C-10 ( $\delta_C$  110.1) and from H-10 ( $\delta_H$  7.05) to C-6a, C-8 ( $\delta_C$  122.5), C-9, C-10a ( $\delta_C$  142.9) and C-11 ( $\delta_C$  116.1) indicated a 3,4,5-trisubstituted phenol (ring D). The cross peaks from H-11  $(\delta_{\rm H}$  7.36) to C-5a ( $\delta_{\rm C}$  107.6), C-6a and C-12 and from 6-OH  $(\delta_{\rm H}$  14.53) to C-5a, C-6  $(\delta_{\rm C}$  160.1) and C-6a indicated a 2,3,5,6tetrasubstituted phenol (ring C), which condensed with ring D and connected to ring A via C-12. Comparing NMR data and structures of 12 with 1, it is appropriate to form 4,4-dimethylcyclohexadienone (ring B) by C-4a, 5, 5a, 11a, 12 and 12a, though C-5 had no coupling with neighboring protons. The resemblance of the NMR data also suggested that a hydroxyl residue is attached to C-2 of 12 as in 1. The cross peaks from H-17 ( $\delta_{\rm H}$  6.23) to C-15 ( $\delta_{\rm C}$  124.5), C-16 ( $\delta_{\rm C}$  155.4), C-18 ( $\delta_C$  157.5) and C-19 ( $\delta_C$  108.8), from H-19 ( $\delta_H$  6.26) to C-15, C-17 ( $\delta_C$  100.8) and C-18, and from H\_3-21 ( $\delta_H$  1.88) to C-15, C-19 and C-20 ( $\delta_C$  138.3) indicated 2-monosubstituted



Figure 2 Structure elucidation of 9.



Figure 3 ORTEP (Oak Ridge thermal ellipsoid plot) plot of the X-ray crystallographic structure of  ${\bf 9}.$ 

3,5-dihydroxytoluene (ring E). The coupling between H-8 and C-15 indicated that ring E connects to ring D at C-7. Thus, the structure of **12** was elucidated as 6,11-didehydro- $O^{16}$ -demethyl-6,11-dideoxo-11-hydroxy-**1**.

#### Structure elucidation of the naphthacemycin C series

The molecular formula of naphthacemycin  $C_2$  (17) was established as  $C_{33}H_{31}ClO_9$  by HR-FAB-MS, which indicated 17 was 58 mass units ( $C_3H_6O$ ) more than 11. <sup>1</sup>H and <sup>13</sup>C NMR spectra and HSQC analysis revealed the presence of 33 carbons, including seven methyl, one



Figure 4 Structure elucidation of 8 and 12 by HMBC.

 $sp^3$  methylene, five  $sp^2$  methine, two  $sp^3$  quaternary and eighteen  $sp^2$  quaternary carbons (Table 1). The structure was elucidated by long-range couplings of HMBC correlation (Figure 6). The cross peaks from H-1 ( $\delta_{\rm H}$  6.46) to C-2 ( $\delta_{\rm C}$  162.6), C-3 ( $\delta_{\rm C}$  102.1) and C-4a  $(\delta_C 108.6)$ , from H-3  $(\delta_H 6.26)$  to C-1  $(\delta_C 105.4)$ , C-2, C-4  $(\delta_C 165.4)$ and C-4a, and from 4-OH ( $\delta_H$  12.52) to C-3, C-4 and C-4a indicated a 2,3,5-trisubstituted phenol (ring A). The cross peaks from H<sub>3</sub>-13  $(\delta_H$  1.59) and H\_3-14  $(\delta_H$  1.01) to C-11a  $(\delta_C$  53.2), C-12  $(\delta_C$  43.3) and C-12a ( $\delta_{C}$  152.9) indicated a dimethyl moiety was connected to C-12a via C-12. This was confirmed by the coupling between H-1 and C-12. The cross peaks from H-8 ( $\delta_{H}$  6.98) to C-6a ( $\delta_{C}$ 123.8), C-9 ( $\delta_{C}$  161.9) and C-10 (\delta<sub>C</sub> 110.1), H-10 (\delta<sub>H</sub> 7.73) to C-6a, C-8 (\delta<sub>C</sub> 124.6), C-9, C-10a ( $\delta_C$  137.3) and C-11 ( $\delta_C$  197.8), and from 9-OCH<sub>3</sub> ( $\delta_H$  3.94) to C-9 indicated a 3,4,5-trisubstituted anisole (ring D). The cross peaks from 6-OH ( $\delta_{\rm H}$  14.74) to C-5a ( $\delta_{\rm C}$  107.6), C-6 ( $\delta_{\rm C}$  168.8) and C-6a and from H<sub>2</sub>-22 ( $\delta_{\rm H}$  2.89, 3.52) to C-5a, C-11 and C-11a indicated a 2,3,5,6-tetrasubstituted 4-hydroxy-2,4-cyclohexadienone (ring C), condensed with ring D and connected to ring A via C-12. The cross peaks from H<sub>3</sub>-24 ( $\delta_{\rm H}$  1.89) to C-22 ( $\delta_{\rm C}$  52.4) and C-23 ( $\delta_{\rm C}$  205.4) and from H2-22 to C-23 indicated that a 2-oxopropyl residue was attached to C-11a. Comparing NMR data and structures of 17 with 11, it is appropriate to form 4,4-dimethyl-2-cyclohexenone (ring B) by C-4a, 5, 5a, 11a, 12 and 12a, though C-5 had no coupling with neighboring protons. The chemical shifts of ring E of 17 is quite similar to 11, which suggests ring E of 17 is 2-monosubstituted 4-chloro-3,5-dimethoxytoluene. This was confirmed by the cross peaks from H-19 ( $\delta_{\rm H}$  6.61) to C-15 ( $\delta_{\rm C}$  130.2), C-17 ( $\delta_C$  113.5) and C-18 ( $\delta_C$  154.7), from H\_3-21 ( $\delta_H$  1.96) to C-15, C-19 ( $\delta_C$  108.6) and C-20 ( $\delta_C$  134.9), from 16-OCH3 ( $\delta_H$  3.63) to C-16 ( $\delta_C$  153.1), and from 18-OCH<sub>3</sub> ( $\delta_H$  3.94) to C-18. The coupling between H-8 and C-15 indicated that ring E connects to ring D at C-7. Thus, C-11a of 11 was substituted with a 2-oxopropyl group and the C-6 ketone was reduced in 17.

The molecular formula of naphthacemycin C<sub>1</sub> (**16**) was established as  $C_{32}H_{29}ClO_9$  by HR-FAB-MS, which indicated **16** was 14 mass units (CH<sub>2</sub>) less than **17**. <sup>1</sup>H and <sup>13</sup>C NMR spectra of **16** were quite similar to those of **17**, except for rings D and E (Supplementary Table S3). The H-10 signal of **17** disappeared in **16** and the cross peaks of HMBC from H-8 ( $\delta_H$  7.01) to C-6a ( $\delta_C$  126.7), C-9 ( $\delta_C$  158.6) and C-10 ( $\delta_C$  120.8) and from 9-OCH<sub>3</sub> ( $\delta_H$  4.03) to C-9 were observed, which indicated that a chlorine was attached to the C-10 of ring D (Figure 6). The 18-OCH<sub>3</sub> signal of **17** was also absent in **16** and an H-17 ( $\delta_H$  6.37) signal appeared in **17**. The cross peaks from H-17 ( $\delta_H$  6.37) to C-15 ( $\delta_C$  123.7), C-16 ( $\delta_C$  158.1), C-18 ( $\delta_C$  158.4) and C-19 ( $\delta_C$  109.4), from H-19 ( $\delta_H$  6.39) to C-15, C-17 ( $\delta_C$  97.4) and C-18, from H<sub>3</sub>-21 ( $\delta_H$  3.64) to C-16 indicated a ring E structure of 2-monosubstituted 3,5-dimethoxytoluene. Thus **16** was 10-



Figure 5 ORTEP plot of the X-ray crystallographic structure of 8.



Figure 6 Structure elucidation of 16 and 17 by HMBC.

chloro-17-dechloro- $O^{18}$ -demethyl-17. We have not elucidated the configurations of 16 and 17 yet.

#### Circumvention of β-lactam resistance in MRSA

The circumvention of  $\beta$ -lactam resistance in MRSA was measured by enhancement of imipenem activity on MRSA using the paper disk method. Naphthacemycins alone showed no antibacterial activity against a clinically isolated MRSA strain K24 at 0.01–1 µg per disk. When the agar plates contained 10 µg ml<sup>-1</sup> of imipenem, which also did not affect the growth of MRSA, 0.01–1 µg per disks of naphthacemycins showed

## Table 2 Enhancement of imipenem anti-MRSA activity by naphthacemycins

	Inhibition zone diameter (mm) by 6 mm paper disk Agar plate containing imipenem (10 μg ml <sup>-1</sup> ) Amount of compound (μg per disk)								
Compound	0.01	0.03	0.1	0.3	1				
1	-	-	_	10	16				
2	_	10	16	17	22				
3	9	14	18	19	21				
4	_	10	11	13	19				
5	9	15	17	21	25				
6	11	15	19	21	23				
7	10	11	14	19	21				
8	9	12	13	15	16				
9	7	13	15	17	17				
10	11	12	12	14	15				
11	-	10	11	16	16				
12	-	-	-	10	14				
13	_	8	11	15	17				
14	-	13	14	14	16				
15	-	9	10	15	18				
16	-	9	11	15	17				
17	-	-	7	10	17				

inhibition zones (Table 2). Compounds **3** and **5–10** inhibited the growth of MRSA at 0.01  $\mu$ g per disk in the presence of imipenem. Among them, **6** and **10** showed the largest inhibition zones (11 mm) at 0.01  $\mu$ g per disk. The detailed activity of naphthacemycins against MRSA will be reported in an accompanying paper.<sup>5</sup>

#### CONCLUSION

In conclusion, 17 new compounds designated naphthacemycins were isolated from the culture broth of Streptomyces sp. KB-3346-5. They are circumventors of *β*-lactam resistance and enhanced imipenem activity against β-lactam resistant MRSA. Naphthacemycins are 1-phenylnaphthacene antibiotics produced by Streptomyces sp. Many naphthacene type compounds have been isolated from actinomycetes, but most of them have partially unsaturated rings, such as tetracyclines and anthracyclines. Some highly unsaturated naphthacene compounds have been reported to be produced by actinomycetes, such as tetracenomycin D, galtamycin and tetracenoquinocin.8-10 A biosynthetic intermediate of tetracycline, pretetramid and an aglycone of anthracycline, n-pyrromycinone, are classified in the latter.<sup>11,12</sup> Naphthacemycins A and B series also belong to the latter group, but they are the first compounds having a 1-phenylnaphthacene skeleton isolated from a natural origin.

#### **METHODS**

#### General experimental procedure

<sup>1</sup>H NMR (300 MHz) and <sup>13</sup>C NMR (75 MHz) spectra were recorded on a Varian XL-300 spectrometer. Chemical shifts are shown in δ values (p.p.m.) relative to the solvents (acetone-d<sub>6</sub> at 2.05 p.p.m. for <sup>1</sup>H NMR and at 29.8 p.p. m. for <sup>13</sup>C NMR; CDCl<sub>3</sub> at 7.26 p.p.m. for <sup>1</sup>H NMR and at 77.0 p.p.m. for <sup>13</sup>C NMR; DMSO-d<sub>6</sub> at 2.50 p.p.m. for <sup>1</sup>H NMR and at 39.5 p.p.m. for <sup>13</sup>C NMR). INADEQUATE experiment of **9** (170 mg) was carried out for 100 h by 125 MHz NMR using methanol-d<sub>4</sub> as a solvent. Mass spectrometry was conducted on a JEOL JMS-AX505 HA spectrometer. The UV and IR spectra were measured with a Hitachi U-2810 spectrophotometer and a Horiba FT-710 Fourier transform infrared spectrometer, respectively. Optical rotations were recorded on a JASCO model DIP-1000 polarimeter.

#### Assay of antibacterial activity

Measurement of inhibition zone of naphthacemycins, with or without imipenem, to evaluate circumvention activity of  $\beta$ -lactam resistance were carried out by paper disk method, as reported previously.<sup>3</sup>

#### Data availability

The X-ray crystallographic data have been deposited at the Cambridge Crystallographic Data Centre (CCDC) under deposition numbers CCDC 1536220 (8) and CCDC 1536223 (9).

#### CONFLICT OF INTEREST

The authors declare no conflict of interest.

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1 Diep, B. A. & Otto, M. The role of virulence determinants in community-associated MRSA pathogenesis. *Trends Microbiol.* **16**, 361–369 (2008).

- 2 Fukumoto, A. *et al.* Cyslabdan, a new potentiator of imipenem activity against methicillin-resistant *Staphylococcus aureus*, produced by *Streptomyces* sp. K04-0144. I. Taxonomy, fermentation, isolation and structural elucidation. *J. Antibiot.* **61**, 1–6 (2008).
- 3 Fukumoto, A. *et al.* Cyslabdan, a new potentiator of imipenem activity against methicillinresistant *Staphylococcus aureus*, produced by *Streptomyces* sp. K04-0144. II. Biological activities. *J. Antibiot.* **61**, 7–10 (2008).
- 4 Ōmura, S. *et al.* (Kitasato Institute, Japan; Kyowa Hakko Kirin Co., Ltd., Japan). KB-3346-5 substances, their fermentative manufacture, and antibacterial agents containing them. *Jpn Kokai Tokkyo Koho*, JP2009046404A (2009).
- 5 Fukumoto, A. *et al.* Naphthacemycins, novel circumventors of β-lactam resistance in MRSA, produced by *Streptomyces* sp. KB-3346-5. I. The taxonomy of the producing strain, and the fermentation, isolation and antibacterial activities. *J. Antibiot.* (doi:10.1038/ja.2017.28).
- 6 Feng, Z., Kallifidas, D. & Brady, S. F. Functional analysis of environmental DNA-derived type II polyketide synthases reveals structurally diverse secondary metabolites. *Proc. Natl Acad. USA* **108**, 12629–12634 (2011).
- 7 Feng, Z., Chakraborty, D., Dewell, S. B., Reddy, B. V. B. & Brady, S. F. Environmental DNA-encoded antibiotics fasamycins A and B inhibit FabF in type II fatty acid biosynthesis. J. Am. Chem. Soc. 134, 2981–2987 (2012).
- 8 Yue, S., Motamedi, H., Wendt-Pienkowski, E. & Hutchinson, C. R. Antracycline metabolites of tetracenomycin C-ninproducing *Streptomyces glaucescens* mutants. *J. Bacteriol.* **167**, 581–586 (1986).
- 9 Egorov, L. V., Tetent'eva, T. G., Rudneva, N. A., Egorenko, G. G. & Ivantiskaia, L. P. Experimental study of the antitumor anthracycline antibiotic aclarubicin (aclacinomycin A). *Antibiot. Med. Biotekhnol.* **30**, 918–927 (1985).
- 10 Motohashi, K., Takagi, M. & Shin-ya, K. Tetracenoquinocin and 5-iminoaranciamycin from a sponge-derived *Streptomyces* sp. Sp080513GE-26. *J. Nat. Prod.* 73, 755–758 (2010).
- 11 Zhang, W., Watanabe, K., Wang, C. C. & Tang, Y. Investigation of early tailoring reactions in the oxytetracycline biosynthetic pathway. J. Biol. Chem. 282, 25717–25725 (2007).
- 12 Brockmann, H., Pla, L. C. & Lenk, W. ζ-pyrromycinone. Angew. Chem. 69, 477 (1957).

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