

NOTE

Biphenyl ether derivatives with protein tyrosine phosphatase 1B inhibitory activity from the freshwater fungus *Phoma* sp.

Deiske A Sumilat^{1,2}, Hiroyuki Yamazaki², Syu-ichi Kanno², Ryo Saito², Yuta Watanabe² and Michio Namikoshi²

The Journal of Antibiotics (2017) 70, 331–333; doi:10.1038/ja.2016.147; published online 11 January 2017

Type 2 diabetes mellitus (T2DM) and obesity are serious public health issues in developed countries.^{1,2} One of the potential strategies for the prevention and treatment of T2DM and obesity is to inhibit protein tyrosine phosphatase 1B (PTP1B) activity because this enzyme has been shown to play a significant role as a negative regulator of signal transduction from insulin and leptin receptors.^{3,4} Therefore, PTP1B inhibitors are expected to become new therapeutic agents for T2DM and obesity.^{5,6}

During the course of our screening program on new types of PTP1B inhibitors from microorganisms, we have reported tricoketides,⁷ verruculides⁸ and asperdichrome.⁹ Continuous efforts revealed that a culture broth of the freshwater-derived fungus *Phoma* sp. TPU1222 inhibited PTP1B activity. Bioassay-guided separation of the EtOAc extract from the culture broth led to the isolation of a new biphenyl ether derivative, 1-methoxy-3,5'-dimethyl-2,3'-oxybiphenyl-5,1',2'-triol (**1**), together with three known phenolic compounds, 5-methoxy-3,5'-dimethyl-2,3'-oxybiphenyl-1,1',2'-triol (**2**),¹⁰ cyperine (**3**)¹¹ and 6-methylsalicylic acid (**4**)^{12,13} (Figure 1). We herein describe the isolation, structural elucidation and biological properties of compounds **1–4**.

Fungal strain TPU1222 was isolated from a freshwater sample collected at Hayakake lake, Aomori, Japan, in November 2012. Namely, ~1 ml of the freshwater sample was spread on a potato dextrose agar plate (BD, Franklin Lakes, NJ, USA) containing 0.005% rose bengal (Wako, Osaka, Japan) and 0.01% kanamycin (Wako), and was then incubated at 25 °C for 7 days. Strain TPU1222 grown on the plate was isolated and then maintained on a Miura's medium (LCA) slant (0.1% glucose (Wako), 0.08% KH₂PO₄ (Wako), 0.02% K₂HPO₄ (Wako), 0.02% MgSO₄·7H₂O (Wako), 0.02% KCl (Wako), 0.2% NaNO₃ (Wako), 0.02% yeast extract (BD) and 1.5% agar (Wako) in freshwater and adjusted to pH 6.0 before sterilization). The 196 bp ITS1 rDNA sequence of strain TPU1222 was identical to 11 known species of the genus *Phoma* including *P. herbarum*, and, thus, the strain was identified as *Phoma* sp.

The mycelia grown on Miura's medium (LCA) were inoculated into a 100-ml Erlenmeyer flask containing 50 ml of seed medium (2.0% glucose, 0.50% polypeptone (Wako), 0.050% MgSO₄·7H₂O, 0.20% yeast extract, 0.10% KH₂PO₄ and 0.10% agar in freshwater and adjusted to pH 6.0 before sterilization). The flask was shaken reciprocally for 3 days at 25 °C to obtain the seed culture, which was then transferred to production medium (3.0% sucrose (Wako), 3.0% soluble starch (Wako), 1.0% malt extract (BD), 0.30% Ebios (Asahi Food & Healthcare Co. Ltd., Tokyo, Japan), 0.50% KH₂PO₄ and 0.050% MgSO₄·7H₂O in freshwater and adjusted to pH 6.0 before sterilization). The production culture was performed at 25 °C for 7 days under agitation.

Acetone (2.41) was added to the culture broth (2.41) after 7 days and filtered. The filtrate was concentrated to remove acetone and extracted with EtOAc. The EtOAc extract was concentrated *in vacuo* to dryness, and the brown residue (0.34 g) was then suspended in 30% CH₃OH and adsorbed on an ODS column (100 g). The ODS column was eluted stepwise with 30, 50, 70, 85 and 100% CH₃OH in H₂O (200 ml each × 2) and separated into 10 fractions (Fr. 1–Fr. 10). Fr. 5 (the first 200 ml of the 70% CH₃OH eluate) exhibited inhibitory activity against PTP1B and was concentrated *in vacuo* to dryness in order to give a dark black oil (38.9 mg), which was purified by preparative HPLC (column; PEGASIL ODS (Senchu Scientific Co., Ltd., Tokyo, Japan), 10 × 250 mm; mobile phase, 45% CH₃CN containing 0.05% TFA; detection, UV at 210 nm; flow rate, 2.0 ml min⁻¹) to give compounds **1** (5.7 mg), **2** (15.3 mg) and **3** (4.8 mg). Active Fr. 4 (20.9 mg, second 200 ml of the 50% CH₃OH eluate) was separated by preparative HPLC (column; PEGASIL ODS, 10 × 250 mm; solvent, 40% CH₃CN containing 0.05% TFA; detection, UV at 210 nm; flow rate, 2.0 ml min⁻¹) to obtain compounds **2** (0.8 mg) and **4** (7.9 mg).

Compounds **2–4** were identified by comparing their spectroscopic data with those for 5-methoxy-3,5'-dimethyl-2,3'-oxybiphenyl-1,1',2'-triol,¹⁰ cyperine¹¹ and 6-methylsalicylic acid^{12,13} (Figure 1a).

¹Faculty of Fisheries and Marine Science, Sam Ratulangi University, Kampus Bahu, Manado, Indonesia and ²Faculty of Pharmaceutical Sciences, Tohoku Medical and Pharmaceutical University, Sendai, Japan
Correspondence: Dr H Yamazaki, Faculty of Pharmaceutical Sciences, Tohoku Medical and Pharmaceutical University, 4-4-1 Komatsushima, Aoba-ku, Sendai 981-8558, Japan.
E-mail: yamazaki@tohoku-mpu.ac.jp

Received 20 June 2016; revised 30 September 2016; accepted 28 October 2016; published online 11 January 2017

Compound **1** was obtained as a brown oil, and its molecular formula was deduced to be $C_{15}H_{16}O_5$ from HREIMS data (m/z 276.0999 $[M]^+$, Δ +0.1 m.m.u.). 1H and ^{13}C NMR spectra (in CD_3OD) showed 13 proton and 15 carbon signals (Supplementary Figures S1 and S2 and Supplementary Table S1), which were classified into two methyl, one oxygenated methyl, four sp^2 methine, two sp^2 quaternary and six sp^2 oxygenated quaternary carbons by an analysis of the HMQC and DEPT spectra of **1**. UV absorption at 277 nm ($\log \epsilon$ 3.6) and an IR band at 3400 cm^{-1} revealed the presence of phenyl and OH groups, respectively. These spectral properties of **1** were very similar to those of **2**.¹⁰ Therefore, compound **1** was presumed to have a biphenyl ether skeleton, and its structure was elucidated as 1-methoxy-3,5'-dimethyl-2,3'-oxybiphenyl-5,1',2'-triol by an analysis of its 1H - 1H COSY and HMBC spectra (Figure 1b). In order to confirm the positions of OMe groups in **1** and **2**, the NOESY spectra of **1** and **2** were examined and compared with each other. Compound **1** showed NOE correlations between 1-OCH₃ (δ_H 3.63)/H-6 (6.22),

1-OCH₃/H-4' (6.43) and 3-CH₃ (2.23)/H-4 (6.44) (Figure 1c). On the other hand, NOE correlations were observed between 3-CH₃ (2.03)/H-4 (6.31), H-4/5-OCH₃ (3.74) and 5-OCH₃/H-6 (6.37) in the NOESY spectrum of **2** (Figure 1c). Consequently, the structure of **1** was elucidated as shown in Figure 1a.

Compounds **1**–**4** were evaluated for their PTP1B inhibitory activities using an enzyme assay method.¹⁴ As shown in Figure 2 and Supplementary Table S2, compound **2** showed 32% inhibition at $36\text{ }\mu\text{M}$, and **4** was inactive at $66\text{ }\mu\text{M}$. Compounds **1** and **3** exhibited PTP1B inhibitory activities in a dose-dependent manner with IC_{50} values of $13\text{ }\mu\text{M}$ and $17\text{ }\mu\text{M}$, respectively (Figure 2 and Supplementary Table S2). Oleonic acid (Tokyo Chemical Industry, Tokyo, Japan),¹⁵ a positive control, had an IC_{50} value of $1.3\text{ }\mu\text{M}$ in the same experiment (Supplementary Table S2).

Protein tyrosine phosphatases (PTPs) constitute a major family of more than 100 members including PTP1B and regulate various cellular functions. Therefore, selective activity against PTP1B over the other PTPs is one of the important properties for the development of PTP1B inhibitors. The inhibitory effects of compounds **1**–**3** on T-cell protein tyrosine phosphatase (TCPTP) as one of the non-transmembrane PTPs, CD45 tyrosine phosphatase (CD45) as one of the receptor-like PTPs and *vaccinia* H-1-related phosphatase (VHR) as one of the dual-specificity phosphatases were evaluated. Although compounds **1** and **3** exhibited similar inhibitory activities against PTP1B and VHR, their activities against TCPTP and CD45 were weaker than those against PTP1B (Figure 2 and Supplementary Table S2).

PTP1B is mainly located in insulin-targeted tissues such as liver, muscle and fat cells,^{3,4} and, thus, Huh-7 cells (human hepatoma cell line) are used in cell-based experiments in order to investigate the mechanisms of action of PTP1B inhibitors.¹⁶ Thus, the growth inhibitory effects of compounds **1**–**4** on Huh-7 cells were examined using the WST-1 assay.¹⁷ After a treatment for 72 h, compound **3** exhibited modest cytotoxicity (26% growth inhibition at $50\text{ }\mu\text{M}$), whereas compounds **1**, **2** and **4** did not affect cell proliferation at $50\text{ }\mu\text{M}$. Consequently, compound **1** as a PTP1B inhibitor has potential as a drug candidate.

The polybromobiphenyl ether, 2-(3',5'-dibromo-2'-methoxyphenoxy)-3,5-dibromophenol, was obtained from the Indonesian marine sponge *Lamellodysidea herbacea* as a potent PTP1B inhibitor ($IC_{50} = 0.85\text{ }\mu\text{M}$),¹⁴ and exhibits greater inhibitory activity than those of **1**–**3**. Therefore, Br atom(s) play an important role in the inhibition of PTP1B activity. Cell-based experiments on compound **1** and bromobiphenyl ethers in Huh-7 cells are now underway.

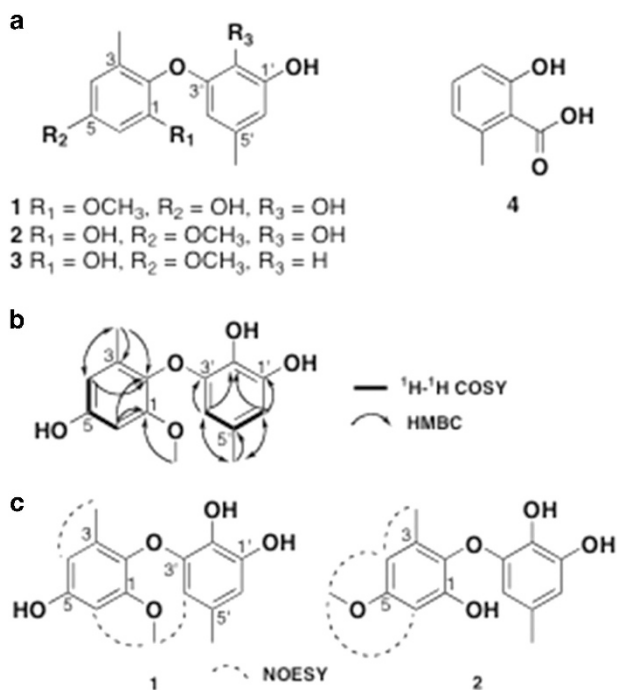


Figure 1 (a) Structures of compounds **1**–**4** produced by the freshwater fungus *Phoma* sp. TPU1222. (b) 1H - 1H COSY and key HMBC correlations for compound **1**. (c) Key NOESY correlations for compounds **1** and **2**.

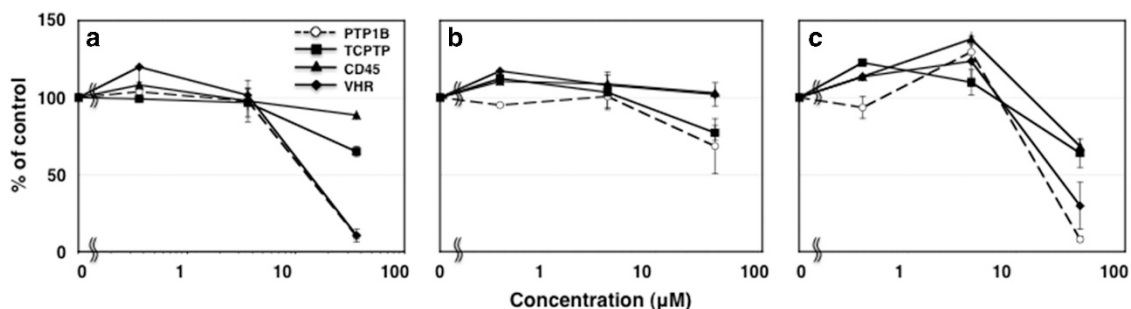


Figure 2 Inhibitory activities of compounds **1** (a), **2** (b) and **3** (c) against PTP1B, TCPTP, CD45 and VHR.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

This work was supported in part by a Grant-in-Aid for Scientific Research (25870660 and 16K21310) from the Ministry of Education, Culture, Sports, Science and Technology (MEXT) of Japan to HY, the Takeda Science Foundation to HY and the Kurita Water and Environment Foundation to HY. We are grateful to the Center for Biomedical Research, Institute of Development, Aging and Cancer, Tohoku University for providing the human cancer cell lines, to Mr T Matsuki and S Sato of Tohoku Medical and Pharmaceutical University for measuring mass spectra, and Mr K Endo, T Ogasawara and Ms M Takahashi of Tohoku Medical and Pharmaceutical University for their technical assistance.

- 1 Shaw, J. E., Sicree, R. A. & Zimmet, P. Z. Global estimates of the prevalence of diabetes for 2010 and 2030. *Diabetes Res. Clin. Pract.* **87**, 4–14 (2010).
- 2 Kitahara, C. M. *et al.* Association between class III obesity (BMI of 40–59 kg m⁻²) and mortality: a pooled analysis of 20 prospective studies. *PLoS Med.* **11**, e1001673 (2014).
- 3 He, R. J., Yu, Z. H., Zhang, R. Y. & Zhang, Z. Y. Protein tyrosine phosphatases as potential therapeutic targets. *Acta Pharmacol. Sin.* **35**, 1227–1246 (2014).
- 4 Zhang, Z. Y., Dodd, G. T. & Tiganis, T. Protein tyrosine phosphatases in hypothalamic insulin and leptin signaling. *Trends Pharmacol. Sci.* **36**, 661–674 (2015).
- 5 Jiang, C. S., Liang, L. F. & Guo, Y. W. Natural products possessing protein tyrosine phosphatase 1B (PTP1B) inhibitory activity found in the last decades. *Acta Pharmacol. Sin.* **33**, 1217–1245 (2012).

- 6 Tamrakar, A. K., Maurya, C. K. & Rai, A. K. PTP1B inhibitors for type 2 diabetes treatment: a patent review (2011–2014). *Expert Opin. Ther. Pat.* **24**, 1101–1115 (2014).
- 7 Yamazaki, H. *et al.* Trichoketides A and B, two new protein tyrosine phosphatase 1B inhibitors from the marine-derived fungus *Trichoderma* sp. *J. Antibiot.* **68**, 628–632 (2015).
- 8 Yamazaki, H. *et al.* Verruculides A and B, two new protein tyrosine phosphatase 1B inhibitors from an Indonesian ascidian-derived *Penicillium verruculosum*. *Bioorg. Med. Chem. Lett.* **25**, 3087–3090 (2015).
- 9 Yamazaki, H., Ukai, K. & Namikoshi, M. Asperdichrome, an unusual dimer of tetrahydroxanthone through an ether bond, with protein tyrosine phosphatase 1B inhibitory activity, from the Okinawan freshwater *Aspergillus* sp. TPU1343. *Tetrahedron Lett.* **57**, 732–735 (2016).
- 10 Du, L. *et al.* Diarylcyclopentendione metabolite obtained from a *Preussia typharum* isolate procured using an unconventional cultivation approach. *J. Nat. Prod.* **75**, 1819–1823 (2012).
- 11 Stierle, A., Upadhyay, R. & Strobel, G. Cyperine, a phytotoxin produced by *Ascochyta cypericola*, a fungal pathogen of *Cyperus rotundus*. *Phytochemistry* **30**, 2191–2192 (1991).
- 12 Venkatasubbaiah, P. & Chilton, W. S. An epoxydon-derived ester from a *Phoma* sp. pathogenic to rhubarb. *J. Nat. Prod.* **55**, 639–643 (1992).
- 13 Venkatasubbaiah, P., Van Dyke, C. G. & Chilton, W. S. Phytotoxic metabolites of *Phoma sorghina*, a new foliar pathogen of pokeweed. *Mycologia* **84**, 715–723 (1992).
- 14 Yamazaki, H. *et al.* A polybromodiphenyl ether from an Indonesian marine sponge *Lamellodysidea herbacea* and its chemical derivatives inhibit protein tyrosine phosphatase 1B, an important target for diabetes treatment. *J. Nat. Med.* **67**, 730–735 (2013).
- 15 Zhang, Y. N. *et al.* Oleonic acid and its derivatives: new inhibitor of protein tyrosine phosphatase 1B with cellular activities. *Bioorg. Med. Chem.* **16**, 8697–8705 (2008).
- 16 Abdjul, D. B., Kanno, S., Yamazaki, H., Ukai, K. & Namikoshi, M. A dimeric urea of the bisabolene sesquiterpene from the Okinawan marine sponge *Axinyssa* sp. inhibits protein tyrosine phosphatase 1B activity in Huh-7 human hepatoma cells. *Bioorg. Med. Chem. Lett.* **26**, 315–317 (2016).
- 17 Berridge, M. V., Herst, P. M. & Tan, A. S. Tetrazolium dyes as tools in cell biology: new insights into their cellular reduction. *Biotechnol. Annu. Rev.* **11**, 127–152 (2005).

Supplementary Information accompanies the paper on The Journal of Antibiotics website (<http://www.nature.com/ja>)